SHORT COMMUNICATION Key role for enkephalinergic tone in cortico-striatalthalamic function

Marylou V. Solbrig,^{1,2} George F. Koob³ and W. Ian Lipkin⁴

¹Department of Neurology, Gillespie Neuroscience Bldg, Rm 3107, University of California-Irvine, Irvine, CA 92697-4292, USA 2 Department of Pharmacology; University of California-Irvine, Irvine, CA 92697, USA

³Department of Neuropharmacology; The Scripps Research Institute, La Jolla, CA 92037, USA

⁴Centre for Immunopathogenesis and Infectious Diseases, Mailman School of Public Health; Columbia University, New York, NY 10032, USA

Abstract

Whereas the role of dopaminergic tone in the cortico-striatal-thalamic system is well-established, the role of endogenous opioids in the function of this system is less understood. We show that Borna disease virus infection of adult rats results in an increase in preproenkephalin transcripts in the striatum of Borna-infected rats, a region important for forming coordinated sequential motor actions and in developing programmes of thought and motivation. Stereotypic behaviours and dyskinesias, the clinical hallmarks of infection in adult Lewis rats (BD rats), are accompanied by a disrupted pattern of immediate early gene c-fos activation in the motor thalamus, with significance for the breakdown in coordinated sequential motor actions. We also find increased preproenkephalin in infected cultured neuroblastoma and rat foetal glial cells. The expression pattern of enkephalin mRNA in vivo and in vitro suggest that increased enkephalin function is one of the neuropharmacological means by which Borna disease virus causes motor disease of animals and possibly cognitive and affective disease in man, and further suggest that enkephalins play a critical role in the maintenance of a balanced tone of activity in the cortico-basal ganglia-thalamo-cortical loops.

Introduction

Borna disease virus (BDV) is a neurotropic negative-strand RNA virus, worldwide in distribution, that causes disturbances of movement and behaviour in a wide range of animal species (Narayan et al., 1983; Ludwig et al., 1988; Solbrig et al., 1994). The virus is a natural pathogen of several domestic mammalian and bird species, and has been linked by serology, detection of viral nucleic acid and virus isolation to neuropsychiatric disorders of man (de la Torre, 2001; Lipkin et al., 2001). Increased central nervous system enkephalin expression was a reported feature of early infection in rats, within 3 weeks of infection (Fu et al., 1993). The purpose of the present study was to characterize the effect of BDV infection on CNS enkephalin expression in specific components of the cortico-striatal-thalamic system in rats at the time their movement and behaviour disorder appeared.

Materials and methods

Adult male Lewis rats infected experimentally with this virus (BD rats) develop a movement and behaviour disorder characterized by stereotypic patterns of grooming, sniffing, rearing, gnawing, selfbiting and dyskinesias (vacuous chewing and retrocollis) 6 weeks after infection (Solbrig et al., 1994). Preproenkephalin (PPE) mRNA expression in brain was examined by in situ hybridization after

Received 9 July 2002, revised 22 August 2002, accepted 28 August 2002

6 weeks of infection, using $[^{35}S]$ cRNA probes synthesized from PPE cDNA clones (Yoshikawa et al., 1984) following published methods (Solbrig et al., 1994). All procedures were performed on animals anaesthetized deeply with inhaled methoxyflurane in compliance with institutional (UCI IACUC-University of California-Irvine Institutional Animal Care and Use Committee) and National Institutes of Health guidelines.

Results

The PPE mRNA signal was higher in the caudate putamen of BD rats than in uninfected rats (Fig. 1A). Autoradiograms were analysed using a computer-based image analysis system (MCID; MicroComputer Imaging Device, Imaging Research Inc., St. Catharines, Ontario, Canada) with calibration curves constructed using \int_1^{14} C]-polymer standards (American Radiolabelled Chemicals, St. Louis, MO, USA). The PPE signal, expressed as d.p.m./mg, was increased significantly in the caudate putamen of BD rats relative to uninfected rats (BD 6633.87 \pm 876.81 vs. normal 3775.98 \pm 743.26; values in d.p.m./mg, mean \pm SEM; $t = 2.486$, d.f. = 10, $P < 0.05$, two-tailed *t*-test, $n = 6$ per group). Increases in signal were also found in nucleus accumbens, and central and basolateral amygdala. Increased caudate putamen signal was attributed to increased in situ hybridization signal per cell. Numbers of cells expressing enkephalin mRNA per high power ($280 \times 176 \mu$ m) field were similar in BD and normal rats (approximately 35–40 per field); however, numbers of silver grains per cell were increased throughout the caudate putamen in BD rats (Fig. 1B).

