Research Article

Does Cattle Milieu Provide a Potential Point to Target Wild Exophilic Anopheles arabiensis (Diptera: Culicidae) with Entomopathogenic Fungus? A Bioinsecticide Zooprophylaxis Strategy for Vector Control

Issa N. Lyimo,¹ Kija R. Ng'habi,¹ Monica W. Mpingwa,¹ Ally A. Daraja,¹ Dickson D. Mwasheshe,¹ Nuru S. Nchimbi,¹ Dickson W. Lwetoijera,^{1,2} and Ladslaus L. Mnyone¹

¹ Biomedical and Environmental Thematic Group, Ifakara Health Institute, P.O. Box 53, Off Mlabani, Ifakara, Morogoro, Tanzania ² Vector Group, Liverpool School of Tropical Medicine, Liverpool L3 5QA, UK

Correspondence should be addressed to Issa N. Lyimo, ilyimo@ihi.or.tz

Received 14 May 2012; Accepted 30 June 2012

Academic Editor: Wej Choochote

Copyright © 2012 Issa N. Lyimo et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Anopheles arabiensis is increasingly dominating malaria transmission in Africa. The exophagy in mosquitoes threatens the effectiveness of indoor vector control strategies. This study aimed to evaluate the effectiveness of fungus against *An. arabiensis* when applied on cattle and their environments. *Methods.* Experiments were conducted under semi-field and small-scale field conditions within Kilombero valley. The semi-field reared females of 5–7 days old *An. arabiensis* were exposed to fungus-treated and untreated calf. Further, wild *An. arabiensis* were exposed to fungus-treated calves, mud-huts, and their controls. Mosquitoes were recaptured the next morning and proportion fed, infected, and survived were evaluated. Experiments were replicated three times using different individuals of calves. *Results.* A high proportion of *An. arabiensis* was fed on calves (>0.90) and become infected (0.94) while resting on fungus-treated mud walls than on other surfaces. However, fungus treatments reduced fecundity and survival of mosquitoes. *Conclusion.* This study demonstrates for the first time the potential of cattle and their milieu for controlling *An. arabiensis.* Most of *An. arabiensis* were fed and infected while resting on fungus-treated mud walls than on other surfaces. Fungus treatments reduced fecundity and survival of mosquitoes. These results suggest deployment of bioinsecticide zooprophylaxis against exophilic *An. arabiensis.*

1. Background

The feeding and resting behaviours of African malaria vectors are the key determinants of a high malaria transmission intensity [1]. Among African malaria vectors, *Anopheles gambiae* s.s and *An. funestus* are well adapted over many generations feeding and resting inside human houses (i.e. endophagy and endophily, resp. [1, 2]). In contrast, *Anopheles arabiensis* is opportunistically feeding on humans [3, 4] or cattle [1, 2, 5–7] and resting outside (exophily) [8] or inside (endophily) houses [3, 4], based on availability of their preferred host species. The endophilic vector species have been well controlled by insecticide-treated nets (ITNs)

[9, 10], long-lasting insecticide-treated nets (LLINs) [11], and indoor residual spraying (IRS) [12, 13] than exophilic population of *An. arabiensis* in most parts of Africa. These suggest that behaviours of malaria vectors are crucially important on designing effective control strategies.

Outdoor feeding behaviour of exophilic *An. arabiensis* minimizes the risk of being killed by ITN/LLIN and/or IRS since these measures are exclusively applied indoors. A recent study predicted that these interventions increase extrinsic mortality of endophilic, anthrophilic *An. gambiae s.s.*, and *An. funestus* and consequently generate selection pressures for insecticide resistance [14]. These explain the phenomena of declining endophilic vectors, increase of exophilic

An. arabiensis in most parts of Africa [10, 11], and shift of *An. gambiae s.s* from endophagy to exophagy in some locations [15]. The widespread insecticide resistance in population of African malaria vectors [16, 17] suggests an urgent need for alternative strategies to complement the universal coverage of LLIN.

Entomopathogenic fungi of the group hyphomycetes, notably Metarhizium anisopliae and Beauveria bassiana, hold a great promise as complementary mosquito control bioinsecticides [18-23]. These are slow acting [19, 24] and nonrepellent bio-insecticides to Anopheles and Culex mosquitoes [25]. These bio-insecticides kill mosquitoes between 4–10 d after exposure [18, 26-29], before malaria parasites become transmissible as such parasites require $\geq 12 d$ to develop within a mosquito [30]. Also fungus infection reduces blood feeding propensity, life-time fecundity, flight propensity, and flight stamina [19, 27, 31, 32]. Also fungus infection inhibits development of Plasmodium parasites within mosquitoes [20, 32] and kills both insecticide susceptible and resistant mosquitoes [22, 33, 34]. Based on these merits, fungi provide a potential candidate for bio-insecticide zooprophylaxis against exophilic An. arabiensis.

However, optimal methods for delivering fungi against outdoor feeding and resting malaria vectors are yet to be developed. An effective and practical fungus delivery method requires spores to be applied on sites whilst maximizing exposure, maintaining spore viability, and minimizing the required dose of conidia. Few point source delivery methods have been tested: (a) eave curtains, baffles to target host seeking mosquitoes [35], (b) odour-baited stations, clay pots, and cotton cloth attached on the ceiling roof to target resting mosquitoes [29, 36, 37]. Only two of these methods, however, achieved high infection rates (>75%) in a population of wild An. arabiensis through use of human sleepers [35], and synthetic human odours [29] to attract these exophilic, zoophilic mosquitoes. However, in rural settings, cattle, the naturally preferred hosts for exophilic An. arabiensis, are kept close to human houses but inside mud houses or wood posts shelters with thatched roof or palm fronds. Such environment (milieu) may favour spore viability and maintain their infectiousness against mosquitoes thereof. Equally important, this milieu may passively attract exophilic An. arabiensis to blood feed on their preferred cattle hosts and subsequently rest on mud walls and/or thatched roof. However, this cattle milieu has never been exploited as an option to apply fungi against exophilic An. arabiensis.

The purpose of the current study was to evaluate the efficacy of entomopathogenic fungus against the local population of exophilic *An. arabiensis* when applied on various delivery surfaces either individually or in combination: calf, mud walls, and cotton-cloth roofs.

