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ORIGINAL RESEARCH

# Defining the phenotypical spectrum associated with variants in *TUBB2A*

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## ABSTRACT

**Background** Variants in genes belonging to the tubulin superfamily account for a heterogeneous spectrum of brain malformations referred to as tubulinopathies. Variants in *TUBB2A* have been reported in 10 patients with a broad spectrum of brain imaging features, ranging from a normal cortex to polymicrogyria, while one patient has been reported with progressive atrophy of the cerebellar vermis.

**Methods** In order to further refine the phenotypical spectrum associated with *TUBB2A*, clinical and imaging features of 12 patients with pathogenic *TUBB2A* variants, recruited via the international network of the authors, were reviewed.

**Results** We report 12 patients with eight novel and one recurrent variants spread throughout the *TUBB2A* gene but encoding for amino acids clustering at the protein surface. Eleven patients (91.7%) developed seizures in early life. All patients suffered from intellectual disability, and 11 patients had severe motor developmental delay, with 4 patients (36.4 %) being non-ambulatory. The cerebral cortex was normal in five individuals and showed dysgyria of variable severity in seven patients. Associated brain malformations were less frequent in *TUBB2A* patients compared with other tubulinopathies. None of the patients had progressive cerebellar atrophy.

**Conclusion** The imaging phenotype associated with pathogenic variants in *TUBB2A* is highly variable, ranging from a normal cortex to extensive dysgyria with associated brain malformations. For recurrent variants, no clear genotype-phenotype correlations could be established, suggesting the role of additional modifiers.

division, migration and intracellular transport.<sup>3</sup> The assembly of polymers requires GTP, and GTP hydrolysis is necessary for the dynamic properties of microtubules. Nevertheless, straightening of the tubules is likely to be the result of the assembly of the microtubule lattice and not of GTP metabolism.<sup>4</sup> Microtubule function also depends on interaction with microtubule-associated proteins (MAPs).<sup>5</sup> Functional heterogeneity is further increased by a number of post-translational modifications, for example, glutamylation and detyrosination.<sup>6</sup>

Variants in *TUBA1A*, *TUBB2A*, *TUBB2B*, *TUBB*, *TUBB3* and *TUBG1* have been linked to a spectrum of cortical malformations and associated brain malformations commonly referred to as tubulinopathies.<sup>7-9</sup> Protein structures within the tubulin superfamily show a high degree of similarity. However, the phenotypical differences associated with variants in the various tubulin isotypes support the hypothesis that each tubulin has a distinctive function.<sup>3 10</sup> Additionally, tubulin isotypes are not interchangeable as shown, for example, for *TUBB3*.<sup>11</sup>

*TUBB2A* is a neuronal-specific isotype of beta-tubulin, but the expression of *TUBB2A* during gestation has been reported to be lower compared with other beta tubulin isotypes. This finding suggests a minor role of *TUBB2A* in brain development.<sup>2 12</sup> Nevertheless, to date, six patients with variants in *TUBB2A* have been reported with a heterogeneous spectrum of brain malformations, described as pachygyria, polymicrogyria, simplified gyral pattern (SGP) and dysgyria. The presence of associated brain malformations, as well as clinical features such as epilepsy and motor and intellectual disability, also varies in severity between the reported patients.<sup>13-16</sup> Three additional patients have been reported, but clinical information is limited to infantile spasms with normal brain imaging in one patient,<sup>17</sup> while for the other two patients, abnormalities of the nervous system have not further been specified.<sup>18</sup> Recently, a variant in *TUBB2A* has also been described in a patient with progressive atrophy of the superior cerebellar vermis and thinning of the corpus callosum with the clinical presentation of progressive spastic

## INTRODUCTION

Human cerebral cortical development requires precise regulation of neuronal proliferation, migration and differentiation, as well as axon growth and guidance.<sup>1</sup>

Microtubules play a key role in these processes. Tubulin genes encoding for the different tubulin isotypes are highly expressed during cortical development.<sup>2</sup> Alpha and beta tubulins form heterodimeric polymers that serve as a scaffold for the cytoskeleton dynamics required during cell



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paraparesis and polyneuropathy, consistent with a neurodegenerative process.<sup>19</sup>

The underlying pathogenic effects of variants on protein function are poorly understood. Although effects on polymerisation, integration into the microtubule scaffold or assembly at the spindle pole during mitosis have been suggested, the heterogeneous phenotypes are likely caused by residue-specific effects on cortical development. Additionally, patients with recurrent variants but differing phenotypes indicate the presence of additional modifiers.<sup>16 19</sup>

The aim of this work was to further delineate the clinical and imaging features associated with pathogenic variants in *TUBB2A* in 12 novel patients, in combination with a review of the literature.

## METHODS

Patients were recruited through the international network of the authors, COST Action CA16118 Neuro-MIG.

Clinical data were collected by a review of the patient's medical records and clinical examination. MRI images were reviewed by SB, TV and ACJ, when available. Particular attention was spent on the type, severity and the anteroposterior severity gradient, and anatomical distribution of abnormal cortical lamination and associated brain malformations. Dysgyria is defined as a cortical malformation consisting of variable sulcal depth and/or orientation without blurring of the grey-white matter interface and with normal cortical thickness.<sup>20 21</sup>

Peripheral blood samples were obtained from all patients for genetic testing. DNA was extracted using standard protocols, specific for each centre. Genetic testing was performed using capture-based methods, either through gene panel analyses (samples 1–4) or whole-exome sequencing (samples 5–12), after which the presence of the variant was confirmed by Sanger sequencing in all patients except for patient 7. The absence of the variants in the parents were also confirmed by trio analysis or Sanger sequencing for all patients.

