



# *In vivo* anti-hyperglycemic activity of saliva extract from the tropical leech *Hirudinaria manillensis*

Abdualkader, Abdulrahman Mohammed<sup>1\*</sup>, Ghawi, Abbas Mohammad<sup>2</sup>, Alaama, Mohamed<sup>1</sup>, Awang, Mohamed<sup>1</sup>, Merzouk, Ahmed<sup>3</sup>

<sup>1</sup>Pharmaceutical Chemistry Department, Faculty of Pharmacy, International Islamic University Malaysia, 25200 Kuantan, Pahang, Malaysia;

<sup>2</sup>Basic Medical Science Department, Faculty of Pharmacy, International Islamic University Malaysia, 25200 Kuantan, Pahang, Malaysia;

<sup>3</sup>Biopep Solutions Inc., 235-11590 Cambie Road, Richmond, BC V6X 3Z5, Canada

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**[ABSTRACT]** The anticoagulant effect of leech saliva was traditionally employed in the treatment of diabetes mellitus complications such as peripheral vascular complications. This study was carried out to examine the effect of leech saliva extract (LSE) on blood glucose levels in alloxan-induced diabetic rats. First, LSE was collected from leeches which were fed on a phagostimulatory solution. Second, total protein concentration was estimated using the Bradford assay. Third, diabetic rats were injected subcutaneously (sc) with LSE at doses of 500 and 1 000  $\mu\text{g}\cdot\text{kg}^{-1}$  body weight (bw). Other diabetic rats were injected sc with insulin at doses of 10 and 20  $\text{U}\cdot\text{kg}^{-1}$  bw. Another group was injected simultaneously with LSE (250  $\mu\text{g}\cdot\text{kg}^{-1}$  bw) and insulin (10  $\text{U}\cdot\text{kg}^{-1}$  bw). Fasting blood glucose (FBG) concentrations were monitored during a study period of eight hours at regular intervals. Findings showed that both doses of LSE resulted in a significant and gradual decrease in FBG starting from 10%–18% downfall after two hours of injection reaching the maximal reduction activity of 58% after eight hours. Remarkably, LSE was sufficient to bring the rats to a near norm-glycemic state. The high dose of insulin induced a severe hypoglycemic condition after 2–4 h of injection. The lower dose was able to decline FBG for 2–6 h in rats which became diabetic again after 8 h. On the other hand, the concurrent injection of low doses of LSE and insulin produced a hypoglycemic effect with all rats showing normal FBG levels. Taken together, these findings indicated that the subcutaneous injection of LSE of the medicinal Malaysian leech was able to provide better glycemic control compared with insulin. Moreover, the synergism between LSE and insulin suggests that LSE could be utilized as an adjuvant medication in order to reduce insulin dosage or to achieve better control of blood glucose.

**[KEY WORDS]** Alloxan; Diabetes mellitus; Insulin; Leech; *Hirudinaria manillensis*

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## 1 Introduction

Amongst haematophagous animals, leeches have been known as familiar therapeutic tools since the ancient ages until the present day. The word leech is etymologically de-

rived from the Anglo-Saxon word for physician ‘*laece*’ which clearly represents the importance of leeches in medicine [1]. Traditionally, leech application was used for many human body disorders starting from the conventional usage of leeches for bloodletting. Moreover, many reports mentioned the usage of leeches in skin diseases, nervous system abnormalities like brain congestion, urinary and reproductive system problems (nephritis, vaginitis). In addition, ocular inflammation, dental problems, and hemorrhoids were also treated by leech therapy [1–3]. On the other hand, there was no scientific documentation of the medical importance of leeches until 1884, when John B. Haycraft (1857–1922) outlined the existence of a potent anticoagulant agent, hirudin, in

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**[\*Corresponding author]** Abdualkader, Abdulrahman Mohammed: E-mail: [armak1983@gmail.com](mailto:armak1983@gmail.com)

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leech saliva which explained why blood oozing from a leech wound coagulated slower than an ordinary wound [4]. Henceforth, many extensive studies have been undertaken on leech products, especially leech saliva. Consequently, a large number of peptides and proteins with substantial clinical applications have been identified and characterized [5]. Recently, leeching has established itself in microsurgery, reconstructive operations [6], metastasis [7] and cancer [8].

