# HGVS recommendations for the description of sequence variants: 2016 update

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#### ABSTRACT

The consistent and unambiguous description of sequence variants is essential to report and exchange information on the analysis of a genome. In particular, DNA diagnostics critically depends on accurate and standardized description and sharing of the variants detected. The sequence variant nomenclature system proposed in 2000 by the Human Genome Variation Society has been widely adopted and has developed into an internationally accepted standard. The recommendations are currently commissioned through a Sequence Variant Description Working Group (SVD-WG) operating under the auspices of three international organizations: the Human Genome Variation Society (HGVS), the Human Variome Project (HVP), and the Human Genome Organization (HUGO). Requests for modifications and extensions go through the SVD-WG following a standard procedure including a community consultation step. Version numbers are assigned to the nomenclature system to allow users to specify the version used in their variant descriptions. Here we present the current recommendations, HGVS version 15.11, and briefly summarize the changes that were made since the 2000 publication. Most focus has been on removing inconsistencies and tightening definitions allowing automatic data processing. An extensive version of the recommendations is available online, at http://www.HGVS.org/varnomen.

KEY WORDS: standards; sequence variation; mutation; database; nomenclature

#### INTRODUCTION

Sequence variant nomenclature needs to be accurate, unambiguous and stable, but sufficiently flexible to allow description of all known classes of sequence variation. After the publication of some initial guidelines [Ad Hoc Committee on Mutation Nomenclature, 1996; Antonarakis, 1998], the HGVS proposed a more comprehensive set of recommendations [den Dunnen and Antonarakis, 2000], now known as the HGVS recommendations/nomenclature (http://www.HGVS.org/varnomen). These guidelines have gradually acquired world-wide acceptance and are currently acknowledged as the standard nomenclature in molecular diagnostics [Gulley et al., 2007; Richards et al., 2015]. As the recommendations were used, it was recognized that the initial proposal had a few errors, contained some inconsistencies, and did not cover all types of sequence variants (*e.g.* complex changes). This paper will summarize the current recommendations, the result of the evolution of the original recommendations [den Dunnen and Antonarakis, 2000] applied in practice: HGVS recommendations version 15.11.

#### THE HGVS RECOMMENDATIONS

The HGVS recommendations are designed to be stable, meaningful, memorable and unequivocal. Still, modifications may be necessary to remove inconsistencies, clarify confusing conventions and/or to extend the recommendation to represent cases that were not previously covered. To allow users to specify up to what time-point they have followed the HGVS recommendations, a version numbers is now included. Any change in the recommendations will result in a new dated version number and all changes introduced will be specified in the version list. *The recommendations described here represent the HGVS recommendations for the description of sequence variants version 15.11 (i.e. 2015 November)*.

#### SVD-WG

To support overall acceptance of the HGVS recommendations, three organizations (HGVS, HVP, and HUGO) joined forces to establish the Sequence Variant Description Working Group (SVD-WG). The SVD-WG operates following a standard procedure (for details see http://www.variome.org/svd-wg.html), discusses incoming requests to modify or extend the recommendations and, where necessary, makes a proposal for changes. When finalized, the proposal is published on the nomenclature website and, for a two-month period, opened for "Community Consultation". People interested in the work of the SVD-WG can register for a mailing list, ensuring that they will receive a copy of all proposals and decisions made. After two months, all comments are collected and evaluated by the SVD-WG. When no major concerns are received the proposal is

accepted, published on the nomenclature website and a new version number is assigned to the recommendations. Proposals receiving many comments are either modified or considered rejected. The entire procedure described was completed for the first time in October 2015 and resulted in the acceptance of two new proposals (see *Community consultation*).

#### Terminology

In contrast to the original recommendations, the terms "polymorphism" and "mutation" are no longer used because both terms have assumed imprecise meanings in colloquial use. Polymorphism is confusing because in some disciplines it refers to a sequence variation that is not disease causing, whereas in other disciplines it refers to a variant found at a frequency of 1% or higher in a population. Similarly, mutation is confusing since it is used both to indicate a "change" and a "disease-causing change". In addition "mutation" has developed a negative connotation [Cotton, 2002; Condit et al., 2002], while the term "variant" has a positive value in discussions between medical doctors and patients by de-dramatizing the implication of the many, often largely uncharacterized, changes detected. Therefore, following recommendations of the Human Genome Variation Society (HGVS) and American College of Medical Genetics (ACMG) [Richards et al., 2015] we only use neutral terms like "variant", "alteration" and "change".

