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Allozyme variation and the taxonomy of Wolffiella

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Abstract

Allozyme electrophoresis was carried out to estimate genetic diversity within and assess divergence between the 10 recognized species in three sections of the aquatic angiosperm genus Wolffiella. Eleven presumptive loci were used in the calculations. Highest variation was found in W. lingulata and W. oblonga, two common species with widespread distributions in North and South America. Four of the species showing low allozyme variation include W. caudata, W. denticulata, W. neotropica, and W. rotunda, all of which have restricted distributions. W. hyalina exhibits low allozyme diversity despite being widely distributed in Africa. Three species with intermediate levels of diversity include: W. welwitschii, which is widely distributed on two continents; W. gladiata, which occurs widely in North America; and W. repanda, which has a restricted distribution in Africa. Genetic identities between species of Wolffiella vary from 0.00 (no alleles in common) to over 0.94. W. lingulata and W. oblonga share the highest identity of any two species. These two species are viewed as most closely related and are difficult to distinguish in some instances. Species within the large sect. Wolffiella (incl. W. caudata, W. denticulata, W. gladiata, W. lingulata, W. neotropica and W. welwitschii) have identities ranging from 0.00 to 0.940, whereas identities with species in this section and the two species of sect. Stipitatae (incl. W. hyalina and W. repanda) are mostly 0.000, and the same applies for W. rotunda, the only species in sect. Rotundae. The two species of sect. Stipitatae, W. hyalina and W. repanda, have an identity of 0.800, which is higher than they share with any other species. Species of sect. Stipitatae have higher identities with W. rotunda (0.538, 0.504) than they do with any species of sect. Wolffiella, and W. rotunda is more closely related to sect. Stipitatae than to sect. Wolffiella.

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Allozyme data support the recognition of sect. *Stipitatae* as now constituted and provide evidence for the circumscription of sect. *Wolffiella* as presently recognized. However, *W. denticulata* is rather isolated within this section. © 1997 Elsevier Science B.V.

Keywords: Lemnaceae; Wolffiella; Allozymes; Genetic identity

1. Introduction

The genus Wolffiella (Hegelm.) Hegelmaier is a member of the Lemnaceae or duckweed family. Landolt (1986) recognized nine species in three sections, and later (Landolt, 1992) described an additional species. The sections and species are: sect. Stipitatae containing W. hyalina (Del.) Monod and W. repanda (Hegelm.) Monod; sect. Rotundae with W. rotunda Landolt; and sect. Wolffiella consisting of W. caudata Landolt, W. denticulata (Hegelm.) Hegelm., W. gladiata (Hegelm.) Hegelm., W. lingulata (Hegelm.) Hegelm., W. neotropica Landolt, W. oblonga (Phil.) Hegelm., and W. welwitschii (Hegelm.) Monod. A recent cladistic analysis based on morphological, micromolecular and anatomical features suggests that the sections of Wolffiella represent monophyletic groups, but the genus Wolffiella is paraphyletic. However, constraining the trees to make the genus monophyletic adds only a few steps to their length (Les et al., 1997).

The present electrophoretic study of Wolffiella was undertaken to ascertain whether allozymic divergence among the species is concordant with their taxonomic disposition in different sections. Also, we wished to see if species within sections viewed as most closely related on the basis of morphology (Landolt, 1986, 1992) show the highest allozymic similarity. Prior electrophoretic studies of two other duckweed genera, Spirodela (Crawford and Landolt, 1993) and Wolffia (Crawford and Landolt, 1995), indicate that species considered most closely related may show genetic identities lower than 0.50. In many pair-wise species comparisons in these two genera, no alleles were shared at any of the loci examined, and thus the data were not useful for assessing relationships other than to indicate that the taxa are much more divergent allozymically than most congeneric species of flowering plants (Gottlieb, 1977; Crawford, 1990). A secondary objective was to assess genetic variation within each species.

