

## Marathoners' Breathing Pattern Protects Against Lung Injury by Mechanical Ventilation: An Ex Vivo Study Using Rabbit Lungs

Yoshiaki Oshima,\*† Naoto Okazaki,\* Kazumi Funaki,\* Akihiro Otsuki,\* Shunsaku Takahashi,\*‡ Tomomi Harada\*§ and Yoshimi Inagaki\*

\*Division of Anesthesiology and Critical Care Medicine, Department of Surgery, School of Medicine, Faculty of Medicine, Tottori University, Yonago 683-8504, Japan, †Department of Anesthesiology, Yonago Medical Center, Yonago 683-0006, Japan, ‡Department of Anesthesiology, Tottori Prefectural Central Hospital, Tottori 680-0901, Japan, and §Department of Anesthesiology, Ehime Prefectural Central Hospital, Matsuyama 790-0024, Japan

### ABSTRACT

**Background** Breathing during a marathon is often empirically conducted in a so-called “2:2 breathing rhythm,” which is based on a four-phase cycle, consisting of the 1st and 2nd inspiratory and the 1st and 2nd expiratory phases. We developed a prototype ventilator that can perform intermittent positive pressure ventilation, mimicking the breathing cycle of the 2:2 breathing rhythm. This mode of ventilation was named the marathoners' breathing rhythm ventilation (MBV). We hypothesized that MBV may have a lung protective effect.

**Methods** We examined the effects of the MBV on the pulmonary pre-edema model in isolated perfused rabbit lungs. The pulmonary pre-edema state was induced using bloodless perfusate with low colloid osmotic pressure. The 14 isolated rabbit lung preparations were randomly divided into the conventional mechanical ventilation (CMV) group and MBV group, (both had an inspiratory/expiratory ratio of 1/1). In the CMV group, seven rabbit lungs were ventilated using the Harvard Ventilator 683 with a tidal volume (TV) of 8 mL/kg, a respiratory rate (RR) of 30 cycles/min, and a positive end-expiratory pressure (PEEP) of 2 cmH<sub>2</sub>O for 60 min. In the MBV group, seven rabbit lungs were ventilated using the prototype ventilator with a TV of 6 mL/kg, an RR of 30 cycles/min, and a PEEP of 4 cmH<sub>2</sub>O (first step) and 2 cmH<sub>2</sub>O (second step) for 60 min. The time allocation of the MBV for one cycle was 0.3 s for each of the 1st and 2nd inspiratory and expiratory phases with 0.2 s of intermittent resting between each phase.

**Results** Peak airway pressure and lung wet-to-dry ratio after 60 min of ventilation were lower in the MBV group than in the CMV group.

**Conclusion** MBV was considered to have a lung-protective effect compared to CMV.

**Key words** 2:2 breathing rhythm; lung-protective ventilation strategy; marathon; perfused lung; ventilator-induced lung injury

Breathing during a marathon is often empirically conducted in a “2:2 breathing rhythm,”<sup>1</sup> which is based on cycles composed of four phases: the 1st and 2nd inspiratory phases, and the 1st and 2nd expiratory phases, which are synchronized with the step rate. For example, a left footstep coincides with the 1st inspiration and expiration phase, and a right footstep coincides with the 2nd inspiration and expiration phase. This continues in a sequential manner. Matching rhythmic steps with inspiration and expiration means that the inspiratory/expiratory (I/E) ratio is 1/1 for the 2:2 breathing rhythm.

Mechanical ventilation causes lung damage, a pathological condition called ventilator-induced lung injury (VILI).<sup>2, 3</sup> VILI is a factor that increases the mortality rate in acute respiratory distress syndrome (ARDS).<sup>2</sup> The main factor that causes VILI is alveolar overdistension (volutrauma) and cyclic reopening of collapsed alveoli (atelectrauma).<sup>4</sup> The ventilation strategy to protect the lungs against deterioration in ARDS is to prevent additional volutrauma and atelectrauma of the remaining normal lung tissue.<sup>5</sup> Recently, low tidal volume (TV) ventilation has been established as a lung-protective ventilation strategy; however, it has the disadvantage of PaCO<sub>2</sub> retention.<sup>6</sup>

During exercise, there is a risk of hypoxemia and pulmonary edema.<sup>7–9</sup> In intense exercise, cardiac output increases as the demand for O<sub>2</sub> increases, and the right ventricular pressure and pulmonary artery pressure

Corresponding author: Yoshiaki Oshima, MD, PhD

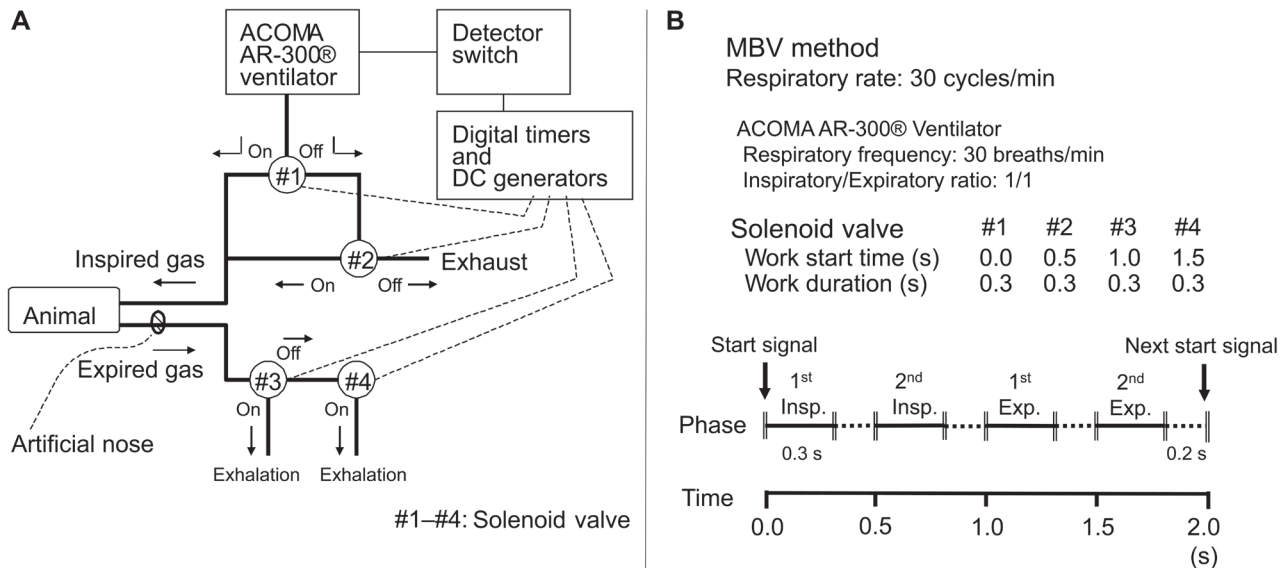
yoshimatomo@gmail.com

Received 2020 July 27

Accepted 2020 September 1

Online published 2020 October 5

Abbreviations: ARDS, acute respiratory distress syndrome; BALF, bronchoalveolar lavage fluid; CMV, conventional mechanical ventilation;  $\Delta$ OD, change in optical density; HFOV, high-frequency oscillatory ventilation; I/E, inspiratory/expiratory; IRV, inverse ratio ventilation; MBV, marathoners' breathing rhythm ventilation; mPaw, mean airway pressure; MPO, myeloperoxidase; PEEP, positive end-expiratory pressure; pPaw, peak airway pressure; PV, pressure-volume; RR, respiratory rate; TV, tidal volume; VILI, ventilator-induced lung injury; W/D, lung wet-to-dry ratio