#### **Definitions**

To enhance clarity as well as to facilitate computational analysis and description of sequence variants, the basic types of variants had to be defined more strictly. In addition, descriptions have been prioritized, meaning that when a description is possible according to several classes, *e.g.* as a duplication or an insertion, one specific class is preferred. The priority assigned is (1) deletion, (2) inversion, (3) duplication, (4) conversion, (5) insertion. These changes made it possible to generate a formalized description of the HGVS standard in Extended Backus-Naur Form [Laros et al., 2011] and to develop software tools that can check and/or generate HGVS descriptions [Wildeman et al., 2008; Hart et al., 2014].

The definitions are shown in Table 1. The consequences of these definitions are *e.g.* that an A>T substitution should not be described as an inversion. Similarly, a change where one nucleotide is replaced by more than one other nucleotide is not a substitution but a deletion-insertion (delins/indel). A duplication is defined as a tandem copy of an upstream sequence. The position of a duplication is represented by defining the position of the nucleotide, or the range of nucleotides, that is duplicated (see Tab.4 and *Variability in repeated sequences* below). When the "duplicated" sequence is not directly 3' to the original copy it should be described as an insertion. Insertions are in general short and *de novo*, *i.e.*, not a copy of an existing sequence from elsewhere in the genome. For larger duplicating insertions, *i.e.*, having a copy elsewhere in the genome, one should define the original source sequence with respect to a reference sequence and a nucleotide range.

#### Reference sequences

Discussions regarding the preferred use of reference sequences (Table 2) remain lively. The most fundamental recommendation is that one has to describe what *was observed*, not what has been *inferred*. i.e., report original observations using an appropriate reference sequence. When genomic DNA is sequenced, a genomic reference sequence should be the preferred choice. However, especially in diagnostics, reporting based on a coding DNA reference sequence is far more popular. The reason is simple, from the description one immediately gets some information regarding the location of the variant; namely exonic or intronic, 5' of the ATG or 3' of the stop codon and, by dividing the nucleotide number by 3, the number of the amino acid residue that is affected (see Fig.1, Table 3). For diagnostic applications, a new reference was recently introduced, the Locus Reference Genomic sequence (LRG) [Dalgleish et al., 2010; MacArthur et al. 2014]. The HGVS recommendations strongly advise the use of an LRG. When for a gene of interest no LRG is available, one should be requested as soon as possible. "Pending" LRGs should not be used (they might change before being approved). If there is no current LRG, the use of a RefSeq sequence (O'Leary et al., 2016), with its version (RefSeqGene or transcript) is recommended.

#### Existing standards

To become compliant with prior standards and conventions, a few changes to the original recommendations were required. The most prominent of these concerned termination variants at the protein level, where "X" had previously been used to describe a stop

codon, to be changed to "Ter" or "\*"; e.g. p.Trp123Ter or p.W123\* (previously p.Trp123X). With this change the recommendations now follow the IUPAC-IUB nomenclature and its use of symbols for amino acids and peptides published in 1984 [IUAPC-IUB, 1984] where "X" is defined to indicate "any amino acid". "Ter" (termination codon) is used in three-letter amino acid code, and "\*" can be used in both one- and three-letter code.

It should be noted that HGVS recommendations follows the full IUPAC-IUBMB "Nomenclature for Incompletely Specified Bases in Nucleic Acid Sequences" (Table 2). This facilitates, for example, the description at DNA level of the uncertainty which remains when publications list variants only at protein level; e.g. p.(Ile321Leu) on DNA level as c.961A>Y (Y being C or T).

#### Modifications to original recommendations

The 2000 recommendations [Den Dunnen and Antonarakis, 2000] contained some minor inconsistencies that demanded modification. A leading feature, to reduce confusion, was to use specific symbols for one purpose only. In the 2000 recommendations the "+" character was used both in nucleotide numbering (for intronic nucleotides) and to list combined variants or alleles. The recommendation is now to "+" only in nucleotide numbering and to use ";" to list combined variants or alleles: c.[76C>T; 283G>C] for two variants known to be on one molecule (in cis), c.[76C>T];[283G>C] for the same two variants known to be on two different molecules (in trans) and c.[76C>T(;)283G>C] for two variants where the phase in unknown. Similarly, "," was introduced to replace the

"+" to describe several RNA transcripts deriving from one variant at DNA level. In the example from Den Dunnen and Antonarakis [2000] r.[76a>c, 70\_77del] now replaces r. [76a>c + 70\_77del].