Correspondence: Dr M. Solbrig, ¹Department of Neurology, as above. E-mail: msolbrig@uci.edu

PPE

 $\alpha \mathbf{BD}$

 α met-enk

merge

Double-labelling immunofluorescence histochemistry showed colocalization of virus and methionine (met)-enkephalin, a biologically active product of the PPE gene, in caudate putamen (Fig. 1C). A total of 63 \pm 4% of cells expressing BDV phosphoprotein also expressed met-enkephalin; $37 \pm 4\%$ of met-enkephalin-expressing cells also expressed BDV phosphoprotein (mean \pm SEM of counts obtained from 306 BDV-infected cells, 544 met-enkephalin cells in three animals). The increase in PPE expression in BD rat brain (Fig. 1A and B) and coincidence of BDV and met-enkephalin immunoreactivity (Fig. 1C) is consistent with the hypothesis that infection directly influences met-enkephalin expression.

The effect of infection on c -fos expression was examined by in situ hybridization as an index to integrity of circuitry. Instead of the wellcircumscribed ventrolateral nuclear signal in the thalamus of uninfected rats, BD rats had a chaotic pattern of gene activation in the ventrolateral region, with irregular clustering of c -fos signal in rostral and central thalamic nuclei, and increased pallidal signal (Fig. 1D). Although cortico-basal ganglia-thalamo-cortical loops participate in the learning and maintenance of sequential motor actions dependent on sensorimotor integration (Graybiel & Rauch, 2000a), our results are consistent with enkephalin overexpression effecting a desynchronization of signal through the motor thalamus, breaking up motor programmes of coordinated sequential actions.

In vitro effects of BDV on PPE transcription in BDV (strain V) infected cell lines (Pauli & Ludwig, 1985) were examined by Northern hybridization. Fifteen micrograms total RNA extracted from cells in Tri-Reagent (Molecular Research Centre Inc., Cincinnati, OH, USA) were size-fractionated in 2.2 M formaldehyde/1% agarose gels, transferred to nylon membranes, UV crosslinked and hybridized to random primed $[32P]$ DNA fragments generated from cloned DNA representing enkephalin or BDV sequence. Levels of PPE mRNA, the enkephalin precursor, were increased in two neural cell lines infected with BDV: LAN (human neuroblastoma) and C6 (rat astroglia). The differential effect on enkephalin was more pronounced in LAN cells than C6 cells (Fig. 2A). Total RNA loaded per lane was standardized using a probe for the cellular gene glyceraldephyde-3-phosphate dehydrogenase (GAPDH) and autoradiographic signal quantified by phosphorimaging (Storm 840 Phosphorimager; Molecular Dynamics, Sunnyvale, CA, USA). Persistent infection of LAN cells caused an increase in PPE mRNA of 2.4-times control (uninfected) values (Fig. 2B).

Specificity of viral effects for PPE in striatum was assessed by measuring levels of dopamine D2 receptor, c-fos and preprodynorphin (PPD) mRNA in caudate putamen by in situ hybridization using $\sqrt{35}$ S]cRNA probes synthesized from dopamine D2 receptor (Bunzow et al., 1988), c-fos (Curran et al., 1987), and PPD (Civelli et al., 1985) clones. Levels were similar in BD and uninfected rats for D2 receptor (BD 3282.96 \pm 362.53 d.p.m./mg vs. normal 3489.67 \pm 556.24 p;d.p.m./mg; $t = 0.3113$, $P > 0.05$), c -fos (BD 84.14 \pm 1.01 d.p.m./ mg vs. 74.97 \pm 11.94 d.p.m./mg, $t = 0.7653$, $P > 0.05$) and dynor-

phin (BD 2997.21 \pm 44.78 d.p.m./mg vs. 3266.65 \pm 254.61 d.p.m./ mg, $t = 1.042$, $P > 0.05$).