**Fig. 1.** Schematic diagram of the prototype ventilator (A). The setting of the Acoma AR-300 and an example of the time allocation regarding the four solenoid valves, when the respiratory rate is set to 30 cycles/min in the MBV method (B). In the MBV method, the 1st and 2nd inspiratory, and the 1st and 2nd expiratory phases taken together, are defined as one cycle. DC, direct current; Exp, expiratory; Insp, inspiratory; MBV, marathoners' breathing rhythm ventilation.

rise.<sup>10</sup> The effectiveness of the 2:2 breathing rhythm on cardiopulmonary function during a marathon has not yet been proven in a study; however, the 2:2 breathing rhythm is empirically favored in marathon runners, presumably because it prevents the progression of additional hypoxemia and pulmonary edema during the marathon. Moreover, since CO<sub>2</sub> is not retained during a marathon, where the metabolism is increased over an extended time, it has been suggested that the 2:2 breathing rhythm provides sufficient capacity for CO<sub>2</sub> elimination.<sup>11</sup>

In this study, we developed a prototype ventilator that can perform intermittent positive pressure ventilation, mimicking the breathing cycle of the 2:2 breathing rhythm (Fig. 1). This mode of ventilation, named the marathoners' breathing rhythm ventilation (MBV), is a mechanical ventilation method that divides the TV required for conventional mechanical ventilation (CMV) into the inspiratory volume of the 1st and 2nd inspiratory phases. Since the prototype ventilator was designed to deliver a constant inspiratory flow rate, and the durations of both the 1st and 2nd inspiratory phases were set to be the same, the inspiratory volumes of the two phases were equal. Thus, each inspiratory volume can be reduced in the MBV. Therefore, MBV can be considered a ventilation mode that has the features of low TV ventilation.

This study focused on the effects of MBV on the pulmonary pre-edema model in isolated perfused rabbit

lungs. The pulmonary pre-edema state was induced by using bloodless perfusate with low colloid osmotic pressure.

## MATERIALS AND METHODS

All experimental protocols regarding the use and care of animals in this study were approved by the Laboratory Animal Care Committee of the Faculty of Medicine at Tottori University. These experimental protocols were performed from 2004 to 2006. Adult female Japanese white rabbits (2.2–3.1 kg) were purchased from Oriental Yeast Co. (Tokyo, Japan). The rabbits were kept under standard housing conditions with free access to food and water.

### Materials

#### Development of the prototype ventilator

We developed a prototype ventilator that can perform intermittent positive pressure ventilation and mimic the breathing cycle of the 2:2 breathing rhythm during a marathon (Fig. 1). The prototype ventilator is a combination of digital timers, solenoid valves, and existing respiratory equipment (ACOMA AR-300, Acoma Co., Tokyo, Japan). The ACOMA AR-300 is a volume-limited, time-cycling ventilator, in which the I/E ratio can be adjusted from 1/3 to 3/1, and the TV can be set from 20 to 300 mL. The ACOMA AR-300 is a piston-driven ventilator, designed to deliver a constant inspiratory flow rate. The I/E ratio of the ACOMA AR-300

was set to 1/1. The detector switch was attached to the piston rod of the ACOMA AR-300 and the switch was set to emit a signal each time it detected the start of the inspiratory phase of the ACOMA AR-300. The signal was sent to four digital timers (H5BR-B, Omron Co., Tokyo, Japan). Each of these timers was connected to a solenoid valve: three tridirectional valves (VDW350-6G-3-01, SMC Co., Tokyo, Japan) and one bidirectional valve (VDW31-6G-3-01, SMC Co.). These four solenoid valves were powered by 12-Volt DC supplied by the connected digital timer. All digital timers were reset as soon as the ACOMA AR-300 inspiratory start signal was received, at which point they started to measure elapsed time. The start time and duration of the 12-Volt DC output (controlling the solenoid valves) could be set individually for each of the four digital timers.

A schematic diagram of the prototype ventilator is shown in Fig. 1A; Fig. 1B shows the setting of the Acoma AR-300 and an example of the time allocation of the four solenoid valves when the respiratory rate (RR) is set to 30 cycles/min in the MBV method. An example of the solenoid valve operation and the corresponding inspired and expired gas dynamics follow. One cycle was defined as the sum of eight separate phases, including the 1st inspiratory phase, a resting phase, the 2nd inspiratory phase, a resting phase, the 1st expiratory phase, a resting phase, the 2nd expiratory phase, and a final resting phase. In this example, the respiratory frequency of the ACOMA AR-300 was set to 30 breaths/min. In response to the inspiratory start signal of the ACOMA AR-300, all digital timers were reset and began to record the time. First, the solenoid valve #1 opened for 0.3 s, and gas from the ventilator flowed directly into the inspiratory circuit (1st inspiratory phase: 0.3 s). When solenoid valve #1 closed, the gas moved towards solenoid valve #2, but since solenoid valve #2 was closed, the gas was released into the atmosphere from the exhaust pipe (resting phase). After 0.5 s from the start signal, solenoid valve #2 was opened for 0.3 s, and the gas from the ventilator went into the inspiratory circuit again (2nd inspiratory phase: 0.3 s). When solenoid valve #2 was closed, the gas was released into the atmosphere from the exhaust pipe of solenoid valve #2 (resting phase). After 1.0 s from the start signal, solenoid valve #3 was opened for 0.3 s, and expired gas was released from the exhalation pipe of this valve into the atmosphere (1st expiratory phase: 0.3 s). When solenoid valve #3 was closed, the expired gas moved towards solenoid valve #4, but since solenoid valve #4 was closed, the emission of the expired gas into the atmosphere was interrupted (resting phase). After 1.5 s from the start signal, solenoid valve #4 was opened for 0.3 s, and the

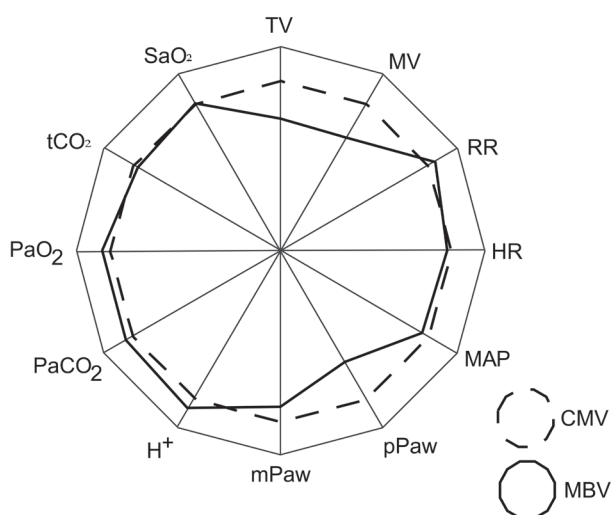
expired gas was released from the exhalation pipe of solenoid valve #4 into the atmosphere (2nd expiratory phase: 0.3 s). When solenoid valve #4 was closed, the emission of the expired gas into the atmosphere was interrupted (resting phase). Two seconds after the start signal, the detector switch detected the next inspiratory cycle of the ACOMA AR-300, the digital timer was reset, and the MBV breathing cycle was repeated. In this example, it did not matter if the open duration of solenoid valve #4 was set to values above 0.5 s, because the detector switch detected the start of the next inspiratory cycle in the ACOMA AR-300. This terminated the open duration of valve #4 since the reset of the digital timers was prioritized; thus, the actual open duration of solenoid valve #4 was 0.5 s.