The alternative use of c.IVS# and c.EX# in the description of nucleotide positions has been retracted. Such descriptions are indirect and cause confusion when different exon/intron numbering schemes are used for a gene. They also compromise the development of software tools. Some tools, e.g. the Mutalyzer Name Checker, accept the c.IVS format, supporting conversion to the recommended format [Wildeman et al., 2008]. Nucleotide positions should be specific and include numbers only (Fig.1, Table 3).

Recommendations regarding the description of variants that have not been fully characterized, e.g. deletion or duplication breakpoints, have been clarified. In such cases the description should indicate the region of uncertainty, using the format (5'border\_3'border). The suggestion to describe exon deletions and duplications using the format c.77-?\_923+?del (or dup) was therefore retracted. Descriptions of genomic deletions therefore have formats such as g.2345\_6789del (break point sequenced), g. (1234\_3456)\_(5678\_7890)del (breakpoints defined but not sequenced), g.(? 3456) 6789del (5' break point undefined, 3' break point sequenced).

Reporting one variant using different descriptions creates confusion and underrepresentation of its frequency. Preventing this is especially important in stretches of repeated DNA sequences. Although not stated prominently, the 2000 guidelines specified normalization (shuffling) to the 3' end. This so called 3' rule states that for variants in stretches of repeated DNA sequences the *most 3' position possible* is arbitrarily deemed to have been changed. Consequently the change of TTT to TT is described as g.3del (not g.1del or g.2del). A corollary of the 3' rule is that predicted duplications and deletions of amino acids are similarly normalized to the most C-terminal position.

The description of variability in repeated sequences has been slightly modified and specified more precisely. Such changes are described by defining the first nucleotide of the repeat unit, or the range of the first repeat unit, with the number of repeat units specified between square brackets, e.g. g.123\_124[4]. For short/simple repeats it is allowable to include the content of the repeated unit, using the format "position-first-nucleotide-repeat-content[number]" such as g.123TG[4]. When the size of the repeat unit is uncertain, this should be specified using parentheses; g.-128GGC[(600\_800)].

For descriptions at the protein level, it is strongly recommended to use the threeletter amino acid code. The three-letter code retains better compatibility with IUPAC and leads to fewer errors when used, especially for those amino acids where the first letter differs from their one-letter code (e.g. Aspartic acid (D), Asparagine (N) and Arginine (R)). The use of the one-letter code should be restricted to the description of long sequences.

At the protein level, besides changing the description of a translational stop codon variant from X to Ter/\*, the description of frame shifts has been specified and additions have been made to describe variants affecting the translation start and stop codon. In addition the recommendation has been made that descriptions at the protein level should make clear whether experimental proof was available or not. When not, one should list

the predicted consequences in parentheses. Variants that are predicted to shift the translational reading frame should be described using either a short or a long form; p. (Arg97fs) and p.(Arg97Profs\*23) respectively. For "fsTer#"/"fs\*#" it is specified that "#" indicates at which codon number the new reading frame ends with a stop codon. The number of the stop in the new reading frame is calculated starting at the first amino acid that is changed by the frame shift, ending at the stop codon (\*#).

A newly added recommendation is to describe changes directly affecting the start or stop codon as an N- or C-terminal extension. Amino acids upstream of the original start site are numbered using a minus sign. For example, p.Met1ValextMet-12 describes the observed N-terminal extension of 12 amino acids (Met-12 to Thr-1) of the protein as the consequence of a variant (DNA c.1A>G) that changes amino acid Met1 to Val. Similarly, p.Ter110GlnextTer17 describes the observed extension of the C-terminus of the protein with 17 new amino acids as a consequence of a variant (DNA c.331T>C) that changes Ter110 to Gln.

#### Community consultation

The Sequence Variant Description Working Group (SVD-WG) currently commissioning the variant description recommendations under the auspices of HGVS, HVP and HUGO recently completed the first round of proposals. Proposal SVD-WG001 was accepted allowing the description g.123G= to indicate that a variant screen was performed but no change detected. Similarly, when a variant g.456G>A has no predicted consequences at the protein level this can be described as p.(Arg152=). Proposal SVD-WG002 was

accepted allowing the use of a non-coding DNA reference sequence using the prefix "n." (n.963G>C, Table X). Proposal SVD-WG003 to further specify the description of exon deletions/duplications detected using MLPA is still under discussion. Discussions are ongoing to achieve a harmonization of the nomenclature systems of the HGVS and the International Standing Committee on Human Cytogenetic Nomenclature [ISCN, 2016], but these have not yet been finalized. The HGVS nomenclature as presented in this paper are those of *version 15.11*.

