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Short Communication





Influence of growth regulating chemicals on growth and flowering in Jasmine (Jasminum sambac.Ait.)

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ABSTRACT

Jasmine is an important commercial flower crop in Tamil Nadu. The crop has a main flowering season during March to October and an off-season from November to February. During this off-season, flowering is very poor or there is no flowering in many growing areas. In recent years, growth regulators are valuable in floriculture for manipulating growth and flowering of many crops and hence and attempt has been made to induce flowering during off season using growth regulators in Jasmine in the Floriculture unit of the Department of Horticulture, Faculty of Agriculture, Annamalai University, Tamil Nadu during November, 2016 to February, 2017. The treatment comprises of three concentrations of each of two growth promoting substances viz., NAA and GA, and two growth retardants (Cycocel and Maleic Hydrazide). The experiment was laid out in randomized block design with three replications. Among the various treatments, application of NAA @ 75 ppm (T_c) recorded the highest plant height (130.6 cm and 178.5 cm at 90 and 180 DAP respectively), number of primary shoots (21.68 and 35.68 at 90 and 180 DAP respectively), number of nodes (9.86 and 15.89 cm at 90 and 180 DAP respectively) and number of leaves (1250.0 and 2689.5 at 90 and 180 DAP respectively). Earliness in flowering (26.38 DAP) and maximum duration of flowering (171.00 days) was noticed in (GA₃@ 150 ppm T₂). From the above studies, it is inferred that application of GA, @ 150 ppm could be recommended for enhanced growth and higher flower yield in Jasminum sambac.

Keywords: Jasmine, growth regulators, NAA,GA,CCC, MH,off-season flowering

INTRODUCTION

Jasmine is one of the oldest fragrant flowers. A serious limiting factor is that the flowering of *Jasminum sambac* is seasonal. There are peak and lean productive seasons with consequent gluts and scarcity which affect the price trends greatly. Regulation of plant growth and development using natural plant hormones for greater production has received the utmost attention (Leetham *et al.*, 1978). Growth and flowering responses of flower crops to these chemical substances have been intensively studied with a view to have compact plants with greater number of flowers and also to hasten or delay flowering according to the needs of the grower (Cathey, 1980).

Regulation of flowering in jasmine has immense practical value. Timing of the peak flowering to coincide with the time of greatest demand and generally modifying the flowering sequence to avoid peak production at about the same time would confer great advantage to the grower and consumers. It is in this respect that the possibility of using plant growth regulators for regulation assumes significance. Keeping this in view, the present investigation was therefore taken to study the effect of growth regulators and their concentration on growth and flowering of Jasmine (*Jasminum sambac* Ait.).

An experiment was formulated with two growth promoters (GA₃ and NAA) and two growth retardants (CCC, MH) at three concentrations each were taken up in the floriculture unit during November, 2016 to February, 2017. Thirteen treatments comprising GA3 at 50, 100 and 150 ppm, NAA at 25, 50, 75 ppm, Cycocel at 500, 1000 and 1500 ppm and Maleic hydrazide at 1000, 2000 and 3000 ppm concentration and a control was replicated thrice. Three year old



jasmine plants were selected from the floriculture unit of the Department of Horticulture, Faculty of Agriculture, Annamalai University, Tamil Nadu for the experiment. Four plants were maintained for each treatment per replication along with a control and conventional pruning during second fortnight of October, 2016. After pruning, when the new shoots appeared with sufficient number of leaves, the freshly prepared Gibberellic acid (GA₃), Naphthalene acetic acid (NAA), 2-chloroethyl-trimethyl ammonium chloride; Chlorocholine chloride (Cycocel CCC) and Maleic hydrazide (MH) were sprayed once as per the treatments. The data on vegetative parameters viz., plant height, number of primary shoots, number of nodes, intermodal length, number of leaves and flowering parameters viz., days taken for flowering, duration of flowering, flower yield and hundred flower weight were recorded. The statistical analysis of the data was done by adopting the standard procedure given by Panse and Sukhatme (1967).

