



Seroepidemiological Studies on Poultry Salmonellosis and its Public Health Importance

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ABSTRACT

Non-typhoid *Salmonella* serovars remain a potential threat to human health, and poultry species are possible sources of these organisms. In this study, trials for *Salmonella* isolation from poultry and humans were conducted in the period April 2009 through March 2010 in Beni-Suef Governorate, Egypt. Cloacal swabs were collected from different live poultry species including 150 broilers, 50 breeders, 50 layers, 50 turkeys, and 50 ducks, beside 30 litter samples from various poultry farms. Regarding the humans, stool samples as well as hand swabs were collected from 90 workers and poultry contacts of the examined farms. All poultry and human samples were subjected to bacteriological examination and serological identification for *Salmonella* spp. The recovered *Salmonella* strains were found belonging to *S. Kentucky*, *S. Typhimurium* and *S. SaintPaul*. The obtained results demonstrated that the occurrence of *Salmonella* spp. accounted for 16.66, 10.0, 2.0, 6.0 and 2.0% in broilers, breeders, layers, ducks and turkeys respectively. Investigation of litter samples revealed that the occurrence of *S. Kentucky* was 53.33, 66.66 and 28.57% in broiler's, breeder's and duck's litters respectively. Examination of human samples declared that 8 out of 90 hand swabs were found positive for *S. Kentucky* whereas all stool samples reacted negatively to all *Salmonella* spp. In the present study, conclusively *Salmonella* serovars (*S. Kentucky*) isolated from chickens were frequently isolated from hand swabs of the examined poultry contacts, this provided evidence that direct contact with poultry or poultry environment may pose health hazards for humans.

Key words: *Salmonella* spp.; Poultry; Humans; Litter

INTRODUCTION

Salmonellosis is considered to be one of the major bacterial disease problems in the poultry industry world-wide. *Salmonella* spp. cause asymptomatic intestinal infections in birds but acute outbreaks exhibiting clinical disease along with high levels of mortality occur in chicks younger than 2 weeks old (Duchet-Suchaux et al., 1995). In laying hens *Salmonella* can be highly invasive leading to systemic infections that can potentially be deposited in the internal contents of eggs by trans ovarian transmission following colonization of the intestinal tract (Woodward et al., 2005). Due to its endemic nature, high morbidity and association with a wide range of foods, salmonellosis is of high public health concern (Aarestrup et al., 2007 and Kottwitz et al., 2008).

There are more than 2500 serovars of *Salmonella enterica* that have been identified. The majority of human cases of non-typhoidal salmonellosis are caused by a limited number of serovars, which may vary from country to country and over time (Hendriksen et al., 2011). *S. Enteritidis* and *S. Typhimurium* have traditionally been the most frequently isolated serovars

from humans worldwide (Fashae et al., 2010 and Hendriksen et al., 2011). However, other serovars have been reported to be more prevalent in specific regions or within countries.

Salmonella spp. in humans, are primarily a cause of self-limiting acute enteritis (diarrhea, abdominal pain, and fever, with a typical duration of 4–7 days). However, invasive *Salmonella* spp. can spread beyond the gastrointestinal mucosa to infect other sites such as the blood stream, the meninges, bone or joint spaces (Crump et al., 2011).

People generally acquire salmonellosis through food-borne exposure, although direct contact with infected animals is another possible route (Mead et al., 1999 and L Plym and Wierup, 2006). A variety of investigations of outbreaks and sporadic cases have indicated that food vehicles identified as the most common source of *Salmonella* infections in humans are poultry and poultry products, including raw and uncooked eggs (Hennessy et al., 2004). The objective of the present work was to determine the zoonotic potential of *Salmonella* strains prevalent among various

different poultry species (chickens, ducks and turkeys) and their human contacts.