#### Dissemination

The HGVS nomenclature pages lists an email address for questions (VarNomen @ HGVS.org). The chair of the SVD-WG collects all requests for information and/or clarification of the recommendations. In most cases questions can be answered easily, in rare cases the SVD-WG is consulted first before an answer is sent. To promote the recommendations a Facebook page (http://www.facebook.com/HGVSmutnomen) has been started where, on a regular basis, topics of interest are discussed. These include simplified Q&As, meetings where the recommendations are discussed and the release of new proposals for Community consultation. Current action points for the SVD-WG are the development of educational material and a restructuring of the HGVS nomenclature pages.

Although the guidelines have gradually acquired world-wide acceptance, there is still room for improvement. Initiated by Human Mutation, the first journal to demand their use, strong support is coming from many journals making the use of HGVS

nomenclature for sequence variant descriptions mandatory. All major variant databases support HGVS nomenclature and professional organizations have started to demand its use in clinical diagnostic reporting [Gulley et al., 2007; Richards et al., 2015]. However, in specific disciplines HGVS nomenclature is used less frequently and alternative nomenclature systems survive [e.g. Berwouts et al., 2011; Kalman et al., 2015]. EQA providers have detected the problem [REFERENCE, 2016] and started to more actively promote HGVS nomenclature by asking participants to follow the recommendations and subtracting marks when labs fail to do so correctly. In this respect it should be noted that excellent open source support tools have been developed that make it very easy to check whether correct HGVS descriptions are reported [Wildeman et al., 2008; Hart et al., 2014]. The latest version of the Mutalyzer suite includes a Variant Description Extractor that, based on a reference and sample sequence, will report all changes present in HGVS nomenclature (https://www.mutalyzer.nl/description-extractor). Finally, the HGVS nomenclature website is currently being completely reconstructed to increase usability and a start is being made to add educational and training material.

#### ACKNOWLEDGEMENTS

We gratefully acknowledge everyone who has, over the years, contacted the HGVS to point out problems with the recommendations, errors/inconsistencies on the web pages and those who have participated in the recent round of Community Consultation of the SVD-WG. Special thanks go to the late Dick Cotton, a pioneer in the uniform nomenclature movement, a strong supporter and warm motivator for the committee and, as Editor of Human Mutation, the first to make HGVS nomenclature mandatory in publications. Finally, we thank the Human Variome Project (HVP, International Coordinating Office) for administrative support establishing and operating the Sequence Variant Description Working Group (SVD-WG).

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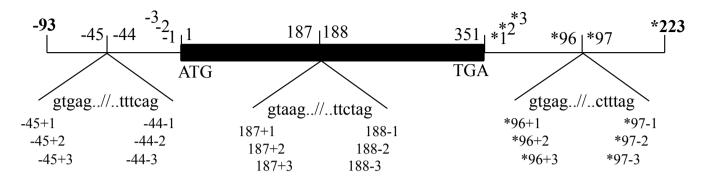
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#### **LEGENDS TO THE FIGURES**

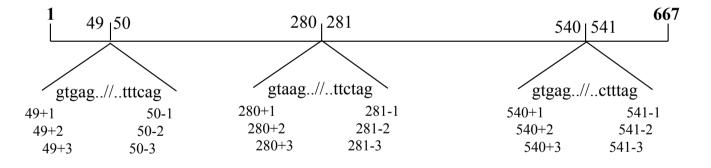
#### Figure 1

Nucleotide numbering for a coding (top) and non-coding (bottom) DNA reference sequence; black box = the protein coding sequence. In the coding DNA reference sequence nucleotide numbering starts with c.1 at the A of the ATG translation initiation codon. Numbering proceeds until the last nucleotide of the translation termination codon (TG<u>A</u>, TA<u>A</u> or TA<u>G</u>). Nucleotides 5' of the ATG are numbered c.-1, c. -2, etc., nucleotides 3' of the stop codon c.\*1, c.\*2, etc. Intronic nucleotides are numbered based on the closest flanking exon nucleotide, on the 5' side going into the intron like c.187+1, c.187+2, etc., on the 3' side going in to the intron like c.188-1, c.188-2, etc. When introns have an uneven number of nucleotides, the central nucleotide (N) in linked to the upstream exon, like c.187+N. Nucleotide numbering for a non-coding DNA reference sequence starts with nucleotide c. and ends at the end of the reference sequence. Intronic nucleotides are numbered as for a coding DNA reference sequence.