2. Materials and Methods

2.1. Study Site. The study was conducted at the Ifakara Health Institute (IHI) in the Kilombero valley, south eastern Tanzania. The predominant malaria vectors in this valley include *An. gambiae s.s, An. funestus,* and *An. arabiensis* [38–41]. Recent field studies have indicated the decline of *An. gambiae s.s* population within Kilombero valley [11]. However, a population of exophilic *An. arabiensis* is increasing within this valley and other parts of Africa [10, 11]. The preferred hosts for this vector, cattle, are commonly kept in or near human houses within Kilombero valley [42].

2.2. Mosquitoes

2.2.1. Semifield Reared Anopheles arabiensis. Semifield experiments were conducted using female *An. arabiensis* reared under semifield conditions at the IHI. The colony of *An. arabiensis* was established with individuals from the village of Sagamaganga in 2007 and 2008 and is maintained at an ambient temperature varying from 25 to 32°C and a relative humidity of 51 to 90% within the semifield system [43, 44].

2.2.2. Wild Population of Anopheles arabiensis. Small-scale field trials were conducted against freely flying wild population of An. gambiae s.l at Lupiro village in Ulanga district, Kilombero valley ($8.385^{\circ}S$ and $36.670^{\circ}E$). The recent species identification using molecular biology techniques of polymerase chain reaction (PCR) demonstrated that 98% of An. gambiae s.l wild population in this village is composed of An. arabiensis [45]. This confirms that An. arabiensis is the most predominant malaria vector in this village. Generally, these mosquitoes are known to feed on cattle and rest outside human houses in most of villages within Kilombero valley [42].

2.3. Fungal Isolates, Formulation and Application. Two species of entomopathogenic fungi of the group hyphomycetes were used: *Beauveria bassiana* isolate 193-825 and *Metarhizium anisopliae* isolate IP 46. The former and the latter were used in semifield and small-scale field experiments, respectively. These experiments were intended to demonstrate delivery methods of fungus and not a comparison between species. Therefore, before each experiment, conidia viability (>85% germination on Sabouraud dextrose agar) was confirmed.

Fungal conidia were suspended in a 1:1 mixture of highly refined Enerpar oil (Enerpar M002, BP Southern Africa Ltd) and Shellsol oil (Shellsol T, South Africa Chemicals). The test suspensions of conidia were prepared and applied to delivery surfaces based on procedures described by Mnyone et al. [18]. For semifield experiments, a calf was treated with 23 mL suspension of 2.3×10^{10} conidia (1×10^{9} conidia mL^{-1}). The calf was sprayed with the suspension of conidia using a handheld pressure sprayer (Minijet SATA, Germany) at a constant pressure of 2 bars over its whole body including tail and legs. For small-scale field trials, hut walls and cotton-cloth roof were also sprayed with 23 mL conidia suspensions (5 \times 10¹⁰ conidia m⁻²). As in the semifield conditions, 23 mL suspension of 5 \times 10¹⁰ conidia (2.2 \times 10^9 mL^{-1}) was sprayed per calf in the small-scale field trials. The control hut and calves were treated with equal volumes of oil mixture alone. Treatments were done at the study site under tree shade to avoid the effect of intense sunlight on conidia. All surfaces, except cottoncloth, were treated 5 h prior to the experiments to allow proper drying. Cottoncloth was treated 24 h a prior. Calves were restrained and left to dry under tree shade.

2.4. Experimental Setup and Design

2.4.1. Net Huts. The rectangular net hut $(1.5 \times 1.8 \times 2.1 \text{ m})$ Figure 1) was constructed from a regular bed net (Safi net) and placed within a netting enclosed tunnel (100 \times 3.5 \times 2.70 m) of the IHI semifield system. The rectangular shape was maintained by fixing wooden rectangular frame from inside the bed net. The rectangular net hut was partitioned into three chambers using pieces of white clothes: chamber 1 on the left-hand side, chamber 2 on the right-hand side (2 releasing chambers), and a middle chamber (a host chamber). Releasing chambers had a round opening with a sleeve through which mosquitoes were introduced. The top of the vertical white clothes partitioning releasing chambers from host chamber were slanted towards the middle chamber to form baffles with open eaves of 1.5 cm. These openings between top side of a net and the vertical white cloth mimic open eaves that allowed mosquitoes from the releasing chamber to enter the host chamber. Releasing chambers had two zipped slits. The first slit formed entrance into the releasing chamber from outside and the second slit allowed entering into the host chamber from releasing chamber. These slits also allowed introducing a calf into the host chamber. In this chamber, there was a small wooden cage $(1.10 \times 0.59 \times 1 \text{ m})$ for restraining a calf not to damage the net hut. The floor of the net hut was made of nylon carpet for easy cleaning of calf urine and faeces and observing for dead mosquitoes. A strip of grease was kept on the nylon carpet surrounding the net hut from outside to ensure no ants enter the hut and eat (scavenging on) dead mosquitoes.

2.4.2. Mud Huts. Mud huts $(2.2 \times 1.6 \times 1.77 \text{ m}, \text{Figure 2})$ were constructed in the same way people are building their local houses at Lupiro village. Hut walls were made from bamboo sticks and soil from the same village. The walls were plastered by mud. The roofs were made from thatches. The space between the roof and wall was 14 cm from outside. The baffles towards inside the hut were constructed to progressively reduce an eave of 14 cm to 3 cm. This tapered eave space allowed host-seeking mosquitoes to enter the hut but preventing them from exiting through hut. Therefore, wild population of *An. arabiensis* could be attracted to feed on a calf inside the hut and rest on mud plastered walls and/or cotton-cloth attached on thatched roofs.

2.4.3. Experimental Procedures. The semifield experiments were conducted following 2×2 Latin square design (LSD) in two rectangular net huts constructed within a unique Ifakara tunnel system. Two calves were randomly selected from Ifakara communities. One calf was treated with fungal conidia and the second calf remained untreated (control). One net hut contained fungal-treated calf (treatment) and





FIGURE 1: Picture of a rectangular net hut and its schematic representation. The sections of net hut are (A) left-hand side releasing chamber, (B) host or middle chamber, (C) right-hand side releasing chamber, and (D) open eaves with baffles to allow host-seeking mosquitoes to enter host chamber.



