

Effect of Salt Stress on Germination and Early Seedling Growth of Savory (*Satureja hortensis*)

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Abstract: Soil salinity is one of the most important constraints that limit crop production in arid and semi arid regions. Seed germination is a critical stage in the history of plants and salt tolerance during germination is crucial for the establishment of plants that grow in saline soils. This research was carried out in order to test the effects of different salinity levels on germination and early growth of savory (*Satureja hortensis*). The experiment was carried out using completely randomized design in four replication in 2011 Zabol University laboratory Iran. Experimental treatment includes 4 levels of NaCl concentration (0, 50, 100 and 150 mM). Result showed that the most percentage and speed of germination, plumule length, radicle length and heaviest wet and dry seedling weight referred to control treatment. In 150 mM and more concentration, germination decreased significantly. This reduction in germination indicates this plant's extreme insensitivity to salinity, so it isn't advisable for cultivating in salinity soil. All the result data analyzed by SAS software and comparison of means had been done with Duncan test in 0.05% probable level.

Key words: germination, NaCl, salinity stress, seedling, *Satureja hortensis*.

INTRODUCTION

More than 900 million hectares of land world-wide, approx. 20 % of the total agricultural land, are affected by salt, accounting for more than 6% of the world's total land area. NaCl is the predominant salt causing salinization, and it is unsurprising that plants have evolved mechanisms to regulate its accumulation (Munns and Tester, 2008).

Seed germination is an important and vulnerable stage in the life cycle of terrestrial angiosperms and determines seedling establishment and plant growth. Despite the importance of seed germination under salt stress (Ungar, 1995), the mechanism (s) of salt tolerance in seeds is relatively poorly understood, especially when compared with the amount of information currently available about salt tolerance physiology and biochemistry in vegetative plants (Hester *et al.*, 2001; Hu *et al.*, 2005; Garthwaite *et al.*, 2005; Kanai *et al.*, 2007). In vegetative plants, salt stress causes reduced cell turgor and depressed rates of root and leaf elongation (Werner and Finkelstein, 1995; Fricke *et al.*, 2006), suggesting that environmental salinity acts primarily on water uptake. Furthermore, high intracellular concentrations of both Na⁺ and Cl⁻ can inhibit the metabolism of dividing and expanding cells (Neumann, 1997), retarding germination and even leading to seed death.

The different results were dedicated from the effect of salinity stress on the quantitative and qualitative parameters. For instance, it was found that, increasing of salinity stress decreased almost all of growth parameters in *Nigella sativa*, some growth parameters and essential oil amount in chamomile (Razmjoo *et al.*, 2008). Also Younis *et al.* (2008) reported that enhancing salinity treatments lead to growth reduction. It also reduces germination amounts and seedling weight. Ashraf and Orooj (2006) reported that salinity treatment lead to reduction of growth and plant developments.

Overall, salinity through enhancement of osmotic pressure leads reduction of water absorbance and metabolically and physiological processes will be under its effect. So it cause more delay in germination beginning following by enhancing seed germination duration (Kang and Saltveit, 2002).

Satureja hortensis is an annual, herbaceous plant belonging to the family Labiatae. It is known as summer savory, native to southern Europe and naturalized in parts of North America. The main constituents of the essential oil of Shortness are phenols, carvacrol and thymol, as well as p-cymene, β -caryophyllene, linalool and other terpenoids (Sefidkon *et al.*, 2006).

METHODS AND MATERIALS

The experiment was carried out using completely randomized design in four replication and 4 salinity levels (0, 50, 100 and 150 mM) in 2011 at Zabol University laboratory in Iran. Each experimental unit includes 1 Petri dish with 100 × 150 mm dimension each contains 15 healthy and homogenous seeds which were on the No1 Watman filter paper. First of all, to disinfect seeds, we put them in 10% Hypochlorite Sodium solvent then we washed them 3 times by distilled water. Next, we added 6 ml NaCl solvent to each Petri dish in this way filter

water was wettered by NaCl completely. Eventually, their lids were closed by parafilm and had been located in growth room. The temperature adjusted in 25° C. This experiment took 7 days.

1. The Following Characteristics Were Studied:

1.1. Germination Percentage (GP):

From second day, the germinated seeds were counted daily in specific time. At that time, those seeds were considered germinated which their radical length was more than 3 mm.

