# KINETIC MODEL FOR TRIGLYCERIDE HYDROLYSIS USING LIPASE: REVIEW

Heri Hermansyah<sup>1</sup>, A. Wijanarko<sup>1</sup>, Dianursanti<sup>1</sup>, M. Gozan<sup>1</sup>, P.P.D.K. Wulan<sup>1</sup>, R. Arbianti<sup>1</sup>, R.W. Soemantojo<sup>1</sup>, T.S. Utami<sup>1</sup>, Yuliusman<sup>1</sup>, M. Kubo<sup>2</sup>, N. Shibasaki-Kitakawa<sup>2</sup>, and T. Yonemoto<sup>2</sup>

Department of Chemical Engineering, University of Indonesia, Depok 16424, Indonesia
Department of Chemical Engineering, Tohoku University, Sendai 980-8579, Japan

E-mail: heri@chemeng.ui.ac.id

### Abstract

Triglyceride hydrolysis using lipase has been proposed as a novel method to produce raw materials in food and cosmetic industries such as diacylglycerol, monoacylglycerol, glycerol and fatty acid. In order to design a reactor for utilizing this reaction on industrial scale, constructing a kinetic model is important. Since the substrates are oil and water, the hydrolysis takes place at oil-water interface. Furthermore, the triglyceride has three ester bonds, so that the hydrolysis stepwise proceeds. Thus, the reaction mechanism is very complicated. The difference between the interfacial and bulk concentrations of the enzyme, substrates and products, and the interfacial enzymatic reaction mechanism should be considered in the model.

Keywords: Lipase, kinetic model, enzymatic reaction mechanism, hydrolysis, triglyceride

### **1. Introduction**

Triglyceride, the main component of natural oil or fat, is stepwise converted into diacylglycerol, monoacylglycerol and glycerol by hydrolysis accompanied with the liberation of a fatty acid at each step [1]. Glycerol and fatty acid are widely used as raw materials, and monoacylglycerol is used as an emulsifying agent in the food, cosmetic and pharmaceutical industries [2]. Recently, diacylglycerol has received much attention as a healthy cooking oil because it has a biological activity to prevent the accumulation of body fat and to lower the level of cholesterol in the blood [3-8].

At present, the Colgate-Emery method has been industrially used for the hydrolysis of triglycerides [9]. This process utilizes steam of high-temperature (523 K) and high-pressure  $(5.00 \times 10^6 \text{ Pa})$ , resulting in high energy consumption and thermal damage of the products. Recently, a hydrolysis method using lipase has been proposed instead of the Colgate-Emery method [10,11]. The enzymatic hydrolysis is conducted under mild condition (at room temperature and atmospheric pressure). Therefore, the above problems can be overcome by this method. Furthermore, the enzymes have substrate and positional specificities [12-17], so that the side reactions such as saponification, polymerization and oxidation are prevented to enhance the yield of the desired product.

In the triglyceride hydrolysis using lipase, the substrates are oil and water, and the hydrolysis takes place at the oil-water interface. In order to industrially utilize this reaction, it is important to elucidate the following subjects.

- 1) Screening lipases having a high activity.
- 2) Selecting solvents never lowering the enzyme activity.
- 3) Investigating effects of various operating factors on the hydrolysis behavior.
- Constructing kinetic model for enzymatic hydrolysis.

### 2. Present Status of the Kinetic Model

A large number of studies have been made on the enzymatic hydrolysis of triglycerides. Biochemical studies on screening lipases from various origins were sufficiently conducted, so that their characteristics such as hydrolysis activity and substrate/positional specificity have been clearly understood [12-23]. Several organic solvents never lowering the enzyme activity have been also reported [20-24]. The effects of the operating factors such as temperature, pH and concentrations of enzyme and substrate on the hydrolysis behavior have been experimentally investigated [18-49]. Many kinetic models have been proposed [50-71], but those simplified models were still not enough to describe the complicated mechanism of the enzymatic triglyceride hydrolysis under wide range of operating conditions.

This is because differences between the interfacial and bulk concentrations of the enzyme, substrates and products and the interfacial enzymatic reaction mechanism were not rigorously considered in the models.