Despite the scarce research on using leeches in the treatment of diabetes mellitus (DM), it is being used by traditional practitioners because of the powerful effects of the anticoagulants-containing leech saliva to alleviate DM complications. For instance, some clinicians use leech therapy to improve blood circulation of extremities in order to avoid gangrene and amputation [9]. The current research was designed to evaluate the antihyperglycemic activity of leech saliva obtained from *Hirudinaria manillensis*, along with examining its ability to prevent the experimental induction of DM in an animal model.

## 2 Materials and Methodology

### 2.1 Chemicals, reagents and instrument

Sodium chloride and arginine hydrochloride were the products of Merck (Germany). Bradford reagent kit was procured from Amresco (Canada). Parafilm membrane was obtained from American Can Company (USA). Anhydrous glucose was the product of Fisher Scientific (Germany). Alloxan monohydrate and bovine serum albumin and insulin from bovine pancreas ( $\geq 27 \text{ U}\cdot\text{kg}^{-1}$ ) were purchased from Sigma Aldrich (Germany).

Centrifugation was done using a Universal 32R centrifuge produced by Hettich Zentrifugen (Germany). Microplate reader model Infinite M200, NanoQuant TECAN was the product of Tecan (USA). Lyophilization was performed using a Christ freeze-drier model Alpha 1-4LD (Germany). One Touch Ultra glucometer and test strips used for the determination of blood glucose concentration were the product of LifeScan Inc., USA. Microscopic examination was done using a Nikon Eclipse 80i microscope (Japan).

### 2.2 Leech sampling, saliva collection and total protein estimation

Leeches, *Hirudinaria manillensis* (Lesson, 1842), were collected from the natural lake, Cheneh, located in Terengganu, Malaysia. They were maintained in well-aerated plastic containers filled with un-chlorinated tap water. Water was regularly changed every 2–3 d. Leech feeding and saliva extract collection were executed as previously described using a phagostimulatory solution (PHS) of  $0.001 \text{ mol}\cdot\text{L}^{-1}$  arginine in  $0.15 \text{ mol}\cdot\text{L}^{-1}$  sodium chloride [8]. Thereafter, LSE was lyophilized and the resulting powder was dissolved in an appropriate volume of distilled water to be concentrated ten-fold. The resultant solution was termed as the ten-time concentrated leech saliva extract ( $10 \times \text{LSE}$ ) which was used during the following procedures.

The total protein estimation of LSE was performed according to the standard protocols using Amresco Bradford reagent kit [10]. Bovine serum albumin (BSA) was used as a standard protein. The PHS was considered as a blank.

### 2.3 Animals and induction of diabetes mellitus

Male Sprague Dawley rats (80–110 g) were obtained from the Faculty of Veterinary Medicine, University Putra Malaysia. Rats were kept in a well air-conditioned animal room under 12 h/12 h dark and light cycle at room temperature. They were acclimatized for a period of one week prior to the experiment. During the acclimatization period, rats were housed in polypropylene cages lined with pine wood husk, changed every two days, and they were given free access to tap water and commercial dry pellet diet, Gold Coin. All experimental protocols were approved by the Ethics Committee Meeting (No. 1/2011 on 22nd April, 2011) of Kulliyah of Medicine, IIUM (Ref. No. IIUM/305/20/4/ 10).

Experimental diabetes was induced by inter-peritoneal (ip) injection of alloxan to overnight fasted animals at a dose of  $160 \text{ mg}\cdot\text{kg}^{-1}$  body weight [11]. In order to prevent fatal alloxan-induced hypoglycemia, rats were administered with 20% glucose solution intraperitoneally followed by 5% glucose solution orally for the next 12 h [12]. The diabetic state of the rats was assessed via fasting blood glucose level (FBG) of fresh capillary blood taken from a tail vein puncture. All measurements were made using a Touch Ultra glucometer. Rats showing FBG more than  $11.1 \text{ mmol}\cdot\text{L}^{-1}$  were considered diabetic [11] and selected for the study.

### 2.4 Experimental procedures

Forty male rats were divided randomly into eight groups, each comprising five rats as detailed below:

Control groups:

Negative control: normal rats.

Positive control: induced-diabetic untreated rats.

Vehicle control: induced-diabetic rats injected subcutaneously (sc) with the PHS at a dose of  $20 \text{ mL}\cdot\text{kg}^{-1}$  body weight (bw).

Comparative control: induced-diabetic rats injected sc with the standard dose of insulin of  $20 \text{ U}\cdot\text{kg}^{-1}$  bw [13].

The hypoglycemic activity of LSE:

Induced-diabetic rats injected sc with LSE at a dose of  $500 \mu\text{g}\cdot\text{kg}^{-1}$  bw.