MATERIALS AND METHODS

Sample collection

The study was conducted in Beni-Suef Governorate, Egypt (30°13' N, 31°40' E), situated 120 kilometers south to Cairo, Egypt with an average altitude of 46 meters. A total of 440 samples, including both poultry (350) and humans (90) were collected during the period from April 2009 to February 2010. The samples were investigated in the department of Hygiene, Management and Zoonoses, Faculty of Veterinary Medicine, Beni-Suef University for demonstrating the occurrence of *Salmonella* in poultry and man.

Poultry samples: Cloacal swabs were collected from different live poultry species including broilers (150), breeders (50), layers (50), turkeys (50) and ducks (50) obtained from various poultry farms. The swabs were directly immersed into sterile tubes containing Buffered Peptone Water (BPW).

Litter samples: A total of 30 litter samples were obtained from the farms of the examined broilers (15), breeders (3), layers (3), turkeys (2) and ducks (7). About 2 g. from each litter was taken and directly immersed into sterile tubes containing BPW.

Human samples: Stool samples as well as hand swabs were collected from 90 workers and poultry contacts of the examined farms. The stool samples were received in sterile plastic containers where a loopful from each sample was inoculated into a sterile tube containing BPW. The hand swabs were also inoculated into sterile tubes containing BPW.

Isolation, identification and serotyping of *Salmonella*

Salmonella cultures from all samples were performed according to Collee et al. (1996) and Waltman (1999). Briefly, samples that were collected in buffered peptone water (BPW) were taken to the laboratory on the day of collection under refrigeration with minimum of delay and incubated at 37 °C for 24 h. After pre-enrichment, 0.1 ml of the broth culture was transferred into a 10 ml Rappaport–Vasiliadis broth (RV broth, Difco, USA) and was incubated at 42 °C for 24–48 h. The RV broth samples were streaked onto Xylose-Lysine-Desoxycolate agar (XLD, oxoid) plates and incubated overnight at 37 °C. Typical colonies were picked and further tested by standard biochemical methods and serotyped using specific commercial sera according to the Kauffmann–White scheme (Kauffmann, 1974).

RESULTS AND DISCUSSION

The results illustrated in Table 1 demonstrated the occurrence of *Salmonella* spp. among different species of poultry including chickens, ducks and

turkeys. In chickens, only *S. Kentucky* could be isolated from broilers, breeders and layers at a rate of 16.66%, 10.0% and 2.0% respectively. The occurrence of *S. Kentucky* in broiler chickens in this study well in line with the reports of Byrd et al. (1997) and Byrd et al. (1999) who declared that *S. Kentucky* and *S. Heidelberg* accounted for the highest percentage of total serovars isolated from commercial broilers production facilities. However these findings are lower than those detected by Caldwell et al. (1995). Furthermore the obtained data are much higher than those reported by Wedderkopp et al. (2001), Hegazy (2002) and Abd El-Hamid et al. (2004). On the contrary in many countries all over the world as e.g. in Ghana, (Sackey et al., 2001), France, (Lahellec et al., 1986), USA, (Payne et al., 2006) and Japan, (Shahada et al., 2006) a wide range of different *Salmonella* serovars were found to contaminate the broiler houses and flocks.

The high level of *Salmonella* isolation in broilers evaluated in this study may be attributed to horizontal and/or vertical transmission of *Salmonella* to the chicks. The horizontal transmission of *Salmonellae* can be mediated by mechanisms including direct bird-to-bird contact, ingestion of contaminated feeds, water or litter, or using contaminated equipments, while vertical transmission to the progeny of infected breeder flocks can result from internal or external contamination of eggs (Gast, 2003). This investigation highlights the importance of isolation of *Salmonella* from broilers as it was reported that slaughtering broiler flocks infected with *Salmonella* can lead to contamination both of the carcasses and the slaughter line (Corry et al., 2002 and Olsen et al., 2003).

On the other hand the percentage isolation of *Salmonella* spp. from breeders (10.0%) is higher than those found by El-Shamy (1999). Even though the breeders may be infected at the farm level by any way of transmission but the birds can also get infected easily from the transporting boxes which are used for different poultry species more than one time without disinfection or from the surrounding environment. Moreover the isolation of *Salmonella* from breeders could explain the theory of vertical transmission which lead to internal or external contamination of eggs followed by infection of newly hatched chicks (Gast, 2003).