Counting continued till we could count more germinated seeds and the resulted final counting considered as final germination percentage.

GP: $N_i / N \times 100$

N_i: number of germinated seed till ith day)

N= total number of seeds.

1.2. Germination Race (GR):

In order that, from the second day to 7th once a 24 hours we counted germinated seeds and its race was determined by Maguire equation (1962):

$$GR = \sum_{i=1}^n \frac{S_i}{D_i}$$

GR: Germination Race (number of germinated seed in each day)

S_i: number of germination seeds in each numeration

D_i: number of days till nth numeration.

n: number of numeration times.

1.3. Seed Vigor (SV):

This index was determined by the following formulation and with the help Abdul-baki and Anderson (1970) method:

Strong seed index = {germination percentage × means of seedling length (radicle + plumule) cm} / 100

At the end of experiment we chose 10 plants from each Petri dish, separated their radicle and plumule and measure each plat’s radicle and plumule length separately. Then we put each repetition on the filter separately. In order to make them dry and measure its dry weight, we put them in oven with 75°C temperature for 24 hours, after we achieved pure numbers, we used SAS software for analyzing them and used Excel software to draw graphs.

RESULTS AND DISCUSSION

Results of statistical analysis of experimental data have been written in table 1 and results of comparison between considered characteristics means have been written in table 2. As table 1 show, salinity made significant differences on all considered characteristics.

Table 1: Result of variance analysis on some germination and growth of seedling characteristics under NaCl concentration.

Mean Square								
S.O.V	df	GP (%)	GR	RL (cm)	PL (cm)	SV	WW (g)	DW (g)
Treatment	3	5218.23**	20.78**	8.71**	3.61**	27.15**	0.00084**	0.00004**
Error	12	15.1	0.043	0.022	0.012	0.027	0.000009	0.0000002
C.V. (%)		9.17	9.33	7.75	12.04	8.15	19.11	13.62

Note: *and ** indicate significant difference at 5% and 1% probability level, respectively.

GR: Germination rate, GP: Germination percentage, PL: Plumule length, RL: Radicle length, SV: Seed vigor, WW: Wet weight, DW: Dry weight.

Table 2: Effect of different salinity concentration on some germination and growth of seedling characteristics.

Salinity concentration (mM)	GP (%)	GR	RL (cm)	PL (cm)	SV	WW (g)	DW (g)
0 (Control)	96.25a	5.67a	3.51a	2.31a	5.6a	0.034a	0.0068a
50	48.75b	2.34b	1.17b	0.9b	1.01b	0.016b	0.0033b
100	27.5c	1.22c	0.65c	0.47c	0.31c	0.007c	0.0004cc
150	13.75d	0.53d	0.2d	0.15d	0.048d	0.00075d	0.000028c

Note: Similar letters in each column hadn’t any significant statistical difference.

GR: Germination rate, GP: Germination percentage, PL: Plumule length, RL: Radicle length, SV: Seed vigor, WW: Wet weight, DW: Dry weight.

Germination Percentage and Race:

According to results of variance analysis, effect of salinity stress level on germination percentage and race were significant (Table 1). Comparison between means of different level of salinity's effects on germination race and percentage has been showed in table 2. As you see in salinity stress the most germination percentage was (96.25%) and the less germination percentage was related to 150 mM concentration (13.75%) (Figure 1).

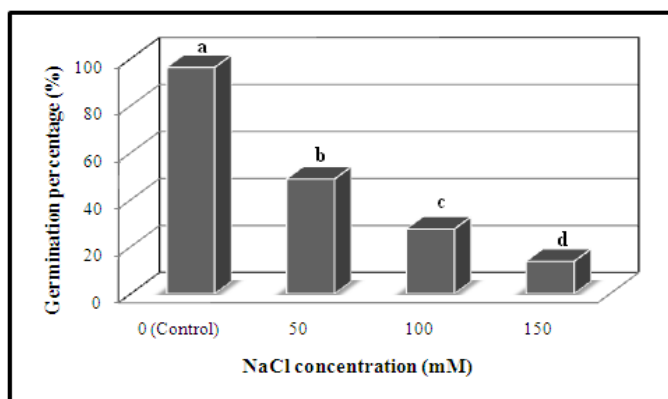


Fig. 1: Effect of different levels of NaCl on germination percentage of *Satureja hortensis*.

