



# Pharmacological and therapeutic effects of A<sub>3</sub> adenosine receptor agonists

### Pnina Fishman<sup>1</sup>, Sara Bar-Yehuda<sup>1</sup>, Bruce T. Liang<sup>2</sup> and Kenneth A. Jacobson<sup>3</sup>

<sup>1</sup> Can-Fite BioPharma Ltd., Kiryat-Matalon, 10 Bareket St., P.O. Box 7537, Petah-Tikva 49170, Israel

<sup>2</sup> Pat and Jim Calhoun Cardiology Center, University of Connecticut Health Center, Farmington, CT 06030, United States

<sup>3</sup> Molecular Recognition Section, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bldg. 8A, Rm. B1A-19, Bethesda, MD 20892-0810, United States

The  $A_3$  adenosine receptor ( $A_3AR$ ) coupled to  $G_i$  (inhibitory regulative guanine nucleotide-binding protein) mediates anti-inflammatory, anticancer and anti-ischemic protective effects. The receptor is overexpressed in inflammatory and cancer cells, while low expression is found in normal cells, rendering the  $A_3AR$  as a potential therapeutic target. Highly selective  $A_3AR$  agonists have been synthesized and molecular recognition in the binding site has been characterized. In this article, we summarize preclinical and clinical human studies that demonstrate that  $A_3AR$  agonists induce specific anti-inflammatory and anticancer effects through a molecular mechanism that entails modulation of the Wnt and the NF- $\kappa$ B signal transduction pathways. At present,  $A_3AR$  agonists are being developed for the treatment of inflammatory diseases, including rheumatoid arthritis (RA) and psoriasis; ophthalmic diseases such as dry eye syndrome and glaucoma; liver diseases such as hepatocellular carcinoma and hepatitis.

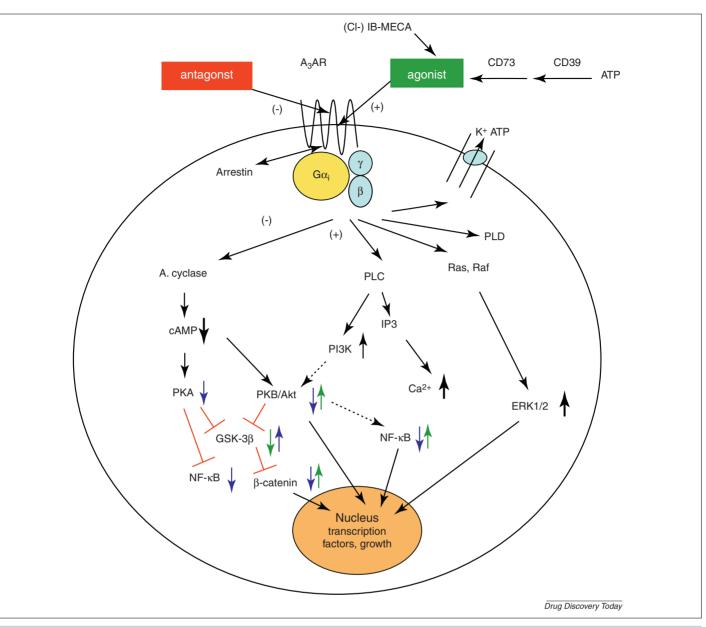
#### Introduction

The  $A_3$  adenosine receptor ( $A_3AR$ ) is a subtype of the adenosine receptor (AR) family, which additionally includes A1, A2A and  $A_{2B}$  receptors [1,2]; each receptor is encoded by a separate gene and has different physiological roles. The G<sub>i</sub>-coupled A<sub>3</sub>AR is less widely distributed than other AR subtypes with expression in human lung, liver, brain, aorta, testis and heart. The utilization of the A<sub>3</sub>AR as a therapeutic target and a biological predictive marker is based on two major findings: (i) the A<sub>3</sub>AR is overexpressed in cancer and inflammatory cells, while low expression is found in normal cells [3–5]. The high receptor expression is also found in peripheral blood mononuclear cells (PBMCs) of patients with cancer or inflammatory diseases [5,6]. (ii) Highly selective A3AR agonists have been synthesized and induce specific anti-inflammatory and anticancer effects through a molecular mechanism that entails modulation of the Wnt and the nuclear factor kappa-light-chain-enhancer of activated B cell (NF-кB) signal transduction pathways [6–8] (Fig. 1). A protective effect of the agonists on normal cells was recorded as well, suggesting that this unique differential effect of the agonists will contribute to a safety profile of these drug candidates in both preclinical and clinical studies. At present, A<sub>3</sub>AR agonists are being developed for the treatment of inflammatory, ophthalmic and liver diseases and demonstrate excellent safety and efficacy in Phase II clinical studies.

#### A<sub>3</sub>AR agonists

The human  $A_3AR$  was cloned in 1993 [1] and soon thereafter found to have cerebroprotective and cardioprotective properties [9,10]. Like other G protein-coupled receptors (GPCRs), it is also known to affect G protein-independent signaling, such as translocation of arrestins, leading to rapid desensitization of the  $A_3AR$  *in vitro* (typically within ~20 min in the presence of agonist) [11,12]. Highly selective  $A_3AR$  agonists have been synthesized, and molecular recognition in the binding site has been characterized using site-directed mutagenesis and molecular modeling. Typical  $A_3AR$ agonists are adenosine derivatives that contain 5'-uronamide and  $N^6$ -benzyl modifications leading to nanomolar receptor

Corresponding author:. Jacobson, K.A. (kajacobs@helix.nih.gov)

