

Indian Journal of Hill Farming 27(1):111-114

# Effect of Presuperovulatory Treatment of Buserelin on Superovulatory Response and Embryo Recovery in Cattle under **Subtropical Conditions of Meghalaya**

## S. KUMAR<sup>1\*</sup>, C. BARMAN<sup>1</sup>, S. DOLEY<sup>1</sup>, G. KADIRVEL<sup>1</sup>, P.K. BHARTI<sup>1</sup>, V. SINGH<sup>2</sup>

Received 23.5.2014, Revised 15.6.2014, Accepted 28.6.2014

### ABSTRACT

An experiment was conducted with 24 healthy cyclic cows between 2<sup>nd</sup> to 5<sup>th</sup> lactation by under intensive system of rearing. The animals were randomly divided into Control (C) and Experiment (E) groups with 12 animals in each The control and experiment groups were again further subdivided in groups C<sub>1</sub> and C<sub>2</sub> & E<sub>1</sub> and E<sub>2</sub> comprising six in each. Onset of oestrus was considered as day 0 for superovulatory treatment. The animals of groups C<sub>1</sub> and C<sub>2</sub> & E<sub>1</sub> and E<sub>2</sub> were injected with 5 ml NSS (i/ml) & 21µg GnRH (i/m) (Buserelin acetate) on day 8 of the oestrous cycle, respectively. Each of the experimental animals of groups C, and E, was superovulated with 400 mg FSH (i/m) in divided and equal doses at 12 h intervals for four days on day 10 to 13 of the oestrous cycle and consequently, each of the experimental animals of groups C, and E, was also superovulated with 2000 I.U.PMSG injection (i/m) as single dose on day 10 of the oestrous cycle. PGF,  $\alpha$  2ml was injected (i/m) at morning time on day 12 of oestrous cycle. At the superovulatory oestrus, all the animals were bred three times at 12 h intervals using frozen semen. The animals were examined per rectally on day 6 of the induced oestrus after first insemination to detect the superovulatory response. Flushing was performed on day 7 of induced oestrus to retrieve embryos from the superovulated cows. In the present experiment, superovulatory response in GnRH treated groups of cows (groups  $E_1$  and  $E_2$ ) were found to be better as compared to that of the animals of control groups but the duration of induced oestrus (h) recorded in groups C<sub>2</sub> (49.02  $\pm$  0.07 h) and E<sub>2</sub> (50.40  $\pm$  0.27 h) were found to be significantly higher (P<0.05) than the value recorded in groups  $C_1$  and  $E_1$ .

Keywords: Follicle Stimulating Hormone, GnRH, PMSG

#### **INTRODUCTION**

Superovulation is a key step in the embryo transfer technology in cattle and requires administration of a gonadotropin preparation that mimics the effect of follicle stimulating hormone (FSH). It is an important and integral part of the fast developing multiple ovulation and embryo transfer (MOET) programmes. Superovulation is done by application of gonadotropin like porcine pituitary FSH (Barman et al. 2012) with or without luteinizing hormone in mithun cows or pregnant mare serum gonadotropin (Saumande and Chupin 1981) with or without luteinizing hormone in cattle. The success of embryo transfer technique depends largely on the reliable method for the induction of superovulation to harvest maximum number of normal fertile ova or embryos. Although various advances have been achieved in superovulation technique, it still remains a weak link in the chain of events influencing the final outcome of embryo transfer in cattle. Beneficial effects of prostaglandins as well as GnRH have been well documented and are now being used for improvement of reproductive efficiency in large ruminants (Kumar et al. 2010). Therefore, the present study was conducted to determine the effect of presuperovulatory treatment of buserelin on

<sup>1</sup>Division of Livestock Production, ICAR Research Complex for NEH Region, Umroi Road, Umiam-793103, Meghalaya <sup>2</sup>Dept. of Animal Reproduction, Gynaecology & Obstetrics, College of Veterinary Sciences & A.H, DUVASU, Mathura-281001, U.P. \*Corresponding author's E-mail: suresh\_vet079@rediffmail.com

superovulatory response and embryo recovery in cattle under subtropical conditions of Meghalaya in order to understand their reproductive physiology.