The most germination race was related to control also with (5.67) and the less related to 150 mM with (0.53) (Figure 2). Its cause could be more than usual presence of anion, cation which in addition to toxication, decreased water potential that is because of its solubability in water. Ion's so plant can't absorb water and encounter to lake of water (Singah *et al.*, 1988). We also can say that this reduction in germination race relies on salinity bad effect on some physiological processes which are effective on seed germination (Khan *et al.*, 2002).

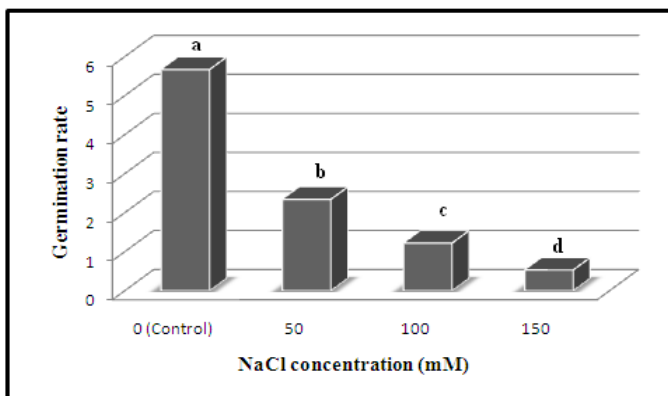


Fig. 2: Effect of different levels of NaCl on germination rate of *Satureja hortensis*.

Radicle and plumule length:

The effects of salinity stress on radicle and plumule length have been showed in table 2 Results a significant difference in radicle and plumule length in 0.05% probable level (Table 1). Comparison of radicle and plumule length means in salinity different level showed that when salinity level increase, seedlings radicle and plumule length decrease.

The most reduction in radicle length (Figure 3) and plumule (Figure 4) related to 150 mM. In this relation Munns and Termaat (1986) suggested that salinity decrease radicle and plumule growth and if we increase salinity level, the amount of reduction will increase. Also Salinity, declines plumule and radicle growth, and by increasing salinity these reduction increase. Salinity which is result of osmotic pressure leads reduction in water absorbance so cell division and differentiation reduce and reduction of plumule and radicle length will be explainable.

Salinity environment have shorter plumule and NaCl more than other salinity factors gas deterrent impact on embryo tissues appearance (Khan and Ungar, 1997).

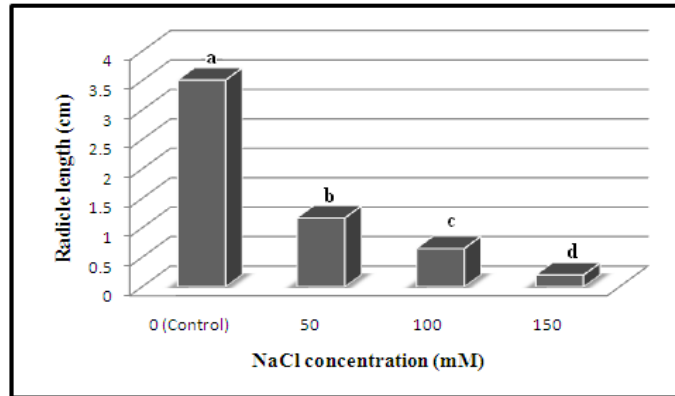


Fig. 3: Effect of different levels of NaCl on radicle length of *Satureja hortensis*.

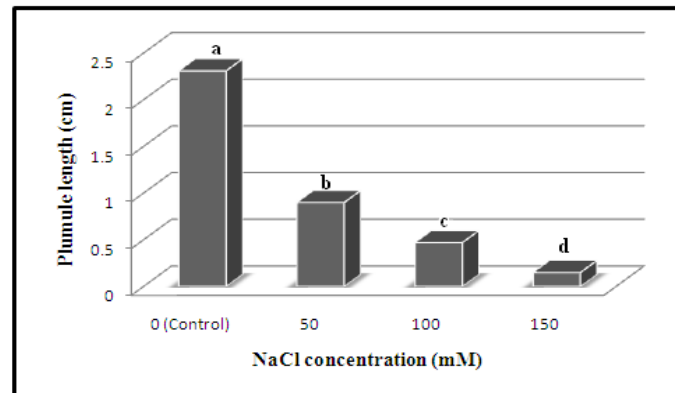


Fig. 4: Effect of different levels of NaCl on plumule length of *Satureja hortensis*.