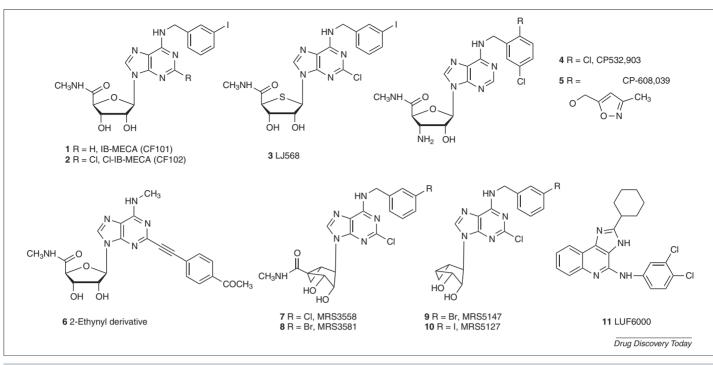


#### FIGURE 1

Signaling pathways involved in the action of A<sub>3</sub>AR agonists. Pathways proposed to be involved in anticancer, anti-inflammatory and cardioprotective effects of A<sub>3</sub>AR agonists. A<sub>3</sub>AR signals through both G protein-dependent (through G<sub>i</sub> or  $\beta$ ,  $\gamma$  subunits) and independent pathways. Translocation of arrestin to the A<sub>3</sub>AR would be associated with receptor downregulation [22] and G protein-independent signaling. Not all pathways are present in all circumstances, and other pathways not shown can also affect cell survival, proliferation and differentiation. In cancer (blue arrows), activation of the A<sub>3</sub>AR corrects an imbalance in the downstream Wnt signaling pathway [6,34]. Administration of an A<sub>3</sub> agonist to activate its cell surface receptor inhibits the formation of cAMP and indirectly decreases phosphorylation (and therefore decreased inactivation) of the serine/threonine kinase GSK-3 $\beta$ . The resulting increased phosphorylation of  $\beta$ -catenin causes it to be removed from the cytoplasm by ubiquitination and therefore preventing its nuclear import. This results in a net suppression of cyclin D1 and c-myc, which leads to cell growth inhibition. With respect to cancer, NF- $\kappa$ B is a potent antiapoptotic agent in malignant cells and its activation is strongly associated with tumors [76]. Additionally, as a major cell survival signal, NF- $\kappa$ B is involved in multiple steps in carcinogenesis and in cancer cells' resistance to chemo- and radio-therapy. Thus, drugs aimed to decrease expression or activity of NF- $\kappa$ B could abrogate its antiapoptotic effect. In inflammatory models [7–9,20,29,71], the reduced activation of NF- $\kappa$ B in splenocytes, neutrophils and other immune cells) has an anti-inflammatory effect, in part by reducing the expression of TNF- $\alpha$ . Opposite effects of A<sub>3</sub>AR activation on some of these pathways (green arrows) are associated with myeloprotective (through increased NF- $\kappa$ B in splenocytes [39]) and cardioprotective (GSK-3 $\beta$  inhibition [47]) responses to A<sub>3</sub>AR agonists. In the heart, there a

affinity (compounds numbered in bold as shown in Fig. 2) [13]. The prototypical agonists IB-MECA **1**,  $N^6$ -(3-iodobenzyl)-5'-*N*-methylcarboxamidoadenosine, CF101, and its 2-chloro analog Cl-IB-MECA **2**, 2-chloro- $N^6$ -(3-iodobenzyl)-5'-*N*-methylcarboxa-midoadenosine, CF102, are 9-ribosides with A<sub>3</sub>AR selectivity. Other selective A<sub>3</sub> agonists include the 4'-thio derivative **3**,

3'-amino-3'-deoxy derivatives CP-608,039 **5**, (2S,3S,4R,5R)-3amino-5-{6-[5-chloro-2-(3-methylisoxazol-5-ylmethoxy)benzylamino]-purin-9-yl}-4-hydroxytetrahydrofuran-2-carboxylic acid methylamide, and its dichlorobenzyl analog CP-532,903 **4**, which were originally developed for cardioprotection, and the  $N^6$ methyl-2-ethynyl derivative **6** [9,14]. Introduction of a fused



#### FIGURE 2

Representative agonists (1–8) and partial agonists (9, 10) of nanomolar affinity at the  $A_3AR$  and a positive allosteric modulator 11. Nucleosides 7–10 contain the (North) methanocarba substitution of the ribose ring, which maintains an  $A_3AR$ -preferred conformation.

bicyclic ring in the rigid analog MRS3558 **7**, (1'R,2'R,3'S,4'R,5'S)-4-{2-chloro-6-[(3-chlorophenylmethyl)amino]-purin-9-yl}-1-(methylaminocarbonyl)-bicyclo[3.1.0]hexane-2,3-diol,

increased A<sub>3</sub>AR potency and selectivity, and identified the North conformation of the ribose ring as the preferred conformation in receptor binding. 7 also shows a preference in potency for the cAMP pathway, in comparsion with arrestin signaling [12]. Truncation of nucleosides at the 4'-position reduces efficacy while retaining affinity of binding to the A<sub>3</sub>AR. Thus, the methanocarba analogs MRS5147 9, (1'R,2'R,3'S,4'R,5'S)-4'-[2-chloro-6-(3-bromobenzylamino)-purine]-2',3'-O-dihydroxybicyclo-[3.1.0]hexane, and its 3-iodo analog MRS5127 10 are low efficacy partial A<sub>3</sub>AR agonists, that are selective in both human and rat [15]. Recently, macromolecular conjugates (e.g. polyamidoamine dendrimers) of chemically functionalized AR agonists were introduced as potent polyvalent activators of the receptors that are qualitatively different in pharmacological characteristics in comparison with the monomeric agonists [16]. Several A<sub>3</sub>AR positron emission tomography (PET) ligands have been introduced for in *vivo* imaging: the antagonist [<sup>18</sup>F]FE@SUPPY 5-(2-fluoroethyl) 2,4diethyl-3-(ethylsulfanylcarbonyl)-6-phenylpyridine-5-carboxylate [16], and a pair of nucleosides (e.g. low efficacy agonist  $[^{76}Br]MRS5147$  **9** and full agonist  $[^{76}Br]MRS3581$  **8**).

The selectivity of  $A_3AR$  agonists differs between *in vitro* and *in vivo* models and between species, although the sequence identity is high (84.4%) within the transmembrane region. The characterization of a given nucleoside derivative as full or partial agonist is highly dependent on the pharmacological system, such that **2** ranges from full agonist to low efficacy partial agonist [17]. LUF6000 **11**, *N*-(3,4-dichloro-phenyl)-2-cyclohexyl-1*H*-imidazo[4,5-*c*]quinolin-4-amine, is a selective positive allosteric modulator of the human  $A_3AR$  [18], increasing the maximal effect of

inhibition of adenylate cyclase. Species-dependence of the affinity and selectivity of  $A_3AR$  antagonists should be carefully considered in preclinical studies. Functional polymorphism of  $A_3AR$  is known; a high-transcript haplotype of the  $A_3AR$  gene was associated with the development of cutaneous hyper-reactivity to aspirin [19].