Induced-diabetic rats injected sc with LSE at a dose of  $1\ 000 \mu\text{g}\cdot\text{kg}^{-1}$  bw.

LSE in combination with insulin:

Induced-diabetic rats injected sc with a half dose of insulin,  $10 \text{ U}\cdot\text{kg}^{-1}$  bw bovine pancreas insulin suspension in distilled water.

Induced-diabetic rats injected sc with  $250 \mu\text{g}\cdot\text{kg}^{-1}$  LSE +  $10 \text{ U}\cdot\text{kg}^{-1}$  bw bovine pancreas insulin.

The antihyperglycemic activity of LSE was assessed by the fall in FBG values within eight hours measured at two-hour intervals. The percentage decrease in fasting blood

glucose concentration was calculated from the following equation<sup>[14]</sup>:

Percentage decrease in FBG after  $x$  hour =

$$\frac{FBG(0, \text{hour}) - FBG(x, \text{hour})}{FBG(0, \text{hour})}$$

### 2.5 Toxicity and histopathological evaluation

All rats were kept under close observation during the trial period of 8 hours. For the whole experimental period, general behavior and mentality patterns (physical activity, weakness, aggressiveness, awareness, motor activity, food refusal, diarrhea, noisy breathing, writhing, etc.), and mortality were monitored<sup>[15-16]</sup>. After the experimental regimen, LSE-treated rats were anesthetized with ether and sacrificed by cervical dislocation. Specimens from their pancreas and liver were fixed in 10% neutral buffered formalin, prepared and stained by hematoxylin–eosin for light microscopic examination<sup>[17]</sup>.

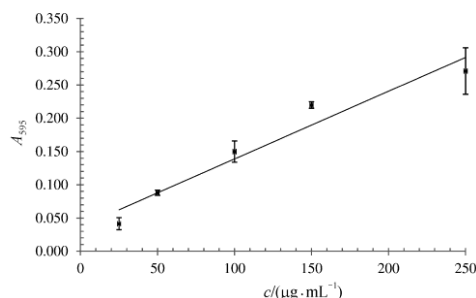
### 2.6 Statistical analysis

The results were expressed as the mean of triplicates  $\pm$  the standard error of the mean (SEM). All data were evaluated with Statistical Package for Social Science (SPSS 18.0) software using general linear model GLM repeated measurement ANOVA followed by Dunnett's (2-sided) post-hoc test considering the positive control group as a reference. Values were considered statistically significant with  $P < 0.05$ .

## 3 Results and Discussion

### 3.1 LSE collection and total protein estimation

The total volume of the colorless fluids collected from twenty starved leeches was 190 mL (pH 6.45), meaning that each leech can produce 9.5 mL of LSE. Protein estimation (Fig. 1) revealed that the collected LSE contained a total protein concentration of  $(54.29 \pm 2.58) \mu\text{g}\cdot\text{mL}^{-1}$ . Lyophilization of LSE yielded a white fine powder which was later



**Fig. 1** Standard curve for total protein estimation in leech saliva extract (LSE). Assay was carried out according to the standard protocols of Bradford method. Bovine serum albumin (BAS) was considered as the standard protein. The phagostimulatory solution (PHS) was used as blank.  $Y = 0.001 X + 0.028$ ,  $R^2 = 0.953$ ;  $Y = \text{absorbance } A_{595}$  and  $X = \text{BSA concentration } \mu\text{g}\cdot\text{mL}^{-1}$

dissolved in a suitable volume of distilled water immediately prior to use to give the  $10 \times$  LSE. This indicates that the pro-

tein concentration of  $10 \times$  LSE was  $542.99 \mu\text{g}\cdot\text{mL}^{-1}$ .

A comparison between the volume of LSE (9.5 mL) given by one leech with the total protein [ $(54.229 \pm 2.589) \mu\text{g}\cdot\text{mL}^{-1}$ ] revealed that each leech can give a total protein of about  $500\mu$  corresponding the minimum dose by which the experimental rats were injected. Consequently, the rats were injected with saliva at doses of 500 and 1 000  $\mu\text{g}\cdot\text{kg}^{-1}$  bw as if they had been bitten by one and two leeches, respectively.

