ata, citation and similar papers at <u>core.ac.uk</u>

brought to yo

provided by Repository of the A

CHEMICALLY MODIFIED CHITOSAN COATINGS: WETTING AND ELECTROCHEMICAL STUDIES

PÉTER MÁRTON^a, EMŐKE ALBERT^a, NORBERT NAGY^b, BORBÁLA TEGZE^a, GABRIELLA STEFÁNIA SZABÓ^{c*}, ZOLTÁN HÓRVÖLGYI^{a*}

ABSTRACT. Native chitosan coatings were prepared on glass and zinc substrates by dip-coating method. The native coatings were chemically modified: crosslinked with glutaraldehyde and sodium-tripolyphosphate and then silvlated with dichlorodimethylsilane. The native layers prepared on zinc substrate were acylated with acetic anhydride and impregnated with indigo carmine (IC). Native coatings on different substrates showed different morphology and physical structure, which were explored by AFM studies. Examining the wetting properties of the coatings, it was found that the chemical modification can form a stable, hydrophobic (advancing contact angle of *ca.* 97°) and water repellent ($H_{\Theta} = 2^{\circ}$) layer on the glass substrate, while the same modifications reduce the hydrophobic nature and stability of the coating on zinc (contact angle decreases from ca. 100° to ca. 60°). Electrochemical studies of the coatings have shown that IC increases the corrosion protection (>90% inhibition efficiency) and reduces the permeability of the coating through ionic crosslinking, while acylation has the opposite effect and the acylated coating protection is worse than the native chitosan. The results can be used in the development of hydrophobic, water-repellent and temporary anti-corrosion coatings.

Keywords: chitosan coating, contact angles, electrochemical measurements

^a Budapest University of Technology and Economics, Faculty of Chemical Technology and Biotechnology, Department of Physical Chemistry and Materials Science, Centre for Colloid Chemistry, H-1521 Budapest, Hungary

^b Institute for Technical Physics and Materials Science, Centre for Energy Research, H-1525 Budapest, Hungary

^c Babeş-Bolyai University, Faculty of Chemistry and Chemical Engineering, 11 Arany Janos str., RO-400028, Cluj-Napoca, Romania

^{*} Corresponding authors: gszabo@chem.ubbcluj.ro, zhorvolgyi@mail.bme.hu

INTRODUCTION

Glucosamine based biomaterials which can be frequently found in the nature are important synthesis materials for numerous products [1] [2]. Chitosan is a glucosamine based biopolymer produced from chitin [3], which is used in many industries due to its biocompatibility, biodegradability and antibacterial properties, such as food industry [4], medicine [5] and agriculture [6].

Chitosan can also be used in composite materials [7] and coatings to develop drug release systems that are very important in pharmacology. The permeability of chitosan coatings and its dependence on the layer thickness can be studied in bilayer systems [8]. The incubated molecule release kinetics is investigated from the chitosan-covered porous layer that is impregnated with the model molecule (usually an ionic dye). In this case, it is important to characterize the permeability of the chitosan coating, and one of the possible ways to do this is by determining its pseudo-porosity value from electrochemical studies, including polarization curves [9]. Due to the structure of chitosan, it can be regarded as a pseudo-porous layer through which the charged species of the electrolyte solution can reach the metal surface. Using the suitable equations, we can deduce the permeability from the rate of ion transport and the resistance of the coating.

Related to this, another area of application for chitosan is in corrosion protection [10]. Although unmodified native chitosan does not have a remarkable protective effect, it can provide effective protection when chemically modified or impregnated with various organic inhibitors. An example of such an inhibitor is indigo carmine (IC) [11], which is a natural dye and thus offers an eco-friendly alternative to more conventional corrosion inhibitors. The protective effect of these chitosan-based coatings were studied on various metallic surfaces, such as aluminum alloys [12], steel [13], zinc [14] and titanium [15]. Due to their low barrier properties, these types of chitosanbased films are often developed for less demanding applications, such as temporary protective layers. These types of coatings are capable of reducing the quantity of corrosion damage during shipment [16].

As was mentioned before, the chemical modification (e.g. impregnation with inhibitors) is a way to improve the protective effect of the coatings. If the hydrophobicity of the surface can be increased by some methods, the corrosion process can also be slowed down, as one of its primary causes is moisture. The production of hydrophobized chitosan has been reported in several articles, mainly silyl- [17] [18] or long-chain carboxylic acid derivatives [19] were used as reagents. Acylation of chitosan, which is basically a process in which chitin is produced, is also common [20] [21].

