Behaviour of 316L Stainless Steel in Simulated Physiological Fluids

AURORA ANCA POINESCU¹, RODICA-MARIANA ION^{1,2}, IONICA IONITA^{3*}, IOANA DANIELA DULAMA⁴, ANDREI CHILIAN⁴

¹ Valahia University of Targoviste, Faculty of Materials Engineering and Mechanics, Department of Material Engineering, 18-24 Unirii Bdvl. Targoviste, 130082, Romania

² The National Institute for Research & Development in Chemistry and Petrochemistry ICECHIM, Analytical Department, 202 Splaiul Independentei, District 6, 060021, Bucharest, Romania

³ Valahia University of Targoviste, Faculty of Science and Arts, Department of Sciences and Advanced Technologies, 18-24 Unirii Bdvl., Targoviste, 130082, Romania

⁴ Valahia University of Targoviste, Multidisciplinary Research Institute for Science and Technologies, Targoviste, 130082, Romania

In contact with biological systems all metals corrode and metal ions freed acts as haptens, forming complexes with native proteins that activate the immune system [1]. For analysis of heavy metals (Fe, Ni, Cr, Mn) that can migrate from 316L stainless steel (plate and powder), under controlled conditions in artificial plasma or serum were performed tests by energy dispersive X-ray fluorescence spectrometer (EDXRF) and absorption spectrometric method in flame atomic absorption spectrometry with electrothermal atomization.

Keywords: artificial plasma, 316L stainless steel, energy dispersive X-ray fluorescence (EDXRF), SFBsimulated fluid body

In the last decades several strategies have been developed for obtaining biomaterials used to make the medical implants [1,5,7,8]. The development of this area is due to the necessity of obtaining biofunctional materials with enhanced features. Developing new materials for medical implants involves methods for characterizing of their properties. Characterization of the materials prior to their use in medical devices is essential in order to reduce the time required for product certification. The prevalence of metal allergy is high in the population; it is estimated that up to 17% of women and 3% of men presenting allergy to nickel and cobalt and chromium the prevalence of allergies is 1-3%. This type of allergy is manifested by allergic contact dermatitis and systemic allergic dermatitis. Moreover, the metal allergy was associated with various implant types of implant failures, hip replacements, knee or coronary stents [2]. In contact with biological systems all metals corrode and metal ion release acts as hapten, forming complexes with native proteins that activate the immune system. The metal-protein complex is recognized by Langerhans cells and would be presented to T cells by T lymphocytes activated T cell receptor (TCR) will undergo a process of proliferation and differentiation into effectors T cells and memory T cells capable of recognizing the metal at a subsequent exposure. The secondary immune response triggered by exposure repetition is fast and cytokines, chemokines and cytotoxin issued by local blood circulation stimulates T cells, macrophages and the eosinophil recruitment of inflammatory process responsible for dermal [1]. Metallic material used for the experiments is 316L stainless steel, known as one of the most commonly used biomaterials for medical applications [2,6,9,10].

Experimental part

Materials and methods Materials

In this respect on this study were made a series of investigations to determine the content of Fe, Cu, Ni and Cr which can migrate from 316L stainless steel (powder and

plate), under controlled conditions in the artificial plasma or serum. These investigations were conducted using energy dispersive X-ray fluorescence (EDXRF) spectroscopy and flame atomic absorption spectrometry/ electrothermal atomic absorption spectrometry. The biomaterials used on this study were:

- 316L stainless steel plate;

- 316L stainless steel powder; The surgical austenitic grade of stainless steel, such as 316L has the composition: 16-18% Cr; 10-15% Ni; Mo 2-3%; Mn 2%; P 0.04%; 0.035% C; Si 0.03%, S 0.03%; Fe the remaining up to 100%.

