Review Article

Ascitic Fluid Analysis in the Differential Diagnosis of Ascites: Focus on Cirrhotic Ascites

Lin-Lin Huang¹, Harry Hua-Xiang Xia² and Sen-Lin Zhu¹

¹Department of Gastroenterology, the First Affiliated Hospital, Sun Yat-sen University, Guangzhou, China; ²Novartis Pharmaceuticals Corporation, East Hanover, NJ, USA

Abstract

Ascites is the pathologic accumulation of fluid within the peritoneal cavity. Because many diseases can cause ascites, in particular cirrhosis, samples of ascitic fluid are commonly analyzed in order to develop a differential diagnosis. The concept of transudate versus exudate, as determined by total protein measurements, is outdated and the use of serumascites albumin gradient as an indicator of portal hypertension is more accurate. Lactate dehydrogenase (LDH), vascular endothelial growth factor (VEGF), and other tumor markers can be helpful in distinguishing between malignant and benign conditions. Glucose and adenosine deaminase levels may support a diagnosis of tuberculous disease, and amylase level may indicate a diagnosis of pancreatitis. Given the specificity and sensitivity of laboratory results, accurate diagnosis should be based on both laboratory data and clinical judgment.

© 2014 The Second Affiliated Hospital of Chongqing Medical University. Published by XIA & HE Publishing Ltd. All rights reserved.

Introduction

Ascites is defined as pathological fluid accumulation within the abdominal cavity. The word ascites is derived from the Greek word 'askos', which means a bag or sack. Clinically, ascites is a consequence or complication of a number of diseases, including hepatic, cardiac, and renal diseases, infection, and malignancy. Ascites usually carries an unfavorable prognosis. For example, the development of ascites in cirrhotic patients is associated with a mortality of 15% and 44% at one-year and five-year follow-up periods, respectively. However, the prognosis largely depends on the underlying cause (*i.e.* the primary disease). Combined analysis of laboratory data of ascitic fluid samples and clinical and pathological data is essential for establishing a

Keywords: Ascitic fluid analysis; Differential diagnosis; Ascites; Cirrhosis. **Abbreviations:** ADA, adenosine deaminase activity; AFP, α-fetoprotein; BHBT, β-hydroxybutyrate; CA, cancer antigen; CEA, carcinoembryonic antigen; 1 H NMR, proton nuclear magnetic resonance; LDH, lactate dehydrogenase; PC, peritoneal carcinomatosis; PCR, polymerase chain reaction; SBP, spontaneous bacterial

peritonitis; TBP, tuberculous peritonitis; TP, total protein; SAAG, serum-ascites albumin gradient; VEGF, vascular endothelial growth factor. Received: 14 July 2013; Revised: 20 January 2014; Accepted: 21 January 2014 DOI of original article: 10.14218/JCTH.2013.00010.

Correspondence to: Sen-Lin Zhu, Department of Gastroenterology, The First Affiliated Hospital, Sun Yat-sen University, 58 Zhongshan 2nd Road, Guangzhou 510080, China. Tel: +86-20-87755766 EXT 8182, E-mail: zhusl@mail.sysu.edu. cn

differential diagnosis. This review aims to assess critically the value of ascitic fluid analysis in the diagnosis of ascites, especially cirrhotic ascites.

Types of ascites and their pathogenesis

Under normal circumstances, the amount of peritoneal fluid depends on a balance between plasma flowing into and out of the blood and lymphatic vessels. It is only when this balance has been disrupted does ascites form. The imbalance in the level of plasma may be due to increased capillary permeability, increased venous pressure, decreased protein (oncotic pressure), or increased lymphatic obstruction. 2,7

Ascites is one of the most frequent complications of cirrhosis and portal hypertension. 1,4,8,9 Up to 50% of cirrhotic patients will develop ascites within a 10 year follow-up period. 10,11 Hepatic cirrhosis accounts for up to 85% of cases of ascites, 12 and malignancies account for approximately 10%. $^{13-16}$ The other types of ascites are categorized as cardiogenic, nephrogenic, infectious, and miscellaneous $^{2,13-16}$ (Table 1).

Ascitic fluid analysis and clinical implications

Gross appearance

The initial evaluation of the gross appearance of ascitic fluid can offer useful information in the differential diagnosis. Under normal conditions, peritoneal fluid is clear to pale yellow.

Milky ascites, also called chylous ascites, is characterized by the presence of chylomicrons, which are lipoprotein particles that consist of large amounts of triglycerides. 2,17,18 There are many known causes of chylous ascites, including cirrhosis, infections (parasitic and tuberculosis), malignancy, congenital defects, traumatism, inflammatory processes, nephropathies, and cardiopathies.^{2,19,20} Abdominal malignancy is a major cause of chylous ascites in adults, whereas congenital lymphatic abnormalities are more likely causes in children.²¹ However, it should be noted that pseudochylous ascites or cloudy/turbid ascites is associated with bacterial infection, peritonitis, pancreatitis, or perforated bowel.²² Therefore, the presence of both chylomicrons and a high concentration of triglycerides is necessary to distinguish chylous ascites from pseudochylous ascites. This is important since the frequency of malignancy is as high as 80% in adults with chylous ascites.2

Bloody ascites is a characteristic of benign or malignant tumors, hemorrhagic pancreatitis, or perforated ulcer,²³