The aim of this paper is to investigate the effect of the mentioned chemical modifications on the wettability and pseudo-porosity (and related permeability) of chitosan coatings on various substrates. The purpose of the chemical modification with dichlorodimethylsilane (DCDMS) was to form a chitosan layer with hydrophobic and water-repellent properties. The change in the pseudo-porosity and corrosion protection of the coatings by reacting with acetic anhydride and impregnation with IC was also studied.

RESULTS AND DISCUSSION

Optical spectroscopy analysis of coatings deposited on glass substrates

The deposited layers on both substrates were transparent and completely covered the surface without any cracks or inhomogeneity. The analysis of the transmittance curves (see Fig. 1) for coatings on glass (GIs) substrates revealed layer thickness of 360 ± 10 nm and a refractive index of 1.54 ± 0.01 .





AFM measurements

Fig. 2 shows the results of AFM measurements of the samples. The left image quartet shows the height (a) and phase image (b) of the bare glass substrate, and the height (c) and phase image (d) of the coated glass. The image quartet on the right is the same for the bare and coated zinc (Zn).

In the case of the bare glass substrate the surface irregularities are in the magnitude of 10 nm, while in the case of the much rougher Zn substrate there are height differences of 100 nm. The latter observation can be explained by the fact that the last step in the preparation of the metal substrates was polishing with 0.3 μ m Al₂O₃ powder. It can be seen, that the surface properties of the substrates also influenced the morphology of the prepared coatings: the roughness of the chitosan layer formed on the Zn surface is an order of magnitude larger. In addition, a finer structure also appears in the coatings on Zn, which is presumably not due to the roughness of the substrate but a result of the chitosan's own structure.

Studying the phase images of the samples, it can be observed that in the case of the film prepared on glass substrate, the surface is smooth, but structurally inhomogeneous: the phase image shows brighter areas (islands with stronger physicochemical interactions, marked with black arrow) with a diameter of 1-2 μ m. In contrast, the rougher surface on the Zn substrate has structural homogeneity. Presumably, the chitosan on the glass substrate separated into two domains with different surface properties before the gelation. A similar phenomenon was observed by Ye and Zao, who also reported in their paper the phase separation of a polymer layer [22]. It seems that the interactions between the molecules of the layer and the substrate greatly influence the structure and morphology of the final coatings.



Figure 2. Results of AFM measurements: Height (a, c) and phase (b, d) images of bare (a, b) and coated (c, d) glass substrates (left) and the same for Zn substrates (right).

Wettability of the coatings

1. Native coatings

The values of the advancing and receding contact angles and the contact angle hysteresis measured on the different substrates for each measuring liquid are given in Table 1.

Table 1. Results of the wettability measurements of native coatings on both
substrates (Θ_A : advancing contact angle, Θ_R : receding contact angle,
 H_{Θ} : contact angle hysteresis)

Sample	Measuring liquid	Θ _Α [°]	Θ _R [°]	Н₀ [°]
Gls/Chit	Distilled water	71 ± 9	68 ± 9	4 ± 1
	0.2 g/L	79 ± 1	57 ± 6	21 ± 5
Zn/Chit	solution	101 ± 8	74 ± 10	27 ± 8

For both cases significant contact angle hysteresis was found due to the chemical heterogeneity (21°, Gls/Chit) or the surface roughness (27°, Zn/Chit). Water contact angles, however, do not show significant hysteresis for samples Gls/Chit. Water is a better wetting agent than the aqueous salt solutions [23] and the quick penetration of water molecules into the layer can result in decreased surface microheterogeneity. However, it seems from the data, that this process manifests itself in a significant macroscopic heterogeneity demonstrated by a relatively high standard deviation of contact angle data (Table 1) and a significant deterioration of water contact angles on Gls/Chit during the investigated time period of 30 minutes (Fig. 3). Macroscopic heterogeneity was also observed with aqueous salt solutions for samples Zn/Chit but in that case the wetting stability of the chitosan layer was found to be much better (Fig. 3), which outlines the complexity of these processes.