Regarding the occurrence of *Salmonella* spp. in layers, the isolation rate (2.0%) is to some extent, in agreement with the data obtained by Ramadan (1997) who demonstrated that *Salmonella* was isolated with a percentage of 2.1% from layer farms. Indeed, *Salmonella* is considered as one of the most important causes of great mortalities and morbidities among laying flocks in addition to its public health importance as it can contaminate table eggs internally or externally (Humphrey, 1998).

The percentage isolation of *Salmonella* spp. from chickens (broilers, breeders or layers) generally in this study is more or less similar to that published by several previous authors as Cardinale et al. (2004), Saad (2007), Badr and Abd El Monaem (2008) and Muhammad et al. (2010) which ranged from 1.7% to 28.6%.

The fore mentioned results in Table 1 clarified that the total occurrence of *Salmonella* spp. in ducks was accounting for 6.0%. The detected *Salmonella* spp. were *S. Typhimurium* (4.0%) and *S. SaintPaul* (2.0%). Such finding is comparable to that achieved by Mohammed (1986). However these data are lower than those recorded by Abd El-Hamid et al. (2004) and Vo et al. (2006). Consequently it was possible in this study to confirm researchers indicating that *S. Typhimurium*

was almost certainly the predominant serovar in ducks, as elsewhere, probably because in addition to horizontal transmission, it was the subject of vertical transmission by all routes (Henry, 2000).

The source of duck infection with *Salmonella* could be attributed to litter contamination, residual environmental contamination or true egg transmission occurred when the ducks' egg were infected before oviposition.

Table 1. Recovery of *Salmonella* spp. from cloacal swabs of the examined poultry species

Poultry species	Examined number	Number of samples positive for:			Total (%)
		<i>S. Kentucky</i> (%)	<i>S. Typhimurium</i> (%)	<i>S. SaintPaul</i> (%)	
Chickens	Broilers	25 (16.66)	0 (0)	0 (0)	25 (16.66)
	Breeders	5 (10.0)	0 (0)	0 (0)	5 (10.0)
	Layers	1 (2.0)	0 (0)	0 (0)	1 (2.0)
Ducks	50	0 (0)	2 (4.0)	1 (2.0)	3 (6.0)
Turkeys	50	0 (0)	1 (2.0)	0 (0)	1 (2.0)
Total	350	31 (8.86)	3 (0.86)	1 (0.29)	35 (10.0)

Table 2. Occurrence of *Salmonella* spp. in the examined poultry litters

Type of litter	Examined number	Number of samples positive for:		
		<i>S. Kentucky</i> (%)	<i>S. Typhimurium</i> (%)	Total (%)
Chickens	Broilers	8 (53.33)	0 (0)	8 (53.33)
	Breeders	2 (66.66)	0 (0)	2 (66.66)
	Layers	0 (0)	0 (0)	0 (0)
Ducks	7	0 (0)	2 (28.57)	2 (28.57)
Turkeys	2	0 (0)	0 (0)	0 (0)
Total	30	10 (33.33)	2 (6.66)	12 (40.0)

Table 3. Occurrence of *Salmonella* spp. in different specimens from the examined humans

Type of sample	No. of samples	Recovered <i>Salmonella</i> strains	
		<i>S. Kentucky</i>	%
Fecal samples	90	0	0
Hand swabs	90	8	8.88

Also in the view of the data illustrated in Table 1 it was clear that *S. Typhimurium* was the only serovar that could be recovered from turkeys at a rate of 2.0%. The current results support the report of Ferris et al. (1999) who revealed that the most commonly identified paratyphoid serovars in turkeys were *S. Senftenberg*, *S. Heidelberg*, *S. hadar*, *S. Muenster*, and *S. Typhimurium*. Furthermore high levels of *Salmonella* spp. were recorded by Aury et al. (2010).