#### MATERIALS AND METHODS

The present experiment was conducted at dairy unit, ICAR Research Complex for NEH Region, Umiam, Meghalaya. Twenty four healthy cyclic cows between 2<sup>nd</sup> and 5<sup>th</sup> lactation, free from genital abnormalities on per rectal examination, having no previous history of infectious diseases were selected for the present experiment. Experimental animals were maintained under intensive system of rearing and they were examined regularly to ascertain their health condition and strict hygienic measures were taken in the shed. The experimental animals were divided in two groups viz. Control (C) and Experiment (E) comprising twelve in each. The control group was again further subdivided in groups C<sub>1</sub> and C<sub>2</sub> comprising six in each and similarly experiment group (E) was also further subdivided into  $E_1$  and  $E_2$  comprising six in each. Onset of oestrus was considered as day 0 for superovulatory treatment. The animals of groups C<sub>1</sub> and C<sub>2</sub> & E<sub>1</sub> and E<sub>2</sub> were injected with 5 ml NSS (i/ml) & 21µg GnRH (i/m) (Buserelin acetate) on day 8 of the oestrous cycle, respectively. Each of the experimental animals of groups C<sub>1</sub> and E<sub>1</sub> was superovulated with 400 mg FSH (i/m) (Folltropin, Veterpharma Inc, Canada) in divided and equal doses at 12 h intervals for four days on day 10 to

13 of the oestrous cycle. The treatment schedule for superovulation with FSH 400 mg was as 50/50, 50/50, 50/50 and 50/50 mg respectively and consequently, each of the experimental animals of group C<sub>2</sub> and E<sub>2</sub> was also superovulated with 2000 I.U.PMSG injection (i/m) (Folligon, Intervet International Gmbh, Germany) as single dose on day 10 of the oestrous cycle. PGF<sub>2</sub>á 2ml (Cloprostenol, Intervet International Gmbh, Germany) was injected (i/m) at morning time on day 12 of oestrous cycle. At the superovulatory oestrus, all the animals were bred three times at 12 h intervals using frozen semen. The animals were examined per rectally on day 6 of the induced oestrus after first insemination to detect the superovulatory response. Flushing of uterine horns for non-surgical embryo collection using modified Dulbecco's phosphate buffer saline (mDPBS) with 1% heat treated oestrus cow serum was performed on day 7 of induced oestrus to retrieve embryos from the superovulated cows according to the method described by Neto et al. (2005). Data obtained from the experiment were statistically analyzed as per the method described by Snedecor and Cochran (2007).

### **RESULTS AND DISCUSSION**

The results of the present experiment are being presented in Table 1 and 2. Animals responding to superovulatory treatment in groups  $C_1$ ,  $C_2$ ,  $E_1$  and  $E_2$  were 100%, 83.33%, 100% and 83.33%,

 Table 1: Effect of presuperovulatory treatment of buserelin on superovulatory response and embryo recovery in cattle

Treatment groups	No. of animals	Oestrus response (%)	Mean ovulation rate (Mean ± SE)	Total embryos recovery (Mean ± SE)	Viable embryos (Mean ± SE)	Anovulatory follicles (Mean ± SE)	Degenerated embryos (Mean ± SE)	Unfertilized ova (Mean ± SE)
C <sub>1</sub> (5 ml NSS+FSH: 400 mg)	6	100.00	6.83 <sup>NS</sup> ±1.01	4.17 <sup>NS</sup> ±0.91	2.5 <sup>NS</sup> ±0.80	0.67 <sup>NS</sup> ±0.49	0.67 <sup>NS</sup> ±0.49	1.0 <sup>NS</sup> ±0.44
C <sub>2</sub> (5 ml NSS+PMSG 2000 I.U.)	6	83.33	5.33 <sup>NS</sup> ±1.11	1.83 <sup>NS</sup> ±0.79	1.67 <sup>NS</sup> ±0.65	1.83 <sup>NS</sup> ±0.79	0.17 <sup>NS</sup> ±0.16	0.5 <sup>NS</sup> ±0.49
$E_1$ (21 µg Buserelin acetate + FSH:	6	100.00	8.17 <sup>NS</sup> ±1.30	4.83 <sup>NS</sup> ±1.22	2.33 <sup>NS</sup> ±0.20	0.67 <sup>NS</sup> ±0.20	1.67 <sup>NS</sup> ±0.40	0.15 <sup>NS</sup> ±0.16
400 mg) E <sub>2</sub> (21 μg Buserelin acetate +PMSG 2000 I.U.)	6	83.33	5.5 <sup>NS</sup> ±1.06	2.83 <sup>NS</sup> ±1.64	1.83 <sup>NS</sup> ±1.16	0.83 <sup>NS</sup> ±0.30	0.83 <sup>NS</sup> ±0.54	0.17 <sup>NS</sup> ±0.16