Although the basis for enkephalin induction by BDV remains unknown, a potential mechanism is suggested by in vitro experiments indicating that BDV biases cells toward kinase reactions through mitogen-activated protein kinase pathway activation (Hans et al., 2001; Planz et al., 2001). These reactions, hypothesized to enhance viral replication and infectivity (Planz et al., 2001), would terminate with phosphoCREB (cyclic AMP response element binding protein) elevations, increasing transcription factors important for striatal enkephalin expression (Konradi et al., 1993), in turn increasing enkephalin mRNA. Induction could be restricted to enkephalin because there is limited infection of striatal dynorphin-expressing cells. Although partial dopamine denervation occurs in BD rats (Solbrig et al., 1994), increases in both striatal dopamine D2 receptor and PPE mRNA that accompany complete 6-hydroxydopamine deafferentation (Gerfen et al., 1990) are not seen. Thus, enkephalin

FIG. 2. Borna disease virus infection results in increased levels of PPE mRNA in neural cell lines. (A) Northern hybridization analysis of total RNA extracted from uninfected (C6) or infected (C6/V) rat glioma cells, uninfected (LAN) or infected (LAN/V) human neuroblastoma cells, or uninfected Lewis rat nucleus accumbens (NA). Membranes were hybridized with random primed $[^{32}P]$ DNA probes for detection of transcripts encoding preproenkephalin (top panel), Borna disease virus phosphoprotein (middle panel), and then stained with Coomassie blue for detection of 28 S rRNA and 18 S rRNA (lower panel) as an index of RNA concentration and integrity. (B) Northern hybridization analysis of total RNA extracted from uninfected (LAN) or infected (LAN/V) cells for detection of transcripts encoding PPE (top panel), Borna disease virus phosphoprotein (second panel), or GAPDH (third panel). Hybridization signal for RNA encoding preproenkephalin in LAN/V and LAN cells was quantified following normalization to signal for GAPDH (lowest panel).

FIG. 1. Functional neuroanatomy of Borna disease virus infection in rat. (A) Borna disease virus infection results in increased levels of preproenkephalin mRNA in rat striatum. In situ hybridization analysis of coronal sections through uninfected or infected striatum. Sections were hybridized with $[^{35}S]$ cRNA probes for detection of preproenkephalin transcripts, opposed to film, quantified using MicroComputer Imaging Device software and [¹⁴C]-standards. (B) Light micrographs of emulsion-dipped, Nissl-stained sections employed in Fig. 1A. Scale bar, 50 µm. (C) Immunofluorescence labelling of striatal coronal sections, using mouse anti-BDV phosphoprotein (gift of L. Stitz) and rabbit anti met-enkephalin antibodies (Chemicon, Temecula, CA, USA), FITCconjugated donkey anti-mouse IgG and Texas Red-conjugated goat anti-rabbit IgG (Rockland, Gilbertsville, PA, USA). Borna disease virus shown in green; met-enkephalin, red; colocalization of virus and met-enkephalin, yellow. (D) Borna disease virus infection results in a disorganized, dispersed pattern of immediate early gene c-fos mRNA expression in motor thalamus. Coronal sections were hybridized with a $[^{35}S]cRNA$ probe for detection of c-fos transcripts, apposed to film and imaged by MCID. VL, ventrolateral thalamic nucleus; GP, globus pallidus.

upregulation as a result of pure dopamine lesion is excluded. Although manipulations that drive phosphorylating kinase reactions to phosphoCREB might induce both striatal PPE (Konradi et al., 1993) and PPD (Cole *et al.*, 1995), only 14 \pm 1% of cells expressing dynorphin B express BDV phosphoprotein (values represent mean \pm SEM of counts obtained from 214 BDV-infected cells, 173 dynorphin B cells in three animals) whereas $37 \pm 4\%$ of metenkephalin-expressing cells also express BDV phosphoprotein. Thus, virus-driven kinase reactions are more likely to influence striatal expression of met-enkephalin than dynorphin.