# **3.** Differences between the interfacial and bulk concentrations

In order to describe the differences between the interfacial and bulk concentrations of the enzyme. substrates and products, linear/nonlinear relationship were incorporated. The enzyme concentration at the interface was initially assumed to be proportional to that in the bulk phase [59-67]. However, this assumption was not applicable to a high enzyme concentration [68]. Saturated enzyme concentration was reported to be reached at high enzyme concentration [39,62,64]. Thus, a nonlinear relation, such as the Langmuir adsorption model, should be introduced. On the other hand, for the substrates and/or products, the linear relationships between the interfacial and bulk concentrations were usually incorporated [63-68]. This is because the molecular sizes of the substrates and products are much smaller than that of the enzyme, so that the interfacial concentrations do not reach saturation.

### 4. Interfacial Reaction Mechanism

In order to describe the interfacial enzymatic reaction, reaction mechanisms such as first order, Michaelist-Menten and Ping Pong Bi Bi mechanism were proposed. In the triglyceride hydrolysis, one mole triglyceride (T) reacts with three moles water (W) to produce one mole glycerol (G) and three moles fatty acids (P) as shown by Eq. (1).



In more detail, the triglyceride is stepwise hydrolyzed by the enzyme to be diglyceride (D), monoglycereide (M) and glycerol (G) while the fatty acid is released at each reaction step. The enzyme-substrate complexes are formed at the respective steps. In the simplest model, however, the formation of the enzyme-substrate complexes was neglected, and the irreversible first order reaction mechanism as shown by Eq. (2) was considered [21,50].



Figure 1. Schematic diagram of Ping Pong Bi Bi mechanism



In the models considering the formation of the enzymesubstrate complexes, Michaelis-Menten mechanism as shown by Eq. (3) was incorporated<sup>51)-68)</sup>.



The substrate, S, reacts with the enzyme, E, to form enzyme-substrate complex, ES. Then, product, P, is released. Since one substrate and one product are considered in this mechanism, one fatty acid residue of triglyceride and free fatty acid were simply assumed to be a substrate and a product, respectively.

Recently, there were a few models<sup>69)-71)</sup> incorporating Ping Pong Bi Bi mechanism with two substrates and two products as schematically shown in Fig.1. The reaction proceeds from left to right side as shown by horizontal arrow. The free enzyme, E, reacts with first substrate, S1, to form the first complex, ES1. The first product, P1, is then released from ES1 to form the second complex, F. This complex reacts with second substrate, S2, to form the third complex, FS2. Finally, the second product, P2, is released and the free enzyme is reformed. In case of triglyceride hydrolysis, the first and second substrates were assumed to be one fatty acid residue of triglyceride and water, respectively, while the first and second products were one alcohol residue of triglyceride and free fatty acid, respectively. Triglyceride is stepwise hydrolyzed by the enzyme to be diglyceride, monoglycereide and glycerol, and three ester bonds of triglyceride are not evenly catalyzed by Although some researchers reported the lipase. produced fatty acid inhibited the hydrolysis [20,72,73] the inhibition by fatty acid has never been incorporated in the models considering Ping Pong Bi Bi mechanism.

#### 5. Summary of the Kinetic Model

The proposed models are categorized based on the assumptions for the interfacial reaction mechanism and the differences between the interfacial and bulk concentrations as shown in Figure 2.



Figure 2. Summary of the kinetic models for enzymatic triglyceride hydrolysis

The assumptions listed as abscissa are first order, Michaelis-Menten (MM) and Ping Pong Bi Bi (PPBB) mechanisms with/without stepwise reaction and/or inhibition by fatty acid and they become more complicated far to the right. The assumptions listed as ordinate are no consideration for the relationship between interfacial and bulk concentration, the linear relationship for substrate/product concentration and the linear/nonlinear relationship for the enzyme concentration and the combination of those relationships also become more complicated upward. In the models without considering the differences between the interfacial and bulk concentrations, the complicated PPBB mechanism has been incorporated by Garcia et al. [69] and Rice et al.[70]. In the models considering the differences between the interfacial and bulk concentrations for not only the enzyme concentration but also the substrate/product concentration, however, only the simple MM mechanism was found to be incorporated. In order to construct a rigorous kinetic model to describe the complicated enzymatic hydrolysis of triglyceride under wide range of operating conditions, therefore, the complicated PPBB mechanism should be considered in addition to the relationships between the interfacial and bulk concentrations used in the Al-Zuhair's model [68]. Furthermore, it is important that the stepwise reaction and the inhibition by fatty acid are taken into consideration in the PPBB mechanism.