<sup>NS</sup>: Non-significant

Treatment groups	No. of animals	Oestrus response (%)	Duration of induced oestrus (hrs.) (Mean±SE)	Interval from induced oestrus to the onset of subsequent natural oestrus (days) (Mean±SE)	Duration of natural oestrus (hrs.) (Mean±SE)
C <sub>1</sub> (5 ml NSS+FSH: 400 mg)	6	100.00	43.05ª±1.15	20.02 <sup>NS</sup> ±0.68	24.85 <sup>NS</sup> ±0.03
C <sub>2</sub> (5 ml NSS+PMSG 2000 I.U.)	6	83.33	49.02 <sup>b</sup> ± 0.07	20.15 <sup>NS</sup> ±0.15	26.06 <sup>NS</sup> ±0.22
E <sub>1</sub> (21 μg Buserelin acetate + FSH: 400 mg)	6	100.00	44.25 <sup>ac</sup> ±0.03	22.5 <sup>NS</sup> ±1.38	25.50 <sup>NS</sup> ±0.25
E <sub>2</sub> (21 µg Buserelin acetate +PMSG 2000 I.U.)	6	83.33	50.40 <sup>b</sup> ± 0.27	21.08 <sup>NS</sup> ±0.27	27.50 <sup>NS</sup> ±0.10

**Table 2:** Effect of presuperovulatory treatment of buserelin on occurrence of induced and natural oestrus in cattle

Means with different superscripts in a column differ significantly (P<0.05), NS-non significant

respectively. 100 percent oestrus response was observed in animals of groups  $C_1$  and  $E_1$ . These findings were in close agreement with the earlier report of Barman et al. (2012). However, the report of Mathur et al. (2006) on oestrus response following superovulation in Frieswal cows was 75 per cent only.

In the present experiment, superovulatory response in GnRH treated groups of cows were found to be better as evidenced by apparently lower number of palpable follicles as well as apparently higher numbers of corpora lutea as compared to that of the animals of control groups. This might be due to increased growth and maturation of follicles by combined effect of GnRH and FSH/ PMSG (Nilchuen et al. 2011) or due to follicle luteinisation and possible ovulation followed by the emergence of new follicular wave (Schmidt et al. 1996) or also due to the involvement of some factors including progesterone profile of donor at the time of superovulation, increased number of follicular population and endogenous hormonal milieu (Arora et al. 1996).

The numbers of unovulated follicles obtained in animals of group  $C_2$  were apparently higher as compared to groups  $C_1$ ,  $E_1$  and  $E_2$  but analysis of variance revealed non-significant differences in respect of anovulatory follicles between GnRH (buserelin acetate) treated and control groups. This might be due to insufficient endogenous LH surge (Taneja et al. 1988). The number of embryos recovered were found to be apparently higher in animals of group  $E_1$  (4.83  $\pm$  1.22) which might be due to increased growth and maturation of follicles caused by combined effect of GnRH (Buserelin acetate) and FSH and also due to genetic variation (Nilchuen et al. 2011) but statistically found to be non-significant (P>0.05) between the treated and control groups.

The duration of induced oestrus (h) recorded in the present experiment were as  $43.05 \pm 1.15$ , 49.02 $\pm 0.07, 44.25 \pm 0.03$  and  $50.40 \pm 0.27$ , respectively for groups  $C_1$ ,  $C_2$ ,  $E_1$  and  $E_2$ . Although apparently higher value was observed in group E<sub>1</sub> as compared to the value recorded in group C<sub>1</sub> but analysis of variance revealed no significant difference (P>0.05) between groups  $C_1$  and  $E_1$ . Similar findings have been also recorded by Bhuyan et al. (2006). The value recorded in group E, were found to be apparently higher than the value recorded in group  $C_2$  but no significant differences (P>0.05) were found between the two respective groups in respect of duration of induced oestrus. The present findings were in close agreement with the earlier report of Abdoon et al. (2000). The duration of induced oestrus (h) recorded in groups C, and E, were found to be significantly higher (P<0.05) than the value recorded in groups  $C_1$  and  $E_1$ . This might be due to variations in duration of oestrus might be influenced by the method of treatment, dose, age, breed, season, cyclicity and stages of reproduction (Mughal et al. 1998) or could be attributed to

increased follicular growth caused by hormone pregnant mare serum gonadotropin (Abdoon et al. 2000).

