



Short communication

## Effect of *panchagavya* and GA<sub>3</sub> on germination and seedling growth in cashew (*Anacardium occidentale* L.)

L.S. Singh, A. Pariari<sup>1</sup> and Gopal Shukla<sup>2</sup>

Central Plantation Crops Research Institute  
Research Centre, Kahikuchi, Guwahati - 781 017, India  
E-mail: singhleichombam@gmail.com

### ABSTRACT

An experiment consisting of three sowing periods (March-May, June-August and September-November) and seven pre-sowing treatments was undertaken to study the effect of these factors on seed germination and initial seedling growth in cashew. Seeds sowing during June - August gave significantly better germination and initial seedling growth. However, maximum germination percentage, maximal seedling growth and minimum days to germination were observed with GA<sub>3</sub> 200ppm during all three sowing periods compared to that in other treatments. As for *panchagavya*, @ 10% and 20%, was found to be beneficial in treated seeds. All the growth parameters studied were also superior with GA<sub>3</sub> application, excepting root growth. Best root growth was recorded with *panchagavya* at 20%.

**Key words:** Cashew, *Anacardium occidentale* L., GA<sub>3</sub>, germination, growth, *panchagavya*

Cashew (*Anacardium occidentale* L.) is native to South America (Mitchell and Mori, 1987) and was introduced into India during the 16<sup>th</sup> Century by the Portuguese (Johnson, 1973). It is now an important commercial-plantation crop in India, grown mainly along the East coast (Tamil Nadu, Andhra Pradesh, Orissa and West Bengal), West coast (Kerala, Karnataka, Maharashtra and Goa) and the North-Eastern Region (Meghalaya, Manipur, Assam, Tripura and Nagaland). It is widely grown in Asia, Africa and South America. Its kernel is highly nutritive (Jain *et al.*, 1954; Morton, 1961; Joseph, 1975) containing about 21% protein, 22% carbohydrates and 41% fats. Production of quality planting material is of utmost important for which the seedlings need to be healthy, free from diseases and pests, and the seed should contain sufficient amount of auxin for good germination, rendering them ideal for softwood grafting. Owing to its significant contribution to the national economy, there is a huge demand for quality planting material both for area expansion and replacement of old and unproductive orchards. Huballi (2009) reported a requirement of about 1.25 crore grafts to cover nearly 50,000 hectares, on annual basis. Therefore, to meet an increasing demand, there is a need to produce quality planting material at a rapid rate. Cashew seed is recalcitrant by nature, and year round production of healthy planting material is difficult, as, viability of the seed deteriorates rapidly upon storage

(Aravindakshan and Gopi Kumar, 1979; Mandal, 2000). To facilitate its germination, the seed must be provided favourable environmental conditions such as adequate moisture supply, appropriate gaseous balance and optimum light. It is necessary to enhance germination while maintaining uniformity of seedlings. With this in view, the present study was undertaken to standardize period of sowing and pre-sowing treatment for optimum germination and good growth in cashew seedlings.

The experiment was conducted over two consecutive years (2009 and 2010), at Horticultural Research Station (HRS), Mondouri, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal. The experimental site is located at 23°N latitude and 80°E longitude, at an elevation of 9.75 meters above mean sea level, with the sub-tropical climate of the region providing average annual rainfall of 154.7cm from the South-West monsoon.

Freshly-harvested seeds of cashew were used for June-August sowing, whereas, stored seeds were used during March-May (9 months) and September-November (4 months) for sowing. Seeds were selected based on sinker and floater method, i.e., seeds that sank in water alone were considered for sowing. A hundred seeds were sown per treatment in polythene bags (size 26cm x 17cm) filled with FYM, sand and soil in 1:1:1 ratio for germination. Seeds

<sup>1</sup>Department of Spices and Plantation Crops, Faculty of Horticulture, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur - 741 252, Nadia, West Bengal,