In the case of Gls/Chit there is not much difference in the advancing contact angle values of the two measuring liquids, however, the value is below 90° in both cases, which indicates the hydrophilic nature of the surface. The contact angle hysteresis is low in the case of water, but much higher in the case of the Na₂SO₄ solution, so the surface has no water-repellent property. In the case of Zn/Chit, the advancing contact angle is well above 90°, but the hysteresis is also a high value, so the surface is hydrophobic, but does not show water-repellent effect.

The change in advancing contact angles over time is shown on Fig. 3. The results show that the contact angle on the coated Zn surface decreases very slowly below 90° , so it can be considered as a hydrophobic layer that is stable in time. However, in the case of the coated glass substrate the stability of the coating is poor, the contact angle decreases significantly over time.

The contact angle data help to interpret information obtained from the analysis of the AFM images (see Fig. 2). Knowing the measured contact angles on each surface, we can assume, that that the phase which produced a larger phase shift on the AFM images belongs to a more nonpolar structure, and on the Zn surface only that structure was formed. However, on the glass substrate a heterogeneous surface was formed (Fig. 2c) with smaller nonpolar domains (producing the same phase shift) surrounded by an area of presumably more polar character (producing a different, lower phase shift). These two phases together determine the wetting properties of the coating formed on glass. Knowing the value of the advancing contact angle of the Na₂SO₄ solution measured on the nonpolar surface (Zn/Chit), we can apply the Cassie-Baxter wetting model to determine the value of the advancing contact angle of the Na_2SO_4 solution on the polar phase formed on the glass substrate. The essence of the model is that on a heterogeneous surface consisting of areas with different wettability, the measurable contact angle can be calculated by a linear combination of the contact angles of each type of area [24].

$$\cos \Theta_{obs} = f \cdot \cos \Theta_1 + (1 - f) \cdot \cos \Theta_2 \tag{1}$$

where Θ_{obs} is the value of the observed contact angle (79°), Θ_1 (101°) and Θ_2 are the contact angles characterizing the nonpolar and polar domains of the surface, and *f* is the percentile area ratio of the nonpolar domains. This *f* parameter can be determined from the AFM phase images: 12%. The resulting contact angle value (Θ_2) for the polar part of the surface is 76°. This is not a big difference from the measurable value but the presence of the nonpolar domains can cause significant contact angle hysteresis as was mentioned above.

Thus, it can be seen that coatings of the same material prepared with the same technique on the various surfaces show greatly different wettability behavior. This may be explained by the different chemical and physical nature of the Zn and glass substrates which can influence the structure (surface polarity) and morphology of the deposited layers.



Figure 3. Change in advancing contact angle values over time for coatings on both substrates (In the case of glass substrates, the measurement was performed with distilled water, in the case of Zn substrates with 0.2 g/L Na₂SO₄ solution).

2. Silylated coatings

Preliminary investigations for the modified chitosan layers deposited on glass substrates

Fig. 4 shows the water droplet shapes and relating advancing contact angles on silylated chitosan coatings on glass substrates, measured 3, 5, 10, 20 and 40 seconds after the drop formation. For the native coating (Fig. 4a), the contact angles reach a relatively constant value (ca. 75°) after 10 s (also see Table 1). For the sample silylated without crosslinking (Fig. 4b), the contact angle initially exceeds 90°, but rapidly decreases below the value measured on the native coating. The same is observed for the coating crosslinked with pentasodium-tripolyphosphate (TPP), then silylated (Fig. 4c), but the decrease is smaller. However, the sample crosslinked with glutaraldehyde (GDA), then silylated (Fig. 4d) keeps a contact angle above 90° in the investigated 5minute time range.

For interpreting the above observations, it should be taken into consideration that the silylation reaction with DCDMS produces HCl, which diffuses into the layer. When the droplet is placed onto the surface, in the aqueous medium the acid can protonate the free amino groups of chitosan forming a much more polar (i.e. hydrophilic) coating. In addition, the diffusion of the hydrophilic parts of the polymer chains to the surface become faster in

the aqueous medium (Fig. 4b). The crosslinkers, however, react with the amino groups of chitosan: TPP forms ionic bonds between the protonated amino groups in aqueous solution, and GDA is covalently bound to the nitrogen atoms. In both cases, the basicity of the amino groups decreases significantly, and the HCI is no longer able to protonate them, so that the contact angle does not decrease to the previously seen extent (Fig. 4c and d). In addition, crosslinking reduces the mobility of the polymer chains upon contact with the aqueous phase, therefore the hydrophobic silyl groups can stay on the surface while maintaining their effect. The results show that the covalently crosslinking agent (GDA) is better than the other one: temporal stability was only experienced for the case of GDA crosslinking (Fig. 4d).