## Differential expression of A<sub>3</sub>AR in pathological and normal cells

 $A_3AR$  was found to be overexpressed in various neoplastic cells, including leukemia, lymphoma, astrocytoma, melanoma and pineal tumor cells, whereas low or almost no receptor expression was found in normal cells [20–25]. Similar data were reported in other studies, the receptor expression levels in tumor tissues derived from patients with colon, breast, small cell lung, pancreatic and hepatocellular carcinomas, and melanoma in direct comparison with adjacent normal tissues [3,4,6]. A direct correlation between  $A_3AR$  tissue expression levels and disease progression was described in breast and colon cancer [3,4].

A similar pattern of receptor overexpression was described in inflammatory cells both in experimental animal models and humans. The most studied inflammatory disease was rheumatoid arthritis (RA) in which  $A_3AR$  overexpression was detected in paw tissue, draining lymph nodes and synovial cells of rats with adjuvant-induced arthritis and in synovial cells from patients with RA [7]. Similar data were observed in colon tissues derived from rats with colitis and in lungs upon inhalation of lipopolysaccharides (LPS) by mice [26,27]. The receptor was also highly expressed in anterior segment tissues derived from eyes with pseudoexfoliation syndrome in comparison with healthy subjects' eyes [28].

The high expression levels of  $A_3AR$  seen in tumor and inflammatory cells were also found in PBMCs derived from tumor-bearing animals and cancer patients [3,6]. Similarly, high receptor expression levels were found in PBMCs derived from experimental animal models of inflammation and from patients with autoimmune inflammatory diseases, such RA, psoriasis and Crohn's disease [5,7,29].

These data suggest that A<sub>3</sub>AR expression levels in PBMCs mirror the receptor expression levels in the remote tumor or inflammatory tissue, rendering the receptor a biological marker. A3AR upregulation is attributed to factors, including elevated adenosine and cytokines, which are characteristic of the microenvironment of cancer and inflammatory cells [29,30]. Under stressed metabolic conditions, extracellular adenosine of intracellular origin accumulates in the surroundings [30,31]. Upon binding to cell surface receptors, adenosine might induce, through an autocrine pathway, the expression of its own receptors. The proinflammatory cytokine tumor necrosis factor-a (TNF-a) initiates downstream signaling by binding to its cell surface receptor, to result in upregulation of protein kinase B (PKB)/Akt, the inhibitor of NFкВ light polypeptide gene enhancer in B cells (ІкВ), ІкВ kinase (IKK) and the transcription factor NF-κB [24,28]. The latter is known to act as an A<sub>3</sub>AR transcription factor.

Bioinformatic analyses revealed that besides NF- $\kappa$ B, other transcription factors, such as c-Rel, MyoD, c-fos, GR, CREB, AP-1, GATA-1, C/EBP, c-Jun and PU.1 bind to the A<sub>3</sub>AR promoter region. It is well established that proinflammatory cytokines regulate the cell expression levels of each of these transcription factors, hence regulating A<sub>3</sub>AR expression levels [5]. Taken together, it seems that receptor overexpression in tumors and inflammatory cells is a consequence and manifestation of the disease state, rather than a causative factor.

Interestingly, *in vivo* pharmacological data revealed that chronic treatment with  $A_3AR$  agonist in various experimental animal models of cancer and inflammation did not desensitize the receptor. This was evidenced by the downregulation of receptor expression levels shortly after the last drug administration in a chronic mode of treatment [6,7,32]. In addition, 24 hours after the last agonist administration,  $A_3AR$  protein expression level was fully recovered to the control level, demonstrating that chronic treatment does not reduce the receptor expression levels [32].

#### In vivo pharmacological profile of A<sub>3</sub>AR agonists

#### Anticancer effect

In experimental animal models, A<sub>3</sub>AR agonists were efficacious in combating growth of solid tumors, including melanoma, prostate, colon and hepatocellular carcinoma (Table 1). The agonists

showed efficacy upon chronic oral treatment, which was initiated after the tumor was already established. Overall, the drugs were much more potent in the syngeneic models rather than in the xenograft models, pointing toward an immunological effect on top of the direct anticancer effect. Supporting this notion are the findings showing that treatment with **2** increased interleukin (IL)-12 and potentiated natural killer (NK) activity in an animal model of melanoma [33].

The direct mechanism of the anticancer effect of A<sub>3</sub>AR agonists entails modulation of the NF- $\kappa$ B and the Wnt signaling pathways. In tumor lesions of A<sub>3</sub>AR agonist-treated animals, the expression levels of PKB/Akt, IKK, NF- $\kappa$ B and TNF- $\alpha$  signaling proteins were downregulated. The expression of glycogen synthase kinase-3 $\beta$ (GSK-3 $\beta$ ) was upregulated, whereas the expression of its downstream proteins,  $\beta$ -catenin, lymphoid enhancer-binding factor-1 (LEF1) and c-Myc, was decreased, leading to inhibition of tumor cell growth [6,32,34]. Apoptosis, an additional mechanism of action, was demonstrated in hepatocellular carcinoma tumors, and manifested by increased expression of the proapoptotic proteins Bcl-2-associated death promoter (BAD), Bcl-2-associated X protien (BAX) and Caspase-3 upon treatment with **2** [6]. These data prompted the selection of **2** as a drug candidate to be developed as an anticancer agent for the treatment of hepatocellular carcinoma.

#### Anti-inflammatory effect

A<sub>3</sub>AR agonists possess a robust anti-inflammatory effect mediated by the inhibition of proinflammatory cytokines [35–37]. **1**, **2** and **3** exert anti-inflammatory effects in experimental animal models of inflammatory bowel disease, systemic toxemia, pulmonary inflammation, RA, osteoarthritis and liver inflammation (Table 2). The molecular mechanism involved with the anti-inflammatory activity entails deregulation of the NF- $\kappa$ B signaling pathway, leading to inhibition of TNF- $\alpha$ , IL-6, IL-12, macrophage inflammatory proteins (MIPs)-1 $\alpha$ , MIP-2 and receptor activator of NF- $\kappa$ B ligand (RANKL), resulting in apoptosis of inflammatory cells [7,38].

