



**University of  
Zurich**<sup>UZH</sup>

**Zurich Open Repository and  
Archive**

University of Zurich  
Main Library  
Strickhofstrasse 39  
CH-8057 Zurich  
[www.zora.uzh.ch](http://www.zora.uzh.ch)

---

Year: 2020

---

## **Description of a fossil camelid from the Pleistocene of Argentina, and a cladistic analysis of the Camelinae**

Lynch, Sinéad ; Sánchez-Villagra, Marcelo R ; Balcarcel, Ana

**Abstract:** We describe a well-preserved South American Lamini partial skeleton (PIMUZ A/V 4165) from the Ensenadan ( 1.95–1.77 to 0.4 Mya) of Argentina. The specimen is comprised of a nearly complete skull and mandible with full tooth rows, multiple elements of anterior and posterior limbs, and a scapula. We tested this specimen's phylogenetic position and hypothesized it to be more closely related to *Lama guanicoe* and *Vicugna vicugna* than to *Hemiauchenia paradoxa*. We formulate a hypothesis for the placement of PIMUZ A/V 4165 within Camelinae in a cladistic analysis based on craniomandibular and dental characters and propose that future systematic studies consider this specimen as representing a new species. For the first time in a morphological phylogeny, we code terminal taxa at the species level for the following genera: *Camelops*, *Aepycamelus*, *Pleiolama*, *Procamelus*, and *Alforjas*. Our results indicate a divergence between Lamini and Camelini predating the Barstovian (16 Mya). *Camelops* appears as monophyletic within the Camelini. *Alforjas taylori* falls out as a basal member of Camelinae—neither as a Lamini nor Camelini. *Pleiolama* is polyphyletic, with *Pleiolama vera* as a basal Lamini and *Pleiolama mckennai* in a more nested position within the Lamini. *Aepycamelus* and *Procamelus* are respectively polyphyletic and paraphyletic. Together, they are part of a group of North American Lamini from the Miocene epoch.

DOI: <https://doi.org/10.1186/s13358-020-00208-6>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-192647>

Journal Article

Published Version



The following work is licensed under a Creative Commons: Attribution 4.0 International (CC BY 4.0) License.

Originally published at:

Lynch, Sinéad; Sánchez-Villagra, Marcelo R; Balcarcel, Ana (2020). Description of a fossil camelid from the Pleistocene of Argentina, and a cladistic analysis of the Camelinae. *Swiss Journal of Palaeontology*:139:6.

DOI: <https://doi.org/10.1186/s13358-020-00208-6>

RESEARCH ARTICLE

Open Access



# Description of a fossil camelid from the Pleistocene of Argentina, and a cladistic analysis of the Camelinae

Sinéad Lynch, Marcelo R. Sánchez-Villagra\* and Ana Balcarcel\*

## Abstract

We describe a well-preserved South American Lamini partial skeleton (PIMUZ A/V 4165) from the Ensenadan (~ 1.95–1.77 to 0.4 Mya) of Argentina. The specimen is comprised of a nearly complete skull and mandible with full tooth rows, multiple elements of anterior and posterior limbs, and a scapula. We tested this specimen's phylogenetic position and hypothesized it to be more closely related to *Lama guanicoe* and *Vicugna vicugna* than to *Hemiauchenia paradoxa*. We formulate a hypothesis for the placement of PIMUZ A/V 4165 within Camelinae in a cladistic analysis based on craniomandibular and dental characters and propose that future systematic studies consider this specimen as representing a new species. For the first time in a morphological phylogeny, we code terminal taxa at the species level for the following genera: *Camelops*, *Aepycamelus*, *Pleiolama*, *Procamelus*, and *Alforjas*. Our results indicate a divergence between Lamini and Camelini predating the Barstovian (16 Mya). *Camelops* appears as monophyletic within the Camelini. *Alforjas taylori* falls out as a basal member of Camelinae—neither as a Lamini nor Camelini. *Pleiolama* is polyphyletic, with *Pleiolama vera* as a basal Lamini and *Pleiolama mckennai* in a more nested position within the Lamini. *Aepycamelus* and *Procamelus* are respectively polyphyletic and paraphyletic. Together, they are part of a group of North American Lamini from the Miocene epoch.

**Keyword:** Lamini, Camelinae, South America, North America, Phylogeny

## Introduction

Exceptional fossils are key to solving taxonomic and phylogenetic questions given the rich and reliable anatomy they preserve. Some of them are still waiting in the field, while some are in museum collections in which proper curation and care allow multiple researchers to re-assess them. We present a case of the latter, with the study of specimen PIMUZ A/V 4165 from Barranca del Parana, San Nicolas (Buenos Aires province, Argentina) (Roth 1889; Schulthess 1920), part of a collection assembled by the celebrated Swiss-Argentinian paleontologist Santiago Roth, 1850–1924 (Bond 1999).

PIMUZ A/V 4165 was initially identified as a *Palaeolama* (Roth 1889; Schulthess 1920), a genus of the 'tribe' Lamini. Its stratigraphic age is Pampeano inferior (Roth 1889), also known as the Ensenadan (Cione et al. 2015). We describe this material for the first time and test its position in a cladistic analysis that also addresses other aspects of camelid evolution in the Americas.

The Camelidae appeared during the middle Eocene in North America (Honey et al. 1998). *Poebrotherium wilsoni*, one of its earliest members, is recorded from the late Eocene through the early Oligocene. According to Honey et al. (1998), the 'family' Camelidae underwent four radiation events. The 'sub-family' Protolabinae appeared in the third radiation, which took place in the late Oligocene and early Miocene. The 'sub-family' Camelinae, which includes the 'tribes' Lamini and Camelini, appeared

Editorial handling: Loïc Costeur.

\*Correspondence: m.sanchez@pim.uzh.ch; ana.balcarcel@gmail.com  
Palaeontological Institute and Museum, University of Zurich,  
Karl-Schmid-Strasse 4, 8006 Zurich, Switzerland



© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.



*Palaeolama weddelli*, *Lama guanicoe*, *Lama castelnaudi*, *Vicugna*, *Vicugna provicugna*, and *Eulamaops parallelus*.

Baskin and Thomas (2016) validated two *Camelops* species: *C. hesternus* and *C. minidokae*. Morphological phylogenies have predominantly assigned *Camelops* to the Lamini clade (Harrison 1979; Honey et al. 1998; Scherer 2013), but recently novel proteomic (Buckley et al. 2019) and genetic (Heintzman et al. 2015) studies indicated a closer connection between this genus and the Camelini. By contrast, no progress has been made to determine the intra- and inter-generic affinities of *Aepycamelus*. Morphological phylogenies have also neglected the intra-generic relationships of *Alforjas*, *Pleiolama*, *Procamelus*, and *Camelops*. Our study provides novel insights into these areas of the Camelid tree, based on revision of original materials in museums, a critical assessment of the literature, and an analysis of these data.