Table 1. Types of ascites and underlying primary diseases

Type of ascites	Primary disease
Hepatic	Cirrhosis Hepatic venous outflow obstruction (Hepatic vein obstruction, Budd-Chiari syndrome, Veno-occlusive disease) Portal vein occlusion Inferior vena cava obstruction Hepatic cancer
Cardiogenic	Congestive cardiac failure Constrictive pericarditis
Nephrogenic	Nephrotic syndrome
Malignant	Ovarian cancer Cervix cancer Endometrial cancer Breast cancer Esophageal cancer Gastric cancer Colorectal cancer Lung cancer Pancreatic cancer Hepatobiliary cancer Primary peritoneal cancer
Infectious ascites	Tuberculous peritonitis Spontaneous bacterial peritonitis Fungal infection Parasite infections Chlamydia infection
Miscellaneous ascites	Chylous ascites Pancreatic ascites Bile ascites Ovarian disease (Meig's syndrome, Struma ovarii, Ovarian hyperstimulation) Systemic lupus erythematosis Whipple's disease Sarcoidosis

whereas clear or straw colored ascites is often associated with cirrhosis. ²⁴ Therefore, the gross appearance of ascites can provide preliminary clues regarding the etiology of the underlying disease.

Biochemical tests

Ascitic fluid total protein and the serum-ascites albumin gradient (SAAG)

For many years, the ascitic total protein concentration has been used to determine whether ascitic fluid was a transudate or exudate.² However, this paradigm was flawed and resulted in frequent misclassifications. Currently, it is accepted that the accuracy of the relationship between ascitic protein concentration and etiology of ascites was overestimated.²⁵ For example, hemodynamic-related cardiac ascites was incorrectly considered to cause low protein concentration. ^{26,27} The same can be applied to cirrhotic and malignant cases. Gupta et al. reported that 24% of patients with uncomplicated cirrhosis had an ascitic total protein concentration greater than 25 g/L,²⁸ and Alexandrakis et al. reported that 20% of malignant ascites cases had a low protein concentration.²⁷ Thus, the use of ascitic total protein is now considered outmoded and was replaced with SAAG. SAAG is a more sensitive and specific measure for the

differentiation of ascites due to portal hypertension from ascites due to other pathophysiological mechanisms (e.g. peritoneal inflammation).

SAAG, which was first proposed by Hoefs et~al. in 1981, is calculated by subtracting the ascites albumin concentration from the serum albumin concentration. In prospective studies, it was shown to be a better discriminant than the older criterion (transudate versus exudate). SAAG is generally low (<1.1 g/dL) in ascites not due to portal hypertension, as in cases of infection or malignancy (not due to portal hypertension-related ascites, as in cases of liver cirrhosis or congestive heart failure. SaAG is high (>1.1~g/dL) in portal hypertension-related ascites, as in cases of liver cirrhosis or congestive heart failure. Has been shown that the causal mechanism was identified in 97% of cases with SAAG, whereas only 55% was identified using ascitic total protein concentration. AAAG has been adopted in the British and the American guidelines as an initial testing strategy. SaAAG has been adopted in the

Lactate dehydrogenase (LDH)

Early studies found uniformly high levels of LDH in malignant effusions and low levels of LDH in non-malignant effusions.^{2,35} Boyer *et al.* observed that the mean ascitic fluid LDH level was much lower in patients with liver disease than in those with malignant ascites (167±9 *vs.* 913±228 SU).³⁵

Similar to the classification of pleural fluid proposed by Light et al., 36 the value of combining LDH with total protein analysis has been explored for ascitic fluid. The cut-off values for three parameters in the ascitic fluid for differentiation between hepatic and non-hepatic ascites are, as follows: LDH of 400 SU, fluid/serum LDH ratio of 0.6, and fluid/serum total protein (TP) ratio of 0.5. Ascitic levels higher than the cutoffs for any two out of three parameters indicate a nonhepatic cause of the ascites, whereas values below the cutoffs for all three parameters strongly suggest a hepatic cause of ascites.² According to Gokturk et al., LDH values were higher in patients with an SAAG of 1.1 g/dL or less than in those with an SAAG greater than 1.1 g/dL. 37 However, Sevinc et al. reported that in patients with malignant ascites, ascitic fluid LDH values had high sensitivity but low specificity for the diagnosis of the disease, and a low value of LDH did not necessarily exclude malignancy.³⁸ Therefore, the value of ascitic LDH levels requires further investigation.

Glucose

Since glucose diffuses readily across membranes, the concentration of glucose in the ascitic fluid, under normal conditions, is similar to that in the serum.³⁹ However, ascitic glucose concentration decreases due to consumption by bacteria, white blood cells or cancer cells in the fluid in tuberculous peritonitis, spontaneous bacterial peritonitis (SBP), and malignancy. ^{39,40} According to Mansour-Ghanaei et al., ascitic glucose concentration is often significantly lower than normal in tuberculous ascites, which makes it an indicator in differentiating tuberculosis from other diseases, such as cirrhosis. 41 This is consistent with Wilkins et al. who recommended that the ascitic/blood glucose ratio is a useful test in the differentiation of tuberculous peritonitis from ascites due to other causes.⁴² However, when considering the value of glucose in patients with SAAG greater or less than 1.1 g/dL, there was no significant difference between them.³⁷ Therefore, due to its low diagnostic sensitivity and specificity, the application of ascitic glucose analysis is limited in routine practice.43

Amylase

Amylase-rich ascitic fluid commonly occurs in cases of pancreatic duct damage or obstruction due to pancreatitis or pancreatic trauma. 44 Elevation of amylase levels above the serum reference range in ascitic fluid was found in up to 90% of patients with acute pancreatitis and pancreatic pseudocyst.² When pancreatic ascites needs to be distinguished from ascites secondary to alcoholic cirrhosis, it can be accomplished by detecting high amylase levels in the ascitic fluid.⁴⁵ During the course of severe acute pancreatitis, the level of ascitic amylase can be 100 times higher than serum. However, increased amylase in ascites can also been found in patients with malignancy, 46 perforated peptic ulcer, upper abdominal surgery, mechanical intestinal obstruction, mesenteric vascular disease, biliary obstruction, and acute cholecystitis. Therefore, hyperamylasemia is not a specific marker for pancreatic damage. 47