# coding DNA reference



# noncoding DNA reference



## Table 1 Nomenclature definitions with example variant descriptions

Substitution (>)	g.1318G>T	a change where <i>one</i> nucleotide is replaced by <i>one</i> other nucleotide
Deletion  (del)	g.3661_3706del	a change where one or more nucleotides are not present (deleted)
Inversion (inv)	g.495_499inv	a change where <b>more than one</b> nucleotide replaces the original sequence and is the reverse-complement of the original sequence (e.g. CTCGA to TCGAG)
Duplication (dup)	g.3661_3706dup	a change where a copy of one or more nucleotides are inserted directly 3' of the original copy of that sequence
Insertion (ins)	g.7339_7340insTAGG	a change where one or more nucleotides are inserted in a sequence <b>and</b> where the insertion is not a copy of a sequence immediately 5'
Conversion (con)	g.333_590con1844_2101	a specific type of deletion-insertion where a range of nucleotides replacing the original sequence are a copy of a sequence from another site in the genome
Deletion- Insertion (delins/indel)	g.112_117delinsTG	a change where one or more nucleotides are replaced by one or more other nucleotides <b>and</b> which is not a substitution, inversion or conversion

In all cases for "*a change where*" read "*a change where in a specific sequence compared* 

to the reference sequence...".

Numbering scheme	Prefix	Position numbering in relation to	
genomic DNA <sup>1</sup>	g.	first nucleotide of the genomic reference sequence	
coding DNA <sup>1</sup>	c.	first nucleotide of the translation start codon of the coding DNA reference sequence	
non-coding DNA <sup>2</sup>	n.	first nucleotide of the non-coding DNA reference sequence <sup>2</sup>	
mitochondrial DNA	m.	first nucleotide of the mitochondrial DNA reference sequence	
RNA	r.	first nucleotide of the translation start codon of the RNA reference sequence <b>or</b> first nucleotide of the non-coding RNA reference sequence	
protein	p.	first amino acid of the protein sequence	

#### Table 2Reference sequences

<sup>1</sup>For diagnostic applications it is strongly recommended to use an LRG, Locus Reference Genomic sequence (LRG) [Dalgleish et al., 2010; MacArthur et al., 2014]. When no LRG is available, one should be requested, "pending" LRGs should not be used. If there is no LRG, a RefSeq sequence (O'Leary et al., 2016) is recommended. <sup>2</sup>Reference sequence recently added after community consultation by the HGVS/HVP/HUGO sequence variant description working group (SVD-WG,

http://www.hgvs.org/mutnomen/accepted002.html).

Location nucleotide	genomic reference sequence		coding DNA reference
	NC_000023.1	LRG_199	sequence LRG 199t1,
	0	(DMD)	NM_004006.2
5' transcription start	g.33231774	g.130953	c2345
in 5' UTR	g.33229552	g.133175	c123
(in intron in 5' UTR) <sup><math>a</math></sup>			(c55+23 / c54-23) <sup>b</sup>
A of the ATG start	g.33229429	g.133298	c.1
codon			
in coding DNA	g.32862930	g.499797	c.234
in intron, 5' side	g.32380903	g.981883	c.5325+2ª
in intron, 3' side	g.32366647	g.996080	c.5326-2ª
G of TAG stop codon	g.31140036	g.2222791	c.11058
in 3' UTR	g.31139691	g.2222836	c.*345
(in intron in $3' UTR$ ) <sup>a</sup>			(c.*54+23 / c.*55-23) <sup>b</sup>
3' transcription end	g.31136580	g.2226247	c.*3456

#### Table 3Nucleotide numbering

Nucleotide numbering using a genomic reference sequence (NC\_000023.10 (genome build GRCh37/hg19), and LRG\_199) and a coding DNA reference sequence (DMD gene, LRG\_199t1 or NM\_004006.2). Nucleotide numbering starts at 1; there is no nucleotide 0. <sup>a</sup>Coding DNA reference sequence NM\_004006.2 does not contain intron sequences, LRG\_199 is required for this description. <sup>b</sup>Hypothetical example, the DMD gene does not contain an intron in the 5' or 3' UTR.