The most rigorous kinetic model considering the difference between the interfacial and bulk concentrations of the enzyme, substrates and products, and the interfacial enzymatic reaction mechanism was proposed in this model [71]. The model describing the stepwise hydrolysis of triglyceride by nonspecific lipase in the biphasic oil-water system was formulated on the basis of the following assumptions:

- 1. Nonlinear relationship between the interfacial and bulk concentrations of the enzyme
- 2. Linear relationship between the interfacial and bulk concentrations of the substrates and products
- 3. Stepwise hydrolysis proceeds via a Ping Pong Bi Bi mechanism
- 4. The inhibition by oleic acid follows the competitive inhibition mechanism
- The non specific lipase evenly cleave the ester bonds at the edge and the center of the glycerol backbone of the substrates (tri-, di- or monoglyceride)

The model well described the hydrolysis behavior under wide range of operating conditions using Candida rugosa lipase, a nonspecific lipase.

## 6. Conclusion

In order to construct a rigorous kinetic model to describe the complicated enzymatic hydrolysis of triglyceride under wide range of operating conditions, the difference between the interfacial and bulk concentrations of the enzyme, substrates and products, and the interfacial enzymatic reaction mechanism should be considered in the model.

### References

- F. Beisson, A. Tiss, C. Riviere, R. Verger, Method for lipase detection and assay: a critical review, Eur. J. Lipid Sci. Technol. (2000) 133-153.
- [2] J.B. Snape, M. Nakajima, Processing of agricultural fats and oils using membrane technology, J. Food. Eng. 30 (1996) 1-41.
- [3] T. Nagao, H. Watanabe, N. Goto, K. Onizawa, H. Taguchi, N. Matsuo, T. Yasukawa, R. Tsushima, H. Shimasaki, H. Itakura, Dietary diacylglycerol suppressed accumulation of body fat compared to triacylglycerol in men in a double-blind controlled trial, J. Nutrition 130 (2000) 792-797.
- [4] M.G. Soni, H. Kimura, G.A. Burdock, Chronic study of diacylglycerol oil in rats, Food Chem. Tox. 39 (2001) 317-329.
- [5] N. Tada, H. Watanabe, N. Matsuo, I. Tokimitsu, M. Okazaki, Dynamics of postprandial remnant-like lipoprotein particles in serum after loading of diacylglycerols, Clin. Chim. Acta 311 (2001) 109-117.
- [6] T. Murase, T. Mizuno, T. Omachi, K. Onizawa, Y. Komine, H. Kondo, T. Hase, I. Tokimitsu, Dietary diacylglycerol suppresses high fat and high sucrose diet-induced body fat accumulation in C57BL/6J mice, J. Lipid Res. 42 (2001) 372-278.
- [7] T. Murase, M. Aoki, T. Wakisaka, T. Hase, I. Tokimitsu, Anti-obesity effect of dietary diacylglycerol in C57BL/6J mice: dietary

diacylglycerol stimulates intestinal lipid metabolism, J. Lipid Res. 43 (2002) 1312.