In the contrary the current data are much higher than those reported by Abd-Allah (2003) who did not recover any *Salmonella* organism from the detected

turkeys. Presumably infection of turkeys with *Salmonella* was transmitted laterally by direct contact between infected birds and uninfected turkeys or by indirect contact with contaminated environments through ingestion of *Salmonella* organisms. Among valuable turkey's breeder flocks, infection with *Salmonella* was generally accompanied by severe economic losses, because of their chronic nature and the difficulty of eradication. In many cases, the infection seriously impairs fertility, hatchability and egg production. The most important aspect, however, is the continuing effect of *Salmonella* contaminated

turkey meat and meat products on public health (Hafez and Jodas, 2000).

On discussing the hazards of poultry litter in the epidemiology of salmonellosis, Table 2 showed that the detected *Salmonella* spp. were *S. Kentucky* (53.33% in broiler's litter and 66.66% in breeder's litter) and *S. Typhimurium* (6.66% in duck's litter) while no *Salmonella* organisms could be recovered from layer and turkey litters. The obtained results are in harmony with that achieved by Orji et al. (2005). However the current data are higher than other findings in previous studies, as those obtained by Samaha et al. (2004) and Bayomi et al. (2006). Anyhow in the present study, *S. Kentucky* was the most prevalent serovar identified in litter samples. The current results are, to some extent, in agreement with the data obtained by Payne et al. (2006) who recorded that *S. Kentucky* was the most commonly isolated serovar from both litter and fecal dropping samples, and Li et al. (2007) who also demonstrated that *S. Kentucky* was the most commonly isolated serovar from processed fecal samples.

In this work, conclusively *Salmonella* serovars isolated from broiler's litter were frequently isolated from cloacal swabs of live birds, this provided evidence that litter play an important role as a source of *Salmonella* infection in poultry farms as the infection may arise from the contaminated litter or the infected birds may contaminate litter via their droppings.

Regarding the occurrence of *Salmonella* spp. in different specimens from the examined humans as shown in Tables 3 it was clearly obvious that all stool samples reacted negatively to all *Salmonella* spp. whereas 8 out of 90 (8.88%) hand swabs from poultry workers were *S. Kentucky*. The occurrence of *Salmonella* spp. in hand swabs are much higher than those obtained by Abd-Allah (2003) who isolated *Salmonella* spp. at the rate of 3.1% from hand swabs. It is worth mentioning that all the examined humans were apparently healthy (absence of fever and diarrhea). The detected isolation rate of *Salmonella* spp. in hand swabs of the examined humans in this study should be considered as a serious threat to public health, because *Salmonella* bacteria are usually transmitted to humans by the fecal-oral route (ingestion of foods contaminated with feces or contaminated by the unwashed hands of an infected food handler).

In the present study, conclusively *Salmonella* serovars (*S. Kentucky*) isolated from broilers, breeders and layers were frequently isolated from hand swabs of the examined individuals (poultry workers), this provided evidence that direct contact with poultry or poultry environment may pose health hazards for humans especially whom their occupation necessitate their contact with poultry.

REFERENCES

Aarestrup, F.M., Hendriksen, R.S., Lockett, J., Gay, K., Teates, K., McDermott, P.F., White, D.G., Hasman, H., Sorensen, G., Bangtrakulnonth, A., Pornreongwong, S., Pulsrikarn, C., Angulo, F.J. and Gerner-Smidt, P. 2007. International spread

of multidrug-resistant *Salmonella* Schwarzengrund in food products. *Emerging Infectious Diseases* 13, 726-731.

Abd-Allah, Heba. A. 2003. Tracing some sources of infection of some zoonotic bacteria among family Enterobacteriaceae. M.V. Sc. Thesis (Zoonoses). Fac. Vet. Med. Zagazig Univ. Egypt.

Abd El-Hamid, H. S.; Torkey, H. A.; Al-Shaboury, F. A.; Meran, A. Sleim.; Ellakany, H. and Awad, A. M. 2004. Epidemiological studies on salmonellosis in poultry. *Alex. J. Vet. Science* 21, 294-313.