## RESULTS

### Clinical features

Clinical findings are summarised in [table 1](#). All patients presented with global developmental delay. Motor development was severely affected in all but one patient (91.6%), and four of these patients (36.4%) were non-ambulatory at ages 5.0, 2.5, 11.0 and 15.0 years, respectively. Spastic paraplegia or cerebellar signs were not reported in this cohort. Eight patients were non-verbal, while four had very poor speech limited to a few single words. One patient was reported to have cortical blindness. Eleven patients (91.6%) had epilepsy, with age of onset varying from the neonatal period up to the age of 5 years. In seven patients, seizures were generalised, four of which presented with infantile spasms. Four patients were refractory to treatment. Five patients (5/10, 50%) had postnatal microcephaly ( $-2.5$  SD and  $-4.5$  SD) and five patients were normocephalic. Six patients (6/10, 60%) presented with mild dysmorphic facial features, while four patients were not dysmorphic. Dysmorphic facial features included a high forehead and deep-set eyes (patient 2) and a round face with a flat nasal bridge, cupid upper lip and full lower lip in patient 3. Dysmorphic features were not further specified for patients 1 and 4.

### Imaging

MRI of the brain was performed in all patients between ages 2 days and 12 years ([table 1](#) and [figures 1 and 2](#)). MRI images

were available for review for nine patients (P1–9). We included three additional patients (P10–12) in which clinical and imaging data were retrieved from the patient files but images were not available for review by the authors. Brain MRI was normal for one patient (P1) and revealed a spectrum of brain malformations in the others. The cerebral cortex was reported to be normal in five patients (41.7%), although focal cortical heterotopia was mentioned in one. Seven patients (58.3%) had tubulinopathy-related dysgyria, closely resembling polymicrogyria in two patients (P6 and P7). The parietal and temporal regions were most severely affected in most patients. No recurrent pattern of the anatomical distribution of the malformations could be observed. The lateral ventricles were enlarged in eight patients (66.7%), and in patients 6 and 7, the lateral ventricles had a hooked appearance. The brainstem was normal in all patients. The corpus callosum was abnormal in eight patients (66.7%) and the basal ganglia were dysmorphic in four patients (33.3%). Four patients (33.3%) had mild hypoplasia of the cerebellar vermis and/or a dysmorphic vermis; the cerebellar hemispheres were hypoplastic in one patient.

### Genetics

Whole-exome sequencing or gene panel analysis was performed in 12 unrelated patients with neurodevelopmental delay with or without brain malformations. The parents were either tested in parallel with the patients (trios) or in a second step by Sanger sequencing. We detected nine pathogenic or likely pathogenic variants in *TUBB2A* (Genomic RefSeq accession: NC\_000006.11; mRNA RefSeq accession: NM\_001069.2), one of which has previously been reported in the literature. All variants in *TUBB2A* occurred *de novo* in the index patients. No additional pathogenic or likely pathogenic variants were detected.

The previously reported p.(Ala248Val) variant detected in patients 8, 9 and 12 has been reported multiple times.<sup>13 15 18</sup> Patients 1 and 3 carried novel missense variants p.(Val49Gly) and p.(Val49Met), respectively, that affect the same amino acid but result in different substitutions. Patients 6 and 7 carry the same not previously reported p.(Pro357Leu) variant.

The variants were spread over the *TUBB2A* gene affecting amino acids throughout the TUBB2A protein ([figure 3](#)) and were all located at highly conserved amino acid positions.<sup>22</sup> Although TUBB2A is largely made up of sheets and helices, the majority of variants are located in the random coils between those secondary structures ([figure 3](#)). Only the p.(His396Tyr) variant in patient 2, the p.(Glu410Lys) in patient 10 and the p.(Asp417Asn) reported by Sferra *et al*<sup>19</sup> are located in the two last helices of the protein, which are part of the c-terminal MAP-binding domain. Nevertheless, all variants were predicted to have a deleterious effect on protein function. When looking at the 3D protein structure, variants appear to cluster at the protein surface, suggesting an impact on protein 3D structure and lateral interactions ([figure 3](#)).

