RESPONSE OF THE PORTAL VEIN OF SPONTANEOUS HYPERTENSIVE RATS TO INTRACELLULAR PH

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Abstract: The effect of intracellular pH perturbations on the portal vein preparations of spontaneously hypertensive rats and their control Wistar Kyoto rats was investigated. Intracellular alkalinity induced by application of 20 mM NH.Cl or 20 mM trimethylamine produced dilatation of both preparations. Intracellular acidity induced by washout of the previous ammonium and trimethylamine solutions or by application of 20 mM sodium propionate solution caused constriction of both preparations. These responses of the portal veins of both animals to intracellular pH variations were qualitatively the same in More nonactivated preparations and in preparations precontracted with 26 more preparations mM K+ or 1 µM norepinephrine. Recovery from acidic constrictions induced by washout of ammonium and trimethylamine solutions was an additional and an additional and a solutions was a solution and a solution and a solution and a solution and a solution are solutions. atualing a Hard as significantly slower in spontaneous hypertensive rats than in Wistar Kyoto rats preparations. Conceivably, a lower intracellular pH in the vascular smooth muscle of the resistance vessels of hypertensive patients, as compared to normotensive individuals, may partly account Engeringerial protocol; The portal vein for the hypertensive phenomena.

Key words: intracellular pH portal vein SHR vascular smooth muscle WKY

INTRODUCTION

Variations in intracellular pH influence vascular tone (1). Intracellular pH can be changed while maintaining constant extracellular pH by applying salts of weak acids and bases (2). The salt most commonly used is NH, Cl. Applying NH, Cl to a preparation causes intracellular alkalinity and its subsequent washout produces intracellular acidity (2-5). Trimethylamine behaves in the same way as NH₄Cl (2, 3). Intracellular alkalinity induced by application of NH,Cl produces dilatation of the rabbit ear artery (6), rabbit aortic strips (7), rat tail artery (8), isolated canine arteries (9) and isolated porcine coronary arteries (10). Most of the previous preparations have been activated with NE. However, NH,Cl

produces constriction in nonactivated preparations of canine pulmonary artery (11) and rat portal vein (12) and in the mesenteric artery activated with 125 mM $\rm K^+$ (5). The discrepancy is less in the effect of intracellular acidity; most of the previous researches (5-11) report constriction caused by washout of NH₄Cl solution leading to intracellular acidity.

The relation between intracellular pH and tone has not been looked at in hypertensive animals. Therefore the aim of this study is to investigate the effect of variations in intracellular pH induced by application of NH₄Cl, trimethylamine and propionate on the portal vein of SHR rat and their controls, WKY rats.

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SHR and WKY rats weighing 200-300 g were killed by a stroke on the head. The portal vein was quickly dissected out, and 5 mm strips of the vessel was mounted in a 40 ml bath. The isometric tension in the preparation was measured with model FT03 Grass transducer and the trace was recorded on a polygraph. In each experiment atleast one preparation of SHR portal vein was mounted with a control preparation of WKY.

Solutions: Normal Ringer's solution contained in mM: 140 NaCl, 6 KCl, 1.5 CaCl₂, 1.5 NaH₂PO₄, 1 MgCl₂ and 10 glucose. The pH of all solutions was kept at 7.4.

Concentrated salt solutions were prepared by total replacement of the NaCl in the normal Ringer's solution with the required salt keeping all other constituents constant. Hence the net concentration of the salt was 140 mM.

Drugs: NE (Arterenol bitartrate) and all other chemicals were obtained from Sigma.

Experimental protocol: The portal vein strips were immersed in 40 ml of normal Ringer's solution at 37° C under a starting tension of 1 g. The preparation was left to stabilize for one hour before any experimental solution was introduced. Wherever appropriate the vessel was activated with 1 μ M NE or 26 mM K* to produce base line tone. Certain measured volumes of the concentrated stock salt solutions were then added to the bath to achieve the required concentration of the salt in the bath without changing the osmolarity. The salt solution was left for 5 min and then washed with the control solution for atleast 5 min before another salt was applied.