2. Methods

A total of 79 clones (strains) representing all ten recognized species of Wolffiella was included in the electrophoretic survey. Because the strains represent single isolates, no assessment of diversity within populations was attempted. The clones studied and the localities of origin are given in Table 1. Clones were selected so that species could be sampled from most of their geographic ranges. The smaller number of samples examined for certain species such as W. caudata, W. denticulata, W. repanda, and W. rotunda is a reflection of their rarity and/or restricted geographic distributions. Plant material, either from agar or liquid culture, was supplied and identified taxonomically by

E.L. The grinding buffer was made up of 10% glycerol and was 0.1 M tris-HCl, pH 7.5, with 14 mM 2 mercaptoethanol, 1.0 mM tetrasodium salt of EDTA, 10 mM MgCl₂, 10 mM KCl, and 5-10 mg polyvinylpolypyrrolidine per 0.5 ml of buffer (Gottlieb, 1981). Several enzymes were separated in polyacrylamide gels according to the methods of Crawford et al. (1987): alcohol dehydrogenase (ADH, E.C. 1.1.1.1); glutamate dehydrogenase (GDH, E.C. 1.4.1.2); and phosphogluconate dehydrogenase (PGD, E.C. 1.1.1.44). The remaining enzymes were resolved in 12.5% starch gels using two buffer systems. Forms of malate dehydrogenase (MDH, E.C. 1.1.1.37) were separated with an electrode buffer of 0.04 M citric acid adjusted to pH 6.1 with N-(3-aminopropyl)-morpholine, and the gel buffer was a 1:19 dilution of the electrode buffer. Forms of glucose-6-phosphate isomerase (GPI, E.C. 5.3.1.1) and triosephosphate isomerase (TPI, E.C. 5.3.1.1) were resolved with an electrode buffer of 0.5 M tris, 0.65 M boric acid, 0.02 M EDTA, pH 8.0, and a 1:9 dilution of this was used for the gel buffer. Staining protocols and nomenclature for all enzymes followed Wendel and Weeden (1989). Several lines of evidence were used to infer the genetic bases of the banding patterns for the enzymes. One useful source of data was the known active subunit composition of the enzymes (Weeden and Wendel, 1989). Additional information included variation seen in banding patterns between clones of the same and/or different taxa, and the expected minimal conserved number of isozymes for diploid plants (Gottlieb, 1982; Weeden and Wendel, 1989). Allelic frequencies were determined for each species and were used to calculate Nei's genetic identity and distance (Nei, 1972). The GeneStat-PC (version 3.3) software (Lewis, 1993) was employed to calculate the statistics. An unweighted pair-group method using arithmetic averages (UPGMA) cluster analysis of genetic identity values was performed using version 1.70 NTSYS-pc (Rolf, 1992). Mean number of alleles per locus and per polymorphic locus, and proportion polymorphic loci were also calculated for each species.

3. Results

Eleven presumptive loci were used to calculate the statistics for the 10 species of Wolffiella, Adh-1, Adh-2, Gdh, Gpi-1, Gpi-2, Mdh-1, Mdh-2, Pgd-1, Pgd-2, Tpi-1, and Tpi-2. Not all loci were scored for every strain because of poor staining and/or resolution.

The mean numbers of alleles per locus and polymorphic locus, and the proportion polymorphic loci are shown in Table 2. No variation was detected between the two clones of W. denticulata. The mean numbers of alleles per locus and per polymorphic locus are highest in W. oblonga and W. lingulata with W. gladiata and W. welwitschii next highest. The five species W. caudata, W. hyalina, W. neotropica, W. repanda and W. rotunda show similarly low values for mean numbers of alleles per locus and per polymorphic locus. Proportion polymorphic loci is highest in W. oblonga, followed by W. lingulata; W. repanda and W. welwitschii exhibit similar proportions of polymorphic loci (Table 2). The same four species with low mean numbers of alleles per locus (W. caudata, W. hyalina, W. neotropica and W. rotunda) also have a low proportion polymorphic loci (Table 2).