In addition, Hajar *et al.* (1996) by studying *Nigella sativa L.* different salinity treatment till 300 mM NaCl conclude that, in *Nigella sativa L.* root growth will decrease if salinity increase till 150 mM NaCl. Some studies showed that germinated seeds in salinity environments have short root and shoot and NaCl, has on extreme deterrence effect on embryo tissues' development rather than other salinity materials (Katergi, 1994; Khan and Ungar, 1985).

Seed Vigor:

In strong seed vigor index, had been observed that there exists a significant difference ($P \leq 0.01$) between different salinity levels (table 1). By increasing NaCl concentration, seed vigor index declines (Table 2, Figure 5). The most seed vigor index was related to control treatment (5.6) and the less was related to 150 mM (0.048) (Figure 5).

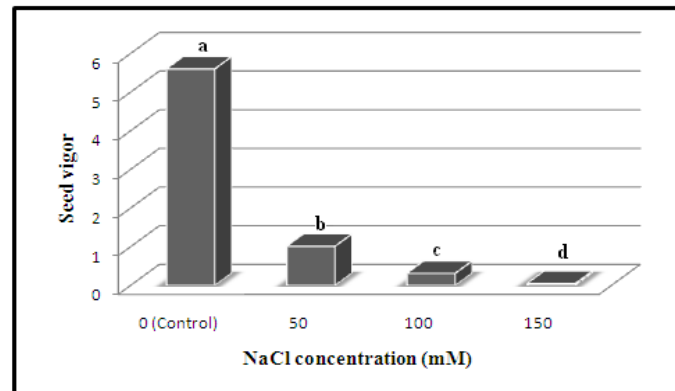


Fig. 5: Effect of different levels of NaCl on seed vigor of *Satureja hortensis*.

Generally, rate and percentage of germination and seed vigor index is related to special impact of ions and reduction of environmental water potential in the presence of salinity. Result showed that if salinity increases (reduction of environmental osmotic potential), seed characteristics will decrease these results is according to Kader and Jutzi (2004) founding's.

Wet and Dry Weight:

Impact of salinity stress treatments, on dry and wet weight of *Satureja hortensis* seedling was significant (Table 1). Impact of salinity stress on dry and wet weight had been showed in Table 2. As you see by enhancing salinity levels, seedlings wet weight amounts decrease extremely. In this case in 150 mM we have 0.00075 gr also in other treatments (100, 50 and 0 mM), dry weights were 0.007, 0.016 and 0.034 in orderly. (Table 2, Figure 6). In addition, dry weight of seedling have similar results which when we increase salinity level till 150 mM dry weight decreases, means that it decrease from 0.0068 gr to 0.000028 gr. (Table 2, Figure 6).

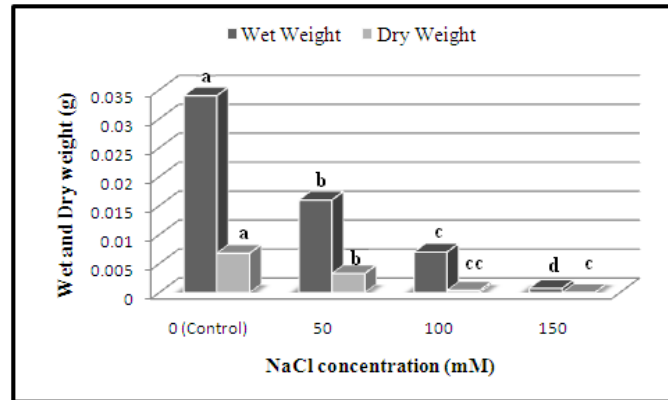


Fig. 6: Effect of different levels of NaCl on dry and wet weight of *Satureja hortensis*.

Etesami and Galeshi (2008) reported that salinity is the cause of reduction in germination percentage, rate and homogeneity of germination and dry weight of barley (*Hordeum vulgare*) seedling. Massai *et al.* (2004) say that salinity is delaying plant growth under reduction of photosynthesis effects, it is cause of closing stomata and reduction of water entrance into the plant and so that it cause duplicate reduction in plant weight.

Redman *et al.* (1994) showed that this reduction in dry weight of plumule and radicle which is results of enhancing the salinity concentration is a normal phenomenon and probably it is the result of low water absorbance by germinated seeds.

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