FIGURE 2: Mud hut at Lupiro village with fixed (A) cotton-cloth roof treated with fungal conidia, (B) baffles to reduce exit of mosquitoes, (C) mud walls either treated with fungal conidia or untreated for resting mosquitoes, and (D) open eave to allow host seeking mosquitoes to enter inside the hut.

the other hut contained untreated calf. These calves were introduced inside the host chamber in the evening (6:30 pm). Then 150–200 female *An. arabiensis* were introduced into each releasing chamber and left to forage overnight on the calf by entering through open eaves as they do in the natural environment. The next morning all mosquitoes were recaptured from releasing and host chambers. The recaptured mosquitoes were identified whether fed or unfed and then held in the semifield insectary to monitor for their subsequent fitness traits including longer-term survival and fecundity. The number of eggs and days survived were counted and recorded. Dead mosquitoes were put onto moist filter paper in petri dishes, sealed with parafilm, and incubated inside a humid chamber for 3–5 d, after which they were examined for fungal sporulation. The efficacy of fungus over time (persistence) was also preliminarily assessed by exposing mosquitoes to fungal-treated calves at 3 d after treatment. These experiments were replicated four times using four different individual calves.

The small-scale experiments were conducted following 3×3 LSD in three mud huts constructed at the periphery of the Lupiro village. Two mud huts were treated with fungal conidia (one was treated on the walls and the other on the roof) and one hut was left untreated. Three calves were randomly selected from the local communities. Only one of these calves was treated with fungal conidia and introduced inside untreated hut, while the other two untreated calves were placed inside wall- and roof-treated huts, respectively. These calves were introduced into the mud huts at evening time (6:00 pm) and left there overnight. The next morning, all mosquitoes were recaptured from inside each mud hut. The mosquitoes were identified whether fed or unfed. All these mosquitoes were individually transferred into a paper cup and held in the field insectary to monitor their survival. Also all blood fed mosquitoes were provided with wet filter paper in the paper cup for them to lay eggs at 4 d after blood feeding on calf (i.e. Anopheles mosquitoes develop eggs within 3-4 days after blood feeding [46]). The number of eggs and days survived was recorded. Also mosquitoes were processed for sporulation as in the semifield system. The calves were rotated between huts to control for the variation between huts. Therefore, the calves spent three days in the same hut before shifting into another hut. These experiments were replicated three times using three different individual calves to control the variation between their in attractiveness.

2.5. Statistical Analyses. Statistical analysis was conducted to evaluate effects of three fungus delivery methods (cattle, mud walls, and cotton-cloth roofs) against a population of exophilic *An. arabiensis*. Three key parameters were analysed: proportion of fed mosquitoes (feeding success) and showing fungal outgrowth (sporulation), number of eggs (fecundity), and postexposure survival of mosquitoes (number of days survived). The first two parameters are binomial response variable, whereas the last two are continuous response variables.

The binomial and continuous response variables were analysed using generalized linear mixed effect models using an appropriate link function in the R statistical package [47], with "treatments" as fixed effects and "replicate" as a random effect. A base model including only the random effect of "replicate" was constructed, to which the main effect of "treatment" was added to form a full model. The significance of additional fixed effects of treatments was tested by sequentially adding this term to a base model and applying likelihood-ratio test (LRTs) to test if they led to a significant improvement (P < 0.05). For semifield experiments where fungus-treated was compared with untreated calf, the chi-square (χ^2) generated from model comparisons was used to test for significant differences between control and treatment. Whereas for small-scale experiments, more than two treatments were compared and when treatment was

identified as being statistically significant, then Tukey's post hoc test (adjusting for multiple comparisons) was used to identify significant two-way differences between control and treatments and within treatments using *z*-values. All *z*-values reported small-scale experiments are those generated from multiple comparisons in *R* statistical software [47].

The continuous response variable of survival data (only infected mosquitoes noninfected were excluded from analysis) are rarely normally distributed and thus were appropriately analysed using Cox proportional hazards model (coxph) [47, 48]. This model tested whether survival of the mosquitoes varies between delivery methods (calf, house wall, and roof) and days after treatment. A frailty function was used to incorporate the random effect of "replicate", and "treatments" were fit as main effect in R statistical software [21]. The coxph model compared the survival curves of different treatments and gave statistical significant differences in overall mortality rates in hazard ratio (HR) values, which indicate the daily risk of dying [48]. These hazard ratios have now replaced direct comparison of mortality rates after a specific point in time using ttests [49–51]. An HR value of 1 indicates equal mortality rates between treatments and control, an HR value > 1 indicates significantly greater mortality rates in treatment than control, and HR < 1 indicates significantly lower overall mortality rates in treatment than control.

3. Results

A total of 1,690 and 547 female *An. arabiensis* were attracted to calf and being recaptured from, respectively inside net huts in the semifield and mud huts in the field experiments. Almost all recaptured mosquitoes were blood fed (a proportion of 0.90 to 0.99) under both semifield and field conditions. This study evaluated the effects of fungus delivery options (calf, mud wall, and cotton-cloth roofs) on infection rates (sporulation), fecundity, and survival of blood fed, exophilic *An. arabiensis*.

3.1. Semifield Experiments

(i) Proportion of Fed Mosquitoes. The proportion of An. arabiensis fed on calf treated with B. bassiana was slightly higher than on untreated calf ($\chi_1^2 = 7.64$, P = 0.01, Figure 3(a)). However, the magnitude of proportion of fed mosquitoes was >0.93 in both cases (Figure 3(a)).

(*ii*) Proportion of Fungus-Infected Mosquitoes. A freshly fungal-treated calf infected a significantly higher proportion of fed *An. arabiensis* than 3-days posttreated calf ($\chi^2_2 = 101.53$, P < 0.001, Figure 3(b)). Calves infected a proportion of 0.90 of fed *An. arabiensis* immediately after treatment, and ~0.70, 3 d after treatment (Figure 3(b)).

(*iii*) Effect of Fungus on Mosquito Survival and Fecundity. The infection of semifield An. arabiensis with B. bassiana significantly reduced their subsequent number of eggs and survival (Figures 3(c) and 3(d)). The number of eggs laid by *An. arabiensis* fed on fungal-treated calf was 17 eggs less than on untreated calves ($\chi_1^2 = 5.17$, P = 0.02, Figure 3(c)). Similarly, the postfeeding survival of fed *An. arabiensis* varied significantly between fungus-treated and untreated calves ($\chi_1^2 = 19.3$, P < 0.001, Figure 3(d)). The risk of death (hazard ratio-HR) of fed *An. arabiensis* on fungus-treated calves was almost twice that on untreated calves (HR = 1.63 (1.31–2.03), Figure 3(d)).