Adenosine deaminase

Ascitic fluid adenosine deaminase activity (ADA) has been reported to be more sensitive and specific for the early

diagnosis of tuberculous ascites than for other types of ascites. 48,49 A recent meta-analysis of four studies that included 264 patients confirmed the high sensitivity (100%) and specificity (97%) of using cut-offs of ADA from 36 to 40 IU/L in the diagnosis of tuberculous ascites. 2 Moreover, a recent study reported that ascitic ADA levels in patients with tuberculous peritonitis (TBP) and peritoneal carcinomatosis (PC) were $66.76 \pm 32.09 \, \text{IU/L}$ and $13.89 \pm 8.95 \, \text{IU/L}$, respectively (P < 0.01), 50 indicating that ascitic ADA analysis is valuable in differentiating between TBP and PC. Furthermore, Liao et~al. found that ADA values of patients with TBP were notably higher than those with cirrhosis, and every patient in the cirrhosis control group had an ascites ADA level lower than the lowest value in the TBP group. 51

Non-biochemical tests

Cell counts, bacterial culture, and polymerase chain reaction (PCR)

Non-biochemical tests of ascitic fluid, including cell counts, bacterial culture, and PCR, play an important role in diagnosing the cause of ascites, especially in infectious ascites. SBP is defined by the presence of neutrophil cells greater than or equal to 250/μL or a positive bacterial culture in the ascitic fluid without evidence of an abdominal source. 8,52 Cell counts using automatized equipment such as a flow cytometer and culture of ascites fluid should be performed simultaneously.8 Despite the use of sensitive methods, ascitic fluid cultures are negative in as many as 60% of patients with increased ascites neutrophil counts and clinical manifestations suggestive of SBP. 53-55 Therefore, if SBP is suggested by an elevated ascitic neutrophil cell counts and clinical signs and symptoms, antibiotic treatment must be initiated without waiting for the culture result. In a recent study of 1,041 patients with cirrhosis, Cadranel et al. performed total and differential leukocyte counts and bacterial cultures of ascitic fluid and observed that SBP occurred in 11.7% of inpatients and 3.1% of outpatients.⁵⁶ Moreover, they reported that the incidence of SBP was 8.3% in symptomatic patients, whereas the rate was 1.2% in asymptomatic patients. Therefore, cell counts and bacterial culture should also be performed in patients with cirrhotic ascites, especially those with symptoms, due to the high incidence of SBP. Moreover, it has been shown that in cirrhotic patients, compared to SBP, tuberculous peritonitis is associated with, in ascites, lower white blood cell counts, a higher proportion of mononuclear leukocytes (lymphocytes and monocytes), a higher protein concentration, and higher $\mathsf{ADA.}^\mathsf{57}$ However, the sensitivity of direct microscopic smear detection of acid-fast bacilli in the ascitic fluid (0%-6%) and ascitic fluid mycobacterial culture (20%-35%) is low, and mortality is high in patients with tuberculous peritonitis and other various medical conditions, such as cirrhosis, renal failure, diabetes mellitus, and malignancy. Because of the delay in obtaining the results of mycobacterial cultures of ascitic fluid, 57-59 the value of these tests in the differential diagnosis of ascites is limited. However, in recent years, advances in molecular techniques have provided a new approach to the rapid diagnosis of bacterial infection, including tuberculosis, by PCR in small volumes of ascitic fluid (50 ml). 60 PCR can detect minimal amounts of bacterial DNA and improves the rates and velocity of bacterial identification⁶¹⁻⁶³ from four to six weeks for microbiological cultures to 24 hours.⁶⁴ Sorianoet al. detected bacterial DNA in ascitic fluid in 60% of cirrhotic patients with sterile ascites, ⁶¹ and this was associated with an increase in inflammatory response ^{65,66} and a worse prognosis. ⁶⁷ In diagnosing tuberculous effusions, PCR appears to be an ideal tool, with 94% sensitivity and 88% specificity. ^{68,69} Therefore, PCR can be a rapid and reliable method for identification of infectious ascites and accelerates the diagnostic decision making process relative to microbiological cultures.

Viscosity

Ascitic fluid viscosity is a newly proposed indicator in differentiating ascites. A recent study by Gokturk et al. evaluated the role of ascitic fluid viscosity in discriminating between ascites due to portal hypertension-related and nonportal hypertension-related causes, and compared the results with SAAG.³⁷ In that study, ascitic fluid viscosity was determined in a programmable rotational viscometer using 0.5 mL ascitic samples from 142 patients with newly diagnosed ascites due to various causes.³⁷ The mean ascitic fluid viscosities were 0.86±0.12 cP and 1.22±0.25 cP in patients with an SAAG greater than 11 g/L and an SAAG of 11g/L or less, respectively, indicating a close correlation between viscosity and SAAG. Moreover, with a cut-off value of 1.03 cP, ascitic fluid viscosity measurement exhibited high sensitivity (98%), specificity (80%), and positive and negative predictive values (79% and 94%, respectively) for the etiological discrimination of ascites.³⁷ Although there are only a few studies evaluating the viscosity of ascites, ^{70,71} the speed, simplicity, inexpensiveness, and necessity of only a small sample volume make it a useful, and likely more popular, diagnostic tool for the differential diagnosis of ascites in clinical research and practice.