With this prototype ventilator, mechanical ventilation is produced in a 2:2 breathing rhythm, the RR can be adjusted in 5 cycles/min increments from 10 cycles/min to 65 cycles/min. In addition, the positive end-expiratory pressure (PEEP) can be set at arbitrary pressure values independently of the exhaust pipes of the solenoid valves #3 and #4. The ventilation volume of one MBV cycle is the difference between the TV set in the ACOMA AR-300 and the exhaust volume from the exhaust pipe of solenoid #2. Since this ventilator is in the prototype stage, the ventilation volume of one MBV cycle has to be determined from the average gas volume of several expiratory cycles collected by the classical water displacement method.<sup>12</sup>

### Preliminary study

A preliminary in vivo experiment was conducted assessing the physiological effects of the MBV method on intact lungs in rabbits ( $n = 9$ ). The rabbits were tracheostomized, anesthetized with pentobarbital, and paralyzed with pancuronium bromide. We measured arterial blood gas values, TV, minute volume, RR, airway pressure, etc. during the MBV method and the CMV method. The CMV method was conducted with a TV of 8 mL/kg using the Harvard Ventilator 683. The I/E ratio of the Harvard Ventilator 683 is fixed at 1/1 as the factory default setting. PEEP was not used for either of the ventilation methods to avoid its impact on the comparison of both the ventilation methods. The above parameters in each ventilation method were measured 20min after performing an alveolar recruitment maneuver in advance. In both the ventilation methods, the RR count was reflected in cycles/min.

Figure 2 shows the above-mentioned outcomes of each parameter in a radar chart. Normalization was performed using each average value of the CMV method. The pH was expressed as the hydrogen ion



**Fig. 2.** Radar chart of physiological parameters in the MBV method normalized with each value in the CMV method. The pH is expressed as the hydrogen ion concentration. CMV, conventional mechanical ventilation; H<sup>+</sup>, hydrogen ion concentration; HR, heart rate; MAP, mean arterial pressure; MBV, marathoners' breathing rhythm ventilation; mPaw, mean airway pressure; MV, minute volume; PaCO<sub>2</sub>, arterial carbon dioxide tension; PaO<sub>2</sub>, arterial oxygen tension; pPaw, peak airway pressure; RR, respiratory rate; SaO<sub>2</sub>, arterial oxygen saturation; tCO<sub>2</sub>, total CO<sub>2</sub> content; TV, tidal volume.

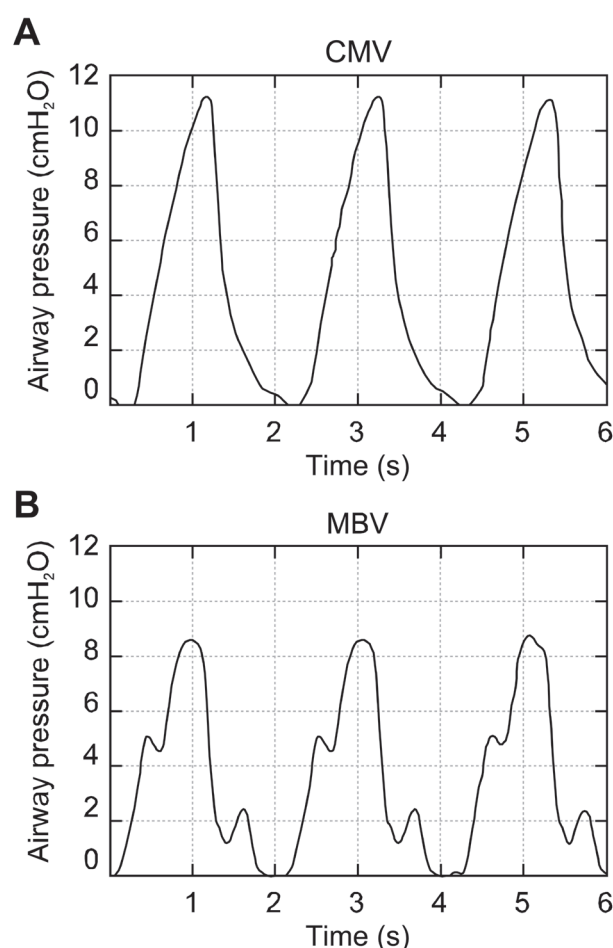
concentration. Compared to the CMV method, the MBV method was expected to effectively eliminate carbon dioxide with a smaller amount of TV and minute volume. This preliminary study was conducted on each rabbit in the form of a crossover design in which the CMV and MBV methods were tested in sequence. The randomization of the order in which the two ventilation methods were tested was not consistent. Therefore, no comparison of the differences in the mean values of each parameter was carried out between the two ventilation methods.

Figure 3 displays typical waveforms of the airway pressure for both mechanical ventilation methods with a PEEP of 0 cmH<sub>2</sub>O in the same rabbit. The waveform in Fig. 3A describes the airway pressure in the CMV method, whereas Fig. 3B presents the airway pressure curve in the MBV method. In the MBV method, the airway pressure transiently decreased immediately after the end of the 1st inspiratory phase and transiently increased immediately after the end of the 1st expiratory phase.

## Methods

### Isolated perfused lung preparation

We compared the effect of MBV with CMV on pre-pulmonary edema in isolated perfused rabbit lungs. The



**Fig. 3.** Typical waveforms of the airway pressure for the CMV (A) and MBV (B) methods in the same rabbit. In the MBV method, the airway pressure transiently decreases immediately after the end of the 1st inspiratory phase and transiently increases immediately after the end of the 1st expiratory phase. CMV, conventional mechanical ventilation; MBV, marathoners' breathing rhythm ventilation.

isolated perfused rabbit lungs were prepared using the method described in detail by Liu et al.<sup>13</sup> with minor modifications. Briefly, the rabbits were anesthetized with pentobarbital 30 mg/kg intravenously, followed by ketamine 25 mg/kg intramuscularly, and anticoagulation with heparin 500 u/kg intravenously. After local anesthesia of the anterior neck and the sternum region with 1% lidocaine, tracheal intubation was performed through a tracheostomy and the rabbits were ventilated mechanically. A median sternotomy was performed, and an incision was made into the right ventricle. The rabbits were euthanized by rapidly exsanguinating whole blood (70 mL) from the incision site in the right ventricle. The pulmonary artery and left atrium were cannulated via the right and left ventriculotomies,

respectively. Finally, the lungs were removed en bloc and enclosed in a humidified chamber.