The determination of Fe, Ni, Cr and Mn from physiological saline (0.9% NaCl) and artificial plasma Methodology

The qualitative analysis is very sensitive and it works for both liquid and solid to ppm for all elements from B to Al. The solid samples intended quantitative analysis are in the form of powder and plate with thickness greater than 2 mm and less than 3 cm discs. The liquid samples were analyzed in special tanks made of plastic, covered with a plastic foil (Mylar) to remove the effects of the vacuum. The analysis of the liquid witness samples (saline, 0.9 % NaCl and artificial plasma) and monitored on a sample period were conducted by energy dispersive X-ray fluorescence spectroscopy using Elvax spectrometer. Spectra were processed using software Elvax. From the qualitative point of view, this method provides information on the initial elemental composition of materials composites, as well as of simulated samples of simulated body fluid (SBF) with biocomposites materials studied. The artificial plasma was prepared according to the literature [3,4] by weighing pure analytically substances necessary for preparing artificial plasma, and the dilution was made with distilled water (table 1). Separately were weighed solid samples of materials.

Four samples were prepared by introducing the solid weighed samples in sterile vial of 100 mL by polyethylene terephthalate; 2 samples solids was treated with 50 mL of commercial normal saline and 2 solid sample with 50 mL

Chemical formula	The quantity of salt dissolved in distilled water [g/L]
NaCl	6.8
CaCl ₂	0.2
KCl	0.4
MgSO ₄	0.1
NaHCO ₃	2.2
Na ₂ HPO ₄	0.126
NaH ₂ PO ₄	0.026

Table 1ARTIFICIAL PLASMA

No	Material	Solution	Acronym	Expo	sure time	e of the	solids	in artifi	cial solu	itions
							[hours]			
1	316L stainless steel	artificial	PLAS1							
	metal plate	plasma								
2	316L stainless steel	physiologic	PLAS2							
	metal plate	al serum		168	336	672	1344	2016	2688	4032
3	316L stainless steel	artificial	PLAS3		550	072	1511	2010	2000	1052
	metallic powder	plasma								
4	316L stainless steel	physiologic	PLAS4	1						
	metallic powder	al serum								

Table 2ACRONYMS OF THE PREPAREDSAMPLES FOR MINERALIZATION

of artificial plasma which was prepared in the laboratory. Samples thus prepared were kept tightly closed at 280-300 nm UV, moderate temperatures of 36-37°C, wellestablished time periods (table 2).

Thus, the first analysis was performed after 168 h, the second analysis after 336 hours of exposure, the third determination after 672 h of exposure period (1 month), the fourth determination after 1344 h (2 months), the fifth determination after 2016 h (3 months), the sixth analysis was done after 2688 h (4 months) and the last measurement was done after 4032 h (six months).

Results and discussions

The qualitative analysis by energy dispersive X-ray fluorescence spectroscopy of the solid and liquid samples after 4032 h of exposure, led to the following conclusions:

In the **PLAS1** sample after 6 months of exposure of the steel 316L in artificial plasma, indicated the presence of the following elements: Fe, P, S, Ni, Cr which comes even from 316L steel (composition: 16-18% Cr, 10-15 Ni %, Mo

2-3%, Mn 2%, P 0.04%, C 0.035%, Si 0.03%, S 0.03% and Fe the remainder to 100%). In spectrum of the initial material introduced in artificial plasma (Alloy_PLAS1) comparative viewed shows the presence of Fe, P, S, Ni, Cr and Mn. The presence of trace amounts of Pb and Cd can be explained by his existence in air or distilled water, or by interfering with other metals. Copper and the lead probably exist even in the solvent used in the preparation of plasma (water).

In sample **PLAS2** after 6 months of exposure of the 316L steel, in the serum has indicated the presence of the following elements: Fe, P, S, Ni, Cr coming even from 316L steel (composition: Cr 16-18%, Ni 10-15%, Mo 2-3%, Mn 2%, P 0.04%, C 0.035%, Si 0.03%, S 0.03% and Fe the remainder to 100%). In the spectrum of initial material introduced in artificial plasma (Alloy_PLAS2), comparative viewed shows the presence of Fe, P, S, Ni, Cr and Mn (fig.1, fig.2).