3.2. Small-Scale Field Experiments

(*i*) Proportion of Fed Wild Mosquitoes. The proportion wild An. arabiensis fed on their natural preferred host (calf) ranged from 0.81 to 1 under field conditions. The proportion of fed wild An. arabiensis from control was similar to all other treatments (P > 0.05, Figure 4(a)). Similarly, within treatments, two-way comparisons revealed that no significant differences in proportion of fed wild An. arabiensis between treatments (P > 0.05, Figure 4(a)).

(*ii*) Proportion of Fungus-Infected Mosquitoes. The proportion of wild *An. arabiensis* infected with *M. anisopliae* isolate IP 46 varied significantly between delivery methods ($\chi_5^2 = 228.05$, P < 0.001, Figure 4(b)). The multiple comparisons revealed that the proportion of infected mosquitoes observed in control was significantly lower than all treatments: treated (cloth + calf), TCTca (z = 7.18, P < 0.001, Figure 4(b)), treated cloth + untreated calf, TCUca (z = 7.78, P < 0.001, Figure 4(b)), treated (wall + calf), TwTca (z = 5.73, P < 0.001, Figure 4(b)), treated wall + untreated calf, TwUca (z = 8.72, P < 0.001, Figure 4(b)), and untreated wall + treated calf, UwTca (z = 6.68, P < 0.001, Figure 4(b)). Within fungal treatments, the proportion of infected mosquitoes was observed to differ significantly between TCUca and TwUca (z = 3.25, P = 0.01, Figure 4(b)).

(iii) Effect of Fungus on Mosquito Survival and Fecundity. The infection of mosquitoes with *M. anisopliae* significantly reduced the number of eggs laid by wild *An. arabiensis* $(\chi_2^2 = 13.61, P = 0.001, Figure 4(c))$. From pairwise multiple comparisons, mosquitoes from the control hut laid significantly more number of eggs than those from treated cottoncloth + untreated calf, TCUca, (z = 4.98, P < 0.001, Figure 4(c)) and treated walls + untreated calf, TwUca (z = -2.43, P = 0.04, Figure 4(c)). However, there was no significant differences between the number of eggs laid by mosquitoes from the hut with TCUca and TwUca (z = 1.54, P = 0.27, Figure 4(c)).

The infection of *An. arabiensis* with *M. anisopliae* had also significantly affected their longer-term survival ($\chi_5^2 = 83$, P < 0.001, Figure 4(d), Table 1). The risks of death (hazard ratio) of these mosquitoes were significantly lower on untreated surfaces than on fungus-treated surfaces (TCTca: $\chi_1^2 = 22.2$, P < 0.001, TCUca: $\chi_1^2 = 29.9$, P < 0.001, TwTca: $\chi_1^2 = 22.0$, P < 0.001, TwUca: $\chi_1^2 = 73$, P < 0.001, UwTca: $\chi_1^2 = 13.6$, P < 0.001, Figure 4(d), Table 1). However, no significant differences of the hazard ratio of fed *An. arabiensis*

TABLE 1: Hazard ratio of *An. arabiensis* after exposure on different fungal-treated surfaces and cattle. The numbers in brackets are 95% confidence intervals. The treatments are abbreviated as TCTca for (treated cloth roof + calf), TCUca for treated cloth + untreated calf, TwTca for (treated mud wall + treated calf), TwUca for (treated mud wall + untreated calf), and UwTca for (untreated mud wall + treated calf).

Fungal treatments	Hazard ratio (HR) Relative to control
Treated (cloth + calf)	2.56 (1.73-3.78)
Treated cloth + untreated calf	2.36 (1.73-3.21)
Treated (wall + calf)	4.05 (2.26-7.26)
Treated wall + untreated calf	4.13 (2.99-5.71)
Untreated wall + Treated calf	2.30 (1.48-3.58)

were observed across fungal treated surfaces (P > 0.05, Figure 4(d)).

4. Discussion

This study demonstrates for the first time the potential of applying fungus to cattle and their milieu (mud walls and roofs) for controlling wild population of exophilic An. arabiensis. A high proportion of these mosquitoes was strongly attracted and fed on cattle (>0.90) and became infected with fungus treatments. Notably, fungus-treated mud walls infected a higher proportion of mosquitoes (~ 0.94) than treated calf and cotton-cloth roof or their combinations under field conditions. The fecundity of fungus-infected mosquitoes on calf, mud wall and cottoncloth roof was, respectively, 17, 40, and 27 eggs less than those on control. Surprisingly, the effects of fungus on survival of An. arabiensis were similar between delivery methods and/or their combinations. However, the magnitude of relative risk of death of mosquito on fungus-treated mud walls was 2 times more than that on treated calf and cotton-cloth roof. Therefore, cattle milieu, mostly mud walls of their houses, could be the best field delivery method of fungus against a population of exophilic, zoophilic An. arabiensis, suggesting a bio-insecticide zooprophylaxis.

Entomopathogenic fungi, notably Metarhizium anisopliae and Beauveria bassiana, are promising potential bioinsecticide against malaria vectors [23, 37, 50]. The mud walls treated with M. anisopliae IP 46 infected a higher proportion of wild An. arabiensis (0.94) than all other treatments or their combinations. This exophilic An. arabiensis is generally known to feed on cattle or on humans if available [5, 7]. Therefore, cattle attracted a high proportion (>0.90) of these mosquitoes that became fed and rest on fungustreated mud walls because the oil formulation of fungal conidia is nonrepellent [25]. Besides, the four sides of the hut provided a bigger surface area of fungus exposure than a calf, and therefore, the likelihood of mosquitoes picking up more conidia on mud walls than on a calf. Furthermore, mud walls provided a natural medium (soil) for better fungal conidia attachment and viability. These results support the previous