Table 1 Collection numbers and geographic origins of Wolffiella clones used for enzyme electrophoresis

Species	Collection number ^a		Geographic origin
W. caudata	9155	Bolivia:	Beni, La Pascane Grande
	9158	Bolivia:	Beni, La Pascane Grande
	9165	Bolivia:	Beni, Rurrenabaques
	9173	Bolivia:	Beni, San Pablo
W. denticulata	7454	South Africa:	Natal, Zululand
	8221	South Africa:	Natal, Sordwana Bay
W. gladiata	7173	USA:	Washington, Tacoma
	7590	USA:	Virginia, Dymer Creek
	7595	USA:	Virginia, Brandon
	7852	USA:	Louisiana, East Baton Rouge Parish
	8066	USA:	Texas, Old Ocean
	8261	USA:	Pennsylvania, Conneaut Lake
	8350	USA:	Illinois, Pine Hills Swamp
	8392	USA:	Texas, Austin
	8768	USA:	Florida, Tallahassee
W. hyalina	7376	Egypt:	Mahallet, El Qubba
	7378	Egypt:	Hafr Shoukr. Naim
	8640	Tanzania:	Arusha: Amboseli
W. lingulata	7289	Brazil:	Amazonas, Neptunia
	7292	Brazil:	Amazonas, Rio Negro
	7330	Trinidad:	St. Augustine
	7360	Surinam:	Saramacca River
	7464	Venezuela:	Yaracuy, Marlin
	7655	Mexico:	Tabasco, Villahermosa
	7725	Argentina:	Corrientes, Mburucuya
	8041	USA:	Louisiana, Pecan Island
	8141	USA:	California, Vandenberg AFB
	8175	USA:	California, Lake Thynan
	8237		Asuncion
	8776	Paraguay: USA:	
			California, Black Lake Canyon
	8823b	Argentina:	Formosa, Clorinda
	8664	Argentina:	Corrienfes, Empedrado
	8886	USA:	California, Monterey Co.
	8898a 9133	Ecuador: Brazil:	Guayaquil Matto Grosso, Corrumba
W. neotropica	7225	Brazil:	Guanabara, Rio de Janeiro
	7279	Brazil:	Rio de Janeiro, Cabo Frio
	7290	Brazil:	Amazonas, Neptunia
	7609	Brazil:	Espirito-Santo, Heliofila
	8848	Brazil:	Rio de Janeiro, Marico
	8849	Brazil:	Rio de Janeiro, Saquarema
W. oblonga	7164	USA:	Louisiana, New Orleans
G	7167	USA:	Louisiana, Norco
	7201	Argentina:	Buenos Aires, Arroyo Burgueño
	7569	Brazil:	Sao Paulo
	7732	Brazil:	Sao Paulo
	7853	USA:	Louisiana, East Baton Rouge Parish

Table 1 (continued)

Species	Collection number ^a		Geographic origin		
W. oblonga	7855	USA:	Louisiana, St. James		
	7923	Argentina:	Buenos Aires, Arroyo Vitel		
	7997	Brazil:	Rio Grande de Sul, Pelotas		
	8031	USA:	Louisiana, Rapides Parish		
	8072	USA:	Texas, Old Ocean		
	8393	USA:	Florida, Immokalee		
	8751	Argentina:	Salta, El Rey		
	8777	USA:	California, Black Lake Canyon		
	8816	Argentina:	Santa Fé, Esperanza		
	8828	Argentina:	Formosa, Clorinda		
	8881b	USA:	California, Black Lake Canyon		
	8984	Columbia:	Cundinamarca, Laguna La Herrera		
	9139	Brazil:	Amazonas, Manaus		
	9140	Chile:	Quillon, Laguna, Allendaño		
	9141	Chile:	Quillon, Laguna Allendaño		
W. repanda	9055	Zimbabwe:	Urungwe Safari Area, Chirundu		
	9062	Zimbabwe:	Urungwe Safari Area, Chirundu		
	9104	Botswana:	South Gate to Moremi		
	9107	Botswana:	85 km NNE of Shorobe		
	9116	Zimbabwe:	Urungwe Safari Area, Chirundu		
	9122	Zimbabwe:	Urungwe Safari Area, Chirundu		
W. rotunda	9048	Zimbabwe:	Urungwe Safari Area, Chirundu		
	9054	Zimbabwe:	Urungwe Safari Area, Chirundu		
	9072	Zimbabwe:	Mana Pools		
	9121	Zimbabwe:	Urungwe Safari Area, Chirundu		
W. welwitschii	7468	Columbia:	Atlantico, Barranquilla		
	7644	Angola:	Benguela, Cubal		
	8863	Senegal:	between Saint Louis and Richard Toll		
	9086	Botswana:	Daonara, Santantadibe		
	9089	Botswana:	Boteti River		
	9093	Botswana:	Moremi Wildlife Reserve		
	9096	Botswana:	Chobe River, Chubu Lodge		

^a Collection numbers those of Landolt.