The lungs were perfused with bicarbonate-buffered physiological salt solution (PSS), which comprised of NaCl, 119; KCl, 4.7; MgSO<sub>4</sub>, 1.17; NaHCO<sub>3</sub>, 22.61; KH<sub>2</sub>PO<sub>4</sub>, 1.18, and CaCl<sub>2</sub>, 3.2 mM in a recirculating manner. To every 100 mL of PSS stock solution, 100 mg dextrose, 20 mU insulin, 3 g Ficoll® PM70 (GE Healthcare Bio-Sciences, Little Chalfont, UK), and 2 mg indomethacin (Sigma Chemical, St. Louis, MO) were added. Ficoll® PM70 is a high molecular weight sucrose polymer with an average molecular weight of 70 000. If an isolated rabbit lung is perfused with Krebs Ringer solution supplemented with 4% (w/v) albumin without the addition of red blood cells to the perfusate, it will develop pulmonary edema within 2 h.<sup>14</sup> Ficoll® PM70 — as well as albumin — provides a normal colloidal osmotic pressure at 4% (w/v).<sup>15</sup> If isolated murine lungs are perfused with Dulbecco's Modified Eagle's Medium containing 4% (w/v) Ficoll® PM70 without the addition of red blood cells to the perfusate, then an edema will form in the lung interstitium within 1 h.<sup>16</sup> In this study, a pulmonary pre-edema model was created in the isolated rabbit lung via perfusion with bicarbonate-buffered PSS containing 3% (w/v) Ficoll® PM70 without the addition of red blood cells to the perfusate.

The perfusate flow rate was gradually increased to 35 mL/kg/min, while the ventilation gas was changed from air to a mixed gas (O<sub>2</sub>, 21%; CO<sub>2</sub>, 5%; N<sub>2</sub>, 74%). The perfusate flow rate was continuously measured with an electromagnetic flowmeter (MF-1200, Nihon Kohden, Tokyo, Japan). The left atrial pressure was set to 4 mmHg by regulating the height of a reservoir connected to the venous circuit. The lungs were ventilated with a TV of 6 mL/kg and a respiratory frequency of 40 breaths/min with a PEEP of 2 cmH<sub>2</sub>O using a Harvard Ventilator 683. Blood gases were measured using iSTAT. The perfusate pH was adjusted to 7.40 using an appropriate amount of 1 mM NaHCO<sub>3</sub> (Meylon, Otsuka Pharmaceutical, Tokyo, Japan), and the temperature of the perfusate was maintained at 38°C using a regulated heating system. The pulmonary arterial pressure, left atrial pressure, and airway pressure were recorded using a PowerLab system (AD Instruments, New South Wales, Australia; software, Chart ver. 5) with a transducer connected to amplifiers. The isolated perfused lungs were allowed to stabilize for 20 min prior to proceeding to the experimental protocol.

### Experimental protocol

The 14 isolated rabbit lung preparations were randomly divided into the CMV and MBV groups.

In the CMV group, rabbit lungs ( $n = 7$ ; body weight,  $2.5 \pm 0.2$  kg) were ventilated using a Harvard Ventilator 683 with a TV of 8 mL/kg, an RR of 30 cycles/min, and a PEEP of 2 cmH<sub>2</sub>O for 60 min.

In the MBV group, rabbit lungs ( $n = 7$ ; body weight,  $2.4 \pm 0.2$  kg) were ventilated using the prototype ventilator with a PEEP of 4 cmH<sub>2</sub>O (first step) and 2 cmH<sub>2</sub>O (second step) for 60 min. Before the experiment, the TV of the MBV was adjusted to 6 mL/kg using a test lung and measured by the water displacement method.<sup>12</sup> The mixed gas containing 5% CO<sub>2</sub> partially dissolves in water, so the volume of the gas decreases. Therefore, we measured in advance that the decrease in volume after exposure to water for 10 minutes was less than 0.05 mL per 10 mL. The MBV group had an RR of 30 cycles/min, and the time allocation for one cycle had the following pattern: 1st inspiratory phase: 0.3 s; resting phase: 0.2 s; 2nd inspiratory phase: 0.3 s; resting phase: 0.2 s; 1st expiratory phase: 0.3 s; resting phase: 0.2 s; 2nd expiratory phase: 0.3 s; and resting phase: 0.2 s.

### Peak airway pressure and mean airway pressure

Both peak airway and mean airway pressure (pPaw and mPaw, respectively) at the stabilization period (baseline) and after 60min of ventilation were calculated based on the values recorded by the PowerLab system.

### Pressure-volume curve measurement

The inflation pressure-volume (PV) curve was measured at the stabilization period (baseline) and after 60 min of ventilation using the quasi-static method and a syringe containing mixed gas, while ventilation and perfusion were temporarily removed.<sup>4</sup> This method involved the measurement of airway pressure as the lungs were gradually inflated in 5-mL steps until a volume of 40 mL was reached. The total inhalation volume was assessed up to 40 mL to prevent injuries caused by the measurement method itself. Each inflation interval was set at 15 s to obtain a plateau pressure. The deflation PV curve was not measured.

### Lung wet-to-dry ratio

After all measurements were completed, the left lung was excised, and its wet weight was measured. It was dried at 60°C in an oven for 2 weeks, and its dry weight was measured to determine the lung wet-to-dry ratio (W/D) using the formula:  $W/D = \text{wet weight} / \text{dry weight}$ .<sup>13</sup>

### Bronchoalveolar lavage fluid analysis

After all measurements were completed, the right lung was used for the bronchoalveolar lavage fluid (BALF)

preparation. Three aliquots (5 mL each) of sterile saline were instilled separately through the trachea and drained. The lavage fluid was centrifuged at  $200 \times g$  for 10 min at  $4^{\circ}\text{C}$ , and the cell-free supernatant was stored at  $-70^{\circ}\text{C}$  as the BALF for further chemical analyses.

The BALF was used to measure total protein concentration and myeloperoxidase (MPO) activity. Total protein concentration was measured using the BAC Protein Assay Reagent Kit (Pierce, Rockford, IL). MPO activity was measured using the method of o-dianisidine dihydrochloride oxidation.<sup>17</sup> MPO activity was expressed as the change in optical density ( $\Delta\text{OD}$ ) per min and per mL of BALF. W/D, and total protein concentration and MPO activity in the BALF were used to determine histochemical lung injury.

### Statistical analyses

All data are expressed as the mean  $\pm$  standard deviation. Prism® ver. 4 (GraphPad Software, San Diego, CA) software was used for statistical analyses and figure preparations. Data were compared using Welch's *t*-test; *P*-values  $< 0.05$  were considered to be statistically significant.

## RESULTS

### Peak airway pressure and mean airway pressure

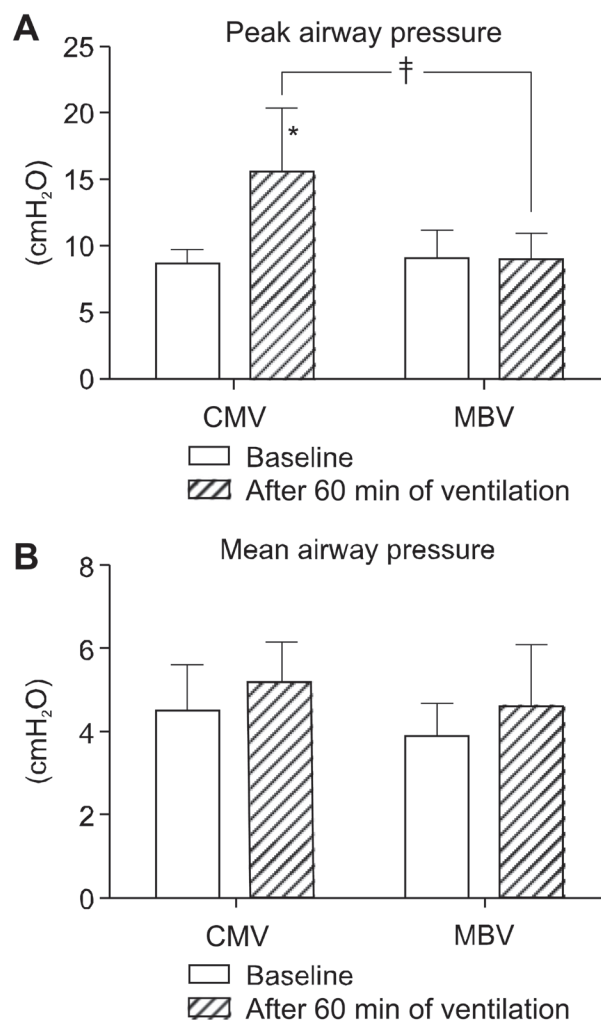
Figure 4 shows pPaw and mPaw both at baseline and after 60 min of ventilation. The pPaw in the CMV group was significantly increased after 60 min of ventilation ( $P < 0.05$ ). A statistically significant difference in pPaw after 60 min of ventilation was also observed between the two groups ( $P < 0.05$ ); conversely, pPaw scarcely changed in the MBV group after 60 min of ventilation ( $P = 0.93$ ). No significant differences in mPaw were observed between the two groups either at baseline ( $P = 0.27$ ) or after 60 min of ventilation ( $P = 0.40$ ).