## Materials and methods

### Materials

Specimen PIMUZ A/V 4165 is from the Roth Collection at the Palaeontological Museum of the University of Zurich. It was collected in Barranca del Parana, San Nicolas (Buenos Aires province, Argentina) (Roth 1889; Schulthess 1920). The stratigraphic age is Ensenadan (early to middle Pleistocene) (Roth 1889; Schulthess 1920; Cione et al. 2015), which dates approximately from ~1.95–1.77 to 0.4 Mya (Verzi et al. 2004; Soibelzon et al. 2008). To investigate the taxonomic and systematic allocation of this fossil, we conducted a phylogenetic analysis of several South and North American taxa. We used *Poebrotherium wilsoni*, a basal camelid (Honey et al. 1998), as the outgroup. For the ingroup, we included members of the Camelinae (see Additional files 1, 2). Several North American taxa (*Pleiolama*, *Camelops*, *Aepycamelus*, *Procamelus*, and *Alforjas*), previously scored as genera by Scherer (2013), Honey et al. (1998) and Harrison (1979, 1985), are now scored as species. For *Pleiolama* and *Camelops*, we included all species currently accepted. We also included four species of *Aepycamelus* and a single one of the genus *Alforjas*. All Protolabinae genera (*Michenia*, *Tanymyktekter*, *Protolabis*) are also in the ingroup.

Most of the terminal taxa are scored from a single, well-preserved specimen. A few taxa, such as *Camelops minidokae*, *Megatylopus matthewi*, and *Palaeolama mirifica*, were scored with more fragmentary material. For *Lama* and *Vicugna*, we included only the wild forms (*L. guanicoe*, *V. vicugna*). For *Camelus*, we scored material from the domestic *C. bactrianus*.

### Characters

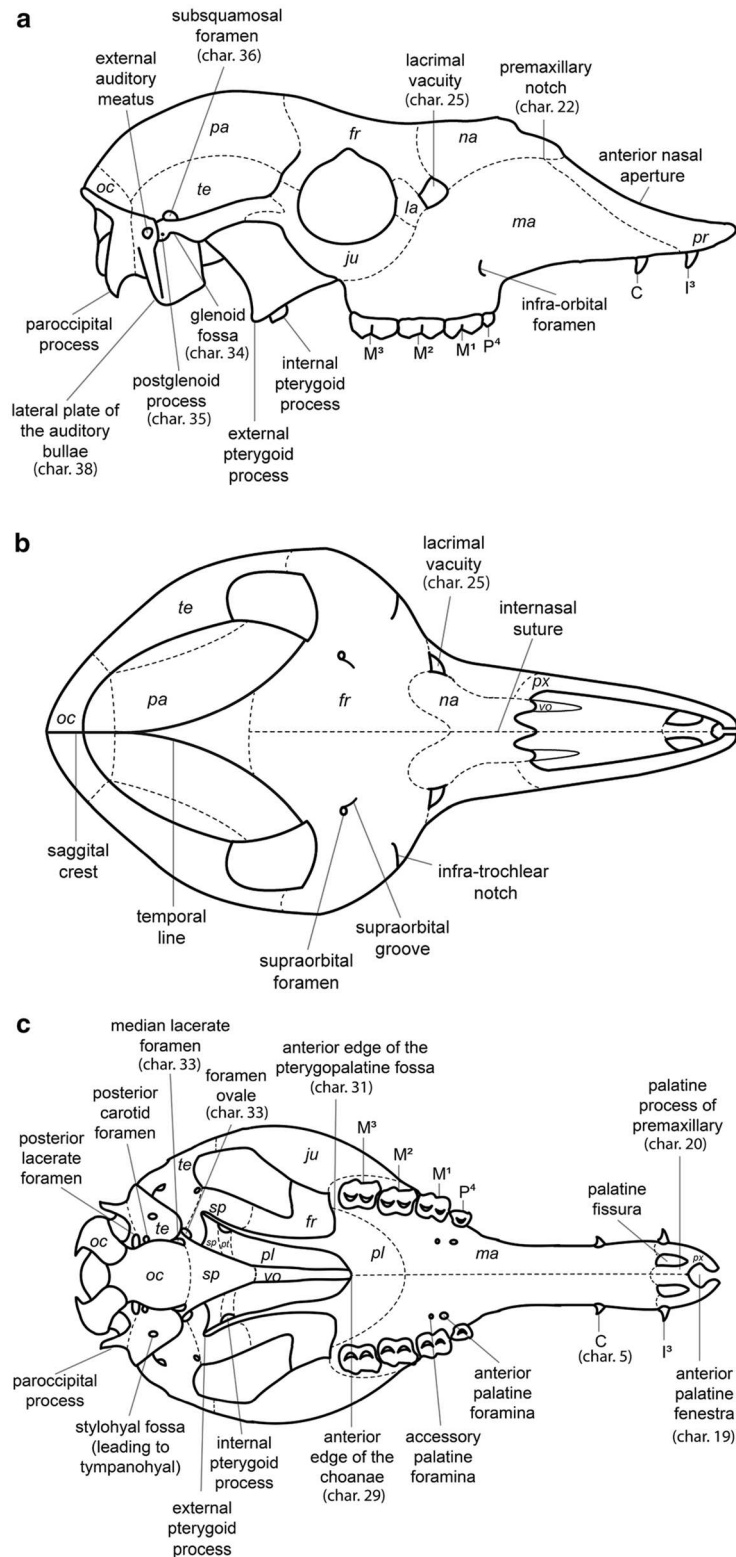
We concentrated on craniomandibular characters exclusively; no postcranial characters were included, since many taxa in the analysis had no (or only fragmentary) postcranial elements. Terminology for skull and mandible morphology follows Webb (1965), Pacheco Torres et al. (1986) and El Allali et al. (2017) (Fig. 2). Dental terminology follows Hershkovitz (1982) (Fig. 3).

We reviewed previous systematic analyses by Scherer (2013), Honey et al. (1998), Harrison (1979, 1985), and Webb (1965). Characters were updated and adapted, as noted. Five characters, not previously used in cladistic analyses, were based on information provided by Webb (1965) and by Honey (2007). In total, we present 17 new characters. We measured the hypsodonty index as per Shockey (1997). We found it to vary greatly, particularly in our sample of *V. vicugna*, likely due to differences in the level of wear. Therefore, we omitted this character from our matrix. We followed the scoring of Scherer (2009, 2013) for P<sup>3</sup> (char. 12) and P<sub>3</sub> (char. 13) in *L. guanicoe* and *V. vicugna*, as she had access to more material from these species and because their states were ambiguous in our sample.

For ratio characters (chars. 44–49), we defined states by gap-coding (figures of the measurements are in Additional file 3). A detailed list of the characters included in this study is presented below.