Table 2 Number of clones examined and allozymic variation in species of Wolffiella

Species	Clones	Alleles per locus (Mean no.)	Alleles per polymorphic locus (Mean no.)	Proportion poly- morphic loci	
W. caudata 4 1.18		1.18	2.00	0.18	
W. denticulata	2	1.00		0.00	
W. gladiata	9	1.46	2.25	0.36	
W. hyalina	3	1.10	2.00	0.10	
W. lingulata	17	1.64	2.40	0.46 0.09	
W. neotropica	6	1.09	2.00		
W. oblonga	21	2.00	2.83	0.55	
W. repanda	la 6 1.20		2.00	0.32	
W. rotunda	4	1.10	2.00	0.10	
W. welwitschii	7	1.44	2.33	0.33	

Table 3	
Nei's genetic identity between species of Wolffiella. Species designations are the	first three letters of names
given in Table 1	

Sect. Wolffiella						sect. Stipitatae		sect. Rotundae		
Species	cau	den	gla	lin	neo	obl	wel	hya	rep	rot
Nei's ger	netic iden	tity betwe	een pairs	of specie	s					
cau	X									
den	0.000	X								
gla	0.438	0.000	X							
lin	0.467	0.000	0.816	X						
neo	0.113	0.096	0.226	0.116	X					
obl	0.461	0.000	0.845	0.940	0.131	X				
wel	0.338	0.142	0.426	0.430	0.316	0.443	X			
hya	0.000	0.000	0.000	0.000	0.012	0.000	0.000	X		
rep	0.000	0.000	0.000	0.000	0.013	0.000	0.000	0.800	X	
rot	0.000	0.000	0.000	0.000	0.012	0.000	0.000	0.504	0.538	X

The pair-wise comparisons of genetic identities between species are shown in Table 3, and the dendrogram produced from UPGMA clustering of these identities is shown in Fig. 1. The values range from 0.00 in 22 pair-wise comparisons to 0.940 for W. lingulata and W. oblonga; these two species also share high identities with W. gladiata (Table 3, Fig. 1). Within sect. Wolffiella identities vary from the high values for the three aforementioned species down to 0.000 (Table 3). The mean identity value for all species in sect. Wolffiella is 0.32. W. hyalina and W. repanda of sect. Stipitatae have an identity of 0.800, which is higher than they share with species in other sections (Table 3, Fig. 1). The highest intersectional identity mean (0.521) occurs between sect. Stipitatae and sect. Rotundae, the latter section consisting only of W. rotunda (Fig. 1). By contrast, these two sections have a mean identity of only 0.001 with sect. Wolffiella; except for W. neotropica, no species of sect. Wolffiella shares any alleles with species in the other two sections (Table 3, Fig. 1).

The two species W. lingulata and W. oblonga, which are widespread geographically

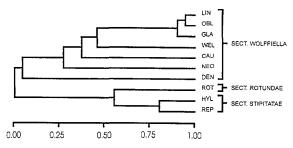


Fig. 1. Unweighted pair-group method using arithmetic averages (UPGMA) dendrogram showing clustering of Nei's genetic identities (shown along bottom) among the ten *Wolffiella* species (abbreviations same as in Table 3). Integrity of taxonomic sections is retained, although several species of sect. *Wolffiella* show very low identities.

and are distributed in both North and South America, also show a particular distribution of alleles at Gpi-1. In W. lingulata, the allele f is found only in South American populations, whereas the two North American populations have allele g and one population from Mexico has both alleles. In W. oblonga, six of the nine clones from North America have allele Gpi-1 f and the three other strains have Gpi-1 g. Eight of the 10 clones from South America have Gpi-1 g, two have Gpi-1 f and another is heterozygous for the latter two alleles.