Aury, K.; Chemaly, M.; Petetin, I.; Rouxel, S.; Picherot, M.; Michel, V. and Le Bouquin, S. 2010. Prevalence and risk factors for *Salmonella enterica* subsp. *Enterica* contamination in French breeding and fattening turkey flocks at the end of the rearing period. *Preventive Veterinary Medicine* 94, 84-93.

Badr, Jihan M. and Abd El Monaem, H. 2008. Antigenic variations between different *Salmonella* serotypes isolated from chickens. *Assiut Vet. Med. J.* 54, 373-388.

Bayomi, A. M.; Radwan, G. S. and Trabees, R. Z. 2006. Existence of *Salmonellae* in the environment of some egg laying farms and its hygienic importance. *SCVMJ X* (1), 303-310.

Byrd, J. A.; Corrier, D. E.; DeLoach, J. R. and Nisbet, D. J., 1997. Comparison of drag-swab environmental protocols for the isolation of *Salmonella* in poultry houses. *Avian Dis.* 41, 709-713.

Byrd, J. A.; DeLoach, J. R.; Corrier, D. E.; Nisbet, D. J. and Stanker, L. H. 1999. Evaluation of *Salmonella* serotype distributions from commercial broiler hatcheries and grower houses. *Avian Dis.* 43, 39-47.

Caldwell, D. J.; Hargis, B. M.; Corrier, D. E.; Vidal, L. and DeLoach, J. R. 1995. Evaluation of persistence and distribution of *Salmonella* serotype isolation from poultry farms using drag-swab sampling. *Avian Dis.* 39, 617- 621.

Cardinale, E.; Tall, F.; Guèye, E. F.; Cisse, M. and Salvat, G., 2004. Risk factors for *Salmonella enterica* subsp. *Enterica* infection in senegalese broiler-chicken flocks. *Preventive Veterinary Medicine* 63, 151-161.

Collee, J. G.; Miles, R. S. and Watt, B. 1996. Tests for the identification of bacteria. In: *Practical Medical Microbiology* (Eds. Collee, J. G.; Marmion B. P.; Fraser, A.G. and Simmons, A.) Churchill Livingstone, New York, Edinburgh and London 131-149.

Corry, J. E. L.; Allen, V. M.; Hudson, W. R.; Breslin, M. F. and Davies, R. H., 2002. Sources of *Salmonella* on broiler carcasses during transportation and processing: modes of contamination and methods of control. *Journal of Applied Microbiology* 92, 424-432.

Crump, J.A., Medalla, F.M., Joyce, K.W., Krueger, A.L., Hoekstra, R.M., Whichard, J.M. and