Proton nuclear magnetic resonance (¹H NMR) spectroscopy

High-resolution ¹H NMR spectroscopy of body fluids has emerged as an important tool for differential diagnosis of diseases. In this technique, a few biochemical agents, such as $\beta\text{-hydroxybutyrate}$ (BHBT), lactate, acetone, and acetoacetate, are used. ¹H NMR spectroscopy can be used to differentiate benign cirrhotic ascites from malignant ascites. In one study, the ascitic concentrations of BHBT, lactate, acetone, and acetoacetate were significantly higher in patients with malignant ascites than in those with cirrhotic ascites.³⁹ In contrast, the ascitic concentrations of glutamine, citrate, glucose, tyrosine, and phenylalanine were significantly lower in patients with malignant ascites than in those with cirrhotic ascites.³⁹ Using a model where BHBT, lactate, citrate, and tyrosine were considered together as markers, ¹H NMR spectroscopy differentiated malignant ascites from cirrhotic ascites with 100% sensitivity and 97.9% specificity, whereas the rates were 53.3% and 76.6% for total ascitic protein, and 60% and 87.2% for SAAG, respectively.³⁹

Vascular endothelial growth factor (VEGF)

VEGF, initially known as vascular permeability factor, has a recognized role in the accumulation of ascitic fluid.^{72–75} Several studies have confirmed, using enzyme immunoassay, the presence of higher VEGF concentrations in malignant ascites than in non-malignant (cirrhotic, tuberculous, inflammatory) ascites.^{76–85} Although VEGF concentrations are significantly higher in malignant ascites, the overlap in the

concentrations of VEGF between malignant and non-malignant ascites is rather large. For example, relative to non-malignant ascites, when using its VEGF mean levels 119.44 pg/ml (70.90 \pm 48.54) as the minimum cut-off limit, the sensitivity and specificity of VEGF to diagnose malignant ascites were 91.3% and 90.9%, respectively. Nascimento et al. and Bamias et al. used 662 pg/ml⁷⁹ and 400 pg/ml⁷⁴ as cut-off values to discriminate between malignant ascites and non-malignant ascites, respectively.

Therefore, VEGF, a noninvasive and simple marker available in clinical pathology laboratories, may be useful as a parameter for the differential diagnosis of malignant and nonmalignant ascites. However, further investigation is necessary to confirm an optimum cut-off value.

Tumor markers

Tumor markers can be used to determine cancer risk, screen for early cancers, confirm diagnosis, predict prognosis, and monitor metastasis, recurrence, or progression of cancers. 13 Well-established tumor markers, including α -fetoprotein (AFP), carcinoembryonic antigen (CEA), cancer antigen (CA) 19-9, and CA125, have been evaluated for their utility in differentiating malignant ascites from non-malignant ascites. It has been shown that ascitic levels of AFP, CEA, CA19-9, and CA125 are significantly higher in patients with malignancies such as hepatocellular cancer, colorectal cancer, pancreatic cancer, and ovarian cancer than in those with non-malignant etiologies. ^{2,14,50,86–88} It should be noted, however, that other non-malignant conditions such as gastritis, diverticulitis, cirrhosis, and other cholestatic, pancreatic, and hepatic diseases are known to cause elevations in these tumor markers.86 For example, increased ascitic CEA and CA 19-9 can be present in cirrhosis, ^{89–91} and high levels of CA125 in ascitic fluid can occur in patients with tuberculous peritonitis or with cirrhosis. ^{13,50,92} These findings indicate that elevated tumor marker levels in ascitic fluid must be interpreted with caution when differentiating malignant ascites from other types of ascites. Although these tumor markers are potentially diagnostic, the gold standard for the diagnosis of malignant ascites is detection of tumor cells in the ascitic fluid.14

Usefulness of ascitic fluid analysis in patients with cirrhosis

Ascites is one of the most frequent complications of cirrhosis. ^{1,4,8,9} Up to 60% of patients with compensated cirrhosis will develop ascites within 10 years of the disease course. ^{10,11,53,93} After the development of ascites, survival rate is only 50% at two to five years. ⁹³ Therefore, differential diagnosis is essential for better management of cirrhosis, and ascitic fluid analysis plays an important role in this purpose. Table 2 outlines the typical characteristics of the ascites in patients with cirrhosis relative to other diseases.

Conclusions

Ascites can be a consequence or complication of many primary diseases and carries an unfavorable prognosis that largely depends on the underlying causes. Cirrhotic ascites accounts for most cases of ascites, and it can be complicated by subsequent infections that also lead to ascites. Ascitic fluid analyses indicating gross appearance, biochemical tests (e.g.

Table 2. Typical characteristics of ascites in patients with cirrhosis compared with other diseases

	Causes of ascites						
Test	Cirrhosis	Congestive cardiac failure	Malignancy	Tuberculosis	SBP	Pancreatitis	
Gross appearance	clear straw or milky	clear to pale yellow	milky or bloody	milky or N	cloudy or turbid	milky or cloudy or turbid	
TP	< 25 g/L	< 25 g/L	≥ 25 g/L	≥ 25 g/L	≥ 25 g/L	≥ 25 g/L	
SAAG	\geqslant 1.1 g/dL	\geqslant 1.1 g/dL	< 1.1 g/dL	< 1.1 g/dL	< 1.1 g/dL	< 1.1 g/dL	
LDH	\downarrow	↓ or N	↑	↑ or N	↑ or N	↑ or N	
Glucose	N	N	\downarrow	\downarrow	\downarrow	\downarrow	
Amylase	N		↑ or N			\uparrow	
ADA	↓ or N		↓ or N	↑			
Cell counts	\geqslant 250/ μ L or N			\geqslant 250/uL or N	≥ 250/uL		
Bacterial culture	+ or -			+ or -	+		
Viscosity	< 1.03 cP	< 1.03 cP	≥ 1.03 cP	≥ 1.03 cP	≥ 1.03 cP	≥ 1.03 cP	
¹ H NMR	↑ or ↓	↑ or ↓					
VEGF	\downarrow	N	↑	\downarrow	\downarrow	\downarrow	
Tumor markers	↑ or N	N	↑	↑ or N		↑ or N	

 $[\]uparrow$ -increase, \downarrow - decrease, N -normal, + -positive, - - negative.