The results of the investigation undertaken with a view to study the effect of certain plant growth regulators on flowering and yield of Jasmine (Jasminum sambac Ait.) are presented in Table1 and 2. Among the various treatments, application of NAA @ 75 ppm (T₆) recorded the highest plant height (130.6 cm and 178.5 cm at 90 and 180 DAP, respectively) and it was followed by (T_2) Gibberllic acid @150 ppm which recorded the value of 127.8 cm and 175.5 cm at 90 and 180 DAP, respectively. The PGR'S when applied as foliar spray, were absorbed by the leaves and readily translocated in both xylem and phloem tissues resulting in distribution throughout the plant system. This might be the reason for the enhancement in plant height. The least plant height (98.9 cm and 142.1 cm at 90 and 180 DAP respectively was observed in T₈ (Cycocel @ 1000 ppm). Cycocel has dwarfening effect on the plant (Dole and Wilkins, 1999). This causes the internodes to shorten and stems to harden due to thickness.

Among the various treatments the maximum number of primary shoots was registered in NAA @ 75 ppm (T_6) which recorded the highest values (21.68 and 35.68 at 90 and 180 DAP, respectively). This was followed by the treatment NAA @ 50 ppm (T_5) which recorded the value of 19.88 and 33.18 at 90 and 180 DAP, respectively.

Significant differences were observed among the growth regulators tried in Jasmine. Foliar spray of Gibberellic acid (GA₃ @150 ppm (T₃) resulted in more number of internodes (17.98 and 43.59 at 90 and 180 DAP respectively), which is followed by (T₉) Cycocel @1500 ppm (17.00 and 41.69 at 90 and 180 DAP respectively). However least number of nodes was registered in T₁₃ (Control) with a value of 8.27 and 23.09 at 90 and 180 DAP respectively.

The data presented in Table 1 clearly shows that the length of internodes was significantly increased by the application of NAA @ 75 ppm (T_c) (9.86 and 15.89 cm at 90 and 180 DAP respectively). This was followed by (T_2) Gibberellic acid @150 ppm (8.96 and 14.59 at 90 and 180 DAP respectively). The maximum number of leaves per plant was recorded in the treatment T₃ - GA₃ @150 ppm (1250.0 and 2689.5 at 90 and 180 DAP, respectively). This might be attributed to the enhanced vegetative growth in early phase due to exogenous application of GA₂ which would have favoured the increased photosynthesis and Co₂ fixation. The least number of leaves was registered in T_{13} (Control) a value of 794.4 and 2070.2 at 90 and 180 DAP respectively. The increased effect could be corroborated with the findings of Sridhar et al (2013) and Pappaiah and Muthusamy (1978) in Jasminum auriculatum.

The increased effect of NAA @ 75 ppm for the above characters might be due to the fact that NAA would have promoted vegetative growth by inducing active cell division in the apical meristem. Increase in growth attributes due to the application of the NAA in the present study is in consonance with the findings of Sridhar *et al* (2013). Similar results were also derived by Pal *et al.*, (1980) in Jasmine and Grisha *et al* (2012) in Daisy.

Earliness in flowering (26.38 DAP) was noticed in plants sprayed with GA₃ @ 150 ppm (T₃) followed by GA₃ @100 ppm (T₂) with took 30.38 days. This might be attributed to the enhanced vegetative growth in early phase attributed by exogenous application of GA₃ which would have favoured the increased photo synthesis and CO₂ fixation. Further, it would have favoured convenience of factors influencing floral initiation *ie.*, carbohydrate pathway and photo periodic pathway with GA₃ pathway. These results are in accordance with findings of Baskaran *et al.*, (2007) and Devadanam *et al.*, (2007) in Gladiolus. Delayed