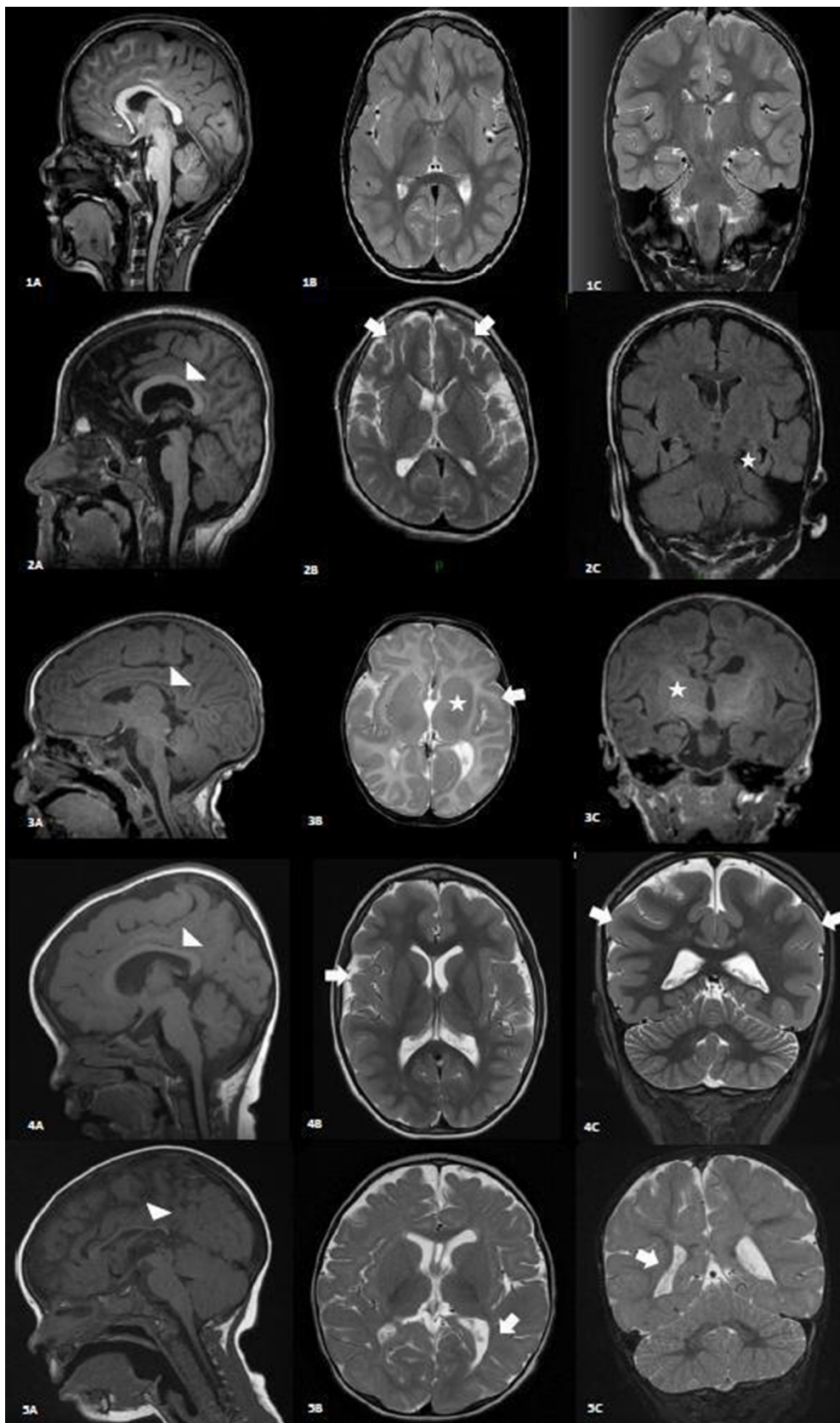
### DISCUSSION

Tubulinopathies have been associated with a spectrum of cortical malformations with associated brain malformations often including dysmorphic basal ganglia, thin corpus callosum and brainstem and/or cerebellar vermis hypoplasia.<sup>7 20</sup> Variants in *TUBB2A* have previously been reported in patients with variable clinical and cortical imaging features, ranging from a normal cortex to polymicrogyria.<sup>13–18</sup> Additionally, one patient has been reported with progressive neurodegeneration and loss of independent walking but without cortical malformations.<sup>19</sup> Furthermore, neurodegeneration has been reported for variants

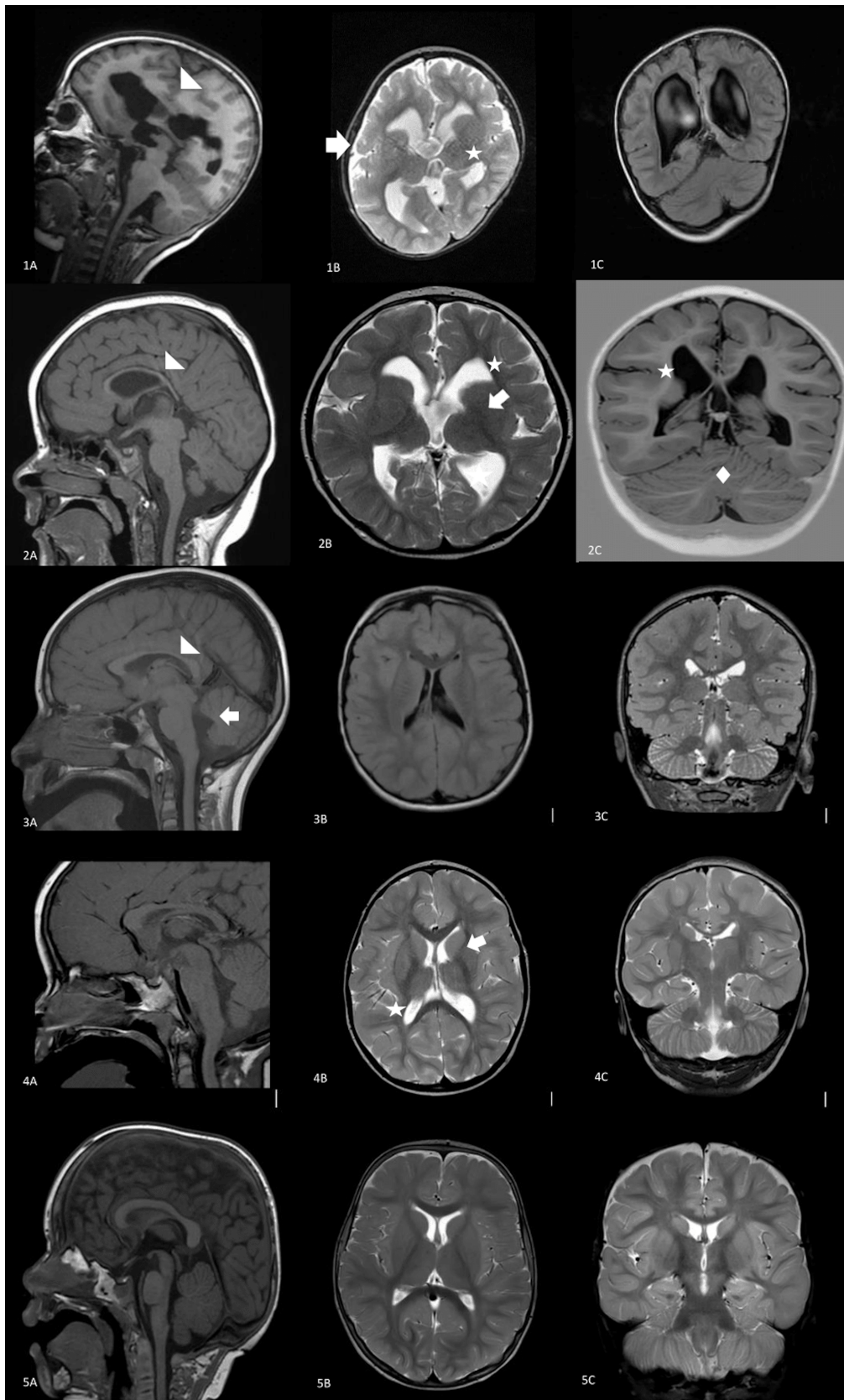
**Table 1** Clinical and imaging features of patients with variants in *TUBB2A*