SAAG, LDH, glucose, amylase, and ADA), and non-biochemical tests (e.g. cell counts, bacterial culture and PCR, viscosity, ¹H NMR spectroscopy, VEGF, and tumor markers) can provide useful clues in the differential diagnosis of ascites and help in establishing a diagnosis. It should be emphasized that physicians should use the ascitic fluid analysis in combination with clinical, pathological, and imaging data, in order to make an accurate diagnosis of the cause of ascites.

Acknowledgements

This project was sponsored in part by grants from the National Natural Science Foundation of China (#81072044) and the Guangdong Natural Science Foundation (#S2011010004653).

Conflict of interest

None

Author contributions

Writing the article (LLH), organizing the article (HXX), writing and modifing the article (SLZ).

References

- Senousy BE, Dragnov PV. Evaluation and management of patients with refractory ascites. World J Gastroenterol 2009;15:67–80.
- [2] Tarn AC, Lapworth R. Biochemical analysis of ascitic (peritoneal) fluid: what should we measure? Ann Clin Biochem 2010;47:397-407.
- [3] Reynolds TB. Ascites. Clin Liver Dis 2000;4:151–168.
- [4] Biecker E. Diagnosis and therapy of ascites in liver cirrhosis. World J Gastroenterol 2011;17:1237–1248.
- [5] Planas R, Montoliu S, Ballesté B, Rivera M, Miquel M, Masnou H, et al. Natural history of patients hospitalized for management of cirrhotic ascites. Clin Gastroenterol Hepatol 2006;4:1385–1394.
- [6] Wimberger P, Gilet H, Gonschior AK, Heiss MM, Moehler M, Oskay-Oezcelik G, et al. Deterioration in quality of life (QoL) in patients with malignant ascites:

- results from a phase II/III study comparing paracentesis plus catumaxomab with paracentesis alone. Ann Oncol 2012;23:1979–1985.
- [7] Ginès P, Cárdenas A, Arroyo V, Rodés J. Management of cirrhosis and ascites. N Engl J Med 2004;350:1646–1654.
- [8] Bendtsen F, Grønbaek H, Hansen JB, Aagaard NK, Schmidt L, Møller S. Treatment of ascites and spontaneous bacterial peritonitis - Part I. Dan Med J 2012;59:C4371.
- [9] Moore KP, Wong F, Gines P, Bernardi M, Ochs A, Salerno F, et al. The management of ascites in cirrhosis: report on the consensus conference of the International Ascites Club. Hepatology 2003;38: 258–266.
- [10] Ginès P, Cárdenas A. The management of ascites and hyponatremia in cirrhosis. Semin Liver Dis 2008;28:43–58.
- [11] Cárdenas A, Ginès P. Management of refractory ascites. Clin Gastroenterol Hepatol 2005;3:1187–1191.
- [12] Runyon BA, Montano AA, Akriviadis EA, Antillon MR, Irving MA, McHutchison JG. The serum-ascites albumin gradient is superior to the exudatetransudate concept in the differential diagnosis of ascites. Ann Intern Med 1992;117:215–220.
- [13] Tuzun Y, Celik Y, Bayan K, Yilmaz S, Dursun M, Canoruc F. Correlation of tumour markers in ascitic fluid and serum: are measurements of ascitic tumour markers a futile attempt? J Int Med Res 2009;37:79–86.
- [14] Shimada M, Berjohn C, Tanen DA. Ascites as the initial presentation of gastrointestinal carcinoma. J Emerg Med 2013;44:e195–e198.
- [15] Hou W, Sanyal AJ. Ascites: diagnosis and management. Med Clin North Am 2009;93:801–817.
- [16] Saif MW, Siddiqui IA, Sohail MA. Management of ascites due to gastrointestinal malignancy. Ann Saudi Med 2009;29:369–377.
- [17] Fukunaga N, Shomura Y, Nasu M, Okada Y. Chylous ascites as a rare complication after abdominal aortic aneurysm surgery. South Med J 2011; 104:365–367.
- [18] Nishigori H, Ito M, Nishizawa Y, Koyama A, Koda T, Nakajima K, et al. Postoperative chylous ascites after colorectal cancer surgery. Surg Today 2012;42:724–728.
- [19] Gómez-Martín JM, Martínez-Molina E, Sanjuanbenito A, Martín-Illana E, Arrieta F, Balsa JA, et al. Chylousascytes secondary to acute pancreatitis: a case report and review of literature. Nutr Hosp 2012;27:314–318.
- [20] Al-Ghamdi MY, Bedi A, Reddy SB, Tanton RT, Peltekian KM. Chylous ascites secondary to pancreatitis: management of an uncommon entity using parenteral nutrition and octreotide. Dig Dis Sci 2007;52:2261–2264.
- [21] Karagol BS, Zenciroglu A, Gokce S, Kundak AA, Ipek MS. Therapeutic management of neonatal chylous ascites: report of a case and review of the literature. Acta Paediatr 2010;99:1307–1310.
- [22] Runyon BA, Akriviadis EA, Keyser AJ. The opacity of portal hypertensionrelated ascites correlates with the fluid's triglyceride concentration. Am J Clin Pathol 1991;96:142–143.
- [23] Runyon BA, Hoefs JC, Morgan TR. Ascitic fluid analysis in malignancy-related ascites. Hepatology 1988;8:1104–1109.