### Dental characters

1. First upper incisor (I<sup>1</sup>): *present* (0); *absent* (1). (Ch.10, Scherer (2013); ch.1, Harrison (1979); Honey et al. (1998); Harrison (1985); Honey and Taylor (1978)).
2. Second upper incisor (I<sup>2</sup>): *present* (0); *absent* (1). (Ch.11, Scherer (2013); mod. ch.2, Harrison (1979); Honey et al. (1998); Harrison (1985); Honey and Taylor (1978)).
3. Enamel layer on the lingual side of lower incisors (I<sub>1-3</sub>): *thick* (0); *thin or absent* (1). (Modified from ch.12, Scherer (2013); Honey et al. (1998); Harrison (1985)).
4. Crown of lower incisors (I<sub>1-3</sub>): *spatulated* (0); *cylindrical* (1). (Modified from ch.12, Scherer (2013); Honey et al. (1998); Harrison (1985)).
5. Upper and lower canine transverse section (C, c): *rounded* (0); *laterally compressed* (1). (Modified from ch.13, Scherer (2013); mod. ch.4+5, Harrison (1979); Honey et al. (1998); Harrison (1985)).
6. Upper canine position (C): *close to third incisor* (I<sup>3</sup>) (< 1 cm) (0); *distant to third incisor* (I<sup>3</sup>) (≥ 1 cm) (1).



**Fig. 2** Schematic drawings of *L. guanicoe*. **a** Skull, lateral view. **b** Skull, dorsal view. **c** Skull, occlusal view. **d** Mandible, lateral view. Abbreviations: pr = premaxillary bone; ma = maxillary bone; fr = frontal bone; la = lacrimal bone; ju = jugal bone; te = temporal bone; oc = occipital bone; sp = sphenoid bone; pt = pterygoid bone; vo = vomer bone; pl = palatine bone; pa = parietal bone; I<sup>3</sup> = third upper incisor; I<sub>1</sub> = first lower incisor; I<sub>2</sub> = second lower incisor; I<sub>3</sub> = third lower incisor; C = Upper canine; c = Lower canine; P<sup>1</sup> = first upper premolar; P<sub>1</sub> = first lower premolar; P<sub>2</sub> = second lower premolar; P<sup>4</sup> = fourth upper premolar; P<sub>4</sub> = fourth lower premolar; M<sup>1</sup> = first upper molar; M<sub>1</sub> = first lower molar; M<sup>2</sup> = second upper molar; M<sub>2</sub> = second lower molar; M<sup>3</sup> = third upper molar; M<sub>3</sub> = third lower molar

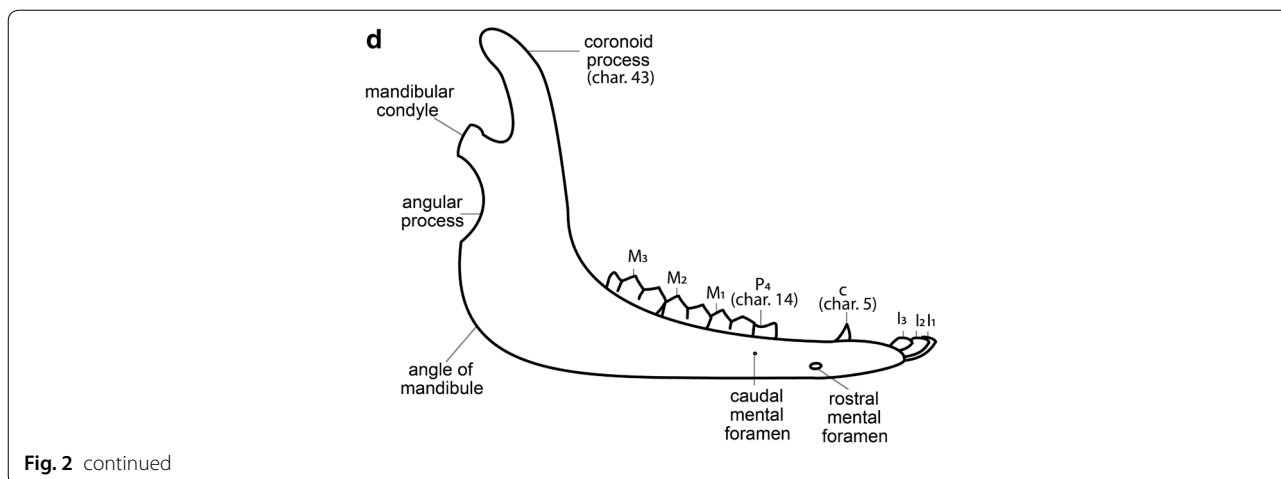
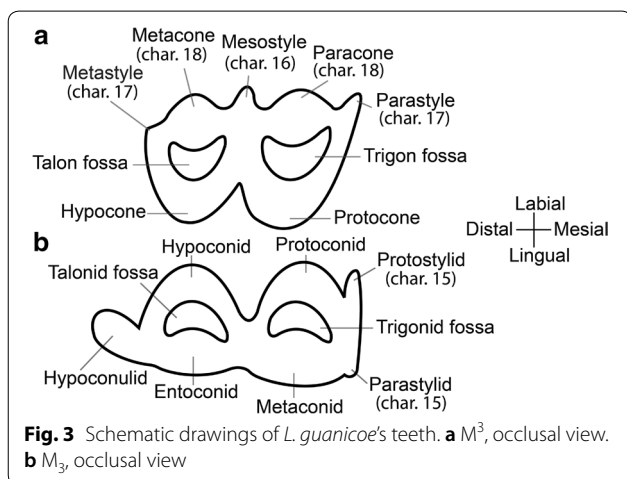