4. Discussion

Numerous studies have examined genetic diversity within a wide taxonomic sample of flowering plants with various life history attributes, and many of these results have been summarized by Hamrick and Godt (1990). Results from the present study may be compared to the compilations of Hamrick and Godt (1990) as well as the species of two other genera of Lemnaceae (Crawford and Landolt, 1993, 1995). The two most widely distributed and common species of Wolffiella are W. lingulata and W. oblonga, and they are the two most variable species allozymically (Table 2). However, the mean numbers of alleles per locus (2.29 versus 2.00 and 1.64) and proportion polymorphic loci (0.59 versus 0.55 and 0.46) for these two taxa are lower than in other widespread species (Hamrick and Godt, 1990). The one species in which no variation was detected (W. denticulata) has one of the smallest geographic distributions of any species of Wolffiella (Landolt, 1986, 1992), but it must be emphasized that only two clones from neighboring localities were examined. Three species with low allozyme variation, W. caudata, W. neotropica, and W. rotunda, have restricted distributions on single continents. W. hyalina exhibits low diversity but is widely distributed in Africa (Landolt, 1986). Two species with 'intermediate' levels of diversity, W. gladiata and W. welwitschii, are widely distributed, with the former restricted to North America and the latter present over wide areas in Africa as well as in South America, Central America and in the Caribbean (Landolt, 1986). Another species with 'intermediate' diversity is W. repanda, which is narrowly distributed in Africa. Thus, in general, more widely distributed and common species of Wolffiella have higher allozyme diversity compared to more restricted taxa.

The lower allozyme variation detected in the rarer species is not an artifact of smaller number of clones sampled. When subsamples of clones of the two most common species, W. lingulata and W. oblonga, were selected randomly and the values calculated for mean number of alleles per locus, per polymorphic locus and proportion polymorphic loci, the values are much higher than those found for the same number of clones of the rare species. This is particularly true when the clones of W. lingulata and W. oblonga originate from different continents (North and South America); in some instances the variation is actually higher than when all clones of each species are included in the calculations.

Diversity in species in two other genera of duckweeds, *Spirodela* and *Wolffia* (Crawford and Landolt, 1993, 1995), are compared with *Wolffiella* in Table 4. The genera have very similar mean levels of variation for each of the measures (Table 4).

Wolffiella

one have been examined					
Genus Number of specinvestigated		Mean (and range) number of alleles per locus	Mean (and range) number of alleles per polymorphic locus	Mean (and range) proportion poly- morphic loci	
Spirodela	3	1.41 (1.13–1.63)	2.17 (2.00-2.25)	0.38 (0.13-0.50)	
Wolffia	10 a	1.40 (1.07-2.29)	2.28 (2.00-3.00)	0.30 (0.07-0.79)	

2.20 (2.00-2.83)

0.25(0.00-0.50)

1.32 (1.00-2.00)

Table 4
Means of genetic variation compared for species in the genera of Lemnaceae; all known extant species except one have been examined

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The range of values is lower in *Spirodela* than in the other two genera for all measures of variation; this may be a reflection of the fact that there are only three species in this genus as compared to 10 investigated in Wolffia and 10 in Wolffiella. Within Spirodela, the most widespread species, S. polyrrhiza, is the least diverse whereas the most restricted species geographically (S. intermedia) has the highest level of genetic variation. In this genus, diversity is correlated with greater frequency of flowering and seed set, and not with geographic range (Crawford and Landolt, 1993). In Wolffia, the three most allozymically variable species (W. arrhiza, W. columbiana and W. globosa) are also the most geographically widespread taxa with distributions on at least two continents (Landolt, 1994; Crawford and Landolt, 1995). All species of Wolffia, except W. microscopica, flower with similar frequencies. Therefore, in Wolffia geographic distribution is associated with the level of allozyme diversity within species. In Wolffiella, flowering frequency is not consistently correlated with allozyme diversity. For example, W. hyalina, W. repanda and W. rotunda are the three species with by far the highest percentage of flowering of any in the genus (Table 5). Yet, W. hyalina and W. rotunda have very low allozyme diversity and W. repanda has just an average diversity (Table 2), although sampling of additional clones of W. hyalina may have revealed higher diversity. By contrast, the three most allozymically diverse species of