- [24] McHutchison JG. Differential diagnosis of ascites. Semin Liver Dis 1997;17: 191–202.
- [25] Rissona JR, Macovei I, Loock M, Paquette B, Martin M, Delabrousse E. Cirrhotic and malignant ascites: differential CT diagnosis. Diagn Interv Imaging 2012;93:365–370.
- [26] Christou L, Economou M, Economou G, Kolettis TM, Tsianos EV. Characteristics of ascitic fluid in cardiac ascites. Scand J Gastroenterol 2007;42:1102–1105.
- [27] Alexandrakis MG, Moschandrea JA, Koulocheri SA, Kouroumalis E, Eliopoulos GD. Discrimination between malignant and nonmalignant ascites using serum and ascitic fluid proteins in a multivariate analysis model. Dig Dis Sci 2000;45:500–508.
- [28] Gupta R, Misra SP, Dwivedi M, Misra V, Kumar S, Gupta SC. Diagnosing ascites: value of ascitic fluid total protein, albumin, cholesterol, their ratios, serum-ascites albumin and cholesterol gradient. J Gastroenterol Hepatol 1995;10:295–299.
- [29] Fincher RK, Green RH. High serum albumin ascites gradient ascites-an atypical presentation of metastatic pancreatic cancer. Mil Med 2012;177: 1117–1118.
- [30] Gotyo N, Hiyama M, Adachi J, Watanabe T, Hirata Y. Respiratory failure with myxedema ascites in a patient with idiopathic myxedema. Intern Med 2010; 49:1991–1996.
- [31] Díaz-Mancebo R, Sánchez-Villanueva R, González-García E, Ossorio-González M, Selgas-Gutiérrez R. Nephrogenic ascites: a thing of the past? Nefrologia 2012;32:406–408.
- [32] Hoefs JC. Serum protein concentration and portal pressure determine the ascitic fluid protein concentration in patients with chronic liver disease. J Lab Clin Med 1983;102:260–273.
- [33] Runyon BA. Management of adult patients with ascites due to cirrhosis. Hepatology 2004;39:841–856.
- [34] Moore KP, Aithal GP. Guidelines on the management of ascites in cirrhosis Gut 2006; (Suppl 6):vi1-vi12.
- [35] Boyer TD, Kahn AM, Reynolds TB. Diagnostic value of ascitic fluid lactic dehydrogenase, protein, and WBC levels. Arch Intern Med 1978;138:1103– 1105.
- [36] Light RW. The Light criteria: the beginning and why they are useful 40 years later. Clin Chest Med 2013;34:21–26.
- [37] Gokturk HS, Demir M, Ozturk NA, Unler GK, Kulaksizoglu S, Kozanoglu I, et al. The role of ascitic fluid viscosity in the differential diagnosis of ascites. Can J Gastroenterol 2010:24:255–259.
- [38] Sevinc A, Sari R, Fadillioglu E. The utility of lactate dehydrogenase isoenzyme pattern in the diagnostic evaluation of malignant and nonmalignant ascites. J Nalt Med Assoc 2005;97:79–84.
- [39] Bala L, Sharma A, Yellapa RK, Roy R, Choudhuri G, Khetrapal CL. ¹H NMR spectroscopy of ascitic fluid: discrimination between malignant and benign ascites and comparison of the results with conventional methods. NMR Biomed 2008;21:606–614.
- [40] Lee HH, Carlson RW, Bull DM. Early diagnosis of spontaneous bacterial peritonitis: value of ascitic fluid variables. Infection 1987;15:232–236.
- [41] Mansour-Ghanaei F, Shafaghi A, Bagherzadeh AH, Fallah MS. Low gradient ascites: a seven year course review. World J Gastroenterol 2005;11:2337– 2339.
- [42] Wilkins EG. Tuberculosis peritonitis: diagnostic value of the ascitic/blood glucose ratio. Tubercle 1984;65:47–52.
- [43] Akriviadis EA, Runyon BA. Utility of an algorithm in differentiating spontaneous from secondary bacterial peritonitis. Gastroenterology 1990;98:127– 133.
- [44] Altuğ K, Bekem S, Yağci M, Akçakil M, Budak D. Pancreatic ascites (author's transl). Zentralbl Chir 1976;101:1376–1378.
- [45] SatzN, AmmannR. Pancreatogenic ascites. Schweiz Med Wochenschr 1978; 108:980–988.
- [46] Yokoyama T, Tanaka A, Kato S, Aizawa H. Multiple myeloma presenting initially with pleural effusion and a unique paraspinal tumor in the thorax. Intern Med 2008;47:1917–1920.
- [47] William B Salt I I, Schenker S. Amylase–its clinical significance: a review of the literature. Medicine (Baltimore) 1976;55:269–289.
- [48] Segura RM, Pascual C, Ocaña I, Martínez-Vázquez JM, Ribera E, Ruiz I, Pelegrí MD. Adenosine deaminase in body fluids: a useful diagnosis tool in tuberculosis. Clin Biochem 1989;22:141–148.
- [49] Riquelme A, Calvo M, Salech F, Valderrama S, Pattillo A, Arellano M, et al. Value of adenosine deaminase (ADA) in ascitic fluid for the diagnosis of tuberculous peritonitis: a meta-analysis. J Clin Gastroenterol 2006;40:705– 710
- [50] Kang SJ, Kim JW, Baek JH, Kim SH, Kim BG, Lee KL, et al. Role of ascites adenosine deaminase in differentiating between tuberculous peritonitis and peritoneal carcinomatosis. World J Gastroenterol 2012;18:2837–2843.
- [51] Liao YJ, Wu CY, Lee SW, Lee CL, Yang SS, Chang CS, et al. Adenosine deaminase activity in tuberculous peritonitis among patients with underlying liver cirrhosis. World J Gastroenterol 2012;18:5260–5265.