#### Protective effects

#### Chemoprotective effect

Myelotoxicity is a severe and dose-limiting complication of chemotherapy. Drug-induced myelosuppression is a major toxic factor that limits the administration of larger, potentially more effective doses of chemotherapy.

A<sub>3</sub>AR agonists administered in combination with chemotherapeutic agents to tumor-bearing mice prevented the myelotoxic

TAE	ЗL	Е	1

Inflammatory condition	Experimental animal model	Refs
RA	Adjuvant and collagen-induced arthritis	[7,38,68–70
Osteoarthritis	Monosodium iodoacetate-induced osteoarthritis	[71]
Inflammatory bowel diseases	Dextran sodium sulfate or 2,4,6-trinitrobenzene sulfonic acid-induced colitis; spontaneous colitis in IL-10 gene deficient mice	[26,37,72]
Uveitis	IRBP induced experimental autoimmune uveitis	[73]
Sepsis/toxemia	CLP and LPS-induced sepsis	[35,74]
Pulmonary inflammation	LPS inhalation	[27]

Abbreviations: CLP: cecal/ligation and puncture; IRBP: interphotoreceptor retinal binding protein.

Effects of A <sub>3</sub> AR agonists in expen	rimental animal models of inflammatory	disease	
Tumor type	Animal strain; cell line	Experimental model	Refs
Melanoma	Mice; B16-F10	Syngeneic; metastatic Syngeneic; subcutaneous	[33,62,63]
Prostate cancer	Rat; AT6.1 Mice; PC3	Xenograft; metastatic Xenograft, subcutaneous	[32,64]
Colon carcinoma	Mice; HCT-116 Mice; CT-26	Xenograft, subcutaneous Syngeneic; metastatic	[34,65,66]
Breast cancer	Mice; SK-BR-3	Xenograft, subcutaneous	[75]
Hepatocellular carcinoma	Rat; N1S1 Mice; Hep3B	Syngeneic; orthotopic Xenograft, subcutaneous	[6,67]

TABLE 2

effects of chemotherapy [39]. Coadministration of **1** prevented a decline in white blood cell (WBC) and neutrophil counts, resulting in full recovery of myeloid system parameters. The A<sub>3</sub>AR agonist induced the production of granulocyte colony-stimulating factor (G-CSF), which stimulates myeloid progenitor cell expansion in the bone marrow and increases the WBC and neutrophil counts in the peripheral blood. The molecular mechanism underlying the events before G-CSF production includes the upregulation of its transcription factor NF- $\kappa$ B and the upstream kinases PI3K, PKB/Akt and IKK [39].

In the cardiovascular system, A<sub>3</sub>AR agonists induce cardioprotection against chemotherapy-induced damage. The anthracycline antibiotic doxorubicin (DOX) or adriamycin has been an effective treatment for leukemias, lymphomas and solid tumors, including breast cancer.

Acute cardiotoxicity of DOX develops during and shortly after the initiation of therapy. However, chronic or late DOX-induced cardiotoxicity has a latency lasting for years before the development of overt heart failure. At present, only dexrazoxane, a freeradical scavenger, shows promise as a cardioprotective agent during DOX treatment [40]. Developing new methods to reduce both acute and chronic cardiotoxicity should increase the effectiveness of this anticancer therapy.

In this context, it is interesting to note that the  $A_3$  agonist **2** can protect against mitochondrial damage and helps preserve ATP production in cultured rat cardiomyocytes. Repeated intravenous injection of **2** before DOX administration in rats helped prevent left ventricular wall thinning and dysfunction [41,42]. It is unknown whether continuous treatment with **2** can delay or prevent the late DOX cardiotoxicity.

#### Cardioprotective anti-ischemic effect

 $A_3AR$  agonists protect against myocardial ischemia/reperfusion (I/R) injury, this has been demonstrated using selective agonists and  $A_3AR$ -knockout mice, which are otherwise physiologically normal [43]. The cardioprotective effect is evident in role of the  $A_3AR$  in ischemic preconditioning and in direct protection during ischemia. The  $A_3AR$  might also be involved in mediating the post-conditioning effect given its ability to reduce infarct size when it is administered during reperfusion [9]. The  $A_3AR$  has the lowest level of myocardial expression among the ARs, at least for the murine heart. However, evidence has accumulated indicating that stimulation of an endogenous cardiac  $A_3AR$ , independent of circulating immune inflammatory cells or resident mast cells, can result in

cardioprotection [44]. An anti-inflammatory action of the  $A_3AR$  *in vivo* might also contribute importantly to the cardioprotective effect of  $A_3$  agonist. Both a direct cardioprotective mechanism and an anti-inflammatory effect exerted at the immune cell level *in vivo* might be important. Future studies are needed to address this question. A cardioprotective role of the  $A_3AR$  was also found in nonrodent mammals, such as rabbits and dogs [45,46].

Mediators that have a direct myocardial protective effect include protein kinase C (PKC), KATP channels, reactive oxygen species, connexin 43, mitochondrial permeability transition pore (MPTP) and GSK-3β. Thus, a signaling cascade might begin with A<sub>3</sub>AR stimulation, PKC activation, phosphorylation (and thus inactivation) of GSK-3β, leading to inhibition of MPTP and reduced cardiac myocyte death [47]. The role of sarcolemmal versus that of mitochondrial KATP channels in mediating the A3 cardioprotection is not clear. There is extensive evidence for a protective role of mitochondrial K<sub>ATP</sub> (mito K<sub>ATP</sub>) channel including the recently elucidated function of connexin 43 in mediating mito KATP opening by PKC. A recent study, however, showed that sarcolemmal KATP deletion abrogated the preconditioning effect of A3AR agonist in murine heart [43]. Given the redundancy of signaling pathways causing cardioprotection, it is possible that species differences exist in the role of such signaling molecules. Genetic background could also modulate cardioprotection not only in mice but also humans.