Our analyses of the BD rat model are consistent with a role for enkephalin upregulation in the production of stereotypic behaviours. Enkephalin analogues are known stimulants of motor behaviour (Stinus et al., 1980; Kalivas et al., 1983). Morphine sensitization produces oral stereotypies (Pollock & Kornetsky, 1996) and endomorphin-1 injected into the globus pallidus induces orofacial dyskinesias in rats (Mehta et al., 2001), behaviours similar to those observed in the context of BDV infection.

In the normal striatum, a greater percentage of neurons in patches/ striosomes than matrix contain enkephalin (Gerfen & Young, 1988). Assuming a similar striosome : matrix distribution for enkephalin in BD rats, the most parsimonious interpretation of data presented here is that viral infection has resulted in increased activity of the striosomal compartment relative to the matrix. A striosome-dominant gene activation pattern is another hypothesized determinant of stereotypy (Graybiel et al., 2000b).

These results with a neurotropic virus demonstrate a key role for enkephalinergic tone in the highly sensitive and exquisitely balanced cortico-basal ganglia-thalamo-cortical loops. By increasing transcription or stability of an enkephalin precursor mRNA, BDV causes a behavioural disease characterized by repetitive or fragmented motor actions. Further evaluation of pathways and mechanisms of the viral effect on neuropeptide expression should lead to enhanced understanding of the role of neuropeptides in movement disorders, and of neuropeptides in the aetiopathology (Gillberg et al., 1985; Engel & Rocha, 1992) and treatment of neuropsychiatric diseases.

Acknowledgements

This work was supported by National Institutes of Health Grant DA 00376 (MVS) and NS 29425 (WIL). We thank Lothar Stitz for BDV antibodies, Wei Tang for technical assistance and Frances Leslie for helpful discussions. Animal care and handling procedures were in compliance with Institutional (UCI IACUC) and National Institutes of Health guidelines. This is publication number 14358-NP from The Scripps Research Institute.

Abbreviations

BDV, Borna disease virus; CREB, cyclic AMP response element binding protein; GAPDH, glyceraldephyde-3-phosphate dehydrogenase; PPD, preprodynorphin; PPE, preproenkephalin.

References

- Bunzow, J.R., van Tol, H.H.M., Grandy, D.K., Albert, P., Salon, J., Christie, M., Machida, C.A., Neve, K. & Civelli, O. (1988) Cloning and expression of a rat D2 dopamine receptor cDNA. Nature, 336, 783-787.
- Civelli, O., Douglass, J., Goldstein, A. & Herbert, E. (1985) Sequence and expression of the rat prodynorphin gene. Proc. Natl. Acad. Sci. USA, 82, 4291±4295.
- Cole, R.L., Konradi, C., Douglass, J. & Hyman, S.E. (1995) Neuronal

adaptation to amphetamine and dopamine: molecular mechanisms of prodynorphin gene regulation in the rat striatum. Neuron, 14, 813-823.