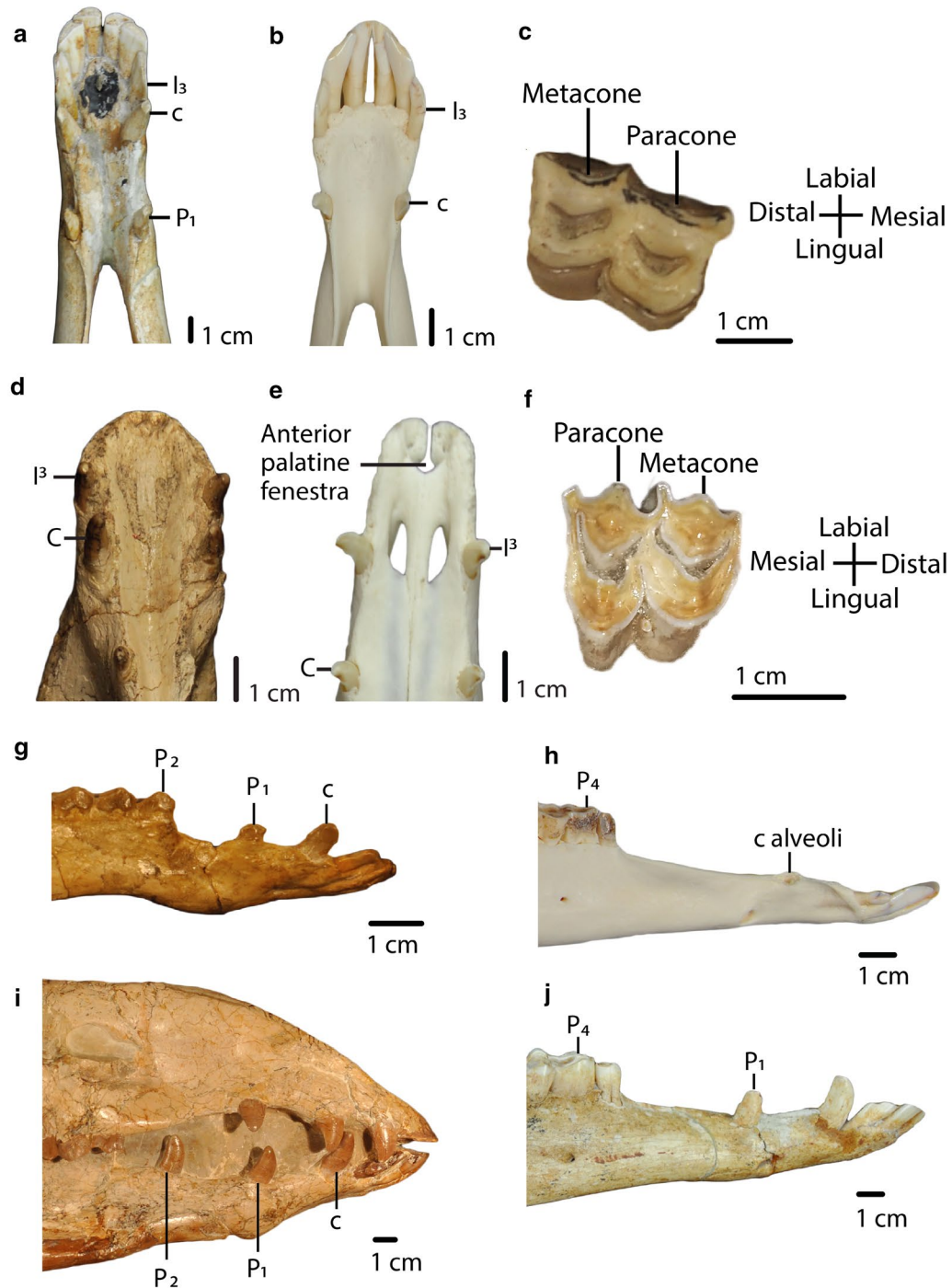
Fig. 2 continued



**Fig. 3** Schematic drawings of *L. guanicoe*'s teeth. **a**  $M^3$ , occlusal view. **b**  $M_3$ , occlusal view

(Distance from the distal border of the third incisor to the mesial border of canine).

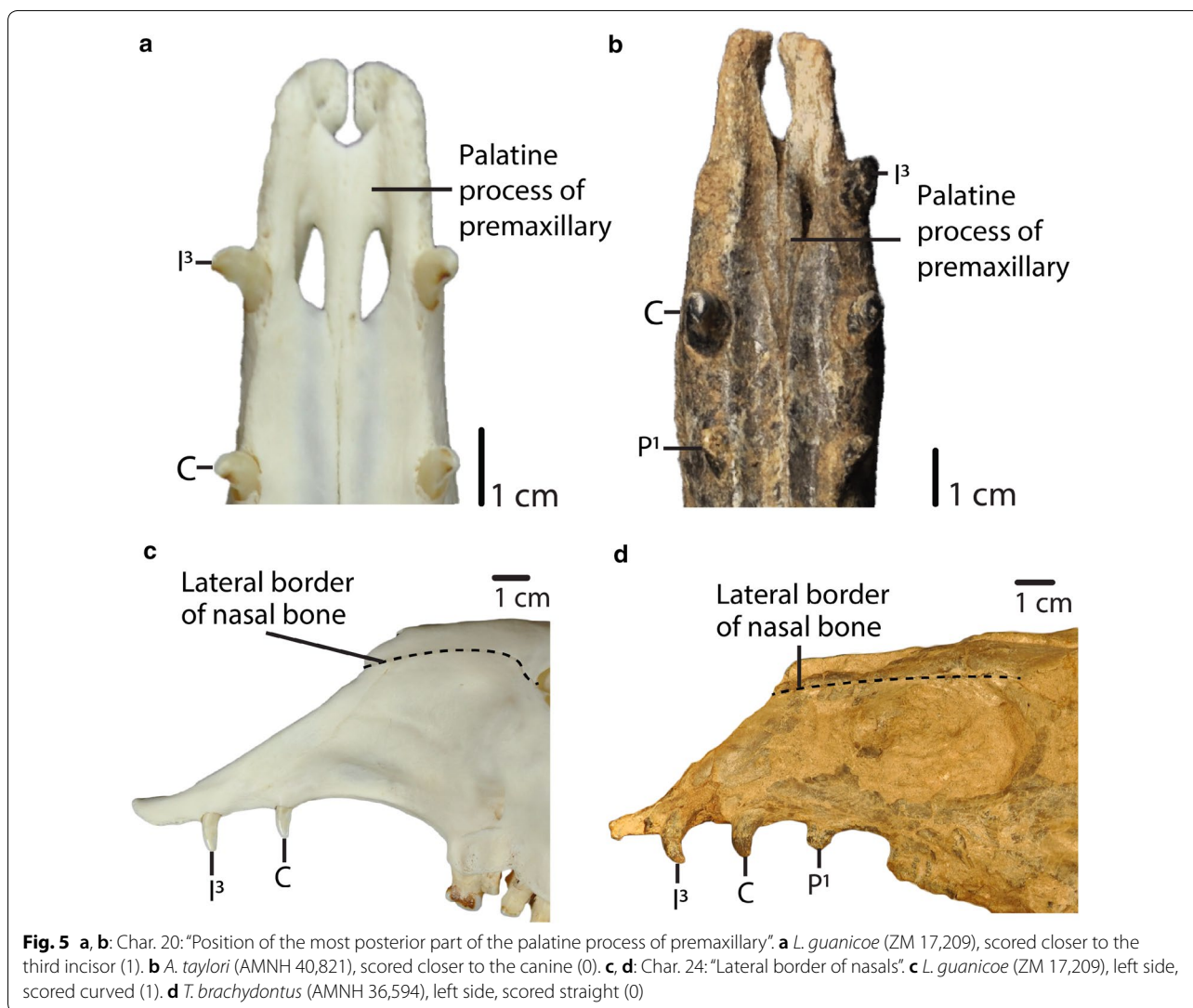
7. Lower canine position (c): *close to incisor* ( $I_3$ ) ( $< 1$  cm) (0); *far from incisor* ( $I_3$ ) ( $\geq 1$  cm) (1). (Distance from the distal border of the third incisor to the mesial border of canine). (Fig. 4).
8. First upper premolar ( $P^1$ ): *premolariform* (0); *caniniform* (1); *absent* (2). (Modified from ch. 14, Scherer (2013); mod. ch.6, Harrison (1979); mod. ch.“P1 and P/1”, Webb (1965, p. 46); Honey et al. (1998); Harrison (1985)).
9. First lower premolar ( $P_1$ ): *premolariform* (0); *caniniform* (1); *absent* (2). (Modified from ch.15, Scherer (2013); mod. ch.7, Harrison (1979); mod. ch.“P1 and P/1”, Webb (1965, p. 46); Honey et al. (1998); Harrison (1985); Fig. 4).
10. Second upper premolar ( $P^2$ ): *present* (0); *absent* (1). (Ch.16, Scherer (2013); mod. ch.8, Harrison (1979); Modified from ch.“P2 and P/2”, Webb (1965, p. 46); Honey et al. (1998); Harrison (1985); Honey and Taylor (1978)).
11. Second lower premolar ( $P_2$ ): *premolariform* (0); *caniniform* (1); *absent* (2). (Modified from ch.17, Scherer (2013); mod. ch.9, Harrison (1979); mod. ch.“P2 and P/2”, Webb (1965, p.46); Honey et al. (1998); Harrison (1985); Honey and Taylor (1978); Fig. 4).
12. Third upper premolar ( $P^3$ ): *present* (0); *absent* (1). (Ch.10, Harrison (1979); ch.18, Scherer (2013); Honey et al. (1998)).
13. Third lower premolar ( $P_3$ ): *present* (0); *absent* (1). (Ch.11, Harrison (1979); ch.19, Scherer (2013); Honey et al. (1998); Harrison (1985)).
14. Fourth lower premolar shape ( $P_4$ ): *triangular, with fossetid only on the distal lobe* (0); *quadrangular, with fossetids on the mesial and distal lobes* (1). (Ch.20, Scherer (2013)).
15. Protostylids and parastylids (“llama buttresses”) on lower molars ( $M_1$ – $M_3$ ): *small or absent* (0); *greatly developed* (1). (Ch.24, Scherer (2013); mod. ch.12, Harrison (1979); mod. ch.“Lower Molars”, Webb (1965, p. 46); Honey et al. (1998); Harrison (1985), see Scherer (2009, Fig. 20, p. 90)).
16. Labial lophids on lower molars ( $M_1$ – $M_3$ ) in occlusal view: *U-shaped or rounded* (0); *triangular* (1). (Mod ch.21, Scherer (2013), see Scherer (2009, Fig. 20, p. 90); see Scherer et al. (2007, Fig. 4, p. 41)).
17. Labial styles (“ribs”) on upper molars ( $M^1$ – $M^3$ ): *weakly developed* (0); *well-developed* (1). (Mod ch.22, Scherer (2013), see Scherer (2009, Fig. 20, p. 90)).
18. Metacone and paracone on upper molars ( $M^1$ – $M^3$ ): *weakly developed* (0); *well-developed* (1). (Fig. 4