Table 5
Percentage of flowering in samples of Wolffiella species

Species	Number of samples investigated	Flowering percentage of Wolffiella species in nature (from herbarium specimens and field observations of EL)		
W. caudata	7	0		
W. denticulata	9	11		
W. gladiata	250	4		
W. hyalina	62	32		
W. lingulata	236	10		
W. neotropica	10	10		
W. oblonga	266	5		
W. repanda	10	40		
W. rotunda	16	56		
W. welwitschii	124	9		

^a W. elongata was not studied.

Wolffiella (W. gladiata, W. lingulata and W. oblonga) have very low flowering frequency (Table 5). Thus, it appears that in Spirodela the frequency of flowering is correlated with higher diversity rather than geographic distribution, whereas in Wolffia and Wolffiella it does not appear that flowering frequency is correlated with (and ostensibly influences) allozyme variation.

Previous studies of allozyme divergence between congeneric species of flowering plants have revealed a wide range of genetic identities, but a mean identity value between 0.65 and 0.70 is common (Gottlieb, 1977; Crawford, 1989, 1990). Given the reduced morphology of Lemnaceae, both in size and number of structures, there are fewer characters to compare than in other terrestrial flowering plants. The taxonomic difficulty in duckweeds could be the result of extreme parallel reduction, or the similarity may reflect close relationships. In the other duckweed genera, Spirodela and Wolffia, all pair-wise species comparisons revealed very low genetic identities between many species (Crawford and Landolt, 1993, 1995). For example, in Spirodela two species share no alleles and the other species pair has an identity of only 0.40. In Wolffia, the highest identity between two species is 0.40, and 37 of the 45 pair-wise species comparisons are 0.00, that is, with no alleles in common (Crawford and Landolt, 1995). The results for Wolffiella are similar in certain respects to the other two genera because several species (22 of the 45 pair-wise comparisons) share no alleles at the loci examined (Table 3). Wolffiella differs from the other two genera, however, because the three species W. gladiata, W. lingulata and W. oblonga have identities of 0.816 or higher, and W. hyalina and W. repanda share an identity of 0.800 (Table 3). The former three taxa are viewed as closely related (Landolt, 1986); W. lingulata and W. oblonga are particularly difficult to distinguish morphologically and the two have the very high identity of 0.940 (Table 3), which is comparable to values often obtained for populations of the same species (Gottlieb, 1977; Crawford, 1989, 1990). W. hyalina and W. repanda are the only two members of sect. Stipitatae and their identity of 0.800 (Table 3) is nearly twice as high as found between any species in either of the other two genera of Lemnaceae. The high identities for the species of Wolffiella could be attributed to a more recent divergence time and/or hybridization. W. gladiata is quite distinct from W. lingulata and W. oblonga, the two other species with which it shares a high identity. In addition, W. gladiata differs from the other two species ecologically and in geographic distribution; it grows in North America in warm temperate regions and overlaps with the other two species only in the very southern United States and in the high plateau of Mexico. W. lingulata and W. oblonga, which share the highest identity of any two species, are not as well differentiated morphologically as each is from W. gladiata. There are some ecological differences, however, with W. oblonga more tolerant of lower temperatures. Therefore, it occurs at higher altitudes in the mountains of South America and also farther south than W. lingulata. Unlike W. lingulata, W. oblonga is very rare in warm tropical regions (Landolt, 1986).