<sup>2</sup>Department of Forestry, Uttar Banga Krishi Viswavidyalaya, Pundibari-736 165 (Cooch Behar) West Bengal

that produced 5mm or longer radicals were taken as germinated. Growth parameters such as seedling height, collar diameter, number of leaves, shoot and root dry weight, were recorded at 60 days after germination. The experiment was laid out in Factorial Randomized Block Design, with seven treatments and three replications. Seeds were sown during the first week each of March, June and September for the three sowing periods (March-May, June-August and September-November). Details of treatments are: 5% *Panchagavya* ( $T_1$ ), 10% *Panchagavya* ( $T_2$ ), 20% *Panchagavya* ( $T_3$ ),  $GA_3$  50ppm ( $T_4$ ),  $GA_3$  100ppm ( $T_5$ ),  $GA_3$  200ppm ( $T_6$ ), and no treatment, i.e., Control ( $T_7$ ). Seeds were soaked in these solutions for three hours. *Panchagavya* was prepared by mixing cow dung (2.5kg), cow urine (1.5 litre), *ghee* i.e. clarified butter (500g), cow milk (1 litre), curd (1 litre) and jaggery (500g). Cow dung, cow urine and *ghee* were mixed in a plastic bucket and stirred continuously for a week to remove methane gas. Then, cow milk, curd and jaggery were added and the mixture stirred and kept aside for a week. The extracts were weighed and diluted in water to prepare 5%, 10% and 20% *panchagavya* solution. Data were statistically analyzed as per Gomez and Gomez (1984).

From seeds sown during three different periods during a year, it was found that June - August was best, yielding the highest germination in a very short time, whereas, March - May was not suitable, as, it resulted in the lowest germination rate (Table 1). Seed germination and growth of cashew seedlings were significantly influenced by pre-sowing treatment (Table 1). Pre-sowing treatment with  $GA_3$  at 200ppm gave 100% germination within 8.5 days, followed by  $GA_3$  at 100ppm (99% germination in 9 days). Lowest germination (90% in 13.5 days) was recorded in Control during June - August sowing. Sowing period September - November also gave germination rates similar to the pre-

sowing treatment of June - August. Results on sowing period confirm that cashew sown during two seasons (March-May and September-November) in West Bengal produces better germination, without substantial deterioration to the crop. This may also be due to the prevalent favourable climatic conditions and high viability retained in a fresh seed during this period. Pre-sowing treatment also gave uniform and quick germination. Similar effect with  $GA_3$  treatment has been reported (Furuta, 1961). It is likely that this treatment removes the waxy layer of the pericarp, thereby facilitating better germination (Harris *et al*, 1994). Three *panchagavya* treatments were also tried and similar effects were recorded on germination in June - August sowing, whereas, lowest values were recorded in March - May sowing. During September - November sowing, germination percentage ranged from 62% to 83%. Maximum germination percentage was recorded with  $GA_3$  200ppm (Fig. 1). Beneficial effect of treatment of seeds with *panchagavya* on germination has also been reported by Pathak and Ram (2004).



Fig. 1. Cashew seedlings treated with  $GA_3$  200ppm during June - August sowing

Table 1. Effect of pre-sowing treatment on germination and number of days to germination in cashew seed

Treatment	March - May		June - August		September - November	
	Germination (%)	Days taken to germination	Germination (%)	Days taken to germination	Germination (%)	Days taken to germination
<i>Panchagavya</i> 5%	38.0 (38.06)	17.50	95.0 (77.08)	13.00	66.0 (54.33)	14.50
<i>Panchagavya</i> 10%	51.0 (45.57)	16.00	97.0 (80.02)	10.00	74.0 (59.34)	11.50
<i>Panchagavya</i> 20%	54.0 (47.29)	15.00	96.0 (78.46)	12.50	70.0 (56.79)	13.50
$GA_3$ 50ppm	47.0 (43.28)	16.50	92.0 (73.57)	11.50	67.0 (54.94)	14.00
$GA_3$ 100ppm	52.0 (46.15)	15.00	99.0 (84.26)	9.00	76.0 (60.67)	11.00
$GA_3$ 200ppm	61.0 (51.35)	13.50	100.0 (90.00)	8.50	83.0 (65.65)	9.00
Control	31.0 (33.83)	18.00	90.0 (71.56)	13.50	62.0 (51.94)	14.50