**Fig. 4** **a, b:** Char. 7: "Lower canine position (c)". **a** *H. macrocephala* (UF 205,750), scored close from incisor ( $I_3$ ) (< 1 cm) (0). **b** *L. guanicoe* (ZM 17,209), scored far from incisor ( $I_3$ ) ( $\geq$  1 cm) (1). **c, f:** Char. 18: "Metacone and paracone on upper molars ( $M^1$ - $M^3$ )". **c** Right  $M^2$ , *P. coartatus* (AMNH 73,377), scored weakly developed (0). **f** Left  $M^2$ , *P. weddelli* (PUN 1), scored well developed (1). **d, e:** Char. 19: "Anterior palatine fenestra". **d** *P. coartatus* (AMNH 73,438), ventral view, scored absent (0). **e** *L. guanicoe* (ZM 17,209), ventral view, scored present (1). **g, h, i, j:** Char. 9: "First lower premolar ( $P_1$ )". **g** *P. wilsoni* (AMNH 47,130), right side, scored premolariform (0). **h** *L. guanicoe* (ZM 17,967), left side, scored absent (2). **i** *A. robustus* (OMNH 016,560), right side, scored caniniform (1). **j** *H. macrocephala* (UF 205,750), right side, scored caniniform (1). **g, h, i, j:** Char. 11: "Second lower premolar ( $P_2$ )". **g** *P. wilsoni* (AMNH 47,130), right side, scored premolariform (0). **h** *L. guanicoe* (ZM 17,967), left side, scored absent (2). **i** *A. robustus* (OMNH 016,560), right side, scored caniniform (1). **j** *H. macrocephala* (UF 205,750), right side, scored absent (2)

## Cranial characters

19. Anterior palatine fenestra: *absent (0)*; *present (1)*. (Based on Honey (2007); Fig. 4).
20. Position of the most posterior part of the palatine process of premaxillary: *closer to the canine (0)*; *closer to the third incisor (1)*. (Fig. 5)
21. Distance between third incisors compared to the distance between canines (distances on the lingual borders): *distance between I3s greater than the distance between Cs (0)*; *distance between Cs greater than the distance between I3s (1)*. (Modified from Honey and Taylor (1978)).
22. Premaxillary notch: *present (0)*; *absent (1)*. (Based on Webb (1965)).
23. Shape of the anterior end of nasal bones in transverse section: *dorso-ventrally flattened (0)*; *arched dorsally (1)*. (Modified from ch.3, Scherer (2013); mod. ch.18, Harrison (1979, Fig. 3, p.11); Honey et al. (1998); Harrison (1985)).
24. Lateral border of nasal bones: *straight (0)*; *curved (1)*. (Based on Webb (1965); Fig. 5).
25. Lacrimal vacuity: *large, bordered by four bones (frontal, maxillary, lacrimal, nasal) (0)*; *small, bordered by four bones (frontal, maxillary, lacrimal, nasal) (1)*; *absent (2)*; *bordered by three bones (frontal, maxillary, lacrimal) (3)*; *bordered by four bones (frontal, maxillary, lacrimal, nasal), lacrimal border greatly reduced, frontal border greatly enlarged (4)*; *bordered by two bones (frontal, maxillary) (5)*. (Modified from ch.1, Scherer (2013); mod. ch.16, Harrison (1979); Honey et al. (1998); Harrison (1985); Fig. 6).