- [8] H. Taguchi, T. Nagao, H. Watanabe, K. Onizawa, T. N. Matsuo, I. Tokimitsu, H. Itakura, Energy value and digestibility of dietary oil containing mainly 1,3-diacylglycerol are similar to those of triacylglycerol, Lipids 36 (2001) 379-382.
- [9] J.A. Kent, Riegel's Handbook of Industrial Chemistry, 7th Edition, Van Nostrand Reinhold, New York, USA (1974) pp.368-371.
- [10] P. Villeneuve, J.M. Muderhwa, J. Graile, M.J. Haas, Customizing lipases for biocatalysis: a survey of chemical, physical and molecular biological approach, J. Mol. Cat. B: Enzymatic 9 (2000) 113-148.
- [11] X. Xu, Engineering of enzymatic reactions and reactors for lipid modification and synthesis, Eur. J. Lipid Sci. Technol. 105 (2003) 289-304.
- [12] P. Villeneuve, M. Pina, D. Monte, J. Graille, Determination of lipase specificities through the use of chiral triglycerides and their racemics, Chem. Phys. Lipids 47 (1995) 109-113.
- [13] S. Benjamin, A. Pandey, Candida rugosa lipases: Molecular biology and versatility in biotechnology, Yeast 14 (1998) 1069-1087.
- [14] F.D. Gunstone, What else besides commodity oils and fats, Fett/Lipid 101 (1999) 124-131.
- [15] X. Xu, Production of specific-structured triacylglycerols by lipase-catalyzed reactions: a review, Eur. J. Lipid Sci. Technol. (2000) 287-303.
- [16] R. Sharma, Y. Chisti, U.C. Banerjee, Production, purification, characterization and applications of lipases, Biotechnol. Adv. 19 (2001) 627-662.
- [17] R.V. Muralidhar, R. Marchant, P. Nigam, Lipases in racemic resolutions, J. Chem. Tech. Biotechnol. 76 (2001) 3-8.
- [18] J. Lavayre, J. Verrier, J. Barrati, Stereospecific hydrolysis by soluble and immobilized lipases, Biotechnol. Bioeng. 14 (1982) 2175-2187.
- [19] H.T. Khor, N.H. Tan, C.L. Chua, Lipase-catalyzed hydrolysis of palm oil, JAOCS 63 (1986) 538-540.
- [20] M. Goto, M. Goto, F. Nakashio, K. Yoshizuka, K. Inoue, Hydrolysis of triolein by lipase in a hollow fiber reactor, J. Membrane Sci. 74 (1992) 207-214.
- [21] F.J. Plou, M. Barandiaran, M.V. Calvo, A. Ballesteros, E. Pastor, High-yield production of mono- and di-oleylglycerol by lipase catalyzed hydrolysis of triolein, Enzyme Microb. Technol. 18 (1996) 66-71.
- [22] X.Y. Wu, S. Jaaskelainen, Y. Linko, An investigation of crude lipases for hydrolysis, esterification, and transesterification, Enzyme Microb. Technol. 19 (1996) 226-231.
- [23] L. Nini, L. Sarda, L.C. Comeau, E. Boitard, J.P. Dubes, H. Chahinian, Lipase-catalyzed hydrolysis of short-chain substrates in solution and in emulsion: a kinetic study, Biochim. Biophys. Acta 1534 (2001) 34-44.

- [24] F. Yang, A.J. Russell, A comparison of lipasecatalyzed ester hydrolysis in reversed micelles, organic solvents and biphasic systems, Biotechnol. Bioeng. 47 (1995) 60-70.
- [25] K.C. O'Connor, J.E. Bailey, Hydrolysis of emulsified tributyrin by porcine pancreatic lipase, Enzyme Microb. Technol. 10 (1987) 352-356.
- [26] Y.J. Wang, J.Y. Sheu, F.F. Wang, J.F. Shaw, Lipase-catalyzed oil hydrolysis in the absence of added emulsifier, Biotechnol. Bioeng. 31 (1988) 628-633.
- [27] J.G.T. Kierkels, L.F.W. Vleugels, J.H.A. Kern, E.M. Meijer, M. Kloosterman, Lipase kinetics: Online measurement of the interfacial area of emulsions, Enzyme Microb. Technol. 12 (1990) 760-763.
- [28] D.M.F. Prazeres, F.A.P. Garcia, J.M.S Cabral, Kinetics and stability of a chromobacterium viscosum lipase in reversed micellar and aqueous media, J. Chem. Tech. Biotechnol. 53 (1992) 159-164.
- [29] M. Tanigaki, M. Sakata, H. Wada, Hydrolysis of soybean oil by lipase with a bioreactor having two different membranes, J. Perment. Bioeng. 75 (1993) 53-57.
- [30] A. Sugihara, T. Senoo, A. Enoki, Y. Shimada, T. Nagao, Y. Tominaga, Purification and characterization of a lipase from Phicia burtonii, Appl. Microbiol. Biotechnol. 43 (1995) 277-281.
- [31] X. Fu, X. Zu, K. Gao, J. Duan, Oil and fat hydrolysis with lipase from Aspergillus sp., JAOCS 72 (1995) 527-531.
- [32] Y. Gargouri, A. Bensalah, I. Doucns het, R. Verger, Kinetic behaviour of pancreatic lipase in five species using emulsion and monomolecular films of synthetic glycerides, Biochim. Biphys. Acta 1257 (1995) 223-229.
- [33] E. Cernia, L. Battinelli, S. Soro, Biocatalysed hydrolysis of triglycerides in emulsion and as monolayers, Thin Solid Film 284-285 (1996) 727-730.
- [34] L. Giorno, E. Drioli, Catalytic behavior of lipase free and immobilized in biphasic membrane reactor with different low water-soluble substrates, J. Chem. Tech. Biotechnol. 69 (1997) 11-14.
- [35] C. Albasi, J.P. Riba, I. Sokolovska, V. Bales, Enzymatic hydrolysis of sunflower oil: characterisation of interface, J. Chem. Tech. Biotechnol. 69 (1997) 329-336.
- [36] L.R. Weatherley, D.W. Rooney, M.V. Niekerk, Clean synthesis of fatty acids in an intensive lipasecatalyzed bioreactor, J. Chem. Tech. Biotechnol. 68 (1997) 437-441.
- [37] Y. Shimada, N. Fukushima, H. Fujita, Y. Honda, A. Sugihara, Y. Tominaga, Selective hydrolysis of borage oil with Candida rugosa lipase: two factors affecting the reaction, JAOCS 75 (1998) 1581-1586.