Patient	1	2	3	4	5	6	7	8	9	10	11	12
Sex	M	M	F	M	F	M	M	F	F	F	M	M
Nucleotide sequence variation	c.145G>A	c.1186C>T	c.146T>G	c.267C>A	c.56>A	c.1070C>T	c.1070C>T	c.749C>T	c.743C>T	c.1228G>A	c.689C>T	c.743C>T
Protein sequence variation	p.(Val49Met)	p.(His396Tyr)	p.(Val49Gly)	p.(Asn89Lys)	p.(Arg2His)	p.(Pro357Leu)	p.(Pro357Leu)	p.(Ala248Val)	p.(Ala248Val)	p.(Glu410Lys)	p.(Ser230Leu)	p.(Ala248Val)
Inheritance	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo	De novo
Age at examination	5 years	7 years	2 years 6 months	6 years 8 months	2 years 8 months	15 years	11 years	8 years 2 months	3 years	4 years 2 months	32 years	4 years 8 months
Microcephaly	No	Yes	Yes	No	Yes	Yes	No	n/a	n/a	No	No	Yes
Dysmorphic features	Yes	n/a	No	Facial dysmorphism, abnormal dermatoglyphs	Facial dysmorphism, clinodactyly of toes	Mild dysmorphism	No	No	n/a	Epicanthus	Facial asymmetry	No
ID	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Motor impairment	Ambulant>2 years	Non-ambulatory	Non-ambulatory	Broad-based gait, stereotypes	Ambulant at 26 months, broad-based gait	Severe impairment	Non-ambulatory, GMFCS5	Walks with support	Ambulated at 38 months	No	Ambulated at 2 years	Ambulated at 25 months
Speech	Non-verbal	Non-verbal	Non-verbal	Non-verbal	Single words	Non-verbal	Non-verbal	Non-verbal	Non-verbal	Delayed and impaired	Delayed and impaired	Delayed and impaired
Social	Autistic traits	n/a	Poor contact	ASD, short attention span, impulsive behaviour	Short attention span, stertile response to noises	Poor eye contact	n/a	Autistic traits, short attention span	Poor eye contact	Well-integrated in a Montessori kindergarten	Hyperactive behaviour, autistic features as adult	Normal
Epilepsy	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Clinically suspected, normal EEG	Yes	Yes
Age at seizure onset	8 months	5 years	5 months	7 months	10 months	6 months	2 months	6 months	Neonatal	n/a	3 years	4 years
Seizure type	Convulsive febrile and non-febrile seizures	Generalised tonic-clonic seizures, myoclonia	n/a	Infantile spasms, myoclonic seizures	Febrile seizures, multifocal seizures	Infantile spasms, tonic seizures	Infantile spasms, tonic seizures	Infantile spasms, intractable epilepsy	Crytogenic generalised epilepsy	/	Complex focal seizures with tonic-clonic generalisation	Focal seizures
Refractory	No	No	No	Yes	Yes	No	Yes	Yes	n/a	/	No	No
Brain imaging												
Age at MRI	5 years	n/a	2 months 3 weeks	1 years 11 months and 7 years	10 months	n/a	10 months	8 years 1 month	3 years 7 months	4 years 5 months and 12 years	8 years 4 months	17 months/25 months/4.5 years
Gyral pattern	Normal	Dysgyria	Dysgyria	Dysgyria	Normal	Dysgyria	Dysgyria	Normal	Normal	Dysgyria	Cortical heterotopia	Dysgyria
Severity	/	Mild	Severe	Mild	/	Severe	Severe	/	/	Mild	/	n/a
Gradient	/	Temporoparietal	Insular, parietal	Anterior-posterior	/	Diffuse	Diffuse	/	/	Normal	Right parietal region	Insular, parietal
White matter	Normal	Normal	Normal	Prominent Virchow-Robin spaces	Reduced	Severely reduced	Reduced	Normal	Mildly reduced, posterior>anterior	Normal	Normal	Prominent Virchow-Robin spaces
Lateral ventricles	Normal	Normal	Enlarged (left>right)	Enlarged	Enlarged	Enlarged, hooked frontal horns	Dilated, hooked frontal horns	Enlarged, asymmetric septal region	Enlarged occipital horns (left>right)	Normal	Enlarged occipital horns (left>right)	Normal
Corpus callosum	Normal	Dysmorphic	Partial agenesis	Mildly dysmorphic	Hypoplasia	Partial agenesis	Diffuse hypoplasia	Hypoplasia	Normal	Normal	Normal	Thin body
Basal ganglia	Normal	Normal	Dysplasia	Normal	Normal	Dysmorphic	Dysmorphic	Normal	Mildly dysmorphic	Normal	Normal	Normal
Hippocampus	Normal	Normal	Normal	Normal	Normal	Dysplastic	Normal	Normal	Normal	Normal	Normal	Normal
Brainstem	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal	Normal
Cerebellum	Normal	Normal	Small haemorrhage (left hemisphere)	Normal	Normal	Vermis hypoplasia and asymmetry, mild cortical atrophy	Vermis hypoplasia and asymmetry, mild hypoplasia	Mild vermis hypoplasia	Normal	Normal	Dysplastic vermis	Normal