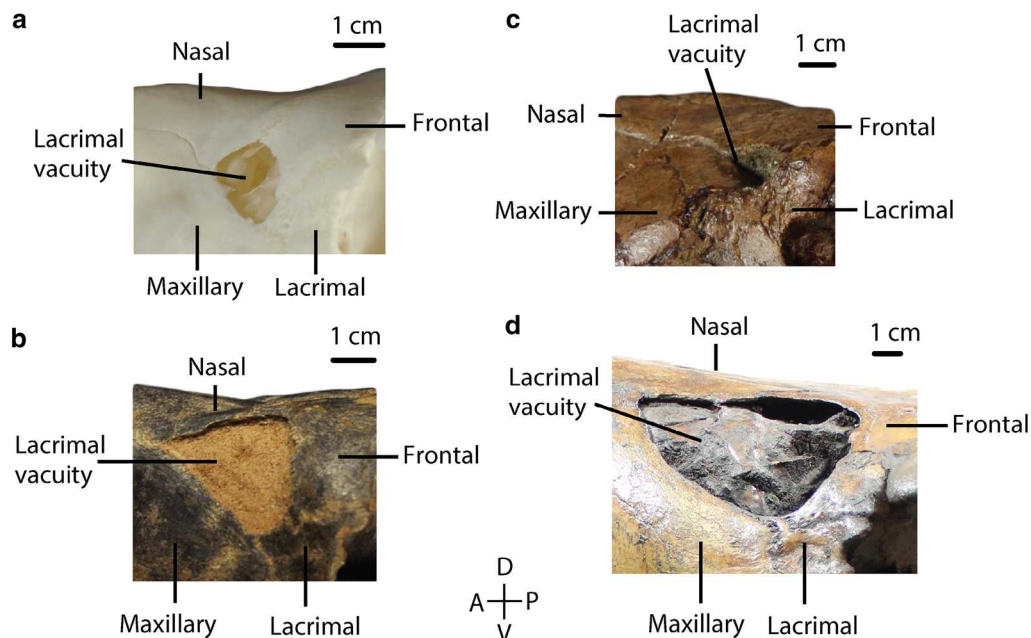




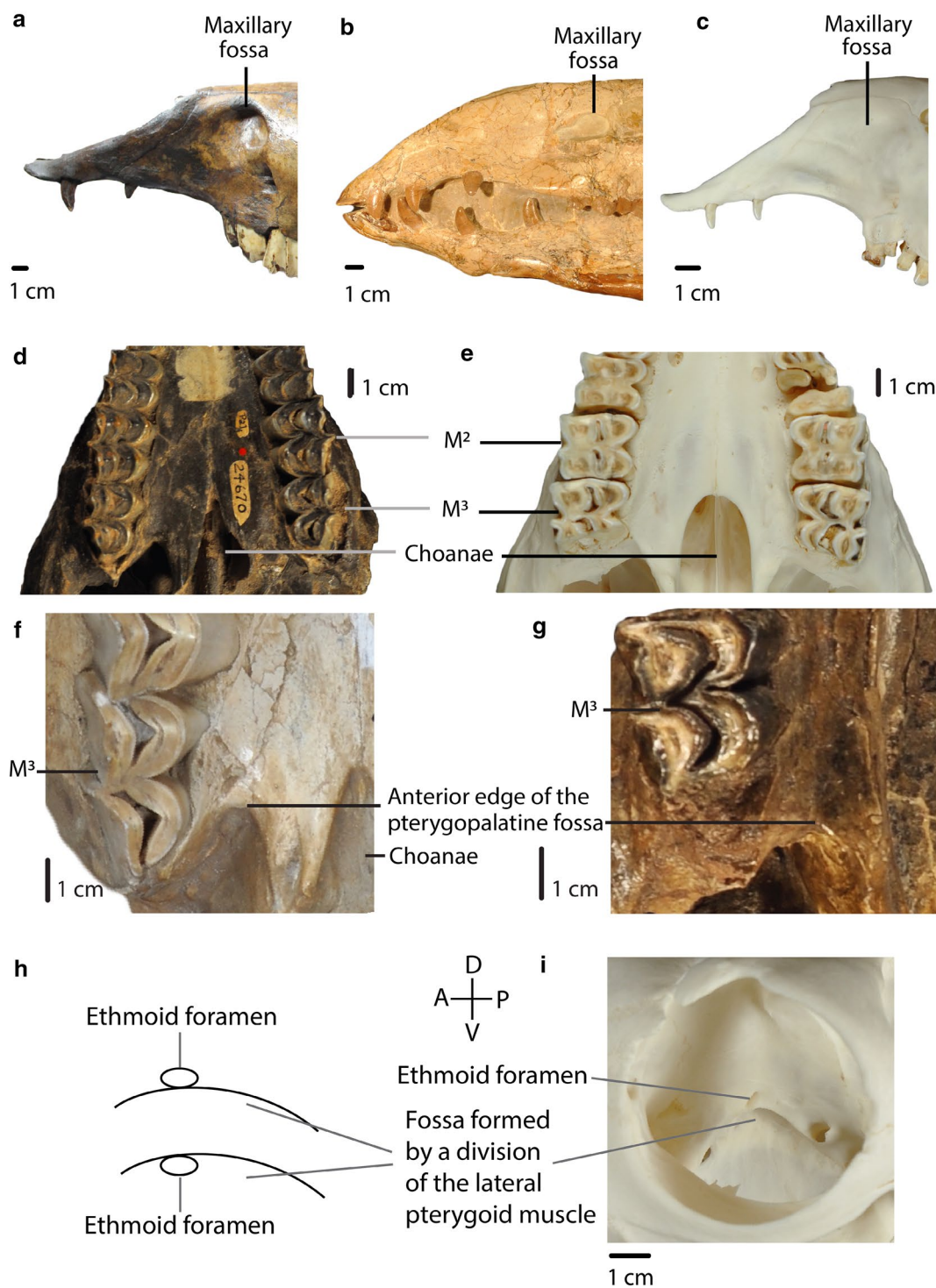
26. Maxillary fossa: *well-developed, large pocket (0); well-developed, small pocket (1); shallow or absent (2)*. (Modified from ch.2, Scherer (2013); mod. ch.“Maxillary Fossa”, p.46, Webb (1965); mod. ch.17, Harrison (1979); Honey et al. (1998); Harrison (1985); Fig. 7).
27. Zygomatic arch in lateral view: *curved (0); straight (1)*. (Ch.4, Scherer (2013); ch.20, Harrison (1979); mod. ch.“Zygoma”, Webb (1965, p.46); Harrison (1985)).
28. Orbital process of palatine: *present (0); narrow or absent (1)*. (Based on Webb (1965)).
29. Anterior edge of the choanae: *posterior to  $M^3$  (0), at the level of  $M^3$  or between  $M^3$  and  $M^2$  (1); at the level of  $M^2$  or between  $M^2$  and  $M^1$  (2)*. (Modified from ch.7, Scherer (2013), see Scherer (2009, Fig. 20, p.90)).
30. Shape of the anterior edge of the choanae: *V-shaped (0); U-shaped (1)*. (Fig. 7).
31. Anterior edge of the pterygopalatine fossa: *posterior to  $M^3$  (0); at the level of  $M^3$  (1)*. (Fig. 7)
32. Position of the ethmoid foramen: *above the fossa formed by a division of the lateral pterygoid muscle (0); in the fossa formed by a division of the lateral pterygoid muscle (1)*. (Based on Webb (1965)). (Fig. 7)