- [38] Q. Gan, H. Rahmat, L.R. Weatherley, Simultaneous reaction and separation in enzymatic hydrolysis of high oleat sunflower oil – evaluation of ultrafiltration performance and process energy, Chem. Eng. J. 71 (1998) 87-96.
- [39] C. Albasi, N. Bertrand, J.P. Riba, Enzymatic hydrolysis of sunflower oil in a standardized agitated tank reactor, Bioproc. Eng. 20 (1999) 77-81.
- [40] Q. Gan, F. Baykara, H. Rahmat, L.R. Weatherley, Analysis of a direct contact membrane reactor for lipase catalyzed oil hydrolysis in a dynamic emulsion system, Catal. Today 56 (2000) 179-190.
- [41] J.C. Wu, Z.M. He, C.Y. Yao, K.T. Yu, Increased activity and stability of Candida rugosa lipase in reverse micelles formed by chemically modified AOT in isooctane, J. Chem. Technol. Biotechnol. 76 (2001) 949-953.
- [42] D. Rooney, L.R. Weatherley, The effect of reaction conditions upon lipase catalyzed hydrolysis of high oleate sunflower oil in a stirred liquid-liquid reactor, Process Biochem. 36 (2001) 947-953.
- [43] S. Al-Zuhair, K.B. Ramachandran, M. Hasan, Investigation of the specific interfacial area of a palm oil-water system, J. Chem. Technol. Biotechnol. 79 (2004) 706-710.
- [44] I.M. Noor, M. Hasan, K.B. Ramachandran, effect of operating variables on the hydrolysis rate of palm oil by lipase, Process Biochem. 39 (2003) 13-20.
- [45] V.R. Murty, J. Bath, P.K.A. Muniswaran, Hydrolysis of rice bran oil using an immobilized lipase from Candida rugosa in isooctane, Biotechnol. Let. 26 (2004) 563-567
- [46] K. Naoe, S. Awatsu, Y. Yamada, M. Kawagoe, K. Nagayama, M. Imai, Solvent condition in triolein hydrolysis by Rhizopus delemar lipase using an AOT reverse micellar system, Biochem. Eng. J. 18 (2004) 49-55.
- [47] H.S. Wu, M.J. Tsai, Kinetics of tributyrin hydrolysis by lipase, Enzyme Microb. Technol. 35 (2004) 488-493.
- [48] H. Haiker, H. Lengsfeld, P. Hadvary, F. Carriere, Rapid exchange of pancreatic lipase between triacylglycerol droplets, Biochim. Biphys. Acta 1682 (2004) 72-79.
- [49] S. Lee, S. Hwang, K. Lee, I.S. Ahn, Microscopic analysis of ester hydrolysis reaction catalyzed by Candida rugosa lipase, Coll. and Surf. B: Biointerfaces. 47 (2006) 78-84.
- [50] C.S. Wang, J.A. Hartsuck, D. Weiser, Kinetics of acylglycerol hydrolysis by human milk protein, Biochim. Biophys. Acta 837 (1985) 111-118.
- [51] R. Verger, M.C.E Mieras, G.H. Haas, Action of phospholipase A at interface, J. Biol. Chem. 248 (1972) 4023-4034.
- [52] J. W. Lagocki, J.H. Law, F. J. Kezdy, The kinetics study of enzyme action on substrate monolayers:

Pancreatic lipase reactions\*, J. Biol. Chem. 248 (1973) 580-587.