- Barzilay, E.J. 2011. Antimicrobial resistance among invasive nontyphoidal *Salmonella* enterica isolates in the United States: National Antimicrobial Resistance Monitoring System, 1996 to 2007. *Antimicrobial Agents and Chemotherapy* 55, 1148–1154.
- Duchet-Suchaux, M.; Le chopier, P.; Marly, J.; Bernardet, P.; Delaunay, R. and Pardon, P. 1995. Quantification of experimental *Salmonella* Enteritidis carrier state in B13 leghorn chicks. *Avian Dis.* 39, 796–803.
- El-Shamy, A. U., 1999. Some studies on bacterial agents causing enteritis in chicken breeder flocks. M.V Sc. Thesis (Poult. Dis.). Fac. Vet. Med. Zagazig Univ. Egypt.
- Fashae, K., Ogunsola, F., Aarestrup, F.M. and Hendriksen, R.S. 2010. Antimicrobial susceptibility and serovars of *Salmonella* from chickens and humans in Ibadan, Nigeria. *Journal of Infection in Developing Countries* 4, 484–494.
- Ferris, K. E.; Fisher, S. D.; Flugrad, B. R. and Timm, J. M. 1999. *Salmonella* serotypes from animals and related sources reported during July 1998-June 1999. Proc. 103rd Ann. Meet U.S. Anim. Health Assoc. U.S. Animal Health Association: Richmond, VA. 488-507.
- Gast, R. K. 2003. Paratyphoid infections. In: *Diseases of poultry*, 11th edn.(Eds Saif, Y.). Iowa State University Press, Ames 583-613.
- Hafez, M. H. and Jodas, S., 2000. *Salmonella* infections in turkeys. In: *Salmonella* in domestic animals (Eds. C. Wray and A. Wray), Kent and Florida 133-149.
- Hegazy, A. E. 2002. Epidemiological studies on salmonellosis in chickens with special reference to *Salmonella* Enteritidis. Ph.D. Thesis, Fac. Vet. Med., Alex. Univ. Egypt.
- Hendriksen, R.S.; Vieira, A.R.; Karlsmose, S.; Lo Fo Wong, D.M.; Jensen, A.B.; Wegener, H.C. and Aarestrup, F.M. 2011. Global monitoring of *Salmonella* serovar distribution from the world health organization global foodborne infections network country data bank: results of quality assured laboratories from 2001 to 2007. *Foodborne Pathogens and Disease* 8, 887–900.
- Henry, R. R. 2000. *Salmonella* infection in ducks. In: *Salmonella* in domestic animals (Eds. C. Wray and A. Wray), Kent and Florida 157-166.
- Hennessy, T.W.; Cheng, L.H.; Kassenborg, H.; Ahuja, S.D.; Mohle-Boetani, J.; Marcus, R.; et al., 2004. Egg consumption is the principal risk factor for sporadic *Salmonella* serotype Heidelberg infections: a case-control study in Food Net sites. *Clin Infect Dis.* 38, 237–43.
- Humphrey, T. J.; Thrlfall, E. J. and Cruickshank, J. G. 1998. Salmonellosis. In: *Zoonoses, Biology, Clinical Practice and Public Health Control*. (Eds. Palmer, S. R.; Soulsby, L. and Simpson, D. I.H.). Oxford University press, New York 191-205.
- Humphrey, A. M.; Radwan, G. S. and Trabees, R. Z. 2006. Existence of *Salmonellae* in the environment of some egg laying farms and its hygienic importance. *SCVMJ X* 1, 303-310.
- Kauffmann, G., 1974. Kauffmann-White Scheme. WHO-BD, 1972, 1, Rev., 1. *Acta. Path. Microbiol. Scand.* 61, 385.
- Kottwitz, L.B. M.; Back, A.; Leão, J.A.; Alcocer, I.; Karan, M. and Oliveira, T.M., 2008. Contaminação por *Salmonella* spp. em uma cadeia de produção de ovos de uma integração de postura commercial. *Arquivos Brasileiro de Med. Vet. Zootecnia* 60 (2), 496-498.
- Labellec, C.; Colin, P.; Bennejean, G.; Pacquin, J.; Guillerme, A. and Debois, J. C., 1986. Influence of resident *Salmonella* on contamination of broiler flocks. *Poult. Sci.* 65, 2034-2039.
- Li, X.; Payne, J.B.; Santos, F. B.; Levine, J. F.; Anderson, K. E. and Sheldon, B. W., 2007. *Salmonella* Populations and Prevalence in Layer Feces from Commercial High-Rise Houses and Characterization of the *Salmonella* Isolates by Serotyping, Antibiotic Resistance Analysis, and Pulsed Field Gel Electrophoresis. *Poultry Science* 86, 591-597.
- L Plym, F. and Wierup, M., 2006. *Salmonella* contamination: a significant challenge to the global marketing of animal food products. *Rev. Sci. Tech.* 25, 541–554.
- Mead, P.S.; Slutsker, L.; Dietz, V.; McCaig, L.F.; Bresee, J.S.; Shapiro, C.; Griffin, P.M. and Tauxe, R.V., 1999. Food-related illness and death in the United States. *Emerg. Infect. Dis.* 5, 607–625.
- Mohammed, G. G., 1986. Epizootological studies on pathogens of migrating ducks in Sinai Peninsula. M.V Sc. Thesis, Fac. Vet. Med. Cairo Univ. Egypt.
- Muhammad, Maryam; Lawal, U.M.; Abdul-Ganiyu, A.; Aliyu, U. M.; Samuel, A. and Lisa, B., 2010. Prevalence of *Salmonella* associated with chick mortality at hatching and their susceptibility to antimicrobial agents. *Veterinary Microbiology* 140, 131-135.
- Olsen, J.E.; Brown, D.J.; Madsen, M. and Bisgaard, M., 2003. Cross contamination on a broiler slaughterhouse line demonstrated by use of epidemiological markers. *Journal of Applied Microbiology* 94, 826-835.
- Orji, M.U.; Henry, C. O. and Theodore, I. M., 2005. Isolation of *Salmonella* from poultry droppings and other environmental sources in Awka, Nigeria. *International Journal of Infectious Diseases* 9, 86-89.
- Payne, J.B.; Li, X.; Santos, F.B. and Sheldon, B.W., 2006. Characterization of *Salmonella* from Three Commercial North Carolina Broiler Farms. *International Journal of Poultry Science* 5 (12), 1102-1109.
- Ramadan, N.M., 1997. Further studies on epidemiology of salmonellosis in poultry in Egypt. Ph. D. Thesis. (Poult. Dis.). Fac. Vet. Med. Cairo Univ. Egypt.
- Saad, A.M.; Almujali, D.M.; Babiker, S.H.; Shuaib, M.