### Basicranial characters

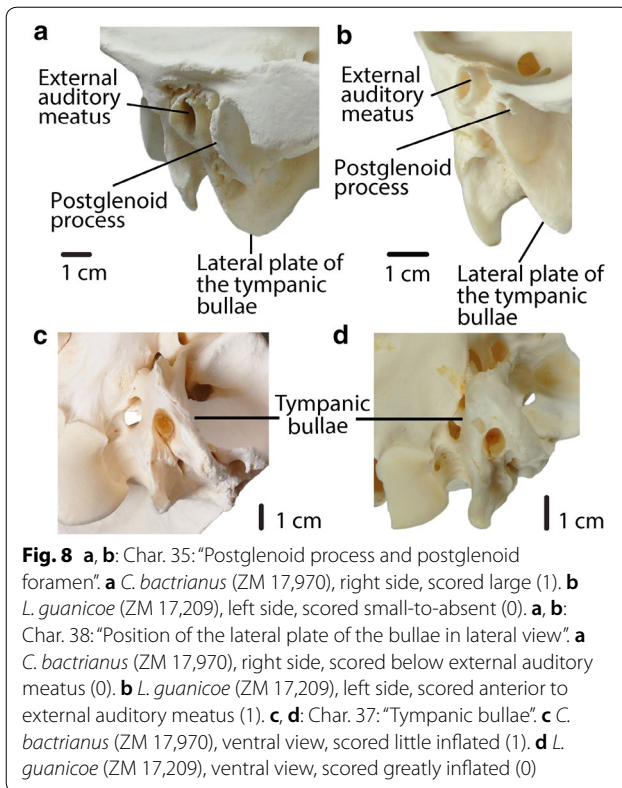
33. Foramen ovale and median lacerate foramen: *completely separated by a thick portion of the alisphenoid (0); completely separated by a thin spine of the alisphenoid (1); confluent (2)*. (Modified from ch.“Median Lacerate Foramen”, Webb (1965, p. 47)).
34. Position of the glenoid fossa: *close to the ventral surface of basisphenoid (0); well above the ventral surface of basisphenoid (1)*. (Modified from ch.“Glenoid Fossa”, Webb (1965, p. 46)).
35. Postglenoid process and postglenoid foramen: *small-to-absent (0); large (1)*. (Modified from ch.22, Harrison (1979); mod. ch.“Postglenoid Process”, Webb (1965, p. 47); Fig. 8).
36. Subsquamosal foramina: *small (0); large (1)*. (Modified from ch.“Temporal canal”, Webb (1965, p. 47)).
37. Tympanic bullae: *greatly inflated (0); little inflated (1)*. (Based on Webb (1965)) (Fig. 8).
38. Position of the lateral plate of the bullae in lateral view: *below external auditory meatus (0); anterior to external auditory meatus (1)*. (Fig. 8).



**Fig. 6** Char. 25: Bones bordering the lacrimal vacuity. **a** *L. guanicoe* (ZM 17,209), left side, scored small, bordered by four bones (frontal, maxillary, lacrimal, nasal) (1) **b** *P. vera* (AMNH 24,670), left side, scored large, bordered by four bones (frontal, maxillary, lacrimal, nasal) (0). **c** *A. alexandrae* (UCMP 26,015), left side, scored bordered by three bones (frontal, maxillary, and lacrimal) (3). **d** *C. hesternus* (UCMP 20,040), left side, scored bordered by four bones (frontal, maxillary, lacrimal, and nasal), lacrimal border greatly reduced, frontal border greatly enlarged (4)



**Fig. 7** A, B, C: Char. 26: "Maxillary fossa": **a** *C. hesternus* (UCMP 20,040), left side, scored well-developed, large pocket (0). **b** *A. robustus* (OMNH 016,560), right side, scored well-developed, small pocket (1). **c** *L. guanicoe* (ZM 17,209), left side, scored shallow or absent (2). **d, e**: Char. 30: "Shape of the anterior edge of the choanae": **d** *P. vera* (AMNH 24,670), ventral view, scored V-shaped (0). **e** *L. guanicoe* (ZM 17,209), ventral view, scored U-shaped (1). **f, g**: Char. 31: "Anterior edge of the pterygopalatine fossa": **f** *M. matthewi* (UCMP 31,100), scored at the level of M3 (1). **g** *T. longirostris* (CM 2498), scored posterior to M3 (0). **h, i**: Char. 32: "Position of the ethmoid foramen": **h** Upper: above the fossa formed by a division of the lateral pterygoid muscle (0). Lower: in the fossa formed by a division of the lateral pterygoid muscle (1). **i** *L. guanicoe* (ZM 17,209), right side, scored above the fossa formed by a division of the lateral pterygoid muscle (0)



### Dentary characters

39. Anterior part of the mandible in lateral view: *straight* (0); *angled dorsally* (1). (Fig. 9).
40. Distance between mesial end of the third lower incisors compared to the distance between the mesial end of fourth lower premolars: *distance between  $P_4$  greater or equal to distance between  $I_3$*  (0); *distance between  $I_3$  greater than distance between  $P_4$*  (1). (Fig. 9).
41. Portion of the mandibular ramus below the mandibular diastema in transverse section: *strongly narrowing dorsally, tear-drop shaped* (0); *little narrowing dorsally, ovoid* (1). (Fig. 9).
42. Angle of the mandible in lateral view: *posteriorly projected* (0); *ventrally produced with lateral flare* (1); *rounded* (2). (Modified from Honey et al. (1998); Honey and Taylor (1978), Fig. 9).
43. Coronoid process at the level of the mandibular condyle in lateral view: *angled posteriorly* (0); *straight* (1). (Fig. 9).
- Ratio characters* (see Additional files 3 and 4).
44. Rostral length (ratio between rostral length and length of the skull): *ratio higher than 0.56* (0); *ratio lower than 0.56* (1).

45. Retraction of nasals (ratio between the internasal suture and rostral length): *ratio higher than 0.6* (0); *retracted ratio from 0.6 to 0.57* (1); *ratio lower than 0.57* (2).
46. Orbit size (ratio between orbital width and length of the skull): *ratio higher than 0.138* (0); *ratio lower or equal to 0.138* (1).
47. Minimum postcanine width (ratio between the minimum postcanine width and width of the skull): *ratio higher than 0.155* (0); *ratio from 0.155 to 0.125* (1); *ratio from 0.125 to 0.055* (2); *ratio lower than 0.055* (3).
48. Length of mandibular symphysis (ratio between length of the symphysis and length of the mandible): *ratio higher than 0.255* (0); *ratio lower than 0.255* (1).
49. Height of the mandible (ratio between height of the mandible and length of the mandible): *ratio higher than 0.58* (0); *ratio lower than 0.58* (1).