J. Hortl. Sci. Vol. 13(2): 88-94, 2018

Treatments	Plant (cm)	height	No. of primary shoots	mary	Number of nodes	of nodes	Intermo	Intermodal length	Number of plant	Number of Leaves per plant
	90 DAP	180 DAP	90 DAP	180 DAP	90 DAP	180 DAP	90 DAP	180 DAP	90 DAP	180 DAP
T_1 - GA ₃ @ 50 ppm	117.1	165.3	15.68	27.38	13.88	35.59	6.06	10.87	1060.0	2425.8
T_2 - GA ₃ @ 100 ppm	124.2	171.4	17.58	29.98	14.66	37.39	7.86	13.17	1195.6	2598.8
T ₃ - GA ₃ @ 150 ppm	124.8	175.5	18.08	30.58	17.98	43.59	8.96	14.59	1250.0	2689.5
T_4 – NAA @ 25 ppm	120.1	168.3	13.78	24.58	10.92	28.59	6.96	11.97	899.5	2212.2
T_5 – NAA @ 50 ppm	127.2	172.4	19.88	33.18	10.27	27.09	8.06	13.27	849.4	2144.2
$T_6-NAA @ 75 ppm$	130.6	178.5	21.68	35.68	11.57	30.09	9.86	15.89	949.7	2280.2
T_{7} -Cycocel @ 500 ppm	111.5	159.1	7.98	16.38	15.52	39.19	4.26	8.27	1159.0	2548.8
$T_8-Cycocel@ 1000 ppm$	98.9	142.1	9.68	18.78	16.02	39.79	1.26	3.01	1229.0	2653.3
$T_9-Cycocel@ 1500 ppm$	102.4	147.9	11.88	21.68	17.02	41.69	1.76	4.27	1204.5	2608.3
T ₁₀ -MH@ 1000 ppm	110.5	157.0	4.58	11.78	12.22	31.59	4.16	8.17	6:666	2348.2
T ₁₁ –MH@ 2000 ppm	105.0	150.9	6.28	14.28	12.32	32.19	2.36	5.57	1007.4	2357.8
T_{12} – MH@ 3000 ppm	107.6	153.9	9.98	19.18	13.1	33.79	3.26	6.87	1110.3	2489.8
T ₁₃ - Control (Water spray)	114.4	162.2	2.08	7.88	8.27	23.09	5.16	9.67	794.4	2070.2
S.E.D	0.96	1.23	0.34	0.81	0.33	0.83	0.42	0.58	5.98	14.63
C.D (<i>P</i> =0.05)	2.01		0.72	1.7	0.69	1.75	0.89	1.22	12.5	30.58

Table 1. Effect of growth regulators on vegetative parameters of Jasmine (Jasminum sambac Ait.)