- [52] Giefer MJ, Murray KF, Colletti RB. Pathophysiology, diagnosis, and management of pediatric ascites. J Pediatr Gastroenterol Nutr 2011;52:503–513.
- [53] European Association for the Study of the liver. EASL clinical practice guidelines on the management of ascites, spontaneous bacterial peritonitis, and hepatorenal syndrome in cirrhosis. J Hepatol 2010;53:397–417.
- [54] Wong F, Bernardi M, Balk R, Christman B, Moreau R, Garcia-Tsao G, et al. Sepsis in cirrhosis: report on the 7th meeting of the International Ascites Club. Gut 2005:54:718–725.
- [55] Fernández J, Navasa M, Gómez J, Colmenero J, Vila J, Arroyo V, et al. Bacterial infections in cirrhosis: epidemiological changes with invasive procedures and norfloxacin prophylaxis. Hepatology 2002;35:140–148.
- [56] Cadranel JF, Nousbaum JB, Bessaguet C, Nahon P, Nguyen-Khac E, Moreau R, et al. Low incidence of spontaneous bacterial peritonitis in asymptomatic cirrhotic outpatients. World J Hepatol 2013;5:104–108.
- [57] Kim NJ, Choo EJ, Kwak YG, Lee SO, Choi SH, Woo JH, et al. Tuberculous peritonitis in cirrhotic patients: comparison of spontaneous bacterial peritonitis caused by Escherichia coli with tuberculous peritonitis. Scand J Infect Dis 2009;41:852–856.
- [58] Chow KM, Chow VC, Hung LC, Wong SM, Szeto CC. Tuberculous peritonitisassociated mortality is high among patients waiting for the results of mycobacterial cultures of ascitic fluid samples. Clin Infect Dis 2002;35: 409–413.
- [59] Cho OH, Park KH, Park SJ, Kim SM, Park SY, Moon SM, et al. Rapid diagnosis of tuberculous peritonitis by T cell-based assays on peripheral blood and peritoneal fluid mononuclear cells. J Infect 2011;62:462–471.
- [60] Tzoanopoulos D, Mimidis K, Giaglis S, Ritis K, Kartalis G. The usefulness of PCR amplification of the IS6110 insertion element of M. tuberculosis complex in ascitic fluid of patients with peritoneal tuberculosis. Eur J Intern Med 2003; 14:367–371.
- [61] Soriano G, Esparcia O, Montemayor M, Guarner-Argente C, Pericas R, Torras X, et al. Bacterial DNA in the diagnosis of spontaneous bacterial peritonitis. Aliment Pharmacol Ther 2011;33:275–284.
- [62] Andrade SS, Bispo PJ, Gales AC. Advances in the microbiological diagnosis of sepsis. Shock 2008;30:41–46.
- [63] Sontakke S, Cadenas MB, Maggi RG, Diniz PP, Breitschwerdt EB. Use of broad range 16S rDNA PCR in clinical microbiology. J Microbiol Methods 2009;76: 217–225.
- [64] Schwake L, von Herbay A, Junghanss T, Stremmel W, Mueller M. Peritoneal tuberculosis with negative polymerase chain reaction results: report of two cases. Scand J Gastroenterol 2003;38:221–224.
- [65] Francés R, Zapater P, González-Navajas JM, Muñoz C, Caño R, Moreu R, et al. Bacterial DNA in patients with cirrhosis and noninfected ascites mimics the soluble immune response established in patients with spontaneous bacterial peritonitis. Hepatology 2008;47:978–985.
- [66] Gonza'lez-Navajas JM, Bellot P, France's R, Zapater P, Muñoz C, García-Pagán JC, et al. Presence of bacterial-DNA in cirrhosis identifies a subgroup of patients with marked inflammatory response not related to endotoxin. J Hepatol 2008;48:61–67.
- [67] Zapater P, France's R, Gonza'lez-Navajas JM, de la Hoz MA, Moreu R, Pascual S, et al. Serum and ascitic fluid bacterial DNA: a new independent prognostic factor in noninfected patients with cirrhosis. Hepatology 2008;48:1924–1931.
- [68] Tan MF, Ng WC, Chan SH, Tan WC. Comparative usefulness of PCR in the detection of Mycobacterium tuberculosis in different clinical specimens. J Med Microbiol 1997;46:164–169.
- [69] Portillo-Gomez L, Morris SL, Pandaro A. Rapid and efficient detection of extrapulmonary Mycobacterium tuberculous by PCR analysis. Int J Tuberc Lung Dis 2000;4:361–370.
- [70] Akay S, Ozutemiz O, Kilic M, Karasu Z, Akyildiz M, Karasulu E, et al. Reabsorption of ascites and the factors that affect this process in cirrhosis. Transl Res 2008:152:157–164.
- [71] Momen-Heravi F, Balaj L, Alian S, Trachtenberg AJ, Hochberg FH, Skog J, et al. Impact of biofluid viscosity on size and sedimentation efficiency of the isolated microvesicles. Front Physiol 2012;3:162.
- [72] Manenti L, Riccardi E, Marchini S, Naumova E, Floriani I, Garofalo A, et al. Circulating plasma vascular endothelial growth factor in mice bearing human ovarian carcinoma xenograftcorrelates with tumor progression and response to therapy. Mol Cancer Ther 2005;4:715–725.
- [73] Dvorak HF, Brown LF, Detmar M, Dvorak AM. Vascular permeability factor/ vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. Am J Pathol 1995;146:1029–1039.
- [74] Bamias A, Koutsoukou V, Terpos E, Tsiatas ML, Liakos C, Tsitsilonis O, et al. Correlation of NK T-like CD3+CD56+ cells and CD4+CD25+(hi) regulatory T cells with VEGF and TNFalpha in ascites from advanced ovarian cancer: Association with platinum resistance and prognosis in patients receiving firstline. platinum-based chemotherapy. Gynecol Oncol 2008:108:421-427.
- [75] Yoshiji H, Kuriyama S, Hicklin DJ, Huber J, Yoshii J, Ikenaka Y, et al. The vascular endothelial growth factor receptor KDR/Flk-1 is a major regulator of malignant ascites formation in the mouse hepatocellular carcinoma model. Hepatology 2001;33:841–847.