FIGURE 3: Effects of *B. bassiana* against semifield reared exophilic *An. arabiensis* populations: (a) Estimated proportion $(\pm 1s.e)$ of fed after exposure to fungus-treated and untreated calf, BB calf indicates a calf sprayed with conidia suspension of *Beauvaria bassiana*. (b) Estimated proportion $(\pm 1s.e)$ of infected mosquitoes after exposure to fungus treated calf on 0 d and 3 d. (c) Estimates $(\pm 1s.e)$ of the mean number of eggs laid by mosquitoes after exposure to fungus-treated and untreated calf. (d) Survival of mosquitoes after exposure to fungus-treated and untreated calf, the lines represent the survival function as estimated from fitting Cox proportional hazard model (controlling for random variation between individual calves).

studies that demonstrate when fungal conidia applied on soil medium (e.g., mud panels [52], clay pots/tiles [19, 36]) the infection rates are higher relative to other substrates. One of these studies demonstrated that a humid and cool environment in a clay pot potentiates high infection rates (>92%) in *An. gambiae* s.s and *An. funestus* in the laboratory and suggested that a human synthetic odour should be incorporated to maximize the number of mosquitoes exposed to fungal conidia [36]. The present study demonstrated that natural preferred hosts (cattle) could attract a high proportion of exophilic *An. arabiensis* that become infected with fungus applied on mud walls of cattle house

Unexpectedly, a significantly low proportion (\sim 0.04) of fed wild *An. arabiensis* was infected by fungus treatments. The possible explanation for this observation could be that fed mosquitoes from fungus-treated huts flew into control



FIGURE 4: Effects of *M. anisopliae* IP 46 against wild exophilic *An. arabiensis*. (a) Estimated proportion $(\pm 1s.e)$ of fed mosquitoes after exposure to fungus-treated and untreated calf inside experimental hut. (b) Estimated proportion $(\pm 1s.e)$ of infected mosquitoes after exposure to fungus-treated and untreated surfaces. (c) Estimates $(\pm 1s.e)$ of the mean number of eggs laid by mosquitoes after exposure to fungus-treated and untreated surfaces. (d) Survival of mosquitoes after exposure to fungus-treated and untreated surfaces. The lines represent the survival function as estimated from the fitting Cox proportional hazard model (controlling for random variation between individual calves).

hut while seeking for the resting places after blood feeding. Alternatively, mosquitoes could have been drifted by wind from fungus-treated huts to control hut.

The fungus is a nonrepellent [25], slow acting bioinsecticide [24], that allows mosquitoes to feed, rest on treated surfaces while developing eggs (e.g., for *Anopheles* mosquitoes take 3-4 days to develop eggs after blood feeding, [46]), and subsequently lay eggs before being killed. This study demonstrated that fungus-infected *An. arabiensis* laid 17–40 eggs less than those mosquitoes in the control group. The possible explanation could be that fungus and mosquitoes compete on the same protein resources and therefore fed mosquitoes that became infected allocated these resources on their survival than egg development. The

observations in this study are consistent with those reported elsewhere [19, 31]. Although a laboratory study revealed that no statistical significant differences between the numbers of eggs laid by fungus-infected and noninfected mosquitoes [53], this is largely linked with similar blood meal size between these two groups such that whatever depleted by fungus left these mosquitoes with threshold volume of blood meal required for egg development [54, 55].

Evolutionary forces act upon reproduction and survival of malaria vectors. The opportunity of surviving fungus infection to lay at least few eggs suggests that they are less likely to generate selection pressures for resistance [24]. The daily survival rates of An. arabiensis were significantly reduced by fungus-treated surfaces by >2 times than on control in both semifield and field conditions. However, the magnitude of the relative risk of death of mosquitoes on fungus-treated mud wall was consistently 4 times higher than on control. These findings are consistent with previous studies that have shown that fungus-treated surfaces reduce daily survival rates of An. gambiae s.l more than untreated surfaces [29, 37]. Similarly, the observations that the efficacy of fungus against mosquitoes was high when applied on mud walls are consistent with those reported under laboratory conditions on mud tiles [19] and mud panels [52]. Most importantly, the present study demonstrated that fungustreated mud walls killed ~70% of the wild population at day 11 after exposure, suggesting that this option has potential of interrupting transmission of Plasmodium falciparum that requires 14 days to develop to transmissible sporozoites within mosquitoes [30].

Fungus kills mosquitoes in 4-10 days depending on exposure dose, viability and virulence of the fungal species/strain, and physiological status of mosquitoes [19, 49, 56]. Therefore, the results reported here have limitations before being compared with other studies or translated into application. First, the present study reported the effect of fungus infection on daily survival rates of blood fed mosquitoes (>95% blood fed mosquitoes) and therefore cannot be directly compared with nonblood fed mosquitoes in other studies. Previous studies have shown that blood resources improve daily survival rates of mosquitoes [57-59], and fungus infection kills nonblood fed mosquitoes much faster than blood fed ones [49]. Second, we briefly measured persistence of fungal conidia on calf under semifield conditions. This study found that ~70% of semifield reared An. arabiensis could be infected by a 3-days-post treatedcalf. However, detailed experiments on persistence of fungal conidia on cattle and mud walls are now planned to be conducted under field conditions: (1) to test for the effect of sunlight (UV) and rainfall on persistence of fungal conidia in grazing cattle, (2) to test for the effect of wet and dry season or smoke on the persistence of fungal conidia on mud walls. Third, this is the first study to apply fungus on cattle for the control of malaria vectors, and therefore we could have underdose mosquitoes. We sprayed the whole body of a calf with a fungal conidia dose of $1.0 - 2.2 \times 10^9$ conidia mL⁻¹ which was slightly higher than the dose tested (1×10^8 conidia mL^{-1}) on cattle to control ticks [60, 61]. However, ticks are sticking on cattle body for days while feeding until become

engorged whereas mosquitoes fly and land on cattle body to feed for few minutes (temporary ectoparasites). This suggests that mosquitoes might have been underdosed. The full-field experiments have been designed to test different doses of fungal conidia applied on either mud walls or calf against wild *An. arabiensis*. Fourth, the use of different fungus species in these experiments: *B bassiana* for semifield experiments and *M. anisopliae* IP 46 for small-scale field experiments. The intention was not to compare their efficacy but was because of the availability of conidia. Although the two fungus species showed similar trend in the semifield and field conditions, now experiments are underway using *M. anisopliae* IP 46 and *B. bassiana* at the same time in the field to confirm if they act on the same direction against malaria vectors in the natural environments.