J. Hortl. Sci. Vol. 13(2): 88-94, 2018



DAP	Treatments	Days taken for	Total duration of	Flower yield per plant (g)	plant (g)	Mean hundred flower
(up to pruning)emergence) A_3° \$50 ppm36.48152.9038.41452.632 A_3° \$100 ppm30.3816.850443.3459.452 A_3° \$100 ppm30.3816.800460.34615.72 A_3° \$150 ppm26.38171.00460.34615.72 A_3° \$50 ppm43.98142.30364.6447.82 $A \oplus 5$ \$5 ppm47.08141.1036.1450.12 $A \oplus 5$ \$5 ppm47.08141.1036.1451.22 $A \oplus 5$ \$5 ppm33.68165.90420.1451.22 $A \oplus 75$ ppm33.38165.90420.1456.32 $A \oplus 75$ ppm33.38165.90420.1456.32 $A \oplus 75$ ppm33.38165.90425.3457.32 $\phi cocle $60 ppm33.38165.90425.4457.32\phi cocle $100 ppm33.38165.90420.1457.32\phi cocle $100 ppm57.48133.4057.32\phi cocle $100 ppm57.48133.4035.5447.32\phi cole $100 ppm57.48133.4035.5447.32\phi cole $100 ppm57.48133.4035.36447.32\phi cole $100 ppm57.48133.4035.5447.32\phi cole $100 ppm57.48133.4025.9447.32\phi cole $100 ppm57.48133.4023.0447.32\phi cole $100 ppm53.2820.0920.2022.5447.22f f f f f f f f f f f f f f f f f f f $		first flower initiation	flowering (days) (After for flower	90 DAP	120DAP	weight (g)
Λ_{g} 650 ppm36.4815.29038.1.45.6.32 Λ_{g} 6100 ppm30.38168.50443.3.45.4.5 Λ_{g} 6150 ppm26.38171.00460.34615.72 Λ_{g} 6150 ppm26.38142.3036.4.6478.32 Λ_{g} 650 ppm47.08141.1036.1.4501.32 $\Lambda @$ 50 ppm39.68142.90402.1.4501.32 $\Lambda @$ 57 ppm39.68164.90402.1.4547.32 $\Lambda @$ 75 ppm33.38165.90420.1.456.82 $\phi cocl@$ 1000 ppm33.38165.90420.1.456.82 $\phi cocl@$ 1500 ppm33.38165.90420.1.456.82 $\phi cocl@$ 1500 ppm50.18147.6039.5.4473.25 $\phi cocl@$ 1500 ppm50.18135.40329.54473.25 $\phi cocl@$ 1500 ppm50.18133.4033.4.14471.25 $\phi cocl@$ 1500 ppm54.28133.4.0323.0.4473.25 $\phi coll 000 ppm53.28120.20282.54473.25f H@ 3000 ppm53.28120.20282.54347.25f H@ 3000 ppm53.28133.4027.889.89f H@ 3000 ppm53.2813.85.859.89f H@ 3000 ppm53.2813.85.859.89f H@ 3000 ppm53.2813.815.859.89f H@ 3000 ppm53.813.85.859.89f H@ 3000 ppm53.813.815.859.89f H@ 3234.$		(up to pruning)	emergence)			
A_{g} (10 ppm30.38168.50443.34594.52 A_{g} (150 ppm26.38171.00460.34615.72 A_{g} © 150 ppm26.38142.30364.64478.32 $A \oplus 35$ ppm47.08141.10366.1478.32 $A \oplus 50$ ppm47.08141.10366.14501.32 $A \oplus 50$ ppm47.08141.10366.14501.32 $A \oplus 50$ ppm47.08164.90402.1456.32 $A \oplus 50$ ppm408159.60420.1456.32 $Vocel \oplus 500 ppm33.38165.90420.1456.32Vocel \oplus 1000 ppm33.38165.90420.1456.32Vocel \oplus 1000 ppm50.18147.6035.54473.22Vocel \oplus 1500 ppm57.48133.40359.54473.22H \oplus 1000 ppm57.48133.40359.54473.22H \oplus 2000 ppm57.48133.40359.54473.22H \oplus 2000 ppm57.48133.40359.54473.22H \oplus 2000 ppm57.48123.70341.54473.22H \oplus 3000 ppm60.58127.70304.54473.22I H \oplus 3000 ppm53.281.837.5898.22I H \oplus 3000 ppm1.321.87.5898.93I H \oplus 3000 ppm5.7458.7458.7458.72I H \oplus 3000 ppm5.7458.7458.7458.72I H \oplus 3000 ppm1.321.187.5898.93I = 20.512.762.7628.74$	T_1 - GA ₃ @ 50 ppm	36.48	152.90	384.14	526.32	588.96
A_{\odot} (6 150 ppm26.38171.00460.34615.72 $A \otimes 0.5$ ppm43.98142.30364.64615.72 $A \otimes 0.5$ ppm47.08141.10364.64478.32 $A \otimes 0.5$ ppm47.08141.10366.14501.32 $A \otimes 0.5$ ppm39.68164.90402.14547.32 $A \otimes 75$ ppm33.38165.90420.14568.32 $vocel \otimes 500 ppm33.38165.90420.14568.32vocel \otimes 1000 ppm33.38147.60359.54473.22vocel \otimes 1500 ppm50.18133.40335.50473.22H \otimes 1000 ppm54.28133.40335.54473.22H \otimes 2000 ppm57.48133.40335.54473.22H \otimes 2000 ppm57.48133.40335.54473.22H \otimes 2000 ppm57.48133.40323.04497.22H \otimes 3000 ppm60.58120.20282.54407.22L \otimes 3000 ppm60.581.187.589.89P = 0.052.763.1815.8519.68$	T_2 - GA ₃ @ 100 ppm	30.38	168.50	443.34	594.52	595.66