Figures in parentheses are logarithmic transformed values

**Table 2. Effect of pre-sowing treatment on initial seedling growth in cashew**

Treatment	March - May			June - August			September - November		
	Plant height (cm)	Seedling diameter (cm)	No. of leaves	Plant height (cm)	Seedling diameter (cm)	No. of leaves	Plant height (cm)	Seedling diameter (cm)	No. of leaves
<i>Panchagavya</i> 5%	18.10	0.74	9.48	22.92	0.88	11.03	22.71	0.85	11.17
<i>Panchagavya</i> 10%	19.01	0.77	9.64	24.31	0.96	12.20	22.80	0.90	11.38
<i>Panchagavya</i> 20%	21.07	0.88	10.56	24.73	1.05	12.55	23.82	0.91	11.94
GA <sub>3</sub> 50ppm	18.38	0.76	9.40	23.06	0.92	11.67	22.65	0.86	11.35
GA <sub>3</sub> 100ppm	20.99	0.86	10.59	25.18	1.05	12.67	24.36	1.02	12.23
GA <sub>3</sub> 200ppm	23.15	1.00	11.64	27.25	1.15	13.91	26.05	1.03	13.12
Control	17.93	0.70	8.83	22.62	0.82	10.83	21.82	0.82	10.98
S.Em. (±)	0.37	0.03	0.25	0.43	0.04	0.17	0.45	0.03	0.30
C.D. ( <i>P</i> =0.05)	1.14	0.09	0.75	1.30	0.12	0.52	1.35	0.11	0.90

**Table 3. Effect of pre-sowing treatment on seedling growth of cashew**

Treatment	March - May			June - August			September - November		
	Leaf area (cm <sup>2</sup> )	Root length (cm)	Shoot: root length	Leaf area (cm <sup>2</sup> )	Root length (cm)	Shoot: root length	Leaf area (cm <sup>2</sup> )	Root length (cm)	Shoot: root length
<i>Panchagavya</i> 5%	27.27	14.61	1.22	27.75	17.80	1.28	29.31	17.62	1.28
<i>Panchagavya</i> 10%	31.01	13.88	1.30	31.84	19.22	1.27	26.96	17.56	1.32
<i>Panchagavya</i> 20%	25.56	15.65	1.34	34.69	18.68	1.32	31.76	18.66	1.27
GA <sub>3</sub> 50ppm	26.19	14.35	1.27	26.58	17.80	1.32	24.36	16.58	1.36
GA <sub>3</sub> 100ppm	26.01	14.82	1.40	35.06	17.81	1.42	35.41	17.23	1.41
GA <sub>3</sub> 200ppm	31.24	15.55	1.48	37.75	18.59	1.46	32.62	17.47	1.48
Control	23.52	14.79	1.28	28.28	18.57	1.21	28.86	16.40	1.32
S.Em. (±)	0.77	0.36	0.03	1.38	0.25	0.03	1.26	0.43	0.01
C.D. ( <i>P</i> =0.05)	2.34	1.10	0.10	4.21	0.76	0.09	3.82	1.31	0.05

Effect of pre-sowing treatment on initial seedling-growth in cashew is presented in Table 2. Initial seedling-growth showed similar trends in germination, and sowing during June - August recorded better growth, followed by September - November. The least growth was recorded in March - May sowing. Seeds pre-soaked in GA<sub>3</sub> 200ppm recorded maximum plant height, collar diameter and number of leaves, while, the least plant height, collar diameter and number of leaves were recorded in Control in all three periods of sowing. This may be due to early and uniform germination supported by GA<sub>3</sub>, hastening initiation of shoot growth, thus leading to better seedling height. Further, application of GA<sub>3</sub> may have also helped increase cell division, leading to better initial shoot-growth. Similar results were reported by Walase *et al* (2007) and Shanmugavelu (1963; 1970). All the three treatments with *panchagavya* gave better seedling growth during June - August sowing, with the highest seedling height, collar diameter and number of leaves, followed by September - November sowing. The lowest values were recorded in March - May sowing. Yelleshkumar *et al* (2008) reported that seeds treated with 3% *panchagavya* were superior in sprout height, seedling diameter, number of sprouts and number of leaves in mango. Leaf area, root length and