#### **Protection of skeletal muscle**

A<sub>3</sub>AR agonists attenuate skeletal muscle injury caused by ischemia and reperfusion or eccentric exercise [48]. Skeletal muscle is susceptible to various forms of injury, including ischemia, trauma and physical exertion. Skeletal muscle is one of the most vulnerable tissues in the extremities. Thus, developing new methods designed to provide cytoprotection to the skeletal muscle is important. Direct infusion of adenosine can mimic the skeletal muscle protective effect of ischemic preconditioning in the extensor digitorum longus muscle before aorta occlusion in the rat in addition to in the pig latissimus dorsi muscle flap model. A<sub>3</sub>AR agonist, when administered in vivo, signals selectively through phospholipase PLC $\beta$ 2/ $\beta$ 3 to cause a reduction in skeletal muscle injury sustained either during I/R or eccentric exercise [48]. Although A<sub>1</sub> and A2AARs can also mediate anti-ischemic protection in skeletal muscle, only the A<sub>3</sub>AR can induce protection against both I/R and eccentric exercise injuries.

Given that the activation of the  $A_3AR$  has a known anti-inflammatory effect; it is possible that skeletal muscle protective effect is mediated, at least, in part at an immune cell level. The following lines of evidence support this hypothesis. First, activated mast cells and neutrophils are important contributors of skeletal muscle ischemia/reperfusion damage. Second, activation of the A<sub>3</sub>AR can block superoxide formation and chemotaxis of murine bone marrow neutrophils [49].

#### Lung ischemia/reperfusion protection

 $A_3AR$  agonist prevents lung injury following ischemia/reperfusion in the cat. Compound **3** produced a sustained protection, which was associated with suppressed p38 protein expression and downregulation of its phosphorylation [50,51].

#### Neuroprotection

Evidence from diverse models suggests that neuroprotective effects might be mediated by the A<sub>3</sub>AR, but differences between acute and chronic agonist administration have been noted [10]. Ischemic brain injury in a model of forebrain ischemia in gerbils is reduced upon chronic treatment with **1** [52]. A<sub>3</sub>AR agonist was found to prevent the loss of retinal ganglion cells following activation of the P2X7 receptor in a rat experimental model [53].

## CF101 for the treatment of inflammatory and ophthalmic diseases

Based on the preclinical pharmacology data and encouraging safety data in Phase I studies [54], the anti-inflammatory effect of **1** was tested in a set of three Phase II clinical studies, including RA, psoriasis and dry eye syndrome (Table 3). Overall, the data obtained from these clinical studies showed excellent safety profile and efficacy, positioning **1** as a disease-modifying anti-inflammatory drug.

#### RA

RA is a chronic, systemic inflammatory disorder attacking joints that results in inflammatory synovitis that could cause destruction of articular cartilage and bone [55]. The mechanisms responsible for causing joint damage and functional impairment in RA are complicated and involve B cell or T-cell products stimulating the release of TNF and other proinflammatory cytokines, including IL-1, IL-6, and TNF- $\alpha$  and degradative enzymes.

In a multicenter Phase II study, blinded to dose (0.1, 1.0 or 4.0 mg), the drug was administered orally, twice daily for 12 weeks to patients with active RA. The primary efficacy endpoint was an improvement of 20% or more according to the classification of RA responses by the American College of Rheumatology (ACR) [55]. Compound **1** was found to be safe and well tolerated, and the maximal responses were observed in patients treated with a 1.0 mg dose. At 12 weeks, 55.6%, 33.3%, and 11.5% of the patients receiving 1.0 mg **1** achieved ACR 20%, 50%, and 70% responses, respectively. In addition, a statistically significant correlation between  $A_3AR$  expression at baseline and patient response to **1** was observed, rendering the  $A_3AR$  as a biological predictive marker [56].

#### Psoriasis

Psoriasis is a chronic inflammatory skin disease characterized by epidermal hyper-proliferation and immature differentiation resulting in multisystem pathology and a negative impact on the quality of life of the patients [57]. Proinflammatory cytokines, such as interferon (INF)- $\gamma$ , TNF- $\alpha$ , IL-23 and T helper (Th)17 are known to have a role in mediating the inflammation and epidermal alterations in psoriasis [58].

The efficacy and safety of **1** were tested in a Phase II, multicenter, randomized, double blind, dose-ranging, placebo-controlled study in patients with moderate to severe chronic plaque-type psoriasis. Compound **1** (1, 2 or 4 mg) or placebo was administered orally twice daily for 12 weeks. Overall, the drug was safe and well tolerated.

The maximal improvement in the mean change from baseline in the psoriasis area and severity index (PASI) score versus placebo and the highest percentage of patients who achieved physician's global assessment (PGA) score of 0 or 1 were observed in the 2 mg **1**-treated group. The improvement was progressive and linear throughout the study period. Thus, **1** was safe and well tolerated.

#### Dry eye syndrome

Dry eye syndrome is an inflammatory condition of the eye that is caused by decreased tear production or increased tear film evaporation. It is characterized by massive production of proinflammatory cytokines. The dryness and the inflammation could result in eye damage leading to impaired vision [59,60].

TABLE 3

Disease	Phase	Primary endpoints	Current status	Ref
RA	lla	ACR20 response at week 12	Phase IIb	[56]
Psoriasis	II	Reduction in PASI score of at least 75% from baseline (PASI 75) to the end of the 12 weeks treatment period	Phase II/III	[57]
Dry eye syndrome	II	Improvement of 25% or more over baseline at week 12 in tear film BUT or in superficial punctate keratitis as assessed by either FS or ST1 results	Phase III	[60]
Hepatocellular carcinoma	1/11	To determine the safety, tolerability, dose-limiting toxicities, maximum tolerated dose, recommended Phase II dose and to assess the repeat-dose pharmacokinetic behavior of orally administered CF102	Ongoing	
Hepatitis C virus infection	1/11	To determine the safety and tolerability of 15 days of orally administered CF102 in patients with chronic hepatitis C genotype 1, to assess the effects on HCV load during 24 weeks and to assess the repeat-dose pharmacokinetic behavior of CF102	Ongoing	

FS: Fluorescein staining; ST1: Schirmer tear test 1.

Anecdotal findings demonstrating that **1** improved indicators of dry eye syndrome in RA patients led to a separate Phase II clinical study (randomized, multicenter, doubled-masked, placebo-controlled and parallel-group) of the safety and efficacy of **1** (1 mg) administered orally daily for 12 weeks to patients with moderate to severe dry eye syndrome. It was found that compound **1** was safe and well tolerated and no serious adverse events were noted throughout the study.