A \odot 25 ppm43.98142.30364.64478.32A \odot 50 ppm47.08141.10366.14478.32A \odot 75 ppm39.68141.10366.14501.32A \odot 75 ppm39.68164.90402.14547.32A \odot 75 ppm33.38165.90420.14568.32vocel \odot 100 ppm33.38165.90425.34573.52vocel \odot 150 ppm50.18147.60359.54473.22vocel \odot 150 ppm57.48133.40359.54473.22H \odot 100 ppm57.48133.40323.04451.22H \odot 2000 ppm57.48133.40323.04473.22H \odot 2000 ppm57.48137.70304.54477.22H \odot 2000 ppm57.581.187.589.89P $= 0.05$ 27.5427.5427.5427.54P $= 0.05$ 27.5427.5427.5427.54P $= 0.05$ 27.5427.5427.54	T ₃ - GA ₃ @ 150 ppm	26.38	171.00	460.34	615.72	600.76
A @ 50 pm 47.08 141.10 366.14 501.32 A @ 75 ppm 39.68 164.90 402.14 501.32 A @ 75 ppm 39.68 164.90 402.14 547.32 vocel@ 500 ppm 40.88 159.60 402.14 543.32 vocel@ 100 ppm 33.38 165.90 426.14 563.32 vocel@ 1500 ppm 50.18 147.60 359.54 473.22 vocel@ 1500 ppm 54.28 135.50 341.54 473.22 H@ 1000 ppm 54.28 135.40 341.54 473.22 H@ 1000 ppm 57.48 133.40 323.04 451.22 IH@ 2000 ppm 60.58 127.70 304.54 473.22 Ontrol (Water spray) 53.28 120.20 282.54 477.22 IH@ 3000 ppm 60.58 127.70 304.54 477.22 IH@ 2000 ppm 53.24 323.04 479.22 IH@ 3000 ppm 53.28 7.58 9.89 IH@ 3000 ppm 53.24 <	T_4 -NAA@ 25 ppm	43.98	142.30	364.64	478.32	563.66
AA@ 75 ppm 39.68 164.90 402.14 547.32 ycocel@ 500 ppm 40.88 159.60 420.14 568.32 ycocel@ 100 ppm 33.38 155.90 425.34 568.32 ycocel@ 1500 ppm 50.18 147.60 359.54 473.22 ycocel@ 1500 ppm 54.28 135.50 341.54 471.32 H@ 1000 ppm 54.28 135.40 323.04 451.22 H@ 2000 ppm 57.48 133.40 323.04 451.22 H@ 2000 ppm 57.32 323.04 323.04 451.22 Ontrol (Water spray) 53.28 127.70 304.54 457.22 Ontrol (Water spray) 53.28 120.20 282.54 407.22 Control (Water spray) 53.28 120.20 282.54 407.22 P=0.05 2.76 34.54 60.58 989	$T_5-NAA @ 50 ppm$	47.08	141.10	366.14	501.32	565.16
ycocel@ 500 ppm 40.88 159.60 420.14 568.32 ycocel@ 1000 ppm 33.38 165.90 425.34 573.52 ycocel@ 1500 ppm 50.18 147.60 359.54 573.52 ycocel@ 1500 ppm 54.28 135.50 341.54 473.22 H@ 1000 ppm 57.48 133.40 323.04 451.22 H@ 2000 ppm 60.58 133.40 323.04 429.22 H@ 3000 ppm 60.58 127.70 304.54 407.22 Control (Water spray) 53.28 120.20 282.54 382.22 Doutrol (Water spray) 53.28 1.18 7.58 9.89	$T_6-NAA @ 75 ppm$	39.68	164.90	402.14	547.32	582.36
ycocel@ 1000 ppm33.38165.90425.34573.52ycocel@ 1500 ppm50.18147.60359.54473.22fH@ 1000 ppm54.28135.50341.54451.22fH@ 2000 ppm57.48133.40323.04451.22fH@ 3000 ppm60.58127.70304.54407.22fU@ 3000 ppm53.28120.20282.54382.22fU@ 3000 fpm1.321.187.589.89	T_7 -Cycocel @ 500 ppm	40.88	159.60	420.14	568.32	576.76
$\phi cocel@$ 1500 ppm50.18147.60359.54473.22 $H@$ 1000 ppm54.28135.50341.54451.22 $H@$ 2000 ppm57.48133.40323.04429.22 $H@$ 3000 ppm60.58127.70304.54407.22Control (Water spray)53.28120.20282.54382.22 $D = 0.05$ 2.763.187.589.89	$T_8-Cycocel @ 1000 ppm$	33.38	165.90	425.34	573.52	587.46
IH@ 1000ptm 54.28 135.50 341.54 451.22 IH@ 2000ptm 57.48 133.40 323.04 429.22 IH@ 3000ptm 60.58 127.70 304.54 407.22 Control (Water spray) 53.28 120.20 282.54 382.22 P=0.05 2.76 3.18 7.58 9.89	$T_9-Cycocel @ 1500 ppm$	50.18	147.60	359.54	473.22	571.06
IH@ 2000 ppm 57.48 133.40 323.04 429.22 IH@ 3000 ppm 60.58 127.70 304.54 407.22 Control (Water spray) 53.28 120.20 282.54 382.22 D=0.05 2.76 3.18 15.85 19.68	T_{10} -MH@ 1000 ppm	54.28	135.50	341.54	451.22	557.96
IH@ 3000 ppm 60.58 127.70 304.54 407.22 Control (Water spray) 53.28 120.20 282.54 382.22 1.32 1.18 7.58 9.89 p=0.05) 2.76 3.18 15.85 19.68	T_{11} – MH@ 2000 ppm	57.48	133.40	323.04	429.22	550.16
Control (Water spray) 53.28 120.20 282.54 382.22 1.32 1.18 7.58 9.89 \$=0.05\$) 2.76 3.18 15.85 19.68	T ₁₂ –MH@ 3000 ppm	60.58	127.70	304.54	407.22	543.96
1.32 1.18 7.58 9.89 $p=0.05$ 2.76 3.18 15.85 19.68	T13 - Control (Water spray)	53.28	120.20	282.54	382.22	533.96
2.76 3.18 15.85 19.68	S.E.D	1.32	1.18	7.58	9.89	0.91
	C.D (<i>P</i> =0.05)	2.76	3.18	15.85	19.68	1.92