### PV curve

The pulmonary PV curves of both groups at baseline and after 60 min of ventilation are shown in Fig. 5, in which the pressure is expressed as the average value for the corresponding volume since no significant differences in pressure values were detected between the two groups. Compared to the PV curves of the MBV group, those of the CMV group were slightly shifted towards higher pressures.

### W/D and BALF analysis

W/D, total protein concentration in the BALF, and MPO activity in the BALF are presented in Fig. 6. A statistically significant difference between the two groups was determined for W/D (CMV:  $7.50 \pm 0.47$ ; MBV:  $6.86 \pm$

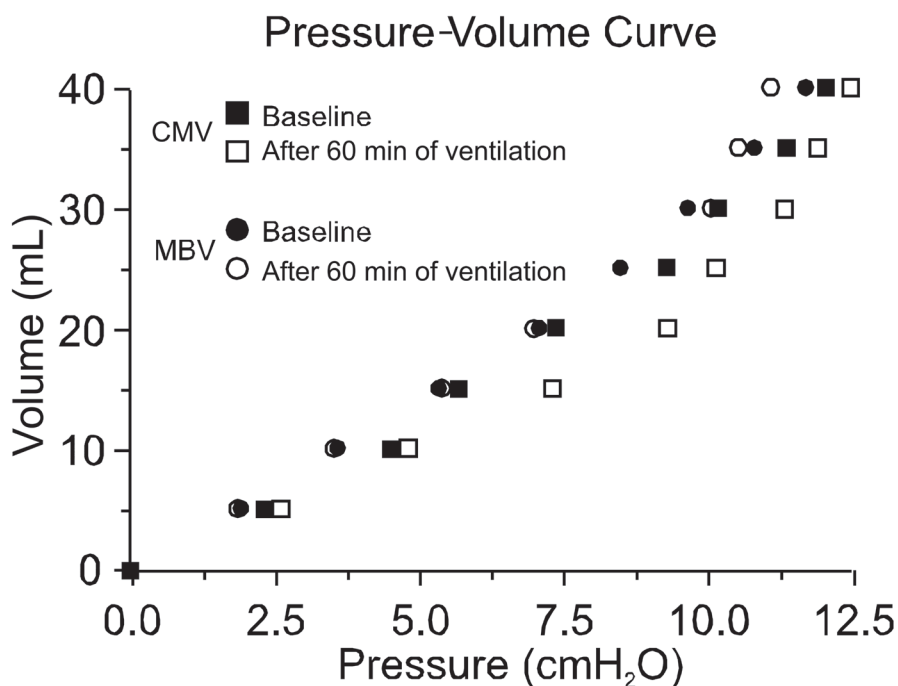


**Fig. 4.** Changes in peak airway pressure (A) and mean airway pressure (B) in the CMV and MBV groups. The peak airway pressure significantly increases after 60 min of ventilation in the CMV group ( $*P < 0.05$ ). A significant difference is observed in the peak airway pressure after 60 min of ventilation between the two groups ( $\dagger P < 0.05$ ). CMV, conventional mechanical ventilation; MBV, marathoners' breathing rhythm ventilation.

$0.18; P < 0.05$ ); however, no significant differences were found in total protein concentration (CMV:  $66.5 \pm 30.2$   $\mu\text{g/mL}$ ; MBV:  $45.8 \pm 32.2$   $\mu\text{g/mL}$ ;  $P = 0.24$ ) or MPO activity (CMV:  $0.56 \pm 0.10$   $\Delta\text{OD/mL/min}$ ; MBV:  $0.42 \pm 0.19$   $\Delta\text{OD/mL/min}$ ;  $P = 0.12$ ).

## DISCUSSION

Many marathon runners have adopted the 2:2 breathing rhythm. We developed an intermittent positive pressure ventilation system that mimics the breathing rhythm of marathon runners. The lung-protective effect of the MBV on the pulmonary pre-edema model was examined using an isolated perfused rabbit lung. Compared



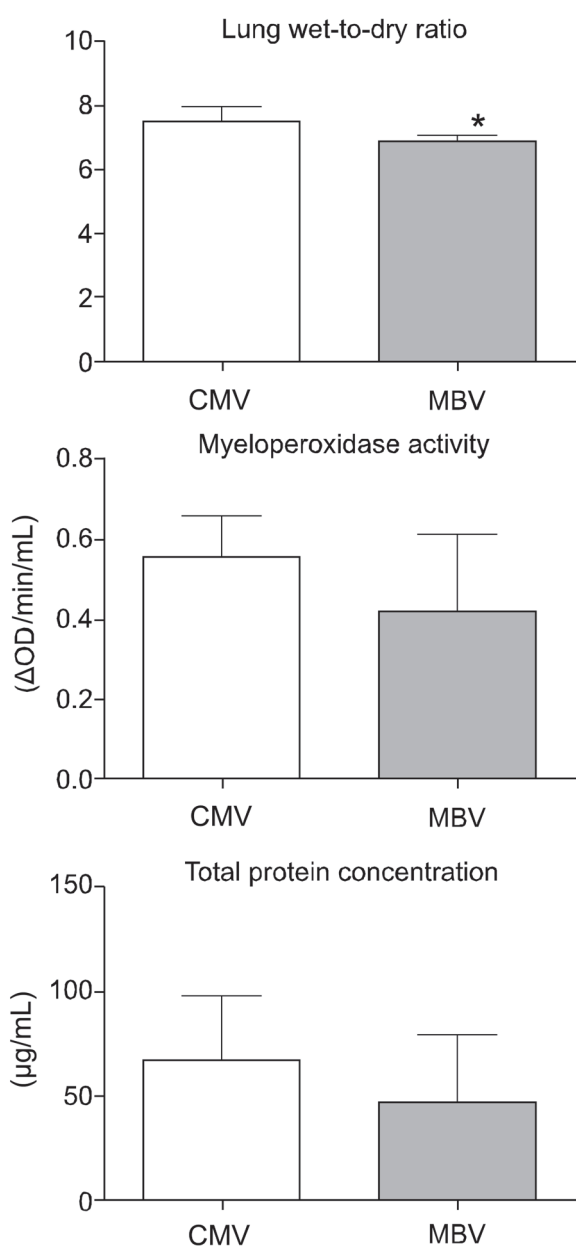
**Fig. 5.** Pulmonary PV curves at baseline and 60 min after ventilation in the CMV and MBV groups. Pressure is expressed as the average value against each volume, since no significant differences are observed between groups at each pressure. Compared with the PV curves in the MBV group, the curves in the CMV group have shifted slightly to higher pressures. CMV, conventional mechanical ventilation; MBV, marathoners' breathing rhythm ventilation.

to the CMV group, the MBV group had lower pPaw and W/D after 60 min of ventilation, and MBV was observed to have a preventive effect on the exacerbation of pulmonary edema.