- [76] Tamsma JT, Keizer HJ, Meinders AE. Pathogenesis of malignant ascites: Starling's law of capillary hemodynamics revisited. Ann Oncol 2001;12: 1353–1357.
- [77] Kraft A, Weindel K, Ochs A, Marth C, Zmija J, Schumacher P, et al. Vascular endothelial growth factor in the sera and effusions of patients with malignant and nonmalignant disease. Cancer 1999;85:178–187.
- [78] Santin AD, Hermonat PL, Ravaggi A, Cannon MJ, Pecorelli S, Parham GP. Secretion of vascular endothelial growth factor in ovarian cancer. Eur J Gynaccol Oncol 1999;20:177–181.
- [79] Nascimento I, Schaer R, Lemaire D, Freire S, Paule B, Carvalho S, et al. Vascular endothelial growth factor (VEGF) levels as a tool to discriminate between malignant and nonmalignant ascites. APMIS 2004;112:585–587.
- [80] Zebrowski BK, Liu W, Ramirez K, Akagi Y, Mills GB, Ellis LM. Markedly elevated levels of vascular endothelial growth factor in malignant ascites. Ann Surg Oncol 1999;6:373–378.
- [81] Verheul HM, Hoekman K, Jorna AS, Smit EF, Pinedo HM. Targeting vascular endothelial growth factor blockade: ascites and pleural effusion formation. Oncologist 2000;5(suppl 1):45–50.
- [82] Thickett DR, Armstrong L, Millar AB. Vascular endothelial growth factor (VEGF) in inflammatory and malignant pleural effusions. Thorax 1999;54: 707-710.
- [83] Cheng D, Rodriguez RM, Perkett EA, Rogers J, Bienvenu G, Lappalainen U, et al. Vascular endothelial growth factor in pleural fluid. Chest 1999;116:760–765.
- [84] Dong WG, Sun XM, Yu BP, Luo HS, Yu JP. Role of VEGF and CD44v6 in differentiating benign from malignant ascites. World J Gastroenterol 2003;9: 2596–2600.

- [85] Lee HK, Chae HS, Kim JS, Kim HK, Cho YS, Rho SY, et al. Vascular endothelial growth factor levels in ascites between chemonaive and chemotreated patients. Yonsei Med J 2008;49:429–435.
- [86] Kaleta EJ, Tolan NV, Ness KA, O'Kane D, Algeciras-Schimnich A. CEA, AFP and CA 19-9 analysis in peritoneal fluid to differentiate causes of ascites formation. Clin Bio chem 2013;46:814–818.
- [87] Brain O, Brown LH, Suvarna S, Chapman R. Markedly elevated CA19-9 associated with benign ovarian cyst and ascites. BMJ Case Rep 2009;pii: bcr11.2008.1219.
- [88] Medeiros LR, Rosa DD, da Rosa MI, Bozzetti MC. Accuracy of CA 125 in the diagnosis of ovarian tumors: a quantitative systematic review. Eur J Obstet Gynecol Reprod Biol 2009;142:99–105.
- [89] Pissaia A Jr, Bernard D, Scatton O, Soubrane O, Conti F, Calmus Y. Significance of serum tumor markers carcinoembryonic antigen, CA 19-9, CA 125, and CA 15-3 in pre-orthotopic liver transplantation evaluation. Transplant Proc 2009;41:682-684.
- [90] Kadayifci A, Simsek H, Savas MC, Toppare M. Serum tumor markers in chronic liver disease. Neoplasma 1996;43:17–21.
- [91] Nowak J, Jakubowska D, Wiczkowski A, Sprzaczkowska K, Stechły T, Zmudziński W, et al. Carbohydrate antigens CA 19-9,CA 242, CA 50 in liver disease. Wiad Lek 1998;51:484–491.
- [92] Ulusoy AN, Karabicak I, Dicle K, Kefeli M, Tosun M, Cetinkaya M, et al. Peritoneal tuberculosis in premenopausal patients with elevated serum CA 125. Arch Gynecol Obstet 2010;282:639–642.
- [93] Singhal S, Baikati KK, Jabbour II, Anand S. Management of refractory ascites. Am J Ther 2012;19:121–132.