Figure 4. Water droplets on different surfaces: advancing contact angles at the indicated time after droplet formation (a: native chitosan, b: chitosan, silylated with DCDMS, c: chitosan, cross-linked with TPP, silylated with DCDMS d: chitosan, cross-linked with GDA, silylated with DCDMS)

Comparative investigation of the wetting properties of modified chitosan layers deposited on glass and Zn substrates

The advancing and receding water and Na₂SO₄ solution contact angles measured for the GDA-crosslinked and DCDMS-modified samples on glass (Gls/Chit-GDA-SilyI) and Zn substrates (Zn/Chit-GDA-SilyI) are shown in Table 2. All samples were silylated in the vapour phase of the n-hexane solution of DCDMS. Comparing the data in Tables 1 and 2, it can be observed that in the case of Gls/Chit-GDA-SilyI, the silylation improved the water-repellent

properties of the coating: the water contact angle is above 90° and the value of the hysteresis is very small, which means that the surface is hydrophobic and water-repellent. The water contact angles show a relatively good stability in the investigated time range (Fig. 5a). It is important to note, however, that the samples lose their advantageous properties by soaking them for 10 minutes in water: the value of the water contact angles decrease below the values measured on the native chitosan. However, the silylation improved the wettability of the chitosan layer on the Zn substrate compared to the wettability of the native chitosan layer, and the wetting stability also deteriorated (Fig. 5b). Interestingly, by increasing the time of silylation the resulting water contact angles will be even smaller (Fig. 5b).

The different behaviour of chitosan coatings on the glass and Zn substrates during the cross-linking and silylation processes can be attributed to the different polarity of the pertinent native coatings. During the cross-linking process the hydrophilicity of the native coatings on glass can further increase due to the chemisorption of GDA [8]. Furthermore, as this process is carried out in aqueous phase, the reorientation of the polymer segments is facilitated and this results in the increase of the surface concentration of hydroxyl groups. The efficiency of the silylation can be attributed to the latter process. The reorientation process is presumably hindered for coatings on Zn substrates due to the hydrophobic character of the surface. Water molecules cannot penetrate the outside of the layer fast enough and so the mobility of the polymer segments is lower. Hence, in this case the surface polarity increase can only happen due to chemisorption of GDA.

Table 2. Wettability results of coatings cross-linked with GDA and silylated with
DCDMS under various conditions (*after 10 min swelling in distilled water and
drying) (Θ_A : advancing contact angle, Θ_R : receding contact angle, H_{Θ} : contact
angle hysteresis)

Sample	Silylation time	Θa [°]	Θr [°]	Н₀ [°]
Cle/Chit CDA Silv/	30 min	97 ± 1	95 ± 3	2 ± 2
Gis/Chit-GDA-Gilyi	30 11111	48 ± 2*	31 ± 8*	16 ± 6*
Zp/Chit CDA Silvi	5 min	77 ± 15	57 ± 24	20 ± 13
Zh/Chit-GDA-Shyl	10 min	64 ± 10	47 ± 13	23 ± 5





Figure 5. Advancing contact angle values measured in time for chemically modified (cross-linked with GDA and silylated with DCDMS) coatings on glass (a) and Zn (b) substrates. (In the case of glass substrates, the measurement was performed with distilled water, in the case of Zn substrates with 0.2 g/L Na₂SO₄ solution).

Electrochemical measurements of the coatings

The linear and semi-logarithmic polarization curves for the investigated samples are represented in Fig. 6.



Figure 6. Linear (a) and semi-logarithmic (b) polarization curves for the bare Zn and the coated samples

In the Table 3. are introduced the OCP, the slopes of the fitted straight lines to the linear polarization curves and the calculated polarization resistance R_{p} . This is the inverse value of the straight line's slope, that was fitted to the linear portion of the curves. Further characteristics of the layers

like pseudo-porosity (P) can be calculated from the obtained data, using the following formula, assuming that the layer is electrochemically inert at low overpotentials [25]:

$$P = \frac{R_{p_s}}{R_p} \cdot 10^{-\left(\frac{|\Delta E_{corr}|}{b_a}\right)} \cdot 100 \ (\%) \tag{2}$$

The R_{ps} and R_p represent the polarization resistance of the bare Zn substrate and the coated Zn, respectively, while ΔE_{corr} symbolizes the difference between the potential of the coated and uncoated samples, and b_a is the anodic Tafel coefficient of the bare substrate.