To consider whether hybridization or lack of divergence may be the primary factors in producing similarity at allozyme loci, three clones of each species from outside the geographic range of the other species were compared. These include clones 7289, 7330 and 7360 of W. lingulata, and 7201, 7923, 9140 of W. oblonga (Table 1). Presumably interspecific hybridization would not be a cause of similarity at allozyme loci in these

regions and if consistent differences exist between 'pure' strains of each species they should be seen when comparing these allopatric clones. This is not the case, however, because the same allelic variation at certain loci occurs between clones of the same species in these areas of allopatry, and thus genetic identities between strains of the two species are just as high between the areas of allopatry as they are for those from sympatric areas. It does not appear, therefore, that allozymically 'pure' clones of each species occur in the sense that particular alleles are restricted to one species or the other. On present evidence, it is not possible to determine whether these taxa represent one variable gene pool or distinct species that my hybridize when they come in contact, but the data cast some doubt on the existence of two separate gene pools. However, despite the allozyme evidence one of us (EL) is consistently able (albeit with difficulty) to place clones into one of the species on the basis of morphology. This situation appears similar to Lemna minima and L. valdiviana, two morphologically similar (nearly indistinguishable) species with a genetic identity of 0.70 at allozyme loci (Crawford et al., 1996). In Wolffiella, those species with very low genetic identities presumably represent taxa of ancient divergence, and this includes the majority of recognized species in the genus. At the same time, there are other recognized species with very high identities, and these are likely either recently diverged taxa, taxa with distinct gene pools but with occasional hybridization that effectively homogenizes allelic frequencies, or the two 'taxa' are in reality minor morphological variants of a single species. Additional studies are needed to elucidate the situation. An important point is that morphological similarity may or may not indicate high similarity at allozyme loci.

The systematics of the subfamily Wolffioideae was treated rather differently by various authors within the last 150 yrs. Until Hegelmaier (1868) all species of this group were incorporated into the genus Wolffia. Hegelmaier (1868) created a subgenus Wolffiella with W. denticulata, W. gladiata, W. lingulata and W. oblonga. The main distinguishing character was the asymmetry of the four species. All the other known species with a symmetrical appearance (including W. hyalina, W. repanda and W. welwitschii) he left with the other species of Wolffia. Later Hegelmaier (1895) upgraded the subgenus to a genus. Monod (1949) placed all species with flat fronds in the genus Wolffiella and the genus comprised in this way also included the three mentioned symmetrical species. In his monograph, Daubs (1965) kept only W. welwitschii within the genus Wolffiella. W. hyalina and W. repanda were transferred to Wolffia again. Finally, Den Hartog and Van der Plas (1970) placed W. hyalina and W. repanda in the separate genus Pseudowolffia, and W. welwitschii in the genus Wolffiopsis. Wolffiopsis was characterized by two flowers per frond in contrast to only one for the other species, and Pseudowolffia was distinguished by the labellum. Landolt (1986), having detected two new species with transitional characters between the three genera of Den Hartog and Van der Plas (1970), included all these species again in the genus Wolffiella creating three sections; sect. Stipitatae with W. hyalina and W. repanda; sect. Rotundae with W. rotunda; and sect. Wolffiella with the rest of the species.

The allozyme data provide some support for the morphological affinities of the species. They show that the genus *Wolffiella* can be divided into two groups, which have identities near zero (Fig. 1, Table 3), one group with the three species *W. hyalina*, *W. repanda* and *W. rotunda*, and one with the rest of the species. Whether these two groups

correspond to two genera, subgenera or sections is a matter of opinion and no decision should be made until DNA sequence data have been analyzed. Allozyme data place some doubt on the justification of placing W. rotunda in its own section separate from W. hyalina and W. repanda, though the lack of a labellum is a very conspicuous characteristic. The three species have in common that they flower frequently, surviving the dry period in the form of seeds. On the other hand, within the large sect. Wolffiella all species except W. denticulata have some level of allozyme similarity (Fig. 1, Table 3). The isolated position of W. denticulata within sect. Wolffiella seems remarkable; it shares only low identities with W. welwitschii and W. neotropica (0.142 and 0.096). Additional investigations are needed to determine if the species is best placed in a separate section. The allozyme data confirm the central position of W. neotropica within the genus; it is the only species having some identity with each of the other species. It has the morphological characters of both the two groups. In addition, it is the only species of Wolffiella which can grow, depending on conditions, either submerged or partly submerged like the species of the sect. Wolffiella or floating entirely on the surface as W. hyalina, W. repanda and W. rotunda.