- A. M.; Abdelgadir, K. A. and Alfadul, Y. A., 2007. Prevalence of *Salmonellae* in Broiler Chicken Carcasses and Poultry Farms in the Central Region, K.S.A. *J. Anim. Vet. Adv.* 6 (2), 164-167.
- Samaha, H.A.; Haggag, Y.N.; Draz, A.A. and Mohamad, L. N., 2004. The role of poultry in transmitting some zoonotic bacterial and fungal diseases. *Alex. J. Vet. Science* 21 (1), 1-13.
- Sackey, B. A.; Mensah, P.; Collison, E. and Sakyi-Dawson, E., 2001. *Campylobacter*, *Salmonella*, *Shigella* and *Escherichia coli* in live and dressed poultry from metropolitan Accra. *International Journal of Food Microbiology* 71, 21-28.
- Shahada, F.; Chuma, T.; Tobata, T.; Okamoto, K.; Sueyoshi, M. and Takase, K., 2006. Molecular epidemiology of antimicrobial resistance among *Salmonella enterica* serovar Infantis from poultry in Kagoshima, Japan. *International Journal of Antimicrobial Agents* 28, 302-307.
- Vo, A. T.; Engeline, D.; Fluit, A. C.; Max, E. C.; Heck, A. V.; Henny, M. M. and Wim, G., 2006. Distribution of *Salmonella enterica* Serovars from humans, livestock and meat in Vietnam and the Dominance of *Salmonella* Typhimurium Phage Type 90. *Veterinary Microbiology* 113, 153-158.
- Waltman W.D., 1999. Methods for Isolating *Salmonellae* from Poultry and the Poultry Environment in: *Salmonella enterica* Serovar Enteritidis in Humans and Animals; epidemiology, pathogenesis, and control. (Eds. Saeed A. M.; Gast R. K.; Potter M. A. and Wall P. G.). Iowa State University Press, Ames, Iowa, USA. 419-432.
- Wedderkopp, A.; Gradel, K.O.; Jørgensen, J. C. and Madsen, M., 2001. Pre-harvest surveillance of *Campylobacter* and *Salmonella* in Danish broiler flocks: a 2-year study. *International Journal of Food Microbiology* 68, 53-59.
- Woodward, C.L.; Kwon, Y.M.; Kubena, L.F.; Byrd, J.A.; Moore, R.W.; Nisbet, D.J.; et al., 2005. Reduction of *Salmonella enterica* serovar Enteritidis colonization and invasion by an alfalfa during molt in leghorn hens. *Poult Sci.* 84, 185-93.