This study demonstrates for the first time the potential of applying fungus on cattle and their milieu (mud walls and cotton-cloth roof) for the control of exophilic wild An. arabiensis. A high proportion of exophilic An. arabiensis was attracted to both fungus-treated and untreated cattle where they fed and became infected by different fungus treatments. The fungus-treated mud walls infected a higher proportion of An. arabiensis than all other treated surfaces and their combinations. Surprisingly, fecundity and survival of infected mosquitoes were similar between treatments and their combinations, but varied from their controls. Although not significantly but the magnitude of risk of death of these mosquitoes on fungus-treated mud walls was 2 times more than on treated calf and cottoncloth roof. These results suggest that a combination of fungus-treated mud walls and untreated cattle in their milieu could be acceptable, cheap, and easy to apply in rural settings, thus making a perfect bioinsecticide zooprophylaxis that may compliment universal coverage of LLIN.

Conflict of Interests

The authors declare that they have no competing interests.

Authors Contribution

I. N. Lyimo and L. L. Mnyone designed and supervised execution of experiments. I. N. Lyimo analysed the data and drafted the paper. L. L. Mnyone, K. R. Ng'habi, and D. W. Lwetoijera critically reviewed the paper. M. W. Mpingwa, A. A. Daraja, N. S. Nchimbi, and D. D. Mwasheshe performed experiments. All authors read and approved the final version of the manuscript.

Acknowledgments

We would like to thank Nina and Thomas (Penn State University, USA) for providing the *B. bassiana* isolate 193-825 and Christian Luz (*Instituto de Patologia, Tropical e Saude Publica, Universidade, Brazil*) for providing *M. anisopliae* isolate IP 46.

References

- A. Kiszewski, A. Mellinger, A. Spielman, P. Malaney, S. E. Sachs, and J. Sachs, "A global index representing the stability of malaria transmission," *American Journal of Tropical Medicine and Hygiene*, vol. 70, no. 5, pp. 486–498, 2004.
- [2] M. E. Sinka, M. J. Bangs, S. Manguin et al., "The dominant Anopheles vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis," *Parasites and Vectors*, vol. 3, no. 1, article 117, 2010.
- [3] C. M. Fornadel and D. E. Norris, "Increased endophily by the malaria vector *Anopheles arabiensis* in southern Zambia and identification of digested blood meals," *American Journal of Tropical Medicine and Hygiene*, vol. 79, no. 6, pp. 876–880, 2008.
- [4] M. Hadis, M. Lulu, Y. Makonnen, and T. Asfaw, "Host choice by indoor-resting *Anopheles arabiensis* in Ethiopia," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 91, no. 4, pp. 376–378, 1997.
- [5] I. Tirados, C. Costantini, G. Gibson, and S. J. Torr, "Blood-feeding behaviour of the malarial mosquito *Anopheles arabiensis*: implications for vector control," *Medical and Veterinary Entomology*, vol. 20, no. 4, pp. 425–437, 2006.
- [6] R. J. Kent, P. E. Thuma, S. Mharakurwa, and D. E. Norris, "Seasonality, blood feeding behavior, and transmission of *Plasmodium falciparum* by *Anopheles arabiensis* after an extended drought in southern Zambia," *American Journal of Tropical Medicine and Hygiene*, vol. 76, no. 2, pp. 267–274, 2007.
- [7] S. M. Muriu, E. J. Muturi, J. I. Shililu et al., "Host choice and multiple blood feeding behaviour of malaria vectors and other anophelines in Mwea rice scheme, Kenya," *Malaria Journal*, vol. 7, no. 43, pp. 1–20, 2008.
- [8] G. B. White, S. A. Magayuka, and P. F. L. Boreham, "Comparative studies on sibling species of the *Anopheles gambiae* Giles complex (Diptera:Culicidae), bionomics and vectorial activity of species A and species B at Segera, Tanzania," *Bulletin of Entomological Research*, vol. 62, pp. 295–317, 1972.
- [9] K. A. Lindblade, J. E. Gimnig, L. Kamau et al., "Impact of sustained use of insecticide-treated bednets on malaria vector species distribution and culicine mosquitoes," *Journal* of *Medical Entomology*, vol. 43, no. 2, pp. 428–432, 2006.
- [10] M. N. Bayoh, D. K. Mathias, M. R. Odiere et al., "Anopheles gambiae: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province, Kenya," Malaria Journal, vol. 9, no. 1, article 62, 2010.
- [11] T. L. Russell, D. W. Lwetoijera, D. Maliti et al., "Impact of promoting longer-lasting insecticide treatment of bed nets upon malaria transmission in a rural Tanzanian setting with pre-existing high coverage of untreated nets," *Malaria Journal*, vol. 9, no. 1, article 187, 2010.
- [12] M. T. Gillies and A. Smith, "The effect of a residual housespraying campaign in East Africa on species balance in the *Anopheles funestus* group. The replacement of *A. funestus* Giles by *A. rivulorum* Leeson," *Bulletin of Entomological Research*, vol. 51, no. 2, pp. 243–252, 1960.
- [13] B. Pluess, F. C. Tanser, C. Lengeler, and B. L. Sharp, "Indoor residual spraying for preventing malaria," *Cochrane Database* of Systematic Reviews, vol. 4, article CD006657, pp. 1–46, 2010.
- [14] H. Ferguson, N. Maire, W. Takken et al., "Selection of mosquito life-histories: a hidden weapon against malaria?" *Malaria Journal*, vol. 11, no. 1, p. 106, 2012.