Table 3. The polarization resistance R_p: determined from the linear polarization curves recorded for the bare Zn and the coated samples. B represents the slope of the fitted line, R² the coefficient of determination, N the number of the points, OCP the open circuit potential and P the pseudo-porosity

Sample	R² [-]	N [-]	10 ^{4.} Β [1/Ω]	R _Ρ [Ω]	P [%]	OCP [V]
Zn	0.999	25	7.31	1368	I	-0.987
Zn/Chit	0.999	27	4.67	2141	44	-0.944
Zn/Chit-IC	0.999	33	0.28	35714	4	-0.977
Zn/Chit-Acyl	0.999	27	6.56	1524	88	-0.973
Zn/Chit-Acyl-IC	0.998	23	1.63	6135	19	-0.999

If the value of the polarization resistance is high, the coating has a good anti-corrosion property. Vice versa if it is low, protective property of the coating is poor. As one can see from Table 3, the highest corrosion resistance R_p is attributed to the Chit-IC layer, indicating the best anti-corrosion effect. The corrosion current densities are proportional to the corrosion rate, which means that this value is low in the case of a good protective coating. In Table 4 the lowest value is attributed to the Chit-IC coating, this has the best anticorrosion property, which is in a very good agreement with the R_p obtained for this layer from the linear polarization.

The low porosity of the Chit-IC coating is correlated with a relatively denser texture.

Sample	i _{corr} [μΑ/cm²]	E _{corr} [V]	b _a [V/dec]	b _c [V/dec]	IE [%]
Zn	8.945	-0.982	0.272	0.029	-
Zn/Chit	5.667	-0.939	0.144	0.057	36.6
Zn/Chit-IC	0.388	-0.992	0.040	0.105	95.7
Zn/Chit-Acyl	7.064	-0.979	0.036	0.097	21.0
Zn/Chit-Acyl-IC	0.688	-0.999	0.067	0.209	92.3

Table 4. Polarization parameters obtained by Tafel interpretation of the
semi-logarithmic polarization curves: icorr: corrosion current density,
Ecorr corrosion potential, ba and bc anodic and cathodic
Tafel coefficients. IE represent the inhibition efficiency

For the calculation of the inhibition efficiency (IE) formula (3) is used:

$$IE = \frac{i_{corr}^s - i_{corr}}{i_{corr}^s} \cdot 100$$
(3)

where i_{corr}^s and i_{corr} are the corrosion current densities of the Zn substrate and the coated sample, respectively. From the IE value calculated for the native chitosan (Chit) coating, 36,6%, it can be deduced that the native chitosan layer has a very poor anti-corrosion effect. In the case of IC impregnated chitosan coating (Chit-IC) the corrosion current density decreased with one order of magnitude while the IE increase significantly (95,7%). This means that this kind of coating has a good anti-corrosion property. The acylation of both the native Chit and Chit-IC causes the decay of the anti-corrosion properties, namely the increase of the corrosion current density and the decrease of the IE. Altough in the case of the acylated, impregnated chitosan coating (Chit-Acyl-IC) the i_{corr} is better than in the case of native chitosan, still the Chit-IC has the best IE.

Considering also the calculated pseudo-porosity values from Table 3, one can say that this kind of difference in behavior is due to the permeability of the thin layers. It can be seen, that the Chit-IC has the smallest porosity and for this reason has the best anti-corrosion property. The increasing order of the porosity in case of the other coatings is Chit-Acyl-IC, Chit and at last the Chit-Acyl with a very high value (88%).