Treatment with **1** resulted in a statistically significant improvement in the mean change from baseline at week 12 of the clearance of corneal staining, tear break-up time and tear meniscus height in the group treated with **1** versus placebo. Compound **1** was well tolerated and exhibited an excellent safety profile with no serious adverse events. Interestingly, a statistically significant decrease from baseline was observed in the intraocular pressure of the **1**treated patients in comparison with the placebo-treated group [61]. No serious adverse events in the RA, psoriasis or dry eye clinical studies were observed. The profile of the adverse events was similar between the placebo and **1**-treated groups.

#### **Concluding remarks**

Based on the experimental animal data and human clinical study results presented in this article,  $A_3AR$  is suggested as a specific and unique therapeutic target to combat proliferative diseases, including inflammation and cancer. The excellent safety profile of  $A_3AR$ agonists, currently tested in human clinical studies, is attributed to the different protective effects mediated through the receptor. The  $A_3AR$  has also been identified as a biological marker to predict a patient's eligibility for treatment with the agonists. Taken together, the utilization of  $A_3AR$  as both a biological predictive marker and as a therapeutic target encompasses a 'personalized medicine' approach and makes the  $A_3AR$  agonists promising small molecule drug candidates.

#### Acknowledgements

We thank the Intramural Research Program of the NIH, National Institute of Diabetes and Digestive and Kidney Diseases and the Ray Neag Distinguished Professorship (to BTL).

#### References

- 1 Salvatore, C.A. *et al.* (1993) Molecular cloning and characterization of the human A<sub>3</sub> adenosine receptor. *Proc. Natl. Acad. Sci.* 90, 10365–10369
- 2 Fredholm, B.B. *et al.* (2011) Nomenclature and classification of adenosine receptors – an update. *Pharmacol. Rev.* 63, 1–34
- 3 Gessi, S. *et al.* (2004) Elevated expression of A<sub>3</sub> adenosine receptors in human colorectal cancer is reflected in peripheral blood cells. *Clin. Cancer Res.* 210, 5895–5901
- 4 Madi, L. *et al.* (2004) The A<sub>3</sub> adenosine receptor is highly expressed in tumor vs. normal cells: potential target for tumor growth inhibition. *Clin. Cancer Res.* 10, 4472–4479
- 5 Ochaion, A. *et al.* (2009) The anti-inflammatory target A<sub>3</sub> adenosine receptor is overexpressed in rheumatoid arthritis, psoriasis and Crohn's disease. *Cell. Immunol.* 258, 115–122
- $6\,$  Bar-Yehuda, S. et al. (2008) The  $A_3$  adenosine receptor agonist CF102 induces apoptosis of hepatocellular carcinoma via de-regulation of the Wnt and NF-κB signal transduction pathways. Int. J. Oncol. 33, 287–295
- 7 Fishman, P. et al. (2006) The PI3K–NF-κB signal transduction pathway is involved in mediating the anti-inflammatory effect of IB-MECA in adjuvant-induced arthritis. *Arthritis Res. Ther.* 8, 1–9
- 8 Varani, K. *et al.* (2010) Expression and functional role of adenosine receptors in regulating inflammatory responses in human synoviocytes. *Br. J. Pharmacol.* 160, 101–115
- 9 Ge, Z.D. *et al.* (2010) A<sub>3</sub> adenosine receptor activation during reperfusion reduces infarct size through actions on bone marrow-derived cells. *J. Mol. Cell. Cardiol.* 49, 280–286
- 10 Jacobson, K.A. (1998) A<sub>3</sub> adenosine receptors: novel ligands and paradoxical effects. *Trends Pharmacol. Sci.* 19, 184–191
- 11 Klaasse, E.C. *et al.* (2008) Internalization and desensitization of adenosine receptors. *Purinerg. Signal.* 4, 21–37
- 12 Gao, Z.G. and Jacobson, K.A. (2008) Translocation of arrestin induced by human A<sub>3</sub> adenosine receptor ligands in an engineered cell line: comparison with G proteindependent pathways. *Pharmacol. Res.* 57, 303–311
- 13 Koscsó, B. et al. (2011) Investigational A<sub>3</sub> adenosine receptor targeting agents. *Expert Opin. Investig. Drugs* 20, 757–768
- 14 Dal Ben, D. *et al.* (2010) Molecular modeling study on potent and selective adenosine A<sub>3</sub> receptor agonists. *Bioorg. Med. Chem.* 18, 7923–7930
- 15 Tosh, D.K. *et al.* (2009) Functionalized congeners of A<sub>3</sub> adenosine receptor-selective nucleosides containing a bicyclo[3.1.0]hexane ring system. *J. Med. Chem.* 52, 7580–7592
- 16 Wadzak, W. et al. (2008) Preparation and first evaluation of [<sup>18</sup>F]FE@SUPPY: a new PET tracer for the adenosine A<sub>3</sub> receptor. Nucl. Med. Biol. 35, 61–66
- 17 Fozard, J.R. (2010) From hypertension (+) to asthma: interactions with the adenosine A<sub>3</sub> receptor from a personal perspective. In A<sub>3</sub> Adenosine Receptors from Cell Biology to Pharmacology and Therapeutics (Borea, P.A., ed.), pp. 3–26, Springer