In sample **PLAS3** after 6 months of exposure of the 316L metal powder in the artificial plasma has indicated the presence of the following elements: Fe, P, S, Ni, Cr coming



http://www.revistadechimie.ro



Fig. 2. Spectrum of PLAS2 from artificial plasma

even from 316L steel (composition: Cr 16-18%, Ni 10-15%, Mo 2-3%, Mn 2%, P 0.04%, C 0.035%, Si 0.03%, S 0.03% Fe the remainder to 100%). In the spectrum of initial material introduced in artificial plasma (Powder PLAS3) comparative viewed shows the presence of Fe, P, S, Ni, Cr and Mn.

In sample **PLAS4** after 6 months of exposure of the 316L metal powder in the physiological serum has indicated the presence of Fe, P, S, Ni, Cr coming even from 316L steel (composition: Cr 16-18%, Ni 10-15%, Mo 2-3%, Mn 2%, P 0.04%, C 0.035%, Si 0.03%, S 0.03% and Fe the remainder to 100%). In the spectrum of initial material introduced in artificial plasma (Powder_PLAS4) comparative viewed shows the presence of Fe, P, S, Ni, Cr and Mn. Unfortunately the device doesn't determine and identify those elements which are after Na in the periodic system, so the presence of carbon and even Si could not be remarked. The chlorine appears from the physiological serum solution.

The determination of Fe, Mn, Ni and Cr from physiological serum and plasma by atomic absorption spectrometry

For analysis of the heavy metals (Fe, Ni, Cr, Mn) which can migrate, under controlled conditions, from the preparations biocomposites in the artificial plasma or physiological serum was used the atomic absorption spectrometry using the method of flame atomic absorption spectroscopy with electrothermal atomization. Regardless of the method chosen, the flame atomic absorption spectrophotometer is calibrated according to the procedure recommended by the company GBS, mixtures standard reference solutions used in the calibration (NIST). The system was washing with reagents blank after each standard used. Before starting measurements on samples, is reanalyzed the solution which is the most concentrated by reference standard, considering like a sample to analyzed.

The concentration value which was found should not differ by more than \pm 5% by the concentration of that standard. It were started the analysis through washing the system with reagent blank. Between consecutive samples, the system is aspirate and washed with reagent blank. After every ten samples the apparatus is checked by analyzing of the blank reagent (calibration) and standard solution.

The digestion method with aqua regia

Microwave digestion system (MWS-2) Berghof is used for the mineralization of the liquid and solid samples, in extreme conditions of pressure and temperature. Solutions which are subject to digestion are heated directly through microwave absorbing by digestion reagent which normally contains the ionic components. The samples subject to microwave digestion are liquids and that does not require second stage of the sampling preparation.

	Phase		1			2	3
	T [⁰ C]		145		1	.80	100
	Microwave power	[%]	80		1	90	40
	Time [min.]		5			15	10
	Samples/Metal	PLAS1	 PLAS2		PLAS3	PLAS4	
			Exposure	time	e: 168 hours		
Ì	Fe	0.001	0.001		0.002	0.001	
		±0.0	±0.0		±0.0	±0.0	
	Ni	0.002	0.002		0.001	0.001	
		±0.0	±0.0		±0.0	±0.0	
	Cr	0.000	0.000		0.002	0.001	
		±0.0	±0.0		±0.0	±0.0	
	Mn	0.001	0.001		0.001	0.001	
		±0.0	±0.0		±0.0	±0.0	

Table 3PROGRAM USED FORMINERALIZATION OF THESAMPLES

Table 4CONCENTRATION OF HEAVYMETALS IN PLASMA/SERUM AFTER168 h [ppb]

PLAS1	PLAS2	PLAS3	PLAS4
Exposure tim	ne: 336 hours		
0.002	0.004	0.002	0.002
±0.0	±0.0	±0.0	±0.0
0.002	0.001	0.001	0.001
±0.0	±0.0	±0.0	±0.0
0.001	0.001	0.000	0.001
±0.0	±0.0	±0.0	±0.0
0.001	0.001	0.001	0.001
±0.0	±0.0	±0.0	±0.0
	PLAS1 Exposure tim 0.002 ±0.0 0.002 ±0.0 0.001 ±0.0 0.001 ±0.0	PLAS1 PLAS2 Exposure time: 336 hours 0.002 0.004 ±0.0 ±0.0 0.002 0.001 ±0.0 ±0.0 0.001 ±0.0 0.001 0.001 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0	PLAS1 PLAS2 PLAS3 Exposure time: 336 hours 336 hours 0.002 0.004 0.002 ±0.0 ±0.0 ±0.0 0.002 0.001 0.001 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0 ±0.0