The explanation of this phenomenon lays in the structure of the coatings. The IC molecules contain negatively charged groups and can crosslink the chitosan, forming ionic bonds with the protonated amino groups. The layer formed in this way has a reduced permeability. By acylation, the number

of the amino groups that can be protonated is significantly reduced, subsequently the extent of the crosslinked portions are reduced too and the permeability against the corrosive environment is increased. In this way the anti-corrosion properties of the coating are diminished. It can be assumed, that as a consequence of the acylation H bonds also cannot be formed between the amino groups. This could be the reason why the acylated coating protection is worse that even that of the native chitosan. The sterical effect of the acyl-groups can also be the cause of weaker structure formation. The acylating reaction conditions can induce the damage of the materials.

CONCLUSIONS

Wetting and electrochemical properties of chemically modified chitosan coatings prepared on glass and zinc substrates were investigated. The purpose of the research was to develop a water-repellent and corrosion protective polymer layer.

Wettability studies of native coatings have shown that the chitosan layer prepared on glass substrate is hydrophilic and unstable in time, while on the zinc substrate a highly hydrophobic and stable in time but non-waterrepellent coating is formed under the same conditions. AFM analysis of the layers detected surfaces with different morphologies and signs of phase separation of chitosan on the glass substrate.

Examination of the wettability of the silylated coatings revealed that of the systems studied, only the samples crosslinked with GDA and then chemically modified with DCDMS provided a surface with satisfactory properties. In this way, a coating with hydrophobic and water-repellent properties, which is very stable in time, has been produced on a glass substrate, however, on zinc substrates the silylation greatly deteriorates the properties of the originally hydrophobic layer.

Electrochemical studies have shown that among the chemical modification methods studied, impregnation with IC improves the anticorrosion effect of chitosan coatings which is explained by the cross-linking effect of IC. The acylation, however, greatly diminished the corrosion protection by increasing the pseudo-porosity of the coatings.

EXPERIMENTAL SECTION

MilliQ water (Millipore Simplicity 185, Resistance: 18,2 MΩcm); sulfuric acid (Carlo Erba, 96%, f. a.); isopropanol (Reanal Labor, 99,7%, f. a.); acetic acid (Lachner, 99,8% f. a.); chitosan (Sigma-Aldrich, Medium molecular weight, Viscosity: 598 cP); glutaraldehyde (Reanal Labor, 25%, purum); pentasodium-tripolyphosphate-pentahydrate (Sigma-Aldrich, ≥ 98%, purum); dichlorodimethylsilane (Reanal Labor, ≥99%, f. a.); n-hexane (Merck, 95%, f. a.); indigo carmine (Reanal Labor, f. a.); acetic anhydride (Reanal, f. a.); methanol (Molar Chemicals Kft.).

The coatings were prepared on glass microscope slides (Menzel-Gläser) and zinc plates (Bronzker Bt.) with a dip coater made by Plósz Engeneering Ltd.

Coatings preparation

The glass substrates were cleaned using aqueous detergent solution, 10% (w/w) sulfuric acid, isopropanol and MilliQ water. The Zn substrates were polished with emery paper (P1200, P2000, and P3000) and Al_2O_3 powder (particle size: 0,3 µm). Then they were washed with isopropanol and MilliQ water and ultrasonicated for 10 min to remove any powder residue. Before the dip-coating, 0.1 M HCl solution was used to clean the surface of any remaining zinc-oxide.

The dip-coating was performed with a withdrawal speed of 5 cm/min from 1% (w/w) chitosan solution. The chitosan was dissolved in 1% (v/v) acetic acid solution, stirred for 24 hours, then centrifuged (30 min, 4000 rpm) to remove any insoluble chitosan residue due to imperfect deacetylation. The prepared coatings were dried overnight at room temperature.

Chemical modification of the coatings

Some samples were chemically modified by acylation and/or impregnation with IC. Other samples were modified by cross-linking agents and/or silylation.

The acylation of the coatings was carried out in 10% (v/v) methanolbased acetic anhydride solution for 60 min. Afterwards, the samples were washed with methanol and dried at room temperature.

The impregnation was performed using the dip-coater: the samples were immersed in 0.01 M IC solution with 10 cm/min immersion speed, kept in the solution for 15 min, then withdrawn with 10 cm/min.

Two different cross-linking agents (covalent, ionic) were tried. The coatings on glass substrates were covalently cross-linked in 5% (w/w) aqueous glutaraldehyde solution for 5 min, while the coatings on Zn substrates were cross-linked in 1% (w/w) glutaraldehyde solution for 30 min, because the more concentrated solution damaged the coating on Zn. The ionic cross-linking by pentasodium-tripolyphosphate took place in a 3% (w/w) aqueous solution for 10 min. Then the samples were washed with distilled water and dried at room temperature.