- 18 Gao, Z.G. et al. (2011) Functionally biased modulation of A<sub>3</sub> adenosine receptor agonist efficacy and potency by imidazoquinolinamine allosteric enhancers. Biochem. Pharmacol. 82, 658–668
- 19 Kim, S.H. *et al.* (2010) Functional variability of the adenosine A3 receptor (ADORA3) gene polymorphism in aspirin-induced urticaria. *Br. J. Dermatol.* 163, 977–985
- 20 Gessi, S. *et al.* (2001) A<sub>3</sub> adenosine receptors in human neutrophils and promyelocytic HL60 cells: a pharmacological and biochemical study. *Br. J. Pharmacol.* 134, 116–126
- 21 Yao, Y. *et al.* (1997) Adenosine A<sub>3</sub> receptor agonists protect HL-60 and U-937 cells from apoptosis induced by A<sub>3</sub> antagonists. *Biochem. Biophys. Res. Commun.* 232, 317–322
- 22 Trincavelli, M.L. et al. (2002) A<sub>3</sub> adenosine receptors in human astrocytoma cells: agonist-mediated desensitization, internalization, and down-regulation. Mol. Pharmacol. 62, 1373–1384
- 23 Merighi, S. *et al.* (2001) Pharmacological and biochemical characterization of adenosine receptors in the human malignant melanoma A375 cell line. *Br. J. Pharmacol.* 134, 1215–1226
- 24 Suh, B.C. et al. (2001) Pharmacological characterization of adenosine receptors in PGT-beta mouse pineal gland tumour cells. Br. J. Pharmacol. 134, 132–142
- 25 Morello, S. *et al.* (2008) Cl-IB-MECA inhibits human thyroid cancer cell proliferation independently of A3 adenosine receptor activation. *Cancer Biol. Ther.* 7, 278–0284
- 26 Guzman, J. *et al.* (2006) ADOA<sub>3</sub>R as a therapeutic target in experimental colitis: proof by validated high-density oligonucleotide microarray analysis. *Inflamm. Bowel Dis.* 12, 766–789
- 27 Wagner, R. et al. (2010) Adenosine receptor A<sub>3</sub> is a critical mediator in LPSinduced pulmonary inflammation. Am. J. Physiol. Lung Cell. Mol. Physiol. 299, L502–L512
- 28 Schlötzer-Schrehardt, U. *et al.* (2005) Selective upregulation of the A<sub>3</sub> adenosine receptor in eyes with pseudoexfoliation syndrome and glaucoma. *Invest. Ophthalmol. Vis. Sci.* 46, 2023–2034
- 29 Madi, L. *et al.* (2007) Over-expression of A<sub>3</sub> adenosine receptor in PBMNC of rheumatoid arthritis patients: Involvement of NF-κB in mediating receptor level. J. Rheumatol. 34, 20–26
- 30 Keibel, A. et al. (2009) Inflammation, microenvironment, and the immune system in cancer progression. Curr. Pharm. Des. 15, 1949–1955
- 31 Ariztia, E.V. *et al.* (2006) The tumor microenvironment: key to early detection. *Crit. Rev. Clin. Lab. Sci.* 43, 393–425
- 32 Fishman, P. *et al.* (2003) Targeting the A<sub>3</sub> adenosine receptor for cancer therapy: inhibition of prostate carcinoma cell growth by A<sub>3</sub> agonist. *Anticancer Res.* 23, 2077–2083
- 33 Harish, A. et al. (2003) A<sub>3</sub> adenosine receptor agonist potentiates natural killer cell activity. Int. J. Oncol. 23, 1245–1249

- 34 Fishman, P. *et al.* (2004) An agonist to the  $A_3$  adenosine receptor inhibits colon carcinoma growth in mice via modulation of GSK-3 $\beta$  and NF- $\kappa$ B. *Oncogene* 23, 2465–2471
- 35 Hasko, G. et al. (1996) Adenosine receptor agonists differentially regulate IL-10, TNF-alpha, and nitric oxide production in RAW 264.7 macrophages and in endotoxemic mice. J. Immunol. 157, 4634–4640
- 36 Hasko, G. et al. (1998) An agonist of adenosine A<sub>3</sub> receptors decreases interleukin-12 and interferon-γ production and prevents lethality in endotoxemic mice. Eur. J. Pharmacol. 358, 261–268
- 37 Mabley, J. *et al.* (2003) The adenosine A<sub>3</sub> receptor agonist, N<sup>6</sup>-(3-iodobenzyl)adenosine-5'-N-methyluronamide, is protective in two murine models of colitis. *Eur. J. Pharmacol.* 466, 323–329
- 38 Szabó, C. et al. (1998) Suppression of macrophage inflammatory protein (MIP)-1alpha production and collagen-induced arthritis by adenosine receptor agonists. Br. J. Pharmacol. 125, 379–387
- 39 Bar-Yehuda, S. et al. (2002) Agonists to the A<sub>3</sub> adenosine receptor induce G-CSF production via NF-kappaB activation: a new class of myeloprotective agents. *Exp. Hematol.* 30, 1390–1398
- 40 Zhang, Y.W. *et al.* (2009) Cardiomyocyte death in doxorubicin-induced cardiotoxicity. *Arch. Immunol. Ther. Exp.* (*Warsz*) 57, 435–445
- 41 Shneyvays, V. *et al.* (2001) Activation of A<sub>3</sub> adenosine receptor protects against doxorubicin-induced cardiotoxicity. *J. Mol. Cell. Cardiol.* 33, 1249–1261
- 42 Emanuelov, A.K. *et al.* (2010) Adenosine A<sub>3</sub> receptor-mediated cardioprotection against doxorubicin-induced mitochondrial damage. *Biochem. Pharmacol.* 79, 180–187
- 43 Wan, T.C. *et al.* (2008) The A<sub>3</sub> adenosine receptor agonist CP-532,903 [N<sup>6</sup>-(2,5dichlorobenzyl)-3'-aminoadenosine-5'-N-methylcarboxamide] protects against myocardial ischemia/reperfusion injury via the sarcolemmal ATP-sensitive potassium channel. *J. Pharmacol. Exp. Ther.* 324, 234–243
- 44 Auchampach, J.A. *et al.* (1997) Canine mast cell adenosine receptors: cloning and expression of the  $A_3$  receptor and evidence that degranulation is mediated by the  $A_{2B}$  receptor. *Mol. Pharmacol.* 52, 846–860
- 45 Auchampach, J.A. *et al.* (2003) A<sub>3</sub> adenosine receptor agonist IB-MECA reduces myocardial ischemia-reperfusion injury in dogs. *Am. J. Physiol. Heart Circ. Physiol.* 285, H607–H613
- 46 Auchampach, J.A. *et al.* (1997) Selective activation of A<sub>3</sub> adenosine receptors with N<sup>6</sup>-(3-iodobenzyl)adenosine-5'-N-methyluronamide protects against myocardial stunning and infarction without hemodynamic changes in conscious rabbits. *Circ. Res.* 80, 800–809
- 47 Park, S.S. *et al.* (2006) N<sup>6</sup>-(3-iodobenzyl)-adenosine-5'-N-methylcarboxamide confers cardioprotection at reperfusion by inhibiting mitochondrial permeability transition pore opening via glycogen synthase kinase 3 beta. *J. Pharmacol. Exp. Ther.* 318, 124–131
- 48 Zheng, J. *et al.* (2007) Protective roles of adenosine A<sub>1</sub>, A<sub>2A</sub>, and A<sub>3</sub> receptors in skeletal muscle ischemia and reperfusion injury. *Am. J. Physiol. Heart Circ. Physiol.* 293, H3685–H3691
- 49 van der Hoeven, D. *et al.* (2008) Activation of the A<sub>3</sub> adenosine receptor suppresses superoxide production and chemotaxis of mouse bone marrow neutrophils. *Mol. Pharmacol.* 74, 685–696
- 50 Matot, I. *et al.* (2006)  $A_3$  adenosine receptors and mitogen-activated protein kinases in lung injury following *in vivo* reperfusion. *Crit. Care* 10, R65
- 51 Matot, I. et al. (2008) Lung injury after in vivo reperfusion: outcome at 27 hours after reperfusion. Anesthesiology 109, 269–278
- 52 von Lubitz, D.K. *et al.* (1999) Chronic administration of adenosine A<sub>3</sub> receptor agonist and cerebral ischemia: neuronal and glial effects. *Eur. J. Pharmacol.* 367, 157–163
- 53 Hu, H. *et al.* (2010) Stimulation of the P2X7 receptor kills rat retinal ganglion cells *in vivo. Exp. Eye Res.* 91, 425–432