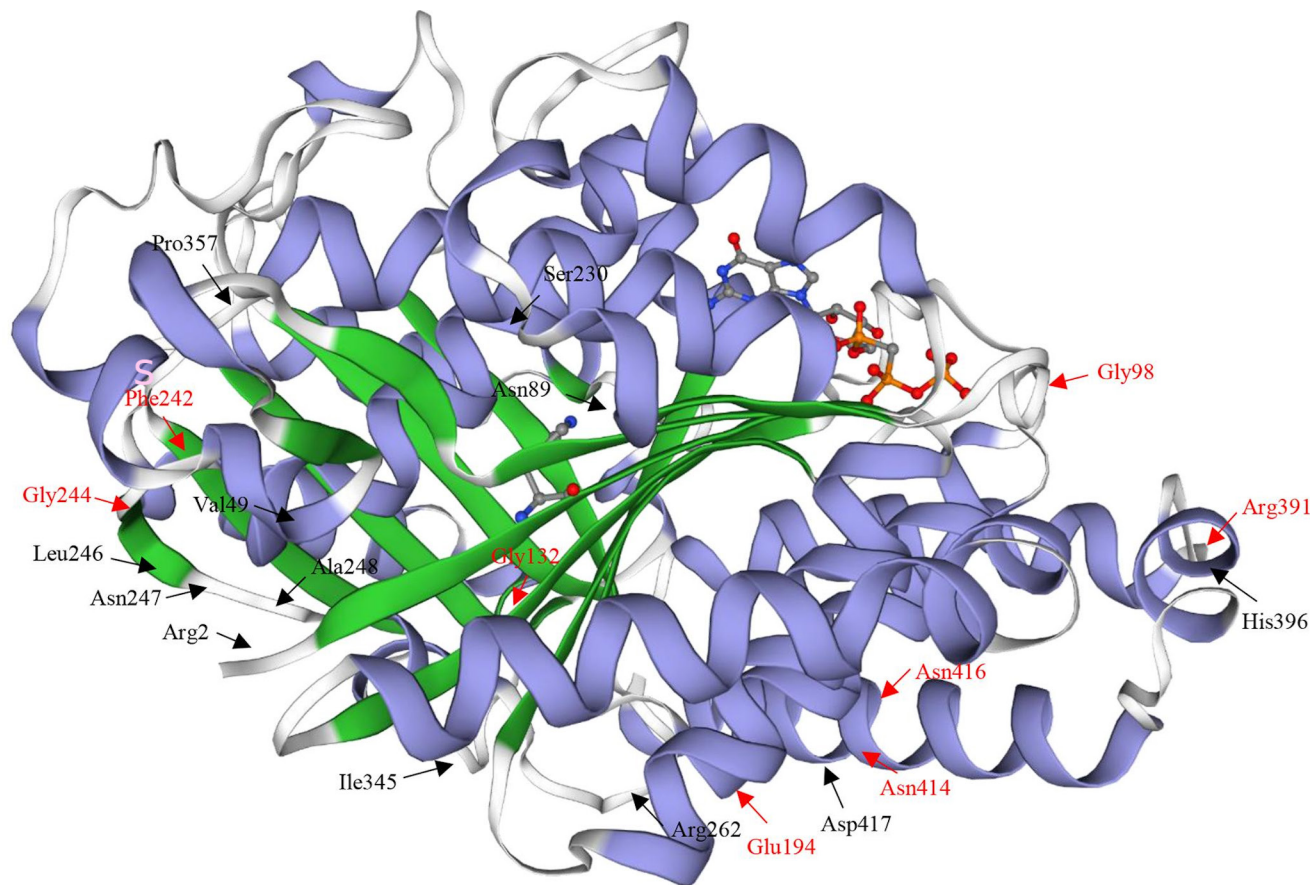
/, not applicable; ASD, autistic spectrum disorder; DCOA, dithionite-diammonium; EEG, electroencephalogram; GMFCS, Gross Motor Function Classification Scale; n/a, not available.



**Figure 1** Brain imaging findings with midline, axial and coronal sections of patients with TUBB2A variants and a normal subject. Patient 1 at age 5 years does not present any brain malformations (1A–C). Patient 2 showing mild dysgyria (arrow denotes fronto-orbital, and asterisk denotes parahippocampal) and a dysmorphic CC (arrowhead) (2A–C). Patient 3 at age 2 months presenting with severe dysgyria (arrows), dysmorphic CC (arrowhead) and BG (asterisk) (3A–C). Patient 4 at age 1 year 11 months (sagittal) and 7 years (axial and coronal) presenting with mild dysgyria (arrows) and a mildly dysmorphic CC (arrowhead) (4A–C). Patient 5 at age 10 months had a normal cortex and hypoplasia of the CC (arrowhead). The white matter was reduced (arrows) (5A–C). BG, basal ganglia; CC, corpus callosum.



**Figure 2** Patient 6 presented with severe bilateral dysgyria (arrow), partial agenesis of the CC (arrowhead) and dysmorphic BG (asterisk) (1A–C). Patient 7 at age 10 months had generalised dysgyria, dysmorphic BG (asterisk), hypoplastic CC (arrowhead), dysgenesis of the cerebellar vermis (diamond), hooked lateral ventricles and reduced white matter volume (arrows) (2A–C). Patient 8 at 8 years 1 month has a normal cortex, mildly enlarged lateral ventricles, hypoplasia of the CC (arrowhead) and vermishypoplasia (arrow) (3A–C). Patient 9 at age 3 years 7 months presenting with a normal cortex, mildly enlarged posterior horns of the lateral ventricles, mildly dysmorphic basal ganglia (asterisk) and reduced white matter (arrow) (4A–C). Healthy control at age 13 months (5A–C). BG, basal ganglia; CC, corpus callosum; M, months; Y, years.