- [15] M. R. Reddy, H. J. Overgaard, S. Abaga et al., "Outdoor host seeking behaviour of *Anopheles gambiae* mosquitoes following initiation of malaria vector control on Bioko Island, Equatorial Guinea," *Malaria Journal*, vol. 10, article 184, 2011.
- [16] H. Ranson, H. Abdallah, A. Badolo et al., "Insecticide resistance in *Anopheles gambiae*: data from the first year of a multi-country study highlight the extent of the problem," *Malaria Journal*, vol. 8, no. 1, article 299, 2009.
- [17] H. Ranson, R. N'Guessan, J. Lines, N. Moiroux, Z. Nkuni, and V. Corbel, "Pyrethroid resistance in African anopheline mosquitoes: what are the implications for malaria control?" *Trends in Parasitology*, vol. 27, no. 2, pp. 91–98, 2011.
- [18] L. L. Mnyone, M. J. Kirby, D. W. Lwetoijera et al., "Infection of the malaria mosquito, *Anopheles gambiae*, with two species of entomopathogenic fungi: effects of concentration, coformulation, exposure time and persistence," *Malaria Journal*, vol. 8, no. 1, article 309, pp. 1–12, 2009.
- [19] S. Blanford, W. Shi, R. Christian et al., "Lethal and pre-lethal effects of a fungal biopesticide contribute to substantial and rapid control of Malaria vectors," *PLoS ONE*, vol. 6, no. 8, Article ID e23591, 2011.
- [20] W. Fang, J. Vega-Rodríguez, A. K. Ghosh, M. Jacobs-Lorena, A. Kang, and R. J. S. Leger, "Development of transgenic fungi that kill human malaria parasites in mosquitoes," *Science*, vol. 331, no. 6020, pp. 1074–1077, 2011.
- [21] M. Farenhorst, A. Hilhorst, M. B. Thomas, and B. G. J. Knols, "Development of fungal applications on netting substrates for malaria vector control," *Journal of Medical Entomology*, vol. 48, no. 2, pp. 305–313, 2011.
- [22] M. Farenhorst, B. G. J. Knols, M. B. Thomas et al., "Synergy in efficacy of fungal entomopathogens and permethrin against west african insecticide-resistant *Anopheles gambiae* mosquitoes," *PLoS ONE*, vol. 5, no. 8, Article ID e12081, 2010.
- [23] E. J. Scholte, J. K. Bart, A. S. Robert, and T. Willem, "Entomopathogenic fungi for mosquito control: a review," *Journal of Insect Science*, vol. 4, no. 19, pp. 1–24, 2004.
- [24] A. F. Read, P. A. Lynch, and M. B. Thomas, "How to make evolution-proof insecticides for malaria control," *PLoS Biology*, vol. 7, no. 4, pp. 1–10, 2009.
- [25] L. L. Mnyone, C. J. Koenraadt, I. N. Lyimo, M. W. Mpingwa, W. Takken, and T. L. Russell, "Anopheline and culicine mosquitoes are not repelled by surfaces treated with the entomopathogenic fungi *Metarhizium anisopliae* and *Beauveria bassiana*," *Parasites and Vectors*, vol. 3, no. 1, article 80, 2010.
- [26] E. Scholte, W. Takken, and B. Knols, "Pathogenicity of five east African entomopathogenic fungi against adult Anopheles gambiae s.s. mosquitoes (Diptera, Culicidae)," in Proceedings of the Section Experimental and Applied Entomology of the Netherlands Entomological Society, vol. 14, pp. 25–29, 2003.
- [27] J. George, S. Blanford, M. J. Domingue, M. B. Thomas, F. R. Andrew, and C. B. Thomas, "Reduction in hostfinding behaviour in fungus-infected mosquitoes is correlated with reduction in olfactory receptor neuron responsiveness," *Malaria Journal*, vol. 10, no. 219, pp. 1–13, 2011.
- [28] L. L. Mnyone, T. L. Russell, I. N. Lyimo, D. W. Lwetoijera, M. J. Kirby, and C. Luz, "First report of *Metarhizium anisopliae* IP 46 pathogenicity in adult *Anopheles gambiae* s.s. and *An. arabiensis* (Diptera; Culicidae)," *Parasites and Vectors*, vol. 2, no. 1, article 59, pp. 1–4, 2009.
- [29] D. W. Lwetoijera, R. D. Sumaye, E. P. Madumla et al., "An extra-domiciliary method of delivering entomopathogenic fungus, *Metharizium anisopliae* IP 46 for controlling adult

populations of the malaria vector, *Anopheles arabiensis*," *Parasites and Vectors*, vol. 3, no. 1, article 18, pp. 1–6, 2010.

- [30] J. C. Beier, "Malaria parasite development in mosquitoes," *Annual Review of Entomology*, vol. 43, pp. 519–543, 1998.
- [31] E. J. Scholte, B. G. J. Knols, and W. Takken, "Infection of the malaria mosquito *Anopheles gambiae* with the entomopathogenic fungus *Metarhizium anisopliae* reduces blood feeding and fecundity," *Journal of Invertebrate Pathology*, vol. 91, no. 1, pp. 43–49, 2006.
- [32] S. Blanford, B. H. K. Chan, N. Jenkins et al., "Fungal pathogen reduces potential for malaria transmission," *Science*, vol. 308, no. 5728, pp. 1638–1641, 2005.
- [33] M. Farenhorst, J. C. Mouatcho, C. K. Kikankie et al., "Fungal infection counters insecticide resistance in African malaria mosquitoes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 41, pp. 17443–17447, 2009.
- [34] A. F. V. Howard, R. N'Guessan, C. J. M. Koenraadt et al., "First report of the infection of insecticide-resistant malaria vector mosquitoes with an entomopathogenic fungus under field conditions," *Malaria Journal*, vol. 10, article 24, 2011.
- [35] L. Mnyone, I. Lyimo, D. Lwetoijera et al., "Exploiting the behaviour of wild malaria vectors to achieve high infection with fungal biocontrol agents," *Malaria Journal*, vol. 11, no. 1, p. 87, 2012.
- [36] M. Farenhorst, D. Farina, E. J. Scholte et al., "African water storage pots for the delivery of the entomopathogenic fungus *Metarhizium anisopliae* to the malaria vectors *Anopheles* gambiae s.s. and *Anopheles funestus*," *American Journal of Tropical Medicine and Hygiene*, vol. 78, no. 6, pp. 910–916, 2008.
- [37] E. J. Scholte, K. Ng'Habi, J. Kihonda et al., "An entomopathogenic fungus for control of adult African malaria mosquitoes," *Science*, vol. 308, no. 5728, pp. 1641–1642, 2005.
- [38] T. Smith, J. D. Charlwood, J. Kihonda et al., "Absence of seasonal variation in malaria parasitaemia in an area of intense seasonal transmission," *Acta Tropica*, vol. 54, no. 1, pp. 55–72, 1993.
- [39] J. D. Charlwood, J. Kihonda, S. Sama et al., "The rise and fall of *Anopheles arabiensis* (Diptera: Culicidae) in a Tanzanian village," *Bulletin of Entomological Research*, vol. 85, no. 1, pp. 37–44, 1995.
- [40] J. D. Charlwood, T. Smith, E. Lyimo et al., "Incidence of *Plasmodium falciparum* infection in infants in relation to exposure to sporozoite-infected anophelines," *American Journal of Tropical Medicine and Hygiene*, vol. 59, no. 2, pp. 243–251, 1998.
- [41] J. D. Charlwood, T. Smith, F. F. Billingsley, W. Takken, E. O. K. Lyimo, and J. H. E. T. Meuwissen, "Survival and infection probabilities of anthropophagie anophelines from an area of high prevalence of *Plasmodium falciparum* in humans," *Bulletin of Entomological Research*, vol. 87, no. 5, pp. 445–455, 1997.
- [42] V. Mayagaya, *The Impact of Livestock on Ecology of Malaria Vectors and Malaria Transmission in the Kilombero Valley*, University of Dar es Salaam, Dar es Salaam, Tanzania, 2010.
- [43] I. Lyimo, Ecological and Evolutionary Determinants of Anopheline Host Species Choice and Its Implications for Malaria Transmission, University of Glasgow, Glasgow, UK, 2010.
- [44] H. M. Ferguson, K. R. Ng'habi, T. Walder et al., "Establishment of a large semi-field system for experimental study of African