The mean time intervals from induced oestrus to the onset of subsequent natural oestrus in the present experiment were  $20.02 \pm 0.68$ ,  $20.15 \pm 0.15$ ,  $22.5 \pm 1.38$  and  $21.08 \pm 0.27$  days, respectively for groups  $C_1$ ,  $C_2$ ,  $E_1$  and  $E_2$ . Although analysis of variance revealed no significant differences for time intervals recorded from induced oestrus to the onset of subsequent natural oestrus among the groups  $C_{1}$ ,  $C_2$ ,  $E_1$  and  $E_2$  but apparently longer time interval was recorded in group  $E_1 (22.5 \pm 1.38 \text{ days})$ followed by groups  $E_2$ ,  $C_2$  and  $C_1$ . This may be attributed to elevated concentrations of circulating P4 due to more numbers of CL. The high concentration of P4 exerts negative feedback effect on LH secretion; thereby causing the attenuation of follicular growth (Barman et al. 2012).

The mean duration of subsequent natural oestrus (h) recorded in the present experiment were 24.85  $\pm 0.03$ ,  $26.06 \pm 0.22$ ,  $25.50 \pm 0.25$  and  $27.50 \pm 0.10$ h, respectively for groups C<sub>1</sub>, C<sub>2</sub>, E<sub>1</sub> and E<sub>2</sub>. Although apparently higher values were observed in group E<sub>2</sub> followed by groups C<sub>2</sub>, E<sub>1</sub> and C<sub>1</sub> but analysis of variance revealed no significant differences for duration of subsequent natural oestrus among different groups. The present findings recorded in respect of duration of subsequent natural oestrus were in close agreement with the earlier report of Bhuyan et al. (2006).

#### ACKNOWLEDGEMENT

The authors wish to acknowledge the Director, ICAR Research Complex for NEH Region, Umiam, Meghalaya, for providing all necessary infrastructure facilities for carrying out the present experiment.

### REFERENCES

- Arora VK, Devanathan TG, Pattabiraman SR, Ealurin MJ (1996). Studies on superovulatory response in cows treated with PMSG or FSH during luteal phase of the estrous cycle. Indian J Anim Reprod 17: 9-12
- Abdoon ASS, Ahmed WM, Kandil OM, Shalaby SI (2000). Evaluation of superovulatory response in anoestrus and cyclic native cows in Egypt with emphasis on hormonal and blood biochemical changes. Vet Med J Giza 48: 185-196
- Bhuyan S, Chakravarty P, Sarmah BC, Goswami J, Dutta JC (2006). Oestrus response following superovulation and embryo recovery in cows. Ind Vet J 83: 168-170
- Barman C, Baruah A, Sarmah BK, Baruah KK, Sarmah BC, Chakravarty P, Dutta A, Deka BC (2012). Occurrence of natural oestrus in mithun cows following superovulatory treatment. Indian Vet J 89: 49-51
  - Kumar S, Chandra R, Haque N, Toppo S, Rahman H (2010). Effect of GnRH and  $PGF_2 \acute{a}$  on Uterine Involution and Post partum Fertility in Cross-bred Cattle. Indian J Anim Sci 80:1175-1178
- Mughal DH, Aleem M, Saeed MA, Khan AH (1998). Comparison of two methods of oestrus synchronization in Sahiwal cows. International J Anim Sci 13: 223-226
- Mathur A K, Kumar PS, Tyagi S (2006). Superovulatory response and embryo recovery in Frieswal cows using FSH and PMSG. Ind J Anim Reprod 27: 80-81
- Neto ASC, Sanches BV, Binelli M, Seneda MM, Perri SH, Garcia JF (2005). Improvement in embryo recovery using double uterine flushing. Theriogenology 63: 1249-1255
- Nilchuen P, Rattanatabtimtong S, Chomchai S (2011). Superovulation with different doses of follicle stimulating hormone in Kamphaeng Saen beef cattle. Songklanakarin J Sci Technol 33: 679-683
- Park SJ, Ryu IS, Son DS, Choi SH, Baek KS, Park SB, Jeon BS, Ahn BS, Kim HS, Kim IH (2007). The effects of consecutive superovulation in Hanwoo and Holstein cattle. J Embryo Trans 22: 101-105
- Saumande J, Chupin D (1981). Production of PMSG antiserum in cattle: Assay of inhibitory activity and use in superovulated heifers. Theriogenology 15: 108-109
- Schmidt EJP, Drost M, Diaz P, Roomes C, Thatcher WW (1996). Effect of a gonadotropin releasing hormone agonist on follicle recruitment and pregnancy rate in cattle. J Anim Sci 74: 154-161
- Snedecor GW, Cochran WG (2007). Statistical Methods. 8<sup>th</sup> edition, Iowa University Press Ames, Iowa 50010, USA
- Taneja VK, Nanda SK, Datta TK, Bhat PN (1988). Embryo transfer in buffalo: present status and future research needs. II World Buffalo Congress, pp 603-609