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M E I V H I Q A G C Q N I G A K F W E V I S D E H G I D P T G S Y H G D S D L Q L E R I N Y Y N E A A G N K Y V P R A I L V D L E P G T M D S V R S G P F G Q I F R P D F V F G Q S G A G N N W A K G H Y T E G A E L V D S V L D V V 120
R K E S E S C D C L Q G F L T H S L G G G T G S G M G T L L I S K I R E E Y P D R I M N T F S V M P S P K V S D V V E P Y N A T L S V H Q L V E N T D E T Y S I D N E A L Y D I C F R T L K L T P T Y G D L N H L V A T M S G V T T C L 240
R F P G Q G S D L R K L L A V N M V P F L H F F M P G F A P L T S R G S Q Q Y R A L T V P E L T Q M F D S K N M M A A C D P R H G R Y L T V A A I F R G R M S M K E V D E Q M L N V Q N K N S S Y F V E W P N N V K T A V C D I P R G 360
L K M S A T F I G N S T A I Q E L F K R I S E Q F T A M F R K A F L W Y T G E G M D E M E F T E A E S N M N L V S E Y Q Q Y Q D A T A D E Q G F E F E E E G E D E A 445
    
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**Figure 3** Distribution of variants in the TUBB2A protein structure. Functional domains are highlighted in the 3D model (A) and the linear model (B): green, helices; blue, sheets. Reported variants from this cohort and variants reported in the literature are written in black in the 3D model and blocked in red in the linear model. Variants reported in ClinVar without further clinical information are written in red in the 3D model and are blocked in yellow in the linear model. The 3D structure is based on Swiss model TBB2A\_HUMAN Q13885 tubulin beta-2A chain.

in *TUBB3* causing adult-onset polyneuropathy and for *TUBA4A* causing familial amyotrophic lateral sclerosis.<sup>23,24</sup> As five patients in our cohort have a normal cortex but are significantly younger (2–12 years at last examination) than the patient reported by Sferra *et al.*,<sup>19</sup> further follow-up is needed to assess for features of early-onset neurodegeneration. There was a strong overlap between the severity of motor developmental delay and imaging findings as patients with severe cortical malformations in our cohort were non-ambulatory, whereas patients with mild or without cortical malformations had severe motor delay but were able to walk with support. Further study and long-term follow-up are needed to assess whether the phenotype can be divided into two groups: (1) severely affected, non-ambulatory patients with severe forms of malformations of cortical development (MCD) and (2) initially ambulating patients, possibly with neurodegeneration causing progressive spastic paraplegia, and with mild or without MCD.

Our findings suggest that absent speech is a common feature of *TUBB2A*. Epilepsy was present in all but one patient in our cohort (P10) and in six patients reported in the literature for

which this information was available (17/20 patients, 85%). Interestingly, epilepsy was clinically suspected in patient 10 in our cohort, presenting with mild dysgyria, but could not be confirmed by long-term EEG. Although the reported series remains small, the incidence of epilepsy seems higher in patients with *TUBB2A* compared with other tubulinopathies.<sup>7,25</sup> Microcephaly and visual impairment are variably present. Mild dysmorphic features were present in six (60%) of our patients where this information was available and in one previously reported patient, which is in line with findings in other tubulinopathies.<sup>16,26,27</sup>

Our cohort includes 12 additional patients with 8 previously unreported variants in *TUBB2A*. Brain imaging confirms the heterogeneous spectrum that occurs in patients with variants in *TUBB2A* ranging from a normal cortex to severe dysgyria. This finding should lead to considering *TUBB2A* variants not only in patients with typical imaging findings suggestive of a tubulinopathy but also in patients with developmental delay, epilepsy and mild brain malformations, with or without involvement of the cortex. Especially subtle features of brain malformations

can easily be missed on MRI. Therefore, we would like to stress the importance of reviewing imaging data when a variant in *TUBB2A* is detected during molecular investigations, a process called reverse phenotyping. Imaging and clinical findings have been reported to show mild asymmetry of white matter reduction and pyramidal findings, respectively.<sup>13 15</sup> However, asymmetry has not been observed in our cohort and is uncommon in other tubulinopathies. The brainstem is normal in all patients included in our cohort, and cerebellar hypoplasia or vermis dysplasia occurred in only four patients (33.3%). This contrasts with the high incidence of cerebellar hypoplasia reported for other tubulinopathies.<sup>20</sup> Dysmorphic basal ganglia, considered a key feature of tubulinopathies, was present in only four patients in our cohort and in one patient reported in the literature, thus contrasting the high incidence reported in other tubulinopathies.<sup>7 13</sup>

Several recurrent variants in the *TUBB2A* gene have been observed, suggesting mutational hotspots. For the recurrent p.(Ala248Val) observed in our cohort and the literature, no clear genotype-phenotype correlations could be established. The imaging phenotype associated with the variant at p.(Ala248Val) is heterogenous and varies from mild brain malformations as seen in patients 8 and 9 and reported by Cushion *et al.*,<sup>13</sup> to severe malformations as in patient 12 and as reported by Rodan *et al.*<sup>15</sup> For the patient reported by Retterer *et al.*,<sup>18</sup> no information of brain imaging is available. Furthermore, variants affecting positions 246 and 247 have been described in patients with infantile spasms and SGP, respectively.<sup>13 17</sup> Clinical and imaging features were heterogenous in patients 1 and 3, both carrying variants affecting the same location within the protein, c.145G>A, p.(Val49Met) and c.146T>G, p.(Val49Gly), respectively. Patient 1 appears to have no brain malformations; patient 3 was severely affected, with MRI revealing severe dysgyria, partial corpus callosum agenesis and dysmorphic basal ganglia. Overlapping clinical and imaging features are present in the severely affected patients 6 and 7 in our cohort, both carrying the p.(Pro357Leu) variant. Both patients have bilateral diffuse dysgyria, abnormal corpus callosum, basal ganglia and cerebellum. The variability of genotype and phenotype is likely linked to additional modifiers such as environmental factors or variable gene expression.

Expression of *TUBB2A* in the developing brain has been reported to be less than that of other beta-tubulin isotypes, suggesting a minor role of *TUBB2A* in brain development.<sup>12</sup> More specifically, the role of *TUBB2A* in neuronal migration has been suggested to be less pronounced compared with *TUBA1A* and *TUBB2B*.<sup>8</sup> As variants in the different isotypes lead to unique as well as overlapping phenotypes, variants are likely to selectively affect particular subsets of neurons.