Analysis of traces: Tone variations due to the salt are expressed as means ± standard error of the mean of their relative values compared to the presalt tone. The significance of tone deviations from unity (presalt tone) and of SHR as compared to WKY rats are evaluated by Student's unpaired t-test. P values <0.05 are considered significant. Recovery rate from acid load is expressed as follows (13).

The % recovery from ammonium constriction (acid load) is taken as

$$\frac{c-d}{c-1}$$
 x 100

Where

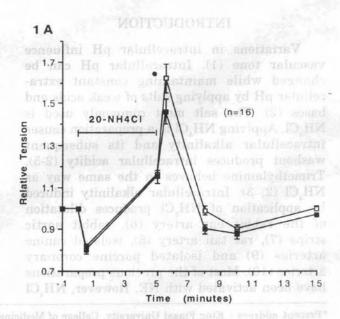
c = maximum tone reached by ammonium washout,

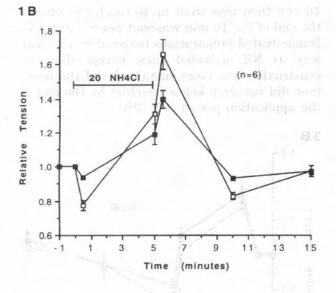
d = tone measured 4 min after 'c' and

1 = presalt tone

RESULTS

Application of 20 mM NH₄Cl dilated NE activated portal vein preparations of both SHR and WKY rats (Fig. 1A). Subsequent washout of the salt produced constriction in both preparations. The same qualitative results were obtained in nonactivated and 26 mM K⁺ activated and preparations (Fig. 1B, 1C). In all of these three traced the maximum dilatation produced by NH₄Cl was obtained in the first half a minute of application, while the maximum washout tension rise was attained within the first min. Recovery of tone towards prepulse level occurred after these maximum points of reduction and increase in tension. Trimethylamine produced the same qualitative results as NH₄Cl in both





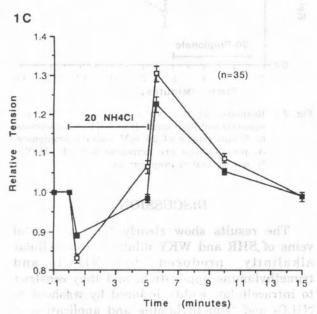


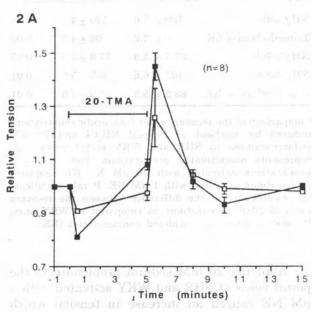
Fig. 1: Response of the portal vein preparations of SHR (open squares) and their controls of WKY (solid squares) to 5 min pulses of 20 mM NH₄Cl.

A: preparations are activated with 1 µM NE.

B: non-activated preparations. C: preparations are activated with 26 mM K⁺.

preparations with and without NE activation (Fig. 2A and 2B). The rate of recovery from NH₄Cl and trimethylamine washout constriction was significantly slower in the SHR portal vein

preparations than in the WKY ones (Table I). This reduction was greater and more significant in the NE activated preparations.



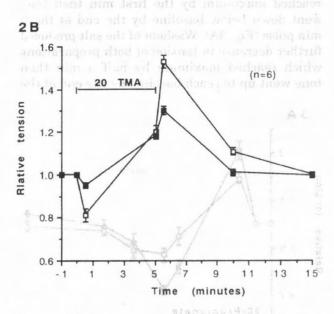


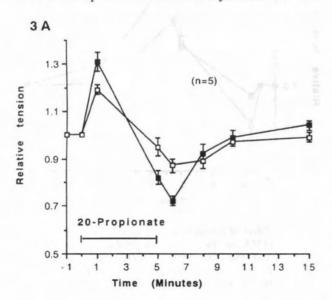
Fig. 2: Effect of 5 min pulses of 20 mM trimethylamine (TMA) on the portal veins of SHR (open squares) and their controls of WKY (solid squares).
 A: preparations are activated with 1 μM NE.
 B: non-activated preparations.