For the silylation reaction the samples were kept in a desiccator. The silylating agent was an n-hexane solution of dichlorodimethylsilane. For the samples on glass 10% (w/w) solution and a reaction time of 30 min were used; while for the samples on Zn 5% (w/w) n-hexane solution with reaction times of 5 and 10 min were used. Afterwards, the samples were kept at room temperature in air for 10 min to remove any left-over reagent.

Transmittance measurements

The layer thickness and refractive index values of the coatings on glass substrates were determined by a thin-layer optical model [26] fitted to the transmittance spectra. The transmittance was measured using an Analytik Jena Specord 200 UV-Vis spectrophotometer in the 350-900 nm wavelength range with 1 nm resolution and a measurement speed of 10 nm/s.

AFM measurements

To explore surface properties, the samples were measured with an AIST-NT SmartSPM 1000 AFM device in tapping mode with a PPP-NCHR-20 needle made by NanoSensors (nominal radius of the needle <20 nm). In the range of 10×10 and 20×20 μ m a height and phase image of the surfaces were recorded.

Wettability measurements

Contact angle measurements were performed on the native and on the silvlated coatings deposited on glass and Zn substrates to get information about the effect of chemical modification on the surface polarity. The measurements were carried out with 0.2 g/L aqueous Na₂SO₄ solution, which was also used as electrolyte during electrochemical investigations. MilliQ water was also used for the characterization of coatings deposited on glass substrates. Wettability was studied by a Drop Shape Analysis System (DSA30, Krűss GmbH). Advancing contact angles were measured by the sessile drop method by the drop-build up technique at 25 °C and 90% humidity within the first minute after the droplet formation (20 µl). Then receding contact angles were also measured after removing ca. half the volume of the droplet. Contact angle hysteresis was determined as the difference of the advancing and receding angles. In certain cases, the advancing contact angles of 20 µl droplets were studied during a 30minute time interval in order to characterize the stability of coatings. On some selected samples the contact angle values were determined after they were soaked in distilled water for 10 min and then dried.

Electrochemical measurements

The corrosion resistance of the coatings was investigated in an undivided electrochemical cell containing three electrodes, connected to the Metrohm Autolab PGSTAT 301N potentiostat. The coated Zn samples were used as working electrode, with a 2 cm² active surface area, while the Platinum wire and the Ag/AgCl, KCl (saturated) were the counter and the reference electrode, respectively. The aggressive environment consisted of a Na₂SO₄ solution with 0.2 g/L concentration. All measurements were carried out at room temperature.

First, for each sample the open circuit potential (OCP) was determined, and every measurement lasted for 60 min. The potentiodynamic polarization was performed in a potential range of OCP±20 mV and OCP±200 mV vs Ag/AgCl, KCl(saturated) electrode and the obtained curves were recorded with 0.166 mV/s rate. By Tafel interpretation of the semi-logarithmic polarization curves, namely by fitting straight lines to the anodic and cathodic branches, kinetic parameters (corrosion current densities, corrosion potentials, Tafel coefficients) were determined.

ACKNOWLEDGMENTS

The research reported in this paper was supported by the BME Nanotechnology and Materials Science TKP2020 IE grant of NKFIH Hungary (BME IE-NAT TKP2020). The research work has been accomplished in the framework of the "BME R+D+I project", supported by the grant TÁMOP 4.2.1/B-09/1/KMR-2010–0002. NRDI TNN-123631, NRDI K-128266, and NRDI FK-128901 grants are acknowledged. Emőke Albert's research work was supported by the European Union and the State of Hungary, co-financed by the European Social Fund in the framework of TÁMOP-4.2.4.A/2–11/1–2012–0001 "National Excellence Program"

REFERENCES

- 1. Z. Sebestyén, E. Jakab, A. Domán, P. Bokrossy, I. Bertóti, J. Madarász, K. László, *J. Therm. Anal. Calorim.* **2020**, 1-8.
- 2. B. Duan, Y. Huang, A. Lu, L. Zhang, Prog. Polym. Sci. 2018, 82, 1–33.
- 3. S. Kumari, S.H. Kumar Annamareddy, S. Abanti, P. Kumar Rath, *Int. J. Biol. Macromol.* **2017**, *104* 1697–1705.