root:shoot ratio were highest when sowing was done during June - August; all the parameters studied were lowest in March - May sowing (Table 3). Pre-sowing treatment also influenced leaf area, root growth and root:shoot ratio. Maximum leaf area (37.75cm<sup>2</sup>) and root:shoot ratio (1.46) was recorded in GA<sub>3</sub> treatment in sowing during June - August, while, maximum root growth was recorded in 10% *panchagavya*. Lowest values were associated with GA<sub>3</sub> 50ppm and 5% *panchagavya*. Root growth was unaffected, as, pre-treated seeds in all the sowings gave similar root growth. It is very interesting to note that application of *panchagavya* produced better root growth irrespective of sowing time. According to Solaiappan (2002), *panchagavya* has beneficial microorganisms like *Azospirillum*, *Azotobacter*, *phosphobacteria* and *Lactobacillus*, which may be the reason for good root growth. The present findings are in conformity with Yabuta and Hayashi (1939) and Sumiki (1952). Fresh and dry root and shoot weights are presented in Table 4. Similar trends were observed in June - August sowing, which showed maximum fresh and dry root and shoot weights, while, lowest values were recorded in March - May sowing. Seeds treated with GA<sub>3</sub> 200ppm produced higher shoot growth and weight in the three different sowing

**Table 4. Effect of pre-sowing treatment on fresh and dry shoot and root weight of cashew seedling**

Treatment	March - May				June - August				September - November			
	Shoot fresh wt. (g)	Shoot dry wt. (g)	Root fresh wt. (g)	Root dry wt. (g)	Shoot fresh wt. (g)	Shoot dry wt. (g)	Root fresh wt. (g)	Root dry wt. (g)	Shoot fresh wt. (g)	Shoot dry wt. (g)	Root fresh wt. (g)	Root dry wt. (g)
<i>Panchagavya</i> 5%	3.70	0.77	1.06	0.15	6.76	1.47	1.72	0.44	5.61	1.05	1.67	0.45
<i>Panchagavya</i> 10%	4.38	0.91	1.24	0.21	6.95	1.57	1.97	0.65	6.06	1.13	1.75	0.43
<i>Panchagavya</i> 20%	5.16	1.04	1.45	0.33	7.41	1.72	1.88	0.59	6.09	1.18	1.78	0.60
GA <sub>3</sub> 50ppm	4.02	0.77	1.13	0.13	6.45	1.48	1.66	0.42	5.11	0.96	1.39	0.24
GA <sub>3</sub> 100ppm	5.20	1.07	1.24	0.21	7.73	1.97	1.70	0.41	6.46	1.50	1.53	0.30
GA <sub>3</sub> 200ppm	5.72	1.20	1.30	0.23	8.11	2.01	1.86	0.58	7.29	1.81	1.58	0.26
Control	3.83	0.74	1.26	0.21	6.19	1.52	1.81	0.57	4.91	0.92	1.40	0.25
S.Em. (±)	0.30	0.10	0.05	0.02	0.49	0.10	0.02	0.01	0.51	0.22	0.07	0.07
C.D. ( <i>P</i> =0.05)	1.01	0.34	0.16	0.08	1.46	0.29	0.07	0.07	1.56	0.61	0.20	0.22

periods. This could be related to broader leaves, thereby enhanced photosynthetic activity and accumulation of nutrients in the leaf tissues. This, in turn, may have helped improve shoot growth and dry biomass.

It may be concluded from the present study that sowing cashew during June -August or September - November was better for germination rate and initial seedling growth. Treatment with GA<sub>3</sub> 200ppm was the most effective for germination and better initial seedling growth in cashew during the three different sowing periods. However, GA<sub>3</sub> may not be a suitable option to a farmer due to its high price and problem with availability. Therefore, pre-sowing treatment with the farmer-friendly *panchagavya* (20%) can be adopted due to its easy availability and economic viability.