- Curran, T., Gordon, M.B., Rubino, K.L. & Sambucetti, L.C. (1987) Isolation and characterization of c-fos rat complementary DNA and analysis of posttranslational modification in vitro. Oncogene, 2, 79-82.
- de la Torre, J.C. (2001) Bornaviridae. In Knipe, D.M. & Howley, P.M. (eds), Fields Virology, 4th edn, Vol. 2. Lippincott, Williams & Wilkins, Philadelphia, pp. 1669-1677.
- Engel, J. Jr & Rocha, L.L. (1992) Interictal behavior disturbances: a search for molecuar substrates. In Engel, J. Jr, Wasterlain, C., Cavalheiro, E.A., Heinemann, U. & Avanzini, G. (eds), Molecular Neurobiology of Epilepsy. Elsevier Science Publishers, New York, pp. 341-350.
- Fu, Z.F., Weihe, E., Zheng, Y.M., Schafer, M.K.H., Sheng, H., Corisdeo, S. & Rauscher, F.J. (1993) 3rd, Koprowoski, H. & Dietzschold, B. Differential effects of rabies and Borna disease viruses on immediate-early and lateresponse gene expression in brain tissues. J. Virol., 67, 6674–6681.
- Gerfen, C.R., Engber, T.M., Mahan, L.C., Susel, Z., Chase, T.N., Monsma, F.J. Jr & Sibley, D.R. (1990) D1 and D2 dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons. Science, 250, 1429±1432.
- Gerfen, C.R. & Young, W.S. III (1988) Distribution of striatonigral and striatopallidal peptidergic neurons in both patch and matrix compartments: an in situ hybridization histochemistry and fluorescent retrograde tracing study. Brain Res., 460, 161-167.
- Gillberg, C., Terenius, L. & Lonnerholm, G. (1985) Endorphin activity in childhood psychosis. Arch. Gen. Psychiatry, 42, 780-783.
- Graybiel, A.M., Canales, J.J. & Capper-Loup, C. (2000b) Levodopa-induced dyskinesias and dopamine-dependent stereotypies: a new hypothesis. Trends Neurosci., 23, S71-S77.
- Graybiel, A.M. & Rauch, S.L. (2000a) Toward a neurobiology of obsessivecompulsive disorder. Neuron, 28, 343-347.
- Hans, A., Syan, S., Crosio, C., Sassone-Corsi, P., Brahic, M. & Gonzalez-Dunia, D. (2001) Borna disease virus persistent infection activates mitogenactivated protein kinase and blocks neuronal differentiation of PC12 cells. J. Biol. Chem., 276, 7258-7265.
- Kalivas, P.W., Widerlov, E., Stanley, D., Breese, G. & Prange, A.J. (1983) Jr Enkephalin action on the mesolimbic system: a dopamine-dependent and a dopamine-independent increase in locomotor activity. J. Pharm. Exp. Ther., 227, 229±237.
- Konradi, C., Kobierski, L.A., Nguyen, T.V., Heckers, S. & Hyman, S.E. (1993) The cAMP-response-element-binding protein interacts, but Fos protein does not interact, with the proenkephalin enhancer in rat striatum. Proc. Natl. Acad. Sci. USA, 90, 7005-7009.
- Lipkin, W.I., Hornig, M. & Briese, T. (2001) Borna disease virus and neuropsychiatric disease-a reappraisal. Trends. Microbiol., 9, 295-298.
- Ludwig, H., Bode, L. & Gosztonyi, G. (1988) Borna disease: a persistent virus infection of the central nervous system. Prog. Med. Virol., 35, 107-151.
- Mehta, A., Bot., G., Reisine, T. & Cheesselet, M.-F. (2001) Endomorphin-1: Induction of motor behavior and lack of receptor desensitization. J. Neurosci., 21, 4436-4442.
- Narayan, O., Herzog, S., Frese, K., Scheefers, H. & Rott, R. (1983) Behavioral disease in rats caused by immunopathological responses to persistent Borna virus in the brain. Science, 220 , $1401-1403$.
- Pauli, G. & Ludwig, H. (1985) Increase of virus yields and releases of Borna disease virus from persistently infected cells. Virus Res., 2, 29-33.
- Planz, O., Pleschka, S. & Ludwig, S. (2001) MEK-speicific inhibitor U0126 blocks spread of Borna disease virus in cultured cells. J. Virol., 75, 4871±4877.
- Pollock, J. & Kornetsky, C. (1996) Reexpression of morphine-induced oral stereotypy six months after last morphine sensitization dose. Pharm. Biochem. Behav., 53, 67-71.
- Solbrig, M.V., Koob, G.F., Fallon, J.H. & Lipkin, W.I. (1994) Tardive dyskinetic syndrome in rats infected with Borna disease virus. Neurobiol. $Dis., 1, 111-119.$
- Stinus, L., Koob, G.F., Ling, N., Bloom, F.E. & LeMoal, M. (1980) Locomotor activation induced by infusion of endorphins into the ventral tegmental area: Evidence for opiate-dopamine interactions. Proc. Natl. Acad. Sci. USA, 77, 2323-2327.
- Yoshikawa, K., Williams, C. & Sabol, S.L. (1984) Rat brain preproenkephalin mRNA: cDNA cloning, primary structure, and distribution in the central nervous system. J. Biol. Chem., 259, 14301-14308.