Variants in *TUBB2A* are distributed through the entire gene. Six of all reported variants are located in the GTPase domain, whereas seven are located in the two-layer sandwich domain. Interestingly, when considering the 3D model, variants seem to cluster close to one surface site of the protein, leading to the hypothesis that the 3D protein structure is altered, causing a change in the dynamic capabilities and lateral interactions by changes in the surface structure. Variants in the carboxy-terminal domain have been shown to alter microtubule function<sup>28</sup> and have been reported in *TUBA1A*,<sup>27</sup> *TUBB3*<sup>23</sup> and *TUBG1*.<sup>10</sup> The p.(Asp417Asn) variant interferes with the interaction of *TUBB2A* with *KIF1A* impairing the assembly of spindle poles during mitosis, which ultimately leads to accumulation of mitotic cells with aberrant spindle figures. However, this has also been observed for the p.(Ala248Val) variant, although to a lesser extent.<sup>19</sup>

In conclusion, this patient cohort expands the phenotypical spectrum associated with variants in *TUBB2A* to both milder and more severe ends, highlighting the heterogeneity of clinical and imaging features. As for counselling, variants in *TUBB2A* therefore need to be considered in patients with severe clinical and brain imaging abnormalities, as well as in patients with neurodevelopmental delay and/or epilepsy and MRI reported as normal.