References

Crawford, D.J., 1989. Enzyme electrophoresis and plant systematics. In Soltis, D.E., Soltis, P.S. (Eds.), Isozymes in Plant Biology. Dioscorides Press, Portland, OR, pp. 146-164.

Crawford, D.J., 1990. Plant Molecular Systematics: Macromolecular Approaches. John Wiley, New York.

Crawford, D.J., Stuessy, T.F., Silva, O.M., 1987. Allozyme divergence and the evolution of *Dendroseris* (Compositae: Lactuceae) on the Juan Fernandez Islands. Syst. Bot. 12, 435-443.

Crawford, D.J., Landolt, E., 1993. Allozyme studies in Spirodela (Lemnaceae): variation among conspecific clones and divergence among the species. Syst. Bot. 18, 389-394.

Crawford, D.J., Landolt, E., 1995. Allozyme divergence among species of Wolffia (Lemnaceae). Pl. Syst. Evol. 197, 59-70.

Crawford, D.J., Landolt, E., Les, D., 1996. An allozyme study of two sibling species of *Lemna* (Lemnaceae) with comments on their morphology, ecology and distribution. Bull. Torrey Bot. Club 123, 1–6.

Daubs, E.H., 1965. A monograph of Lemnaceae. Illinois Biol. Monogr. 34, 1-118.

Den Hartog, C., Van der Plas, F., 1970. A synopsis of the Lemnaceae. Blumea 18, 355-368.

Gottlieb, L.D., 1977. Electrophoretic evidence and plant systematics. Ann. Missouri Bot. Gard. 64, 161-180.

Gottlieb, L.D., 1981. Gene number of species of Astereae that have different chromosome number. Proc. Natl. Acad. Sci. USA 78, 3726–3729.

Gottlieb, L.D., 1982. Conservation and duplication of isozymes in plants. Science 216, 373-380.

Hamrick, J.L., Godt, M.J.W., 1990. Allozyme diversity in plant species. In Brown, A.H.D. et al. (Eds.), Plant Population Genetics, Breeding, and Genetic Resources. Sinauer, Sunderland, MA, pp. 43-63.

Hegelmaier, F., 1868. Die Lemnaceen. Eine monographische Untersuchung. Engelmann, Leipzig.

Hegelmaier, F., 1895. Systematische Uebersicht der Lemnaceen. Bot. Jb. 21, 268-305.

Landolt, E., 1986. The family of Lemnaceae – a monographic study. 1. Veröff. Geobot. Inst. ETH, Stiftung Rübel, Zürich. 71, 1–566.

Landolt, E., 1992. Wolffiella caudata, a new Lemnaceae species from the Bolivian Amazon region. Ber. Geobot. Inst. ETH, Stiftung Rübel, Zürich 58, 121-123.

Landolt, E., 1994. The Lemnaceae of Zimbabwe and Botswana. Ber. Geobot. Inst. ETH, Stiftung Rübel, Zürich 60, 110-136.

Les, D.H., Landolt, E., Crawford, D.J., 1997. Systematics of the Lemnaceae (duckweeds): inferences from micromolecular and morphological data. Pl. Syst. Evol. 204, 161-177.

Lewis, P., 1993. GeneStat-PC3.3. Department of Statistics, North Carolina State University, Raleigh, NC.

Monod, T., 1949. Sur une Lemnaceae africaine: Wolffiella Welwitschii (Hegelmaier, 1865) com. nov. Mém. Soc. Hist. Nat. Afr. Nord. 2, 229-242.

Nei, M., 1972. Genetic distance between populations. Am. Naturalist 106, 283-293.

Rolf, F.J., 1992. NTSYS-pc numerical taxonomy and multivariate analysis system, version, Exeter, New York, NY.

Weeden, N.F., Wendel, J.F., 1989. Genetics of plant isozymes. In: Soltis, D.E., Soltis, P.S. (Eds.), Isozymes in Plant Biology. Dioscorides Press, Portland, OR, pp. 46–72.

Wendel, J.F., Weeden, N.F., 1989. Visualization and interpretation of plant isozymes. In: Soltis, D.E., Soltis, P.S. (Eds.), Isozymes in Plant Biology. Dioscorides Press, Portland, OR, pp. 5-45.