Since the publication of the 2000 ARDS Network trial,<sup>18</sup> the focus of mechanical ventilation in acute respiratory failure has shifted from normalization of arterial blood gas values to lung protection. In recent years, low TV ventilation has secured a leading position among lung-protective ventilation strategies<sup>6</sup>; additionally, high-frequency oscillatory ventilation (HFOV)<sup>19</sup> and inverse ratio ventilation (IRV)<sup>20</sup> are also listed as possible options. Low TV ventilation is significantly disadvantaged, as it sometimes causes respiratory acidosis at a pH < 7.2.<sup>6</sup> In a piglet model of VILI, it was found that even with low TV ventilation, high RR values activate transforming growth factor  $\beta$  pathways and exacerbate pulmonary edema.<sup>21</sup> In HFOV, the lung volume is secured by the mPaw. If the mPaw setting is too low, the lung volume decreases, and oxygenation remains insufficient; conversely, setting the mPaw too high can result in decreased cardiac output and increased pulmonary vascular resistance.<sup>22</sup> In the OSCILLATE trial,<sup>19</sup> a randomized control trial for ARDS, HFOV did not prove to be superior to low TV ventilation; in the same trial, the high mortality rate that is found in

ARDS patients with HFOV was caused by circulatory suppression due to a high mPaw.<sup>23</sup> Moreover, when CO<sub>2</sub> is retained under HFOV, the only possible strategy is to decrease the frequency and increase the TV.<sup>24</sup> IRV is a positive-pressure ventilation method with an I/E ratio > 1.<sup>25</sup> In the IRV, pPaw values are kept low, while mPaw is maintained at high values.<sup>26</sup> The improved oxygenation capacity of the IRV is attributed to the increase in mPaw<sup>24</sup> and the occurrence of intrinsic PEEP<sup>27</sup> due to the decrease in expiratory time; however, in mouse experiments with high TV ventilation, lung injury was more frequently induced with the IRV than the CMV.<sup>28</sup>

During intense exercise, the O<sub>2</sub> demand increases beyond the limit of pulmonary diffusion capacity, thus increasing the alveolar-arterial oxygen difference and inducing hypoxemia.<sup>29</sup> It was shown that the heart rate increases to 145–180/min during a marathon.<sup>30</sup> In an exercise that corresponds to 80% of the maximum oxygen uptake rate, the mean pulmonary artery pressure increases up to 38 mmHg in young people, while the left atrial pressure increases up to 25 mmHg.<sup>31</sup> Subclinical interstitial pulmonary edema is found in 17% of runners after a marathon.<sup>32</sup> The fact that the 2:2 breathing rhythm is favored empirically in situations such as a marathon, which has a long exercise load, suggests that this respiratory technique works effectively to



**Fig. 6.** Comparison of lung wet-to-dry ratio, myeloperoxidase activity and total protein concentration between the CMV and MBV groups. A significant difference was observed in the lung wet-to-dry ratio between the two groups ( $*P < 0.05$ ). CMV, conventional mechanical ventilation;  $\Delta$ OD, change in optical density; MBV, marathoners' breathing rhythm ventilation.

prevent further progression of hypoxia and pulmonary edema. Moreover, the fact that the end-tidal  $\text{CO}_2$  is not retained during a marathon, along with the increased metabolism over a long time,<sup>11</sup> suggests that the 2:2 breathing rhythm provides sufficient capacity for  $\text{CO}_2$  elimination.

In this study, MBV ameliorated the progression of

pulmonary edema in a pre-edema model. We suggest that alveolar overdistension was prevented in the MBV because the total ventilation volume for one respiratory cycle was divided into two fractions. In high TV ventilation of the isolated perfused rat lung, active sodium transport and Na-K-ATPase activity of the alveolar epithelium are impaired, and lung edema clearance is reduced.<sup>33</sup> In this study, the PV curves at the end of the experiment were not very different between for the CMV and MBV groups. The pulmonary pre-edema model we employed can be considered as an early stage of lung injury, since no changes are observed in lung mechanics in the early stages of lung injury, and lung mechanics are not a valid marker for early stage lung injury.<sup>34</sup> In this study, there was no significant difference in MPO activity in the BALF between the CMV and MBV groups, suggesting that polymorphonuclear cells had not yet infiltrated into the alveolus. In small animals, acute extreme lung stretching rapidly develops into increased permeability lung edema. The mechanism is thought to involve the destruction of vascular endothelial cells, which induce direct contact between the basement membrane and polymorphonuclear cells, rather than involving the recruitment of inflammatory cells.<sup>2</sup>

The isolated perfused rabbit lung model enabled the investigation of the effects of ventilatory strategies on the lungs without the influence of confounding factors, e.g., the metabolization of inflammatory mediators by other organ systems. It is a reliable model by which lungs can be investigated under various pathophysiological conditions and represents an accepted model in the study of VILI-related problems.<sup>35–39</sup>

Some limitations of the study include the following factors. The average frequency of steps taken during a marathon is about 180/min.<sup>40</sup> In the 2:2 breathing rhythm, the average RR during a marathon is 45 cycles/min, given that one cycle consists of a 1st inspiratory phase, 2nd inspiratory phase, 1st expiratory phase, and 2nd expiratory phase. The resting RR in humans is 12–20 breaths/min, whereas in rabbits, it is 32–60 breaths/min.<sup>41</sup> In the current study, we examined the effects of MBV with RRs of approximately 30 cycles/min in rabbits. Higher RRs should also be examined. Furthermore, the TV in rabbits in CMV is considered to be 8–10 mL/kg,<sup>42, 43</sup> therefore, the TV of the CMV group was set to 8 mL/kg in this study. The preliminary study indicated that efficiency of  $\text{CO}_2$  elimination could be better with the MBV rather than the CMV method. Therefore, in this study, the TV of the MBV group was set to 6 mL/kg which may be the reason for which the lung edema did not progress in the MBV group.



The 2:2 breathing rhythm in a marathon runner, significantly differs from MBV, as the former is spontaneous ventilation, whereas the latter consists of intermittent positive pressure ventilation. Therefore, before the MBV approach can be applied in the clinic, its effects must be assessed in larger animals that have breathing cycle values closer to those in humans during a marathon. Moreover, MBV implementation requires the administration of sedatives or muscle relaxants, which must be taken into consideration prior to the clinical use of MBV. In severe ARDS with high airway pressure, if spontaneous breathing is preserved, transpulmonary pressure becomes too high, which can increase lung damage. Therefore, in severe ARDS, muscle relaxation may reduce lung damage.<sup>44</sup>

In conclusion, we successfully developed a prototype ventilator that can perform intermittent positive pressure ventilation, mimicking the breathing cycle of the 2:2 breathing rhythm. This mode of ventilation was named the “marathoners’ breathing rhythm ventilation.” The pPaw and W/D after 60 min of ventilation were significantly lower in the MBV than in the CMV group. Future studies may focus on examining the lung-protective effect of long-term MBV in diseased lungs in large animals.