### 3.2 The anti-hyperglycemic activity of leech saliva extract (LSE)

After three days of intraperitoneally alloxan injection ( $160 \text{ mg}\cdot\text{kg}^{-1}$  bw), results revealed that alloxan caused a significant increase ( $P < 0.001$ ) in blood glucose concentration in the positive control group when compared with the negative control. The FBG levels were significantly reduced ( $P < 0.001$ ) after subcutaneous injection of LSE at both doses of 1 000 and 500  $\mu\text{g}\cdot\text{kg}^{-1}$  bw. In addition, a significant reduction in FBG ( $P < 0.05$ ) occurred two hours after injection of LSE, and a more significant decline was noticed after four, six and eight hours ( $P < 0.001$ ). Furthermore, all of the insulin-injected rats experienced a sharp, significant decrease ( $P < 0.001$ ) in FBG after two hours of injection. Rats which received the lower dose of insulin ( $10 \text{ U}\cdot\text{kg}^{-1}$  bw) returned to the diabetic state, with a rapid increase in FBG values, after 8 hours of treatment ( $P = 0.090$ ). Whereas, the rats which received insulin ( $10 \text{ U}\cdot\text{kg}^{-1}$ ) and LSE ( $250 \mu\text{g}\cdot\text{kg}^{-1}$ ) exhibited a significant drop in FBG during the whole eight hour study period ( $P < 0.001$ ). In contrast, the diabetic rats injected with the phagostimulatory solution showed no significant reduction in FBG compared with the positive control rats. Results are presented in Table 1.

Fig. 2 illustrates that the dose of  $1 000 \mu\text{g}\cdot\text{kg}^{-1}$  bw gradually decreased FBG by about 18% after two hours of subcutaneous administration compared with a 10% reduction caused by the dose  $500 \mu\text{g}\cdot\text{kg}^{-1}$  bw. After four to six hours of injection, the biological activity of the injected LSE went through a steady state stage, because both doses exhibited approximate blood glucose lowering activity ranging from 40%–50%. Thereafter, the maximum antihyperglycemic activity of LSE (56.5% reduction in FBG) was found after eight hours of injection. Noteworthy, all of the LSE-injected rats exhibited FBG values less than the diabetic limit of  $11.1 \text{ mmol}\cdot\text{L}^{-1}$ <sup>[11]</sup> after eight hours of injection, meaning that the eight hour treatment session was sufficient to bring the diabetic rats to the normal state.

On the other hand, the subcutaneous injection of insulin at a dose of  $20 \text{ U}\cdot\text{kg}^{-1}$  bw induced a sharp reduction in FBG of about 80%–90% within two to six hours of dosage. During this period, rats which were injected with this dose of insulin experienced a hypoglycemic condition with low FBG values which eventually resulted in the death of one rat. After eight hours of injection, the blood glucose concentrations started to

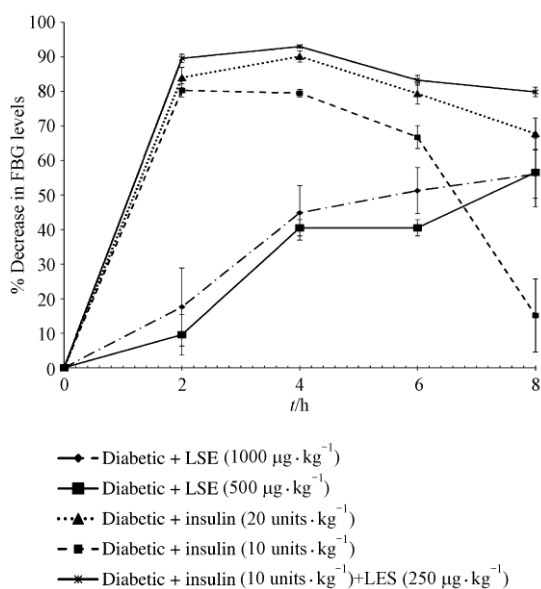
rise but less than the diabetic range. In contrast, a lower dose of insulin ( $10 \text{ U}\cdot\text{kg}^{-1} \text{ bw}$ ) resulted in a dramatic decline in FBG of about 80% during the first four hours of administration and the hypoglycemic activity was reduced slightly after six hours of injection. After eight hours of injection of the lower dose of insulin, all experimental animals tended to become diabetic again showing FBG values of ( $22.4 \pm 4.7$ )  $\text{mmol}\cdot\text{L}^{-1}$ .