- [53] M. Tanigaki, M. Sakata, H. Takaya, K, Mimura, Hydrolysis of palm stearin oil by a thermostable lipase in a draft tube-type reactor, J. Perment. Bioeng. 80 (1995) 340-345.
- [54] R. Arroyo, F. J. Sanchez-Muniz, C. Cuesta, F. J. Burguillo, J. M. Sanchez-Montero, Hydrolysis of used frying palm olein and sunflower oil catalyzed by porcine pancreatic lipase, Lipids 31 (1996) 1133-1139.
- [55] S. Fadiloglu, Z. Soylemez, Kinetic of lipasecatalyzed hydrolysis of olive oil, Food Res. Int. 30 (1997) 171-175.
- [56] R. Arroyo, F. J. Sanchez-Muniz, C. Cuesta, J. V. Sinisterra, J. M. Sanchez-Montero, Thermoxidation of substrate models and their behavior during hydrolysis by porcine pancreatic lipase, JAOCS 74 (1997) 1509-1517.
- [57] S.C. Mohapatra, J.T. Hsu, Lipase kinetics in organic-water solvent with amphipathic substrate for chiral reaction, Biotechnol Bioeng. 55 (1997) 399-407.
- [58] T. Kaambre, V. Tougu, P. Kaambre, H. Vija, P. Sikk, Hydrolysis of emulsified mixtures of triacylglycerols by pancreatic lipase, Biochim. Biphys. Acta 1431 (1999) 97-106.
- [59] O. Martinez, A.M. Wilhelm, J.P. Riba, Kinetic study of an enzymatic liquid-liquid reaction: The hydrolysis of tributyrin by Candida cylindracea lipase, J. Chem. Tech. Biotechnol. 53 (1992) 373-378.
- [60] Y. Kawano, M. Kawasaki, K. Shiomori, Y. Baba, T. Hano, Hydrolysis kinetics of olive oil with lipase in a transfer cell, J. Ferment. Bioeng. 77 (1994) 283-287.
- [61] Y. Kawano, S. Kiyoyama, K. Shiomori, Y. Baba, T. Hano, Hydrolysis of olive oil with lipase in a "VibroMixer", J. Ferment. Bioeng. 78 (1994) 293-297.
- [62] K. Shiomori, T. Hayashi, Y. Baba, Y. Kawano, T. Hano, Hydrolysis rates of olive oil by lipase in a monodispersed O/W emulsion system using membrane emulsification, J. Ferment. Bioeng. 80 (1995) 552-558.
- [63] S. Mukataka, T. Kobayashi, J. Takahashi, Kinetics of enzymatic hydrolysis of lipids in biphasic organic-aqueous system. J. Perment. Bioeng. 63 (1985) 461-466.
- [64] S. Al-Zuhair, M. Hasan, KB.. Ramachandran, Kinetics of the enzymatic hydrolysis of palm oil by lipase, Process Biochem. 38 (2003) 1155-1163.
- [65] S. Al-Zuhair, K.B. Ramachandran, M. Hasan, Unsteady-state kinetics of lipolytic hydrolysis of palm oil in a stirred bioreactor, Biochem. Eng. J. 19 (2004) 81-86.
- [66] S.W. Tsai, C.L. Chiang, Kinetics, mechanism, and time course analysis of lipase-catalyzed hydrolysis

of high concentration olive oil in AOT-Isooctane reversed micelles, Biotechnol. Bioeng. 38 (1991) 206-211.

- [67] S.W. Tsai, G.H. Wu, C.L. Chiang, Kinetics of enzymatic hydrolysis of olive oil in biphasic organic-aqueous systems, Biotechnol. Bioeng. 38 (1991) 761-766.
- [68] S. Al-Zuhair, K.B. Ramachandran, M. Hasan, High enzyme concentration model for the kinetics of hydrolysis of oils by lipase, Chem. Eng. J. 80 (2004) 552-558.
- [69] H.S. Garcia, F.X. Malcata, C.G. Hill, C.H. Amundson, Use of Candida rugosa lipase immobilized in a spiral wound membrane reactor for the hydrolysis of milkfat, Enzyme Microb. Technol. 14 (1992) 535-545.
- [70] K.E. Rice, J. Watkins, C.G. Hill, Hydrolysis of menhaden oil by Candida cylindracea lipase immobilized in a hollow-fiber rector, Biotechnol. Bioeng. 63 (1999) 33-45.
- [71] H. Hermansyah, M. Kubo, N. Shibasaki-Kitakawa, T. Yonemoto: Biochem. Eng. J. 31 (2006) 125-132.