 Table 5

 CONCENTRATION OF HEAVY METALS IN PLASMA/

 SERUM AFTER 336 h [ppb]

Samples/Metal	PLAS1	PLAS2	PLAS3	PLAS4
	Exposure time: 67	2 hours	I	
Fe	0.002	0.003	0.005	0.005
	±0.0	±0.0	±0.0	±0.0
Ni	0.002	0.002	0.003	0.002
	±0.0	±0.0	±0.0	±0.0
Cr	0.001	0.001	0.001	0.001
	±0.0	±0.0	±0.0	±0.0
Mn	0.001	0.002	0.004	0.009
	±0.0	±0.0	±0.0	±0.0

Samples/Metal	PLAS1	PLAS2	PLAS3	PLAS4
	Exposure time: 1	344 hours	I	
Fe	0.007	0.006	0.003	0.004
	±0.0	±0.0	±0.0	±0.0
Ni	0.006	0.008	0.006	0.009
	±0.0	±0.0	±0.0	±0.0
Cr	0.002	0.001	0.003	0.002
	±0.0	±0.0	±0.0	±0.0
Mn	0.011	0.016	0.012	0.013
	±0.0	±0.0	±0.0	±0.0
		1	I I	

Samples/Metal	PLAS1	PLAS2	PLAS3	PLAS4
	Exposure time: 201	6 hours	L	
Fe	0.008	0.009	0.006	0.005
	±0.0	±0.0	±0.0	±0.0
Ni	0.007	0.005	0.019	0.023
	±0.0	±0.0	±0.0	±0.0
Cr	0.001	0.002	0.002	0.001
	±0.0	±0.0	±0.0	±0.0
Mn	0.016	0.019	0.012	0.015
	±0.0	±0.0	±0.0	±0.0

In this respect were prepared the sets of 4 samples for the mineralization as follows: each 15 mL sample of plasma or physiological serum at each of the six periods of analysis was introduced into the vessel DAP- 60K by Teflon TFM of the digestion system (MWS-2) Berghof.

It was added 2.5 mL HNO₃ 67 % (Merck) and 7.5 mL HCl 37 % (Merck) (1:3 volume ratio) in each vessel with sample. The mixture was mechanically stirred for 15-20 min. The Teflon vessels were closed and then were introduced into the digester Berghof MWS-2. The program used for sampling mineralization is shown in table 3.

The samples were cooled to room temperature for \sim 30 min. The clear solution samples were brought to 25 mL

Table 6CONCENTRATION OF HEAVY METALSIN PLASMA/SERUM AFTER 672 h [ppb]

Table 7CONCENTRATION OF HEAVY METALSIN PLASMA/SERUM AFTER 1344 h [ppb]

Table 8CONCENTRATION OF HEAVY METALS INPLASMA/SERUM AFTER 2016 h [ppb]

in bottles without dilution and then were analyzed by flame atomic absorption spectroscopy and analyzed the following elements: Fe, Mn, Cr and Ni.

The determination of Fe, Mn, Ni, Cr from physiological serum and artificial plasma samples are calculated depending on the concentration Fe, Mn, Ni, Cr measured on the spectrometer c (μ g/mL), V the volume at which sample was diluted (25 mL), reported to the volume of sample subject to mineralization, v (mL). After the research period with a view to assessment of the heavy metals migration (Fe, Ni and Cr) the samples were differentially dried in order to be weighed.



and Mn in PLAS3 during monitoring

For establishing the quantity of some relevant elements into own reference materials used for routine quality-control procedures, it is required the use of certified reference materials (CRM). The laboratory results for each item shall be determined by means of control diagrams and is not accepted any result that is outside the agreed limits. CRM will be used regularly to maintain the integrity of their reference own materials and - through this - the system of quality control. The flame atomic absorption spectroscopy (FAAS) during 6 months of monitoring has not been able to determine concentrations at ppm level.