Interestingly, the concurrent administration of both LSE ( $250 \mu\text{g}\cdot\text{kg}^{-1} \text{ bw}$ ) and insulin ( $10 \text{ U}\cdot\text{kg}^{-1} \text{ bw}$ ) produced a sharp decline in the fasting blood glucose of approximately 93% during the first four hours of injection. After six to eight hours, blood glucose concentrations displayed a marginal increment with a hyperglycemic lowering activity of the LSE-insulin mixture effectively sufficient to keep the rats at the normal state.

**Table 1** The effect of different doses of leech saliva extract (LSE) and insulin on fasting blood glucose (FBG) in alloxan-induced diabetic rats at various time intervals (hours) (mean  $\pm$  SEM,  $n = 5$ )

Groups/doses	FBG ( $\text{mmol}\cdot\text{L}^{-1}$ )				
	0 h	2 h	4 h	6 h	8 h
Negative control <sup>a</sup>	$4.6 \pm 0.4^{**}$	$5.1 \pm 0.2^{**}$	$4.7 \pm 0.2^{**}$	$5.4 \pm 0.2^{**}$	$5.1 \pm 0.2^{**}$
Vehicle control	$22.8 \pm 3.6$	$22.0 \pm 3.6$	$22.8 \pm 3.3$	$19.9 \pm 3.5^*$	$22.4 \pm 4.7^{**}$
Comparative control <sup>a</sup>	$26.4 \pm 1.7$	$4.2 \pm 1.3^{**}$	$2.7 \pm 0.6^{**}$	$5.3 \pm 0.5^{**}$	$8.4 \pm 0.9^{**}$
Diabetic + LSE $1000 \mu\text{g}\cdot\text{kg}^{-1}$ <sup>a</sup>	$21.0 \pm 2.8$	$17.0 \pm 2.7^*$	$11.9 \pm 2.7^{**}$	$10.8 \pm 3.0^{**}$	$8.8 \pm 3.1^{**}$
Diabetic + LSE $500 \mu\text{g}\cdot\text{kg}^{-1}$ <sup>a</sup>	$21.3 \pm 3.2$	$19.0 \pm 2.3^*$	$12.6 \pm 1.9^{**}$	$11.4 \pm 1.9^{**}$	$8.0 \pm 1.5^{**}$
Diabetic + insulin $10 \text{ U}\cdot\text{kg}^{-1}$ <sup>a</sup>	$24.9 \pm 1.3$	$4.9 \pm 0.4^{**}$	$5.1 \pm 0.4^{**}$	$8.1 \pm 0.4^{**}$	$21.3 \pm 3.2$
Diabetic + LSE $250 \mu\text{g}\cdot\text{kg}^{-1}$ + insulin $10 \text{ U}\cdot\text{kg}^{-1}$ <sup>a</sup>	$23.2 \pm 1.2$	$2.4 \pm 0.3^{**}$	$1.6 \pm 0.2^{**}$	$3.9 \pm 0.4^{**}$	$4.6 \pm 0.2^{**}$
Positive control	$27.3 \pm 2.1$	$27.6 \pm 2.2$	$26.96 \pm 1.1$	$25.6 \pm 1.2$	$27.5 \pm 1.7$

<sup>a</sup> significant compared to the positive control group; \*  $P < 0.05$  and \*\*  $P < 0.001$



**Fig. 2** Percentage decrease in fasting blood glucose in alloxan-induced rats treated with different doses of leech saliva extract (LSE) and insulin (mean  $\pm$  SEM,  $n = 5$ ). Fasting blood glucose levels (FBG) were taken at two-hour intervals. The percentage decrease values in FBG were calculated according to the equation mentioned in materials and methodology section.

Taken together, these findings indicate that the subcutaneous injection of the salivary gland secretion (LSE) collected from the medicinal Malaysian leech, *H. manillensis*, showed hypoglycemic activity which provided better glycaemic control than that of insulin alone, with no fatal hypogly-

cemic cases during the eight hour study period. Moreover, it was found that LSE and insulin at lower doses had a synergistic antihyperglycemic activity. This type of synergism suggests that LSE could be utilized as an adjuvant medication in order to reduce insulin dosage or to achieve acceptable control of blood glucose concentration instead of using mixtures of short- and long-acting insulin. In fact, LSE antidiabetic activity may be due to the presence of one or more antihyperglycemic factors, thought to be peptides or proteins, and/or their synergistic properties. The mechanism by which LSE exhibited its blood glucose lowering activity still needs to be determined but many mechanisms could be hypothesized. Alloxan is a selective cytotoxic agent to the pancreatic  $\beta$ -cells by accelerating their necrotic cell death by forming reactive oxygen species (ROS), resulting in decreased endogenous insulin release (insulinopenia) and a consequent insulin-dependent type 1 diabetes syndrome [12]. In case that most  $\beta$ -cells have been damaged, leech saliva extract (LSE) could have potential extrapancreatic effects. One suggested mechanism could be termed as an “insulin mimic”, that insulin-like proteins in LSE bind the  $\alpha$ -subunit of insulin receptors in the tissue cells, which results in conformational changes of the  $\beta$ -subunit of the receptors and eventually receptor activation. Consequently, phosphorylation cascades will be triggered within the cell leading to multiple effects, involving glucose uptake by muscle cells due to glucose receptor translocation and more hepatic glycogen formation due to glycogen synthase activation [18].