## ACKNOWLEDGEMENT

The authors are thankful to University Grants Commission (UGC), New Delhi, for providing financial assistance through Rajiv Gandhi National Fellowship, and to Dr. Mini Poduval, Reader (Research) and Officer-in-Charge, AICRP on Cashew, Regional Research Station (RRS), Jhargram, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, for providing necessary facilities and for the co-operation extended during the investigation.

## REFERENCES

- Aravindakshan, M. and Gopikumar, K. 1979. Seed viability in cashew. *Cashew Bulletin*, **16**:6-7
- Furuta, T. 1961. Influence of gibberellins on germination of seeds. *Amer. Camellia Yearbook*, pp. 141-145
- Gomez, K.A. and Gomez, A.A. 1984. *Statistical Procedure for Agricultural Research* (2<sup>nd</sup> Ed.),

- Interscience Publication, Johan Wiley and Sons, New York, pp. 20-30
- Harris, C.V., Pandian, I.R.S. and Thangavelu, S. 1994. Pre-treatment of cashew seeds to improve germination. *South Indian Hort.*, **42**:121-122
- Huballi, V.N. 2009. Cashew in India. *Proceedings of Cashew Field Day*, February 20, Bidhan Chandra Krishi Viswavidyalaya, Jhargram, Paschim Midnapur, West Bengal, pp. 8-14
- Jain, N.L., Das, D.P. and Lal, G. 1958. Utilization of cashew apples. Procs. of the symposium on Fruit and Vegetable Preservation Industry in India, CFTRI, Mysore, pp.75-80
- Johnson, D. 1973. The botany, origin, spread of cashew (*Anacardium occidentale* L.). *J. Pl. Crops*, **1**:1-7
- Joseph, K.T. 1975. Cashew nut: A valuable nutritive food product. *Indian Cashew J.*, **10**:5-6
- Mandal, R.C. 2000. *Cashew Production and Processing Technology*. Agrobios (India) Publishers, Ludhiana, pp. 22-31
- Mitchell, J.D. and Mori, S.A. 1987. The cashew and its relatives (*Anacardium occidentale* L.), Anacardiaceae. *Memoirs of the New York Bot. Gardens*, **42**:1-76
- Morton, J.F. 1961. The cashew's brighter future. *Econ. Bot.*, **15**:57-78
- Pathak, R.K. and Ram, R.A. 2004. *Manual on Vedic Krishi*. Central Institute for Subtropical Horticulture, Rehmankhera, Lucknow, pp. 1-38
- Shanmugavelu, K.G. 1963. Studies on the effect of plant growth regulators on some forest plant species. Ph.D. Thesis, Annamalai Univ., Tamil Nadu, India
- Shanmugavelu, K.G. 1970. Effect of gibberellic acid on seed germination and development of seedlings of some tree plant species. *Madras Agril. J.*, **55**: 311-314
- Solaiappan, A.R. 2002. Microbiological studies in

- panchagavya*. *Bio-control Laboratory - Official Communication*, Chengalpet, Tamil Nadu, India
- Sumiki, Y. 1952. Biochemistry of the Bakane fungus. XXV. The Physiological action of gibberellins. *J. Agril. Chem. Soc. (Japan)*, **26**:393-397
- Walase, S.R., Ghawade, D.M., Panchbhai and Dod, V.N. 2007. Effect of GA<sub>3</sub> and chemicals on seed germination and seedling growth of aonla. *Souvenir and Abstracts*, Second Indian Horticulture Congress, April 18-21, ICAR Complex for North Eastern Region (NER), Barapani, Meghalaya, India, pp. 138
- Yabuta, J. and Hayashi, T. 1939. Biochemical studies - Bakane fungus of the rice. III. Physiological action of gibberellins on the plants. *J. Agril. Chem. Soc. (Japan)*, **15**:403-413
- Yelleshkumar, H.S., Swamy, G.S.K., Patil, C.P., Kanamadi, V.C. and Kumar, P. 2008. Effect of pre-soaking treatments on the success of soft-wood grafting and growth of mango grafts. *Karnataka J. Agril. Sci.*, **21**:471-472

(MS Received 02 September 2013, Revised 30 June 2015, Accepted 29 August 2015)