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#### REFERENCES

- Mukhtar T, Taylor V. Untangling cortical complexity during development. *J Exp Neurosci* 2018;12:1179069518759332.
- Leandro-García LJ, Leskelä S, Landa I, Montero-Conde C, López-Jiménez E, Letón R, Cascón A, Robledo M, Rodríguez-Antona C. Tumoral and tissue-specific expression of the major human beta-tubulin isoforms. *Cytoskeleton* 2010;67:214–23.
- Tischfield MA, Engle EC. Distinct alpha- and beta-tubulin isoforms are required for the positioning, differentiation and survival of neurons: new support for the ‘multi-tubulin’ hypothesis. *Biosci Rep* 2010;30:319–30.
- Rice LM, Montabana EA, Agard DA. The lattice as allosteric effector: structural studies of alpha-beta- and gamma-tubulin clarify the role of GTP in microtubule assembly. *Proc Natl Acad Sci U S A* 2008;105:5378–83.
- Kapitein LC, Hoogenraad CC. Building the neuronal microtubule cytoskeleton. *Neuron* 2015;87:492–506.
- Hammond JW, Cai D, Verhey KJ. Tubulin modifications and their cellular functions. *Curr Opin Cell Biol* 2008;20:71–6.
- Bahi-Buisson N, Poirier K, Fournier F, Saillour Y, Valence S, Lebrun N, Hully M, Bianco CF, Boddaert N, Elie C, Lascelles K, Souville I, Beldjord C, Chelly J, LIS-Tubulinopathies Consortium. The wide spectrum of tubulinopathies: what are the key features for the diagnosis? *Brain* 2014;137:1676–700.
- Chakraborti S, Natarajan K, Curiel J, Janke C, Liu J. The emerging role of the tubulin code: from the tubulin molecule to neuronal function and disease. *Cytoskeleton* 2016;73:521–50.
- Romaniello R, Arrigoni F, Fry AE, Bassi MT, Rees MI, Borgatti R, Pilz DT, Cushion TD. Tubulin genes and malformations of cortical development. *Eur J Med Genet* 2018;61:744–54.
- Brock S, Stouffs K, Scalais E, D’Hooghe M, Keymolen K, Guerrini R, Dobyns WB, Di Donato N, Jansen AC. Tubulinopathies continued: Refining the phenotypic spectrum associated with variants in TUBG1. *Eur J Hum Genet* 2018;26:1132–42.
- Saillour Y, Broix L, Bruel-Jungerman E, Lebrun N, Muraca G, Rucci J, Poirier K, Belvindrah R, Francis F, Chelly J. Beta tubulin isoforms are not interchangeable for rescuing impaired radial migration due to Tubb3 knockdown. *Hum Mol Genet* 2014;23:1516–26.
- Breuss M, Heng JI-T, Poirier K, Tian G, Jaglin XH, Qu Z, Braun A, Gstrein T, Ngo L, Haas M, Bahi-Buisson N, Moutard M-L, Passemard S, Verloes A, Gressens P, Xie Y, Robson KJH, Rani DS, Thangaraj K, Clausen T, Chelly J, Cowan NJ, Keays DA. Mutations in the beta-tubulin gene TUBB5 cause microcephaly with structural brain abnormalities. *Cell Rep* 2012;2:1554–62.
- Cushion TD, Paciorkowski AR, Pilz DT, Mullins JGL, Seltzer LE, Marion RW, Tuttle E, Ghoneim D, Christian SL, Chung S-K, Rees MI, Dobyns WB. De novo mutations in the beta-tubulin gene TUBB2A cause simplified gyral patterning and infantile-onset epilepsy. *Am J Hum Genet* 2014;94:634–41.
- Lee H, Deignan JL, Dorrani N, Strom SP, Kantarci S, Quintero-Rivera F, Das K, Toy T, Harry B, Yourshaw M, Fox M, Fogel BL, Martínez-Agosto JA, Wong DA, Chang VY, Shieh PB, Palmer CGS, Dipple KM, Grody WW, Vilain E, Nelson SF. Clinical exome sequencing for genetic identification of rare Mendelian disorders. *JAMA* 2014;312:1880–7.
- Rodan LH, El Achkar CM, Berry GT, Poduri A, Prabhu SP, Yang E, Anselm I. De novo TUBB2A variant presenting with anterior temporal Pachygyria. *J Child Neurol* 2017;32:127–31.
- Ejaz R, Lionel AC, Blaser S, Walker S, Scherer SW, Babul-Hirji R, Marshall CR, Stavropoulos DJ, Chitayat D. De novo pathogenic variant in TUBB2A presenting with arthrogryposis multiplex congenita, brain abnormalities, and severe developmental delay. *Am J Med Genet A* 2017;173:2725–30.
- Yuskaitis CJ, Ruzhnikov MRZ, Howell KB, Allen IE, Kapur K, Dlugos DJ, Scheffer IE, Poduri A, Sherr EH. Infantile spasms of unknown cause: predictors of outcome and genotype-phenotype correlation. *Pediatr Neurol* 2018;87:48–56.
- Retterer K, Juusola J, Cho MT, Vitazka P, Millan F, Gibellini F, Vertino-Bell A, Smaoui N, Neidich J, Monaghan KG, McKnight D, Bai R, Suchy S, Friedman B, Tahiliani J, Pineda-Alvarez D, Richard G, Brandt T, Haverfield E, Chung WK, Bale S. Clinical application of whole-exome sequencing across clinical indications. *Genet Med* 2016;18:696–704.
- Sferra A, Fattori F, Rizza T, Flex E, Bellacchio E, Bruselles A, Petrini S, Cecchetti S, Teson M, Restaldi F, Cioffi A, Santorelli FM, Zanni G, Barresi S, Castiglioni C, Tartaglia M, Bertini E. Defective kinesin binding of TUBB2A causes progressive spastic ataxia syndrome resembling saccinopathy. *Hum Mol Genet* 2018;27:1892–904.
- Oegema R, Cushion TD, Phelps IG, Chung S-K, Dempsey JC, Collins S, Mullins JGL, Dudding T, Gill H, Green AJ, Dobyns WB, Ishak GE, Rees MI, Doherty D. Recognizable cerebellar dysplasia associated with mutations in multiple tubulin genes. *Hum Mol Genet* 2015;24:5313–25.
- Mutch CA, Poduri A, Sahin M, Barry B, Walsh CA, Barkovich AJ. Disorders of microtubule function in neurons: imaging correlates. *AJNR Am J Neuroradiol* 2016;37:528–35.
- Waterhouse A, Bertoni M, Bienert S, Studer G, Tauriello G, Gumienny R, Heer FT, de Beer TAP, Rempfer C, Bordoli L, Lepore R, Schwede T. SWISS-MODEL: homology modelling of protein structures and complexes. *Nucleic Acids Res* 2018;46:W296–303.
- Tischfield MA, Baris HN, Wu C, Rudolph G, Van Maldergem L, He W, Chan W-M, Andrews C, Demer JL, Robertson RL, Mackey DA, Ruddle JB, Bird TD, Gottlob I, Pieh C, Traboulsi EI, Pomeroy SL, Hunter DG, Soul JS, Newlin A, Sabol LJ, Doherty EJ, de Uzcátegui CE, de Uzcátegui N, Collins MLZ, Sener EC, Wabbels B, Hellebrand H, Meitinger T, de Berardinis T, Magli A, Schiavi C, Pastore-Trossello M, Koc F, Wong AM, Levin AV, Geraghty MT, Descartes M, Flaherty M, Jamieson RV, Möller HU, Meuthen I, Callen DF, Kerwin J, Lindsay S, Meindl A, Gupta ML, Pellman D, Engle EC. Human Tubb3 mutations perturb microtubule dynamics, kinesin interactions, and axon guidance. *Cell* 2010;140:74–87.
- Smith BN, Ticozzi N, Fallini C, Gkazi AS, Topp S, Kenna KP, Scotter EL, Kost J, Keagle P, Miller JW, Calini D, Vance C, Danielson EW, Troakes C, Tiloca C, Al-Sarraj S, Lewis EA, King A, Colombrita C, Pensato V, Castellotti B, de Belleruche J, Baas F, ten Asbroek ALMA, Sapp PC, McKenna-Yasek D, McLaughlin RL, Polak M, Asres S, Esteban-Pérez J, Muñoz-Blanco JL, Simpson M, van Rheenen W, Diekstra FP, Lauria G, Duga S, Corti S, Cereda C, Corrado L, Sorarù G, Morrison KE, Williams KL, Nicholson GA, Blair IP, Dion PA, Leblond CS, Rouleau GA, Hardiman O, Veldink JH, van den Berg LH, Al-Chalabi A, Pall H, Shaw PJ, Turner MR, Talbot K, Taroni F, García-Redondo A, Wu Z, Glass JD, Geller C, Ratti A, Brown RH, Silani V, Shaw CE, Landers JE, SLAGEN Consortium. Exome-wide rare variant analysis identifies TUBA4A mutations associated with familial ALS. *Neuron* 2014;84:324–31.
- Romaniello R, Zucca C, Arrigoni F, Bonanni P, Panzeri E, Bassi MT, Borgatti R. Epilepsy in Tubulinopathy: personal series and literature review. *Cells* 2019;8:E669.
- Fallet-Bianco C, Laquerrière A, Poirier K, Razavi F, Guimiot F, Dias P, Loeuillet L, Lascelles K, Beldjord C, Carion N, Toussaint A, Revencu N, Addor M-C, Lhermitte B, Gonzales M, Martinovich J, Bessieres B, Marcy-Bonnière M, Jossic F, Marcourelles P, Loget P, Chelly J, Bahi-Buisson N. Mutations in tubulin genes are frequent causes of various foetal malformations of cortical development including microlissencephaly. *Acta Neuropathol Commun* 2014;2.
- Hebebrand M, Hüffmeier U, Trollmann R, Hehr U, Uebe S, Ekici AB, Kraus C, Krumbiegel M, Reis A, Thiel CT, Popp B. The mutational and phenotypic spectrum of TUBA1A-associated tubulinopathy. *Orphanet J Rare Dis* 2019;14:38.
- Aiken J, Buscaglia G, Bates EA, Moore JK. The alpha-Tubulin gene TUBA1A in Brain Development: A Key Ingredient in the Neuronal Isoform Blend. *J Dev Biol* 2017;5:30008.





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