TABLE I: Recovery rates from acidic loads of SHR and WKY portal veins.

Treatment	%Recovery from acidic load		
	SHR	WKY	P value
NH ₄ Cl-6k ⁺	130 ± 7.6	120 ± 4	NS
Trimethylamine-6K+	80 ± 3.2	96 ± 4.9	0.02
NH ₄ Cl-26K ⁺	67.5 ± 3.9	77.9 ± 3.4	0.05
NH ₄ Cl-NE	102 ± 6.6	125 ± 5.6	0.01
Trimethylamine-NE	63.2 ±15.3	91.8 ± 7.9	0.01

Comparison of the recovery rates from acidic constrictions induced by washout of 20 mM NH,Cl and 20 mM trimethylamine in SHR and WKY portal veins. 6K+ represents nonactivated preparations. 26K+ indicates preparations activated with 26 µM K+. NE indicates preparations activated with 1 µM NE. P values indicate the significance of the difference between the recovery rates of SHR preparations as compared to WKY ones. P values > 0.05 are considered nonsignificant (NS).

Applying 20 mM sodium propionate to the portal veins of SHR and WKY activated with 1 uM NE caused an increase in tension which reached maximum by the first min then tone went down below baseline by the end of the 5 min pulse (Fig. 3A). Washout of the salt produced further decrease in tension of both preparations which reached maximum by half a min then tone went up to reach baseline by the end of the



10 min then tone went up to reach baseline by the end of the 10 min washout period (Fig. 3A). Nonactivated preparations behaved in a similar way as NE activated ones, except that the constriction was more sustained and the mean tone did not drop below baseline by the end of the application period (Fig. 3B).

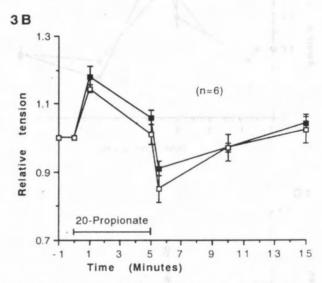


Fig. 3: Response of the portal veins of SHR (open squares) and their controls of WKY (solid squares) to 5 min pulses of 20 mM sodium propionate. A: preparations are activated with 1 µM NE. B: non-activated preparations.

DISCUSSION

The results show clearly that the portal veins of SHR and WKY dilate to intracellular alkalinity produced NH,Cl by and trimethylamine application, and they constrict. to intracellular acidity induced by washout of NH.Cl and trimethylamine and application of sodium propionate. This is in general agreement with previous results on the portal vein of Wistar rats, cats and rabbits (14). But what seems to be contradictory results have been reported recently; NH,Cl application produced constriction (increase in tension) of the nonactivated Wistar rat portal vein preparations which disappeared on washout of the salt (11). These constrictions obtained by NH,Cl application are most probably due to membrane depolarization effect of the K+

like cation NH, rather than intracellular pH effect (14, 15). The qualitative response of the portal veins of SHR and WKY to variations in intracellular pH is independent of the mode of activation. This supports previous proposals that intracellular pH effect is mediated through direct interaction between intracellular protons ([H+]i) and intracellular calcium ions ([Ca2+li) (10, 13, 14, 16, 17). Hence a rise in [H+]i would produce a rise in [Ca2+]i and subsequent constriction of the vessel. If this relation applies to human vessels then a lower intracellular pH in the vascular smooth muscle of the resistance vessels of hypertensive patients when compared to normotensive individuals may partly explain the hypertension phenomena.

Recovery from acid load in vascular smooth muscle is mainly carried out by Na⁺/H⁺ exchanger (5, 6). In the present study the recovery from acidic load induced by washout of NH₄Cl or trimethylamine has been slower in SHR portal vein preparations than in WKY ones. The most probable explanation for this is a slower Na⁺/H⁺ exchanger in SHR portal vein preparations; but a slower activity of Na⁺/Ca²⁺ can not be excluded from the results in this study. In fact it has

been reported that Na⁺/Ca²⁺ exchanger is less active in SHR rat tail arterial preparations than in WKY ones (18). However, Na⁺/Ca²⁺ exchanger plays a minor role in the recovery from NH₄Cl washout constriction (13).