Table 2. Effect of growth regulators on flowering parameters of Jasmine (Jasminum sambac Ait.)



Dhanasekaran



flowering was recorded in T_{12} MH @ 3000 ppm (60.58 days). This might be due to lesser mitotic activity and preservation of bio-synthesis of gibberellic acid like substances. These results are in agreement with the findings of Sen and Maharana (1971) and Dutta *et al.*, (1993) in Chrysanthemum. Dalal *et al.*, (2009) stated that the delayed flowering might be due to influence of growth retardants in reducing the partition of carbohydrates to floral organ when compared to control.

Maximum flowering duration was in GA₃ @150 ppm (T₃), which recorded 171.00 days, followed by GA₃ @ 100 ppm (T₂) with168.5 days. This might be attributed to the enhanced vegetative growth in early phase attributed by exogenous application of GA₃ would have favoured carbohydrate pathway. The longevity of flowering was also observed by Sridhar *et al.*, (2013) in *Jasminum auriculatum* and Dalal *et al.*, (2009) in chrysanthemum.

The minimum duration of flowering was recorded in control (T_{13}) with 120.20 days. Among the various treatments significant increase in flower yield per plant was recorded in GA₃ @ 150 ppm (T_3) with a value of 460.34 and 615.72 g plant⁻¹ at 90 and 120 DAP respectively. This is followed by GA₃ @ 100 ppm (T_2) 443.34 and 594.52 g plant⁻¹ at 90 and 120 DAP respectively. The increase in weight of flowers might be due to the production of more number

of secondary shoots at early stage, which then had sufficient time to accumulate reserve carbohydrates for proper flower bud differentiation. The lowest yield was observed in T₁₃-Control (282.54 and 382.22 g plant⁻¹) at 90 and 180 DAP respectively. Among the treatments, $GA_3 @ 150 \text{ ppm}(T_3)$ recorded the highest value of 600.76 g followed by $GA_3 @ 100 \text{ ppm} (T_2)$ (595.66g). The lowest hundred bud weight was recorded in T_{13} - Control (533.96 g). This might be attributed to the enhanced vegetative growth in early phase attributed by exogenous application of GA₂ which would have favoured the increased photosynthesis and CO₂ fixation. In the present study, the data on extended duration of flowering and hundred bud weight due to the application of GA, @ 150 ppm might also be a cause for increase in yield.

Earlier reports indicated that the growth and yield of *Jasminum grandiflorum* was enhanced by application of GA_3 (Bhattarcharjee, 1983), Narayana Gowda (1985) and Sridhar *et al.*, (2013).Similar findings were also recorded by Amit kumar *et al.*, (2011) in African marigold, Shinde *et al.*, (2010) in chrysanthemum.

From the above studies, it is inferred that application of GA_3 @ 150 ppm could be recommended for enhanced growth and higher flower yield in *Jasminum sambac*.

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