In contrast, because it is already known that the majority of  $\beta$ -cells were destroyed by alloxanisation, other mechanisms that may reduce the blood glucose levels depending on

the presence of circulating insulin and intact  $\beta$ -cells, such as the direct stimulation of insulin secretion via closing ATP-sensitive potassium channels of pancreatic islet  $\beta$ -cells, improving insulin receptor sensitivity, etc. are not applicable<sup>[19]</sup>. Hence, more studies are required to determine the active principle and its actual mechanism of action.

Furthermore, the concomitant existence of antithrombotic<sup>[20]</sup>, antioxidant<sup>[21]</sup> and hypoglycemic activities in the LSE of the medicinal Malaysian leech, *H. manillensis* provides an extraordinary protection from cardiovascular complications in diabetic patients who are at a high risk of developing heart diseases<sup>[22]</sup>. It was reported that the increased risk of cardiovascular complications in patients with diabetes mellitus, especially type 2, could be ascribed to many stress factors, such as hyperglycemia, hyperlipidemia, inflammation, coagulation and platelet adhesion disorders, hypertension and oxidative stress<sup>[23]</sup>. Some researchers argued that oxidative stress plays a pathogenic role in macro- and micro-events in diabetic patients due to hyperglycemia-induced glucose autoxidation and lipoprotein modifications such as glycation and oxidation<sup>[22]</sup>.

### 3.3 Toxicity and histopathology evaluation

Neither mortality nor behavioral changes were observed among the saliva-injected animals during the study. All these animals exhibited typical locomotion and physical activity, with no signs of weakness or aggressiveness. No toxic reactions were observed for instance no anorexia, ataxia, piloerection, loss of weight, diarrhea, urination, breathing difficulty, or noisy breathing. On the other hand, injection of insulin at a dose of 20 U·kg<sup>-1</sup> bw resulted in a hypoglycemic condition in all rats leading to the death of one rat. The other rats which survived showed less physical activity especially during the first two hours after injection. In contrast, injection of lower dose of insulin (10 U·kg<sup>-1</sup> bw) led to hypoglycemic events which persisted for a shorter time of only two hours with no mortality. Additionally, the simultaneous administration of LSE and insulin induced hypoglycemic cases with no mortality.

Although the experiment took a short time, no toxic signs were observed when the tissue specimens taken from the treated animals by subcutaneous injection with 1 000  $\mu\text{g}\cdot\text{kg}^{-1}$  bw were compared with those taken from the normal control group. That means that both doses of LSE for such a period of time have no toxic effects. These observations were reinforced by the traditional leech therapy which did not document toxic events, even though leeches were directly applied to the patients. Nevertheless, there are some reports about post leeching infections<sup>[3]</sup>.

## 4 Conclusions

To the best of our knowledge, this is the first demonstration that the salivary gland secretion of leeches (LSE) has an anti-hyperglycemic effect against type-1 like diabetes mellitus. It was shown that subcutaneous injection of LSE col-

lected from the medicinal Malaysian leech, *H. manillensis*, was sufficient to obtain regular glycemic control, with no fatal post-injection hypoglycemic conditions during the study period. Moreover, it was found that both LSE and insulin at lower doses had a synergistic antihyperglycemic activity. This type of synergism suggests that LSE could be utilized as an adjuvant medication in order to reduce insulin dosage or to achieve acceptable control of blood glucose concentration instead of using mixtures of short- and long-acting insulin. The toxicity study presented in the current research suggested that leech saliva can be considered as a safe medication for diabetic patients. More studies of longer duration are needed to evaluate the effectiveness and toxicity of long-term usage. Moreover, the active compound(s) are still obscure which indicated the need for more researches aimed at isolating and identifying the anti-hyperglycemic agent(s) in leech saliva.

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