*Acknowledgments:* This work was supported in part by a Grant-in-Aid for Scientific Research from the Japanese Ministry of Education, Culture, Sports, Science, and Technology (No. 16591539).

*The authors declare no conflict of interest.*

## REFERENCES

- Coates B, Kowalchik C. Runner’s world running on air: The revolutionary way to run better by breathing smarter. Emmaus (PA): Rodale Books; 2013.
- Dreyfuss D, Saumon G. Ventilator-induced lung injury: lessons from experimental studies. *Am J Respir Crit Care Med.* 1998;157:294-323. DOI: 10.1164/ajrccm.157.1.9604014, PMID: 9445314
- Ochiai R. Mechanical ventilation of acute respiratory distress syndrome. *J Intensive Care.* 2015;3:25. DOI: 10.1186/s40560-015-0091-6, PMID: 26045965
- Muscudere JG, Mullen JB, Gan K, Slutsky AS. Tidal ventilation at low airway pressures can augment lung injury. *Am J Respir Crit Care Med.* 1994;149:1327-34. DOI: 10.1164/ajrccm.149.5.8173774, PMID: 8173774
- Slutsky AS, Ranieri VM. Ventilator-induced lung injury. *N Engl J Med.* 2013;369:2126-36. DOI: 10.1056/NEJMr1208707, PMID: 24283226
- Brower RG, Matthay MA, Morris A, Schoenfeld D, Thompson BT, Wheeler A; Acute Respiratory Distress Syndrome Network. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. *N Engl J Med.* 2000;342:1301-8. DOI: 10.1056/NEJM200005043421801, PMID: 10793162
- Dempsey JA, Hanson PG, Henderson KS. Exercise-induced arterial hypoxaemia in healthy human subjects at sea level. *J Physiol.* 1984;355:161-75. DOI: 10.1113/jphysiol.1984.sp015412, PMID: 6436475
- Hopkins SR, McKenzie DC. Hypoxic ventilatory response and arterial desaturation during heavy work. *J Appl Physiol.* 1989;67:1119-24. DOI: 10.1152/jappl.1989.67.3.1119, PMID: 2793704
- Rice AJ, Thornton AT, Gore CJ, Scroop GC, Greville HW, Wagner H, et al. Pulmonary gas exchange during exercise in highly trained cyclists with arterial hypoxemia. *J Appl Physiol.* 1999;87:1802-12. DOI: 10.1152/jappl.1999.87.5.1802, PMID: 10562625
- Reeves JT, Linehan JH, Stenmark KR. Distensibility of the normal human lung circulation during exercise. *Am J Physiol Lung Cell Mol Physiol.* 2005;288:L419-25. DOI: 10.1152/ajplung.00162.2004, PMID: 15695542
- Thornadsson A, Drca N, Ricciardolo F, Högman M. Increased levels of alveolar and airway exhaled nitric oxide in runners. *Ups J Med Sci.* 2017;122:85-91. DOI: 10.1080/03009734.2017.1317886, PMID: 28481126
- Brack P, Dann S, Wijayantha KGU, Adcock P, Foster S. A simple, low-cost, and robust system to measure the volume of hydrogen evolved by chemical reactions with aqueous solutions. *J Vis Exp.* 2016;114:54383. DOI: 10.3791/54383, PMID: 27584581
- Liu R, Ueda M, Okazaki N, Ishibe Y. Role of potassium channels in isoflurane- and sevoflurane-induced attenuation of hypoxic pulmonary vasoconstriction in isolated perfused rabbit lungs. *Anesthesiology.* 2001;95:939-46. DOI: 10.1097/00000542-200110000-00024, PMID: 11605936
- Nicolaysen G. Perfusate qualities and spontaneous edema formation in an isolated perfused lung preparation. *Acta Physiol Scand.* 1971;83:563-70. DOI: 10.1111/j.1748-1716.1971.tb05114.x, PMID: 5160135
- Folkow B, Gurévig M, Hallbäck M, Lundgren Y, Weiss L. The hemodynamic consequences of regional hypotension in spontaneously hypertensive and normotensive rats. *Acta Physiol Scand.* 1971;83:532-41. DOI: 10.1111/j.1748-1716.1971.tb05111.x, PMID: 5160133
- Rowan SC, Rochfort KD, Piouveau L, Cummins PM, O’Rourke M, McLoughlin P. Pulmonary endothelial permeability and tissue fluid balance depend on the viscosity of the perfusion solution. *Am J Physiol Lung Cell Mol Physiol.* 2018;315:L476-84. DOI: 10.1152/ajplung.00437.2017, PMID: 29792349
- Henson PM, Zanolari B, Schwartzman NA, Hong SR. Intracellular control of human neutrophil secretion. I. C5a-induced stimulus-specific desensitization and the effects of cytochalasin B. *J Immunol.* 1978;121:851-5. PMID: 211165
- Slutsky AS, Marco Ranieri V. Mechanical ventilation: lessons from the ARDSNet trial. *Respir Res.* 2000;1:73-7. DOI: 10.1186/rr15, PMID: 11667968