- 4. S. Tripathi, G.K. Mehrotra, P.K. Dutta, *Int. J. Biol. Macromol.* **2009**, *45*, 372–376.
- 5. T. Dai, M. Tanaka, Y.Y. Huang, M.R. Hamblin, *Expert Rev. Anti. Infect. Ther.* **2011**, *9* 857–879.
- 6. P.L. Kashyap, X. Xiang, P. Heiden, Int. J. Biol. Macromol. 2015, 77, 36-51
- 7. T. V. Podust, T. V. Kulik, B.B. Palyanytsya, V.M. Gun'Ko, A. Tóth, L. Mikhalovska, A. Menyhárd, K. László, *Appl. Surf. Sci.* **2014**, *320*, 563–569.
- 8. M. Dabóczi, E. Albert, E. Agócs, M. Kabai-Faix, Z. Hórvölgyi, *Carbohydr. Polym*, **2016**, *136*, 137–145.
- 9. C. Liu, Q. Bi, A. Leyland, A. Matthews, Corros. Sci. 2003, 45, 1257-1273
- 10. M.L. Zheludkevich, J. Tedim, C.S.R. Freire, S.C.M. Fernandes, S. Kallip, A. Lisenkov, A. Gandini, M.G.S. Ferreira, *J. Mater. Chem.* **2011**, *21*, 4805–4812.
- 11. Á.F. Szőke, G.S. Szabó, Z. Hórvölgyi, E. Albert, L. Gaina, L.M. Muresan, *Carbohydr. Polym.* **2019**, *215*, 63–72.
- 12. O. Lundvall, M. Gulppi, M.A. Paez, E. Gonzalez, J.H. Zagal, J. Pavez, G.E. Thompson, *Surf. Coatings Technol.* **2007**, *201* 5973–5978.
- 13. S.A. Umoren, M.J. Banera, T. Alonso-Garcia, C.A. Gervasi, M. V. Mirífico, *Cellulose*, **2013**, *20* 2529–2545.
- 14. Á.F. Szőke; G. Szabó; Z. Simó; Z. Hórvölgyi; E. Albert; A.G. Végh; L. Zimányi; L.M. Muresan; *Eur. Polym. J.*, **2019**, *118*, 205–212.
- 15. J. Redepenning; G. Venkataraman; J. Chen; N. Stafford; *J. Biomed. Mater. Res. Part A.*, **2003**, *66*, 411–416.
- 16. J. Carneiro; J. Tedim; S.C.M. Fernandes; C.S.R. Freire; A. Gandini; M.G.S. Ferreira; M.L. Zheludkevich; *ECS Electrochem. Lett.*, **2013**, *2*, C19.
- 17. K. Kurita, M. Hirakawa, S. Kikuchi, H. Yamanaka, J. Yang, *Carbohydr. Polym.* **2004**, *56*, 333–337.
- 18. A. Tiwari, J. Inorg. Organomet. Polym. Mater. 2009, 19, 361–366.
- 19. S. Höhne, R. Frenzel, A. Heppe, F. Simon, *Biomacromolecules*. **2007**, *8*, 2051–2058.
- 20. J. Xu, S.P. McCarthy, R.A. Gross, D.L. Kaplan, *Macromolecules*. **1996**, *29*, 3436–3440.
- 21. K.Y. Lee, W.S. Ha, W.H. Park, *Biomaterials*. **1995**, *16*, 1211–1216.
- 22. Z. Ye, X. Zhao, J. Microsc. 2010, 238, 27–35.
- 23. Z. Hórvölgyi, M. Máté, M. Zrínyi, *Colloids Surfaces A Physicochem. Eng. Asp.* **1994**, *84*, 207–216.
- 24. E. Bormashenko, *Colloids Surfaces A Physicochem. Eng. Asp.* **2008**, 324, 47–50.
- 25. B. Matthes, E. Broszeit, J. Aromaa, H. Ronkainen, S.P. Hannula, A. Leyland, A. Matthews, *Surf. Coatings Technol.* **1991**, *49*, 489–495.
- 26. E. Hild, A. Deák, L. Naszályi, Ö. Sepsi, N. Ábrahám, Z. Hórvölgyi, *J. Opt. A Pure Appl. Opt.* **2007**, *9*, 920–930.