## Table 4DNA level descriptions

Variant type	g. description	c. description <sup>a</sup>	Remarks
substitution	g.32662262G>A	c.1318G>A	
deletion	g.32466684_32466698del	c.3661_3706del	specification of deleted nucleotides(s) optional
duplication	g.32466684_32466698dup	c.3661_3706dup	specification duplicated nucleotide(s) optional
insertion	g.31792279_31792280insTAGG	c.7339_7340insTAGG	specification of inserted nucleotides mandatory
inversions	g.32481638_32481654inv	c.3334_3350inv	minimum size: 2 nucleotides
deletion-insertion (indel)	g.32867914_32867919delinsTG	c.112_117delinsTG	specification of inserted nucleotides mandatory
translocation			no recommendation yet <sup>b</sup>
repetitive DNA stretch	g.31836932T[22] g.33170306TAA[9] or g.33170306_33170308[9]	c.7309+1160T[22] c.31+59093TAA[9] or c.31+59093_31+59095[9]	describe first nucleotide and repeat unit or range of first repeat unit with number of repeat units between brackets
2 variants on 1 chromosome ( <i>in cis</i> )	g.[32841486C>T;33038273G>C]	c.[76C>T; 283G>C]	
2 variants on 2 different chromosomes ( <i>in trans</i> )	g.[32841486C>T];[33038273G>C]	c.[76C>T];[283G>C]	
2 variants, phase unknown (on 1 or 2 chromosomes)	g.[32841486C>T(;)33038273G>C]	c.[76C>T(;)283G>C]	

Variants are described in relation to hypothetical genomic and coding DNA reference sequences. A more extensive collection of examples is available from the HGVS nomenclature website. <sup>a</sup>For another location of the nucleotide relative to a coding DNA reference sequence see Table

3. <sup>b</sup>Subject of proposal SVD-WG004.

Variant type	r. description	Remarks
substitution	r.1318g>u	
deletion	r.3661_3706del	no specification of deleted nucleotide(s)
duplication	r.3661_3706dup	no specification of duplicated nucleotide(s)
insertion	r.7339_7340instagg r.456_457ins456+87_456+121	mandatory specification of inserted nucleotide(s)
inversions	r.3334_3350inv	minimum size 2 nucleotides
deletion-insertion (indel)	r.112_117delinsug	mandatory specification of inserted nucleotides
2 variants on 1 chromosome (in cis)	r.[76c>u;283g>c]	
2 variants on different chromosomes (in trans)	r.[76c>u];[283g>c]	
2 variants, phase unknown	r.[76c>u(;)283g>c]	
one DNA change yielding 2 transcripts	r.[76a>c,70_77del]	
predictions	r.spl r.?	affects splicing unknown consequences

### Table 5RNA level descriptions

Variants are described in relation to a hypothetical RNA reference sequence. Compared to DNA descriptions, lower case nucleotides are used and "u" instead of "t". A more extensive collection of examples is available from the HGVS nomenclature website.

Variant type	p. description	Remarks
	p.(Arg490Ser)	the protein change is predicted (no experimental proof)
substitution	p.Arg490Ser / p.R490S p.Trp87Ter / p.Trp78* / p.W87*	both 3- (preferred) and 1-letter amino acid code may be used; * accepted for 1- and 3-letter code
deletion	p.Asp388_Gln393del	no specification of deleted amino acid(s)
duplication	p.Asp388_Gln393dup	no specification of duplicated amino acid(s)
insertion	p.Ala228_Val229insTrpPro p.Ala228_Val229insLys*	mandatory specification of inserted amino acids
inversions		not possible
deletion-insertion (indel)	p.L7_H8delinsWQQFRTG	mandatory specification of inserted amino acids
frame shift	p.(Arg97fs) p.(Arg97Profs*23)	short and long form accepted; long form contains "fsTer" or "fs*"
extension	p.Met1ValextMet-12 p.Ter110GlnextTer17	short and long form accepted; long form contains "fsTer" or "fs*"
repetitive amino acid stretch	p.Gln34[22] p.Ser7_Ala9[6]	describe first amino acid repeat unit
2 protein coding variants on 1 chromosome ( <i>in cis</i> )	p.[Trp78*;Arg490Ser]	
2 protein coding variants on 2 different chromosomes ( <i>in trans</i> )	p.[Trp78*];[Arg490Ser]	
2 protein coding variants, phase unknown	p.[Trp78*(;)Arg490Ser]	

## Table 6Protein level descriptions

predictions	<i>p</i> .?	unknown consequences

Variants are described in relation to a hypothetical protein reference sequence. A more extensive collection of examples is available from the HGVS nomenclature website.