malaria vector ecology and control in Tanzania," *Malaria Journal*, vol. 7, article 158, 2008.

- [45] F. O. Okumu, G. F. Killeen, S. Ogoma et al., "Development and field evaluation of a synthetic mosquito lure that is more attractive than humans," *PloS ONE*, vol. 5, no. 1, article e8951, 2010.
- [46] M. T. Gillies, "The duration of the gonotrophic cycle in Anopheles gambiae and Anopheles funestus, with a note on the efficiency of hand catching," East African Medical Journal, vol. 30, no. 4, pp. 129–135, 1953.
- [47] M. J. Crawley, The R Book, John Wiley & Sons, 2007.
- [48] D. R. Cox, "Regression of models and life tables," *Journal of the Royal Statistical Societ*, vol. 34, no. 2, pp. 187–220, 1972.
- [49] L. L. Mnyone, M. J. Kirby, M. W. Mpingwa et al., "Infection of *Anopheles gambiae* mosquitoes with entomopathogenic fungi: effect of host age and blood-feeding status," *Parasitology Research*, vol. 108, no. 2, pp. 317–322, 2011.
- [50] C. K. Kikankie, B. D. Brooke, B. G. Knols et al., "The infectivity of the entomopathogenic fungus *Beauveria bassiana* to insecticide-resistant and susceptible *Anopheles arabiensis* mosquitoes at two different temperatures," *Malaria Journal*, vol. 9, no. 1, article 71, 2010.
- [51] M. Farenhorst and B. G. Knols, "A novel method for standardized application of fungal spore coatings for mosquito exposure bioassays," *Malaria Journal*, vol. 9, no. 1, article 27, 2010.
- [52] L. L. Mnyone, M. J. Kirby, D. W. Lwetoijera et al., "Tools for delivering entomopathogenic fungi to malaria mosquitoes: effects of delivery surfaces on fungal efficacy and persistence," *Malaria Journal*, vol. 9, no. 1, article 246, 2010.
- [53] S. N. Ondiaka, S. T. Bukhari, M. Farenhorst, W. Takken, and B. G. J. Knols, "Effects of fungal infection on the host-seeking behaviour and fecundity of the malaria mosquito Anopheles gambiae Giles," in Proceedings of the Netherlands Entomological Society Meeting, vol. 19, pp. 121–128, 2008.
- [54] A. O. Lea, H. Briegel, and H. M. Lea, "Arrest, resorption, or maturation of oocytes in *Aedes aegypti*: dependence on the quantity of blood and the interval between blood meals," *Physiological Entomology*, vol. 3, no. 4, pp. 309–316, 1978.
- [55] B. D. Roitberg and I. Gordon, "Does the Anopheles blood meal—fecundity curve, curve?" *Journal of Vector Ecology*, vol. 30, no. 1, pp. 83–86, 2005.
- [56] A. S. Bell, S. Blanford, N. Jenkins, M. B. Thomas, and A. F. Read, "Real-time quantitative PCR for analysis of candidate fungal biopesticides against malaria: technique validation and first applications," *Journal of Invertebrate Pathology*, vol. 100, no. 3, pp. 160–168, 2009.
- [57] I. N. Lyimo and H. M. Ferguson, "Ecological and evolutionary determinants of host species choice in mosquito vectors," *Trends in Parasitology*, vol. 25, no. 4, pp. 189–196, 2009.
- [58] L. C. Harrington, J. D. Edman, and T. W. Scott, "Why do female *Aedes aegypti* (Diptera: Culicidae) feed preferentially and frequently on human blood?" *Journal of Medical Entomol*ogy, vol. 38, no. 3, pp. 411–422, 2001.
- [59] I. N. Lyimo, S. P. Keegan, L. C. Ranford-Cartwright, and H. M. Ferguson, "The impact of uniform and mixed species blood meals on the fitness of the mosquito vector *Anopheles gambiae* s.s: does a specialist pay for diversifying its host species diet?" *Journal of Evolutionary Biology*, vol. 25, no. 3, pp. 452–460, 2012.

Journal of Parasitology Research

- [60] G. P. Kaaya, M. Samish, M. Hedimbi, G. Gindin, and I. Glazer, "Control of tick populations by spraying *Metarhizium anisopliae* conidia on cattle under field conditions," *Experimental and Applied Acarology*, vol. 55, no. 3, pp. 273–281, 2011.
- [61] P. Polar, D. Moore, M. T. K. Kairo, and A. Ramsubhag, "Topically applied myco-acaricides for the control of cattle ticks: overcoming the challenges," *Experimental and Applied Acarology*, vol. 46, no. 1, pp. 119–148, 2008.



BioMed Research International

Zoology





Hindawi

Submit your manuscripts at http://www.hindawi.com





International Journal of Genomics





The Scientific World Journal



Journal of Signal Transduction

Genetics Research International



Anatomy Research International



International Journal of Microbiology



Biochemistry Research International



Advances in Bioinformatics







International Journal of Evolutionary Biology



Molecular Biology International



Journal of Marine Biology