- 19 Ferguson ND, Cook DJ, Guyatt GH, Mehta S, Hand L, Austin P, et al.; OSCILLATE Trial Investigators; Canadian Critical Care Trials Group. High-frequency oscillation in early acute respiratory distress syndrome. *N Engl J Med.* 2013;368:795-805. DOI: [10.1056/NEJMoa1215554](https://doi.org/10.1056/NEJMoa1215554), PMID: 23339639
- 20 Schreiter D, Reske A, Stichert B, Seiwerts M, Bohm SH, Kloeppe R, et al. Alveolar recruitment in combination with sufficient positive end-expiratory pressure increases oxygenation and lung aeration in patients with severe chest trauma. *Crit Care Med.* 2004;32:968-75. DOI: [10.1097/01.CCM.0000120050.85798.38](https://doi.org/10.1097/01.CCM.0000120050.85798.38), PMID: 15071387
- 21 Retamal J, Borges JB, Bruhn A, Cao X, Feinstein R, Hedenstierna G, et al. High respiratory rate is associated with early reduction of lung edema clearance in an experimental model of ARDS. *Acta Anaesthesiol Scand.* 2016;60:79-92. DOI: [10.1111/aas.12596](https://doi.org/10.1111/aas.12596), PMID: 26256848
- 22 Goddon S, Fujino Y, Hromi JM, Kacmarek RM. Optimal mean airway pressure during high-frequency oscillation: predicted by the pressure-volume curve. *Anesthesiology.* 2001;94:862-9. DOI: [10.1097/00000542-200105000-00026](https://doi.org/10.1097/00000542-200105000-00026), PMID: 11388539
- 23 Young D, Lamb SE, Shah S, MacKenzie I, Tunnicliffe W, Lall R, et al.; OSCAR Study Group. High-frequency oscillation for acute respiratory distress syndrome. *N Engl J Med.* 2013;368:806-13. DOI: [10.1056/NEJMoa1215716](https://doi.org/10.1056/NEJMoa1215716), PMID: 23339638
- 24 Hatcher D, Watanabe H, Ashbury T, Vincent S, Fisher J, Froese A. Mechanical performance of clinically available, neonatal, high-frequency, oscillatory-type ventilators. *Crit Care Med.* 1998;26:1081-8. DOI: [10.1097/00003246-199806000-00032](https://doi.org/10.1097/00003246-199806000-00032), PMID: 9635659
- 25 Warner MA, Patel B. Mechanical ventilation. In: Hagberg CA, editor. *Benumof and Hagberg's airway management.* 3rd ed. Philadelphia: Elsevier; 2013. p. 981-97.
- 26 Sari A, Yamashita S, Toriumi T, Nakashima K, Kawata R, Kunihiko M, et al. The effects of inverted ratio ventilation (IRV) on arterial oxygenation during mechanical ventilation in patients with acute respiratory failure. *Resuscitation.* 1991;22:93-101. DOI: [10.1016/0300-9572\(91\)90068-A](https://doi.org/10.1016/0300-9572(91)90068-A), PMID: 1658898
- 27 Cole AGH, Weller SF, Sykes MK. Inverse ratio ventilation compared with PEEP in adult respiratory failure. *Intensive Care Med.* 1984;10:227-32. DOI: [10.1007/BF00256258](https://doi.org/10.1007/BF00256258), PMID: 6386921
- 28 Müller-Redetzky HC, Felten M, Hellwig K, Wienhold SM, Naujoks J, Opitz B, et al. Increasing the inspiratory time and I:E ratio during mechanical ventilation aggravates ventilator-induced lung injury in mice. *Crit Care.* 2015;19:23. DOI: [10.1186/s13054-015-0759-2](https://doi.org/10.1186/s13054-015-0759-2), PMID: 25888164
- 29 Stickland MK, Lindinger MI, Olfert IM, Heigenhauser GJ, Hopkins SR. Pulmonary gas exchange and acid-base balance during exercise. *Compr Physiol.* 2013;3:693-739. DOI: [10.1002/cphy.c110048](https://doi.org/10.1002/cphy.c110048), PMID: 23720327
- 30 Manier G, Moinard J, Téhoueyres P, Varène N, Guénard H. Pulmonary diffusion limitation after prolonged strenuous exercise. *Respir Physiol.* 1991;83:143-53. DOI: [10.1016/0034-5687\(91\)90024-D](https://doi.org/10.1016/0034-5687(91)90024-D), PMID: 2068412
- 31 Reeves JT, Dempsey JA, Grover RF. Pulmonary circulation during exercise. In: Weir EK, Reeves JT, editors. *Lung biology in health and disease, Vol. 38. Pulmonary vascular physiology and pathophysiology.* New York: Marcel Dekker Inc; 1988.
- 32 Zavorsky GS, Milne ENC, Lavorini F, Rienzi JP, Cutrufello PT, Kumar SS, et al. Small changes in lung function in runners with marathon-induced interstitial lung edema. *Physiol Rep.* 2014;2:e12056. DOI: [10.14814/phy2.12056](https://doi.org/10.14814/phy2.12056), PMID: 24973330
- 33 Lecuona E, Saldías F, Comellas A, Ridge K, Guerrero C, Sznajder JI. Ventilator-associated lung injury decreases lung ability to clear edema in rats. *Am J Respir Crit Care Med.* 1999;159:603-9. DOI: [10.1164/ajrccm.159.2.9805050](https://doi.org/10.1164/ajrccm.159.2.9805050), PMID: 9927379
- 34 Vaporidi K, Voloudakis G, Priniannakis G, Kondili E, Koutsopoulos A, Tsatsanis C, et al. Effects of respiratory rate on ventilator-induced lung injury at a constant P<sub>aCO2</sub> in a mouse model of normal lung. *Crit Care Med.* 2008;36:1277-83. DOI: [10.1097/CCM.0b013e318169f30e](https://doi.org/10.1097/CCM.0b013e318169f30e), PMID: 18379255
- 35 Gama de Abreu M, Heintz M, Heller A, Széchényi R, Albrecht DM, Koch T. One-lung ventilation with high tidal volumes and zero positive end-expiratory pressure is injurious in the isolated rabbit lung model. *Anesth Analg.* 2003;96:220-8. DOI: [10.1213/00000539-200301000-00045](https://doi.org/10.1213/00000539-200301000-00045), PMID: 12505956
- 36 Hermle G, Mols G, Zügel A, Benzing A, Lichtwarck-Aschoff M, Geiger K, et al. Intratidal compliance-volume curve as an alternative basis to adjust positive end-expiratory pressure: A study in isolated perfused rabbit lungs. *Crit Care Med.* 2002;30:1589-97. DOI: [10.1097/00003246-200207000-00032](https://doi.org/10.1097/00003246-200207000-00032), PMID: 12130984
- 37 Broccard AF, Hotchkiss JR, Kuwayama N, Olson DA, Jamal S, Wangenstein DO, et al. Consequences of vascular flow on lung injury induced by mechanical ventilation. *Am J Respir Crit Care Med.* 1998;157:1935-42. DOI: [10.1164/ajrccm.157.6.9612006](https://doi.org/10.1164/ajrccm.157.6.9612006), PMID: 9620930
- 38 Broccard AF, Vannay C, Feihl F, Schaller MD. Impact of low pulmonary vascular pressure on ventilator-induced lung injury. *Crit Care Med.* 2002;30:2183-90. DOI: [10.1097/00003246-200210000-00002](https://doi.org/10.1097/00003246-200210000-00002), PMID: 12394942
- 39 Mols G, Hermle G, Fries G, Benzing A, Lichtwarck-Aschoff M, Geiger K, et al. Different strategies to keep the lung open: A study in isolated perfused rabbit lungs. *Crit Care Med.* 2002;30:1598-604. DOI: [10.1097/00003246-200207000-00033](https://doi.org/10.1097/00003246-200207000-00033), PMID: 12130985
- 40 Daniels JT. *Daniels' running formula.* 3rd ed. Champaign (IL): Human Kinetics; 2013.
- 41 Harkness JE, Turner PV, VandeWoude S, Wheler CL. *Harkness and Wagner's biology and medicine of rabbits and rodents.* 5th ed. Hoboken (NJ): John Wiley & Sons; 2010.
- 42 Fuchs H, Mandler MR, Scharneck D, Ebsen M, Hummler HD. Very low tidal volume ventilation with associated hypercapnia--effects on lung injury in a model for acute respiratory distress syndrome. *PLoS One.* 2011;6:e23816. DOI: [10.1371/journal.pone.0023816](https://doi.org/10.1371/journal.pone.0023816), PMID: 21886825
- 43 García-Delgado M, Navarrete-Sánchez I, Chamorro-Marín V, Díaz-Monrové JC, Esquivias J, Fernández-Mondéjar E. Alveolar overdistension as a cause of lung injury: differences among three animal species. *ScientificWorldJournal.* 2012;2012:1-5. DOI: [10.1100/2012/985923](https://doi.org/10.1100/2012/985923), PMID: 22645503
- 44 Yoshida T, Uchiyama A, Matsuura N, Mashimo T, Fujino Y. The comparison of spontaneous breathing and muscle paralysis in two different severities of experimental lung injury. *Crit Care Med.* 2013;41:536-45. DOI: [10.1097/CCM.0b013e3182711972](https://doi.org/10.1097/CCM.0b013e3182711972), PMID: 23263584