- 54 van Troostenburg, A.R. *et al.* (2004) Tolerability, pharmacokinetics, and concentration-dependent hemodynamic effects of oral CF101, an A<sub>3</sub> adenosine receptor agonist, in healthy young men. *Int. J. Clin. Pharmacol. Ther.* 42, 534–542
- 55 Harris, E.D. (1990) Rheumatoid arthritis: pathophysiology and implications for therapy. N. Engl. J. Med. 322, 1277–1289
- 56 Silverman, M.H. *et al.* (2008) Clinical evidence for utilization of the A<sub>3</sub> adenosine receptor as a target to treat rheumatoid arthritis: data from a phase II clinical trial. *J. Rheumatol.* 35, 1–7
- 57 Christophers, E. (2001) Psoriasis epidemiology and clinical spectrum. *Clin. Exp. Dermatol.* 26, 314–320
- 58 Asarch, A. et al. (2008) Th17 cells: a new paradigm for cutaneous inflammation. J. Dermatol. Treat. 19, 259–266
- 59 Latkany, R. (2008) Dry eyes: etiology and management. Curr. Opin. Ophthalmol. 19, 287–291
- 60 Acera, A. et al. (2008) Inflammatory markers in the tears of patients with ocular surface disease. Ophthalmic Res. 40, 315–321
- 61 Avni, I. *et al.* (2010) Treatment of dry eye syndrome with orally administered CF101: data from a phase 2 clinical trial. *Ophthalmology* 117, 1287–2129
- 62 Fishman, P. *et al.* (2001) The A<sub>3</sub> adenosine receptor as a new target for cancer therapy and chemoprotection. *Exp. Cell Res.* 269, 230–236
- 63 Madi, L. et al. (2003) A<sub>3</sub> adenosine receptor activation in melanoma cells: association between receptor fate and tumor growth inhibition. J. Biol. Chem. 278, 42121–42130
- 64 Jajoo, S. *et al.* (2009) Adenosine A<sub>3</sub> receptor suppresses prostate cancer metastasis by inhibiting NADPH oxidase activity. *Neoplasia* 11, 1132–1145
- 65 Ohana, G. *et al.* (2003) Inhibition of primary colon carcinoma growth and liver metastasis by the A<sub>3</sub> adenosine receptor agonist CF101. *Br. J. Cancer* 89, 1552–1558
- 66 Bar-Yehuda, S. *et al.* (2005) CF101, an agonist to the  $A_3$  adenosine receptor enhances the chemotherapeutic effect of 5-flurouracil in a colon carcinoma murine model. *Neoplasia* 7, 85–90
- 67 Cohen, S. et al. (2011) CF102 an A<sub>3</sub> adenosine receptor agonist mediates anti-tumor and anti-inflammatory effects in the liver. J. Cell. Physiol. 226, 2438–2447
- 68 Baharav, E. et al. (2005) The anti-inflammatory effect of A<sub>3</sub> adenosine receptor agonists in murine autoimmune arthritis models. J. Rheumatol. 32, 469–476
- 69 Rath-Wolfson, L. et al. (2006) IB-MECA, an A<sub>3</sub> adenosine receptor agonist prevents bone resorption in rats with adjuvant induced arthritis. *Clin. Exp. Rheumatol.* 24, 400–4068
- 70 Ochaion, A. *et al.* (2006) Methotrexate enhances the anti-inflammatory effect of CF101 via up-regulation of the A<sub>3</sub> adenosine receptor expression. *Arthritis Res. Ther.* 8, R169
- 71 Bar-Yehuda, S. *et al.* (2009) Induction of an antiinflammatory effect and prevention of cartilage damage in rat knee osteoarthritis by CF101 treatment. *Arthritis Rheum.* 60, 3061–3071
- 72 Antonioli, L. *et al.* (2010) Control of enteric neuromuscular functions by purinergic A<sub>3</sub> receptors in normal rat distal colon and experimental bowel inflammation. *Br. J. Pharmacol.* 161, 856–871
- 73 Bar-Yehuda, S. *et al.* (2011) Inhibition of experimental auto-immune uveitis by the A<sub>3</sub> adenosine receptor agonist CF101. *Int. J. Mol. Med.* 28, 727–731
- 74 Lee, H.T. *et al.* (2006) A<sub>3</sub> adenosine receptor activation decreases mortality and renal and hepatic injury in murine septic peritonitis. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 291, R959–R969
- 75 Chung, H. *et al.* (2006) The antitumor effect of LJ-529, a novel agonist to A<sub>3</sub> adenosine receptor, in both estrogen receptor-positive and estrogen receptor-negative human breast cancers. *Mol. Cancer Ther.* 5, 685–692
- 76 Karin, M. (2006) Nuclear factor-kappa B in cancer development and progression. *Nature* 441, 431–436
- 77 Zhai, P. et al. (2011) Differential roles of GSK-3β during myocardial ischemia and ischemia/reperfusion. Circ. Res. 109, 502–511