In conclusion, SHR and WKY portal vein preparations dilate to intracellular alkalinity and constrict to intracellular acidity. The recovery from acidic load is slower for the SHR preparations. This is most probably due to a lower activity of the Na*/H* exchanger is SHR than in WKY preparations. Conceivably, a lower intracellular pH in the vascular smooth muscle of the resistance vessels of hypertensive patients, as compared to normotensive individuals, may partly account for the hypertensive phenomena.

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REFERENCES

- Wray S. Smooth muscle intracellular pH: measurement, regulation, and function. Am J Physiol 1988; 254: C213-C225.
- Roos A, Boron WF. Intracellular pH. Physiol Rev 1981; 61: 296-434.
- Thomas RC. Experimental displacement of intracellular pH and the mechanism of its subsequent recovery. J Physiol (Lond) 1984; 354: 3P-22P.
- Spurway NC, Wray S. A phosphorus nuclear magnetic resonance study of metabolites and intracellular pH in rabbit vascular smooth muscle. J Phsiol (Lond) 1987; 393: 57-71.
- Aalkjaer C, Mulvany MJ. Effect of changes in intracellular pH on the contractility of rat resistance vessels. Prog Biochem Pharmacol 1988; 23: 150-158.
- Ighoroje AD, Spurway NC. Procedures to acidify cytoplasm raise the tone of isolated (rabbit ear) blood vessels. J Physiol (Lond) 1984; 357: 105P.

- Furtado MR. Effects of NH₄Cl on the contractility of isolated vascular smooth muscle. *Life Sci* 1987; 41: 95-102.
- Bamosa AO, Ighoroje AD, Spurway NC, Taggart MJ. Vascular tone variations, initiated by ammonium 'pulses', in an isolated, perfused rat tail preparation. J Physiol (Lond) 1987; 392: 23P.
- Feletou M, Harker CT, Komori K, Shepherd JT, Vanhoutte PM. Ammonium ions cause relaxation of isolated canine arteries. J Pharmacol Exp Ther 1989; 251(1): 82-89.
- Hoang ND. Effects of manipulations of intracellular pH on the mechanical responses of isolated porcine coronary arteries. Arch Int Physiol Biochim Biophys 1993; 101-110.
- Wakabayashi I, Katake K, Sakamoto K. Ammonium ion increases the tone of rat portal vein. Gen Pharmacol 1992; 23(6): 1189-1192.
- 12. Krampetz IK, Rhoades RA. Intracellular pH: effect on

- 28
 - pulmonary arterial smooth muscle. Am J Physiol 1991; 260 : L516-L521.
- 13. Ighoroje AD. pH and vascular tone. PhD Thesis 1987, University of Glasgow.
- 14. Bamosa AO. Generality and mechanism of intracellular pH effect on vascular tone. PhD Thesis 1989, University of Glasgow.
- 15. Bamosa AO, Ighoroje AD, Spurway NC. Tone overshoot during recovery of isolated (rabbit and rat) vascular preparations from ammonium-induced dilations. J Physiol (Lond) 1987; 392: 47P.

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- 16. Spurway NC. Intracellular pH and vascular tone. Proceedings of the XIXth European Conference on muscle and cell motility, ed. Marchal G, Carraro V, 1991, PP 229-233.
- 17. Bamosa AO, Spurway NC. Effect of intracellular pH on pCai in isolated rat tail arteries loaded with Fura-2. J Physiol (Lond) 1990; 423: 64P.
- 18. Thompson LE, Rinaldi GJ, Borh DF. Decreased activity of the sodium-calcium Exchanger in tail artery of stroke-prone spontaneously hypertensive rats. Blood Vessels 1990; 27 : 197-201.