Considering the fact that by atomic absorption spectrometry with direct aspiration in the flame have not been obtained the relevant results about the metals plotted in figures 3-6.

1355

Cr and Mn in PLAS4 during monitoring

concentration which can be analyzed in the artificial

plasma/physiological serum under controlled conditions

at the ppm level, it was examined the heavy metals

concentration by atomic absorption spectrometry, graphite

furnace technique. The results were summarized in tables

(table 4 - 10). Until mow the migration of the heavy metals

were chosen only metal samples labelled by acronyms PLAS1, PLAS2, PLAS3 and PLAS4. The variation of heavy

metal concentrations of Fe, Ni, Cr and Mn for samples was

(such as Fe, Cr, Mn) determined in physiological solution a

migration which is presented in a higher number compared to the elements which migrated in plasma solution.

In figure 4 one can see that the heavy metal particles

The behaviour of 316L stainless steel suported for covering with hydroxiapatite was studied in [11].

Conclusions

Qualitative analysis by Energy Dispersive X-ray Fluorescence (EDXRF) spectroscopy of the liquid and solid samples, after 4032 h of exposure, led to the following conclusions.

No significant values obtained in the plasma which results from the metal materials discussed in the artificial plasma/serum, under controlled in flame/atomic absorption spectrometry with electrothermal atomization.

In the atomic absorption spectrometry for graphite furnace technique was a migration of the analyzed metals (Fe, Mn, Cr and Ni) in simulated body fluid (SBF), which was analyzed after a period of six months, but only in the ppb.

The number of heavy metal particles that migrated from the powder stainless steel was greater in the plasma and physiological solution compared to the number of those ones that migrated from the plate stainless steel.

These types of stainless steel are successfully used in medical field concerned with the design, manufacture and application of material requirements in orthopedic prosthesis (plates, screws and rods) for bone fracture healing and recovery.

References

1. FORTE, G., PETRUCCI, F., BOCCA, B., Inflammation & Allergy – Drug Targets, (2008);

2. KAJZER W., KRAUZE A., WALKE W., MARCINIAK J., Corrosion behaviour of AISI 316L steel in artificial body fluids, Journal of Achievements in Materials and Manufacturing Engineering, 31(2), December 2008, p.247-253;

3. THYSSEN J. P., TORKIL MENN., Metal Allergy – A review on Exposures, Penetration, Genetics, Prevalence and Clinical Implications. In Chem. Res. Toxicol., 23 (2), (2010), p. 309–318;

4. SPERLING, M. B., WELZ, B., Atomic Absorption Spectrometry. Weinheim: Wiley-VCH, 1999.

5. BAHRAMI NASAB M., MOHD ROSHDI HASSAN, Metallic Biomaterials of Knee and Hip - A Review, Biomaterials of Knee and Hip - A Review 1 Trends Biomater. Artif. Organs, Vol 24(1), p. 69-82, (2010);

6. VERMEŞAN H., Cercetări privind comportarea la coroziune a oțelurilor inoxidabile supuse deformării plastice și nitrurarii ionice, teza de doctorat, Universitatea Tehnica Cluj Napoca, (1998);

7. WEI M., UCHIDA M., KIM H., KOKUBO T. ,NAKAMURA T., Biomater. 23 (2001) p.167;

8. RATNER B.D., HOFMAN A.S., SCHOEN F.J., LEMONS J.E., Biomaterials Science: An Introduction to Materials in Medicine, Academic Press, San Diego, (1996), p. 37–50;

9. SINGH R., NARENDRA B. DAHOTRE, Corrosion degradation and prevention by surface modification of biometallic materials, J Mater Sci: Mater Med (2007) **18**, p.725–751;

10. TRUŞCULESCU M., IEREMIA A., Oţeluri inoxidabile şi refractare, Editura Facla Timişoara, (1983)

11. POINESCU, A.A., RADULESCU, C., VASILE, B.St., IONITA, I., Rev. Chim. (Bucharest), **65**, no. 10, 2014, p. 1245

Manuscript received: 16.06.2014