### Systematic analysis

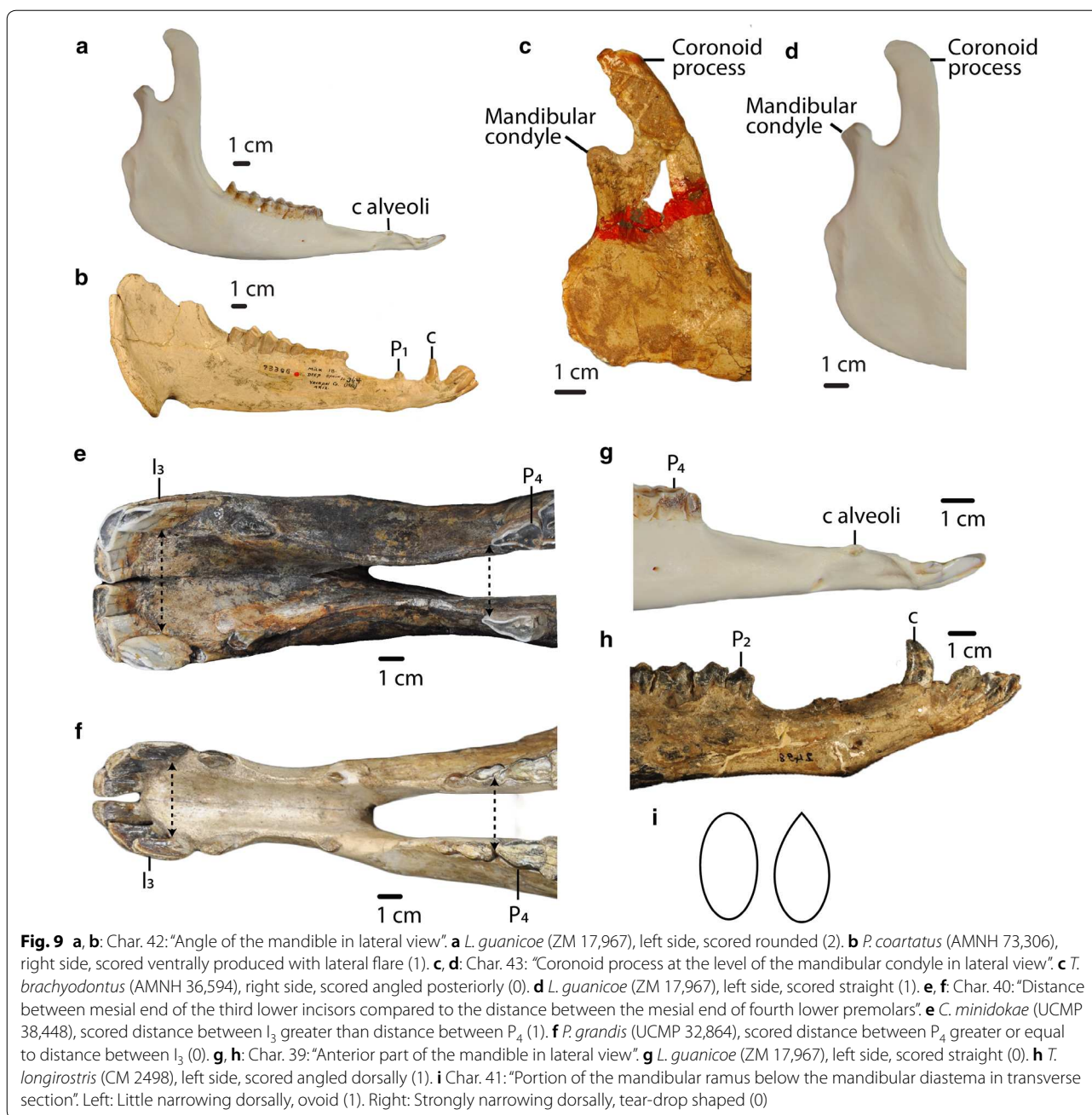
The character matrix is available in the Electronic Supplementary Material (Additional file 5). It comprises 49 morphological characters. Eight of these characters (chars. 8, 9, 11, 26, 29, 33, 45, and 47) were ordered. For the ingroup, we examined 18 species and 3 specimens identified at the genus level distributed among 13 Camelinae genera and 3 Protolabinae genera (Additional file 2).

We performed a cladistic analysis using parsimony with a traditional search in TNT (Goloboff and Catalano 2016). We ran 6000 replicates with 10 trees saved per replication. Bremer supports were obtained with suboptimal trees by 7 steps. Bootstraps values were obtained with a cutoff of 50.

### Description of PIMUZ A/V 4165

#### Locality and horizon

Specimen PIMUZ A/V 4165 was collected near the Parana river in the locality of San Nicolas (Barranca del Parana, San Nicolas, province of Buenos Aires, Argentina) (see Roth 1888, table 23; Voglino 2008, Fig. 1). There is no further information on the specimen or its precise stratigraphic position within las Barrancas. Geological age is Ensenadan (Early-to-Middle Pleistocene) according to museum archives and current curatorial staff at Museo de La Plata (Roth 1889; Schulthess 1920; Cione et al. 2015, A. Carlini, personal communication, July 18, 2020). However, there has been no modern revision of the stratigraphy at Roth's collection sites, so these dates must be interpreted with caution.



### Description

PIMUZ A/V 4165 is a nearly complete skull and mandible with all upper and lower teeth except a fragment of right  $I^3$  and left  $I_3$ . Postcranial elements include all metapodials, carpals, tarsals, radio-ulnae, humeri, and tibiae as well as the left femur, a damaged left scapula, and most phalanges. The axial skeleton (pelvis, vertebrae, and ribs) and the patellae are absent.

### Skull and mandible

The skull dimensions are similar to those of *Lama guanicoe*. Length from the posterior edge of the occipital condyle to the anterior end of the rostrum is 31.5 cm (tip broken) and the width is 13.7 cm measured at the greatest breadth of the skull. The skull displays a high degree of flexion at the boundary between the basicranium and palate. The angle of flexion is  $14^\circ$ —it is among the highest

in *Lama*, according to data provided by Webb (1965). As in all camelids, the rostrum becomes extremely tapered, starting anterior to the orbits.

The skull displays many morphological characters consistent with South American camelids. The zygomatic arch is sigmoid in lateral view, and the maxillary fossa is absent. The nasal bones are strongly retracted, and the anterior edge of the choanae is rounded and situated anteriorly at the level of  $M^2$ . In more basal camelids outside of the Lamini clade, such as *Poebrotherium*, the nasal bones are longer and extend closer to the anterior edge of the premaxillae, resulting in a smaller opening. In these earlier camelids, the anterior edge of the choanae tends to be more posteriorly positioned and “V-shaped” (Fig. 10).

### Teeth

The dental formula is  $I1/3 C1/1 P2/2 M3/3$ , with premolars in the third and fourth positions. Among Lamini, this dental formula is also found in *L. guanicoe* and *H. paradoxa*. PIMUZ A/V 4165 retains the  $P_3$ , which is absent in other camelids, such as *Camelops*. “Llama buttresses” (protostylids and parastylids) are well developed. Upper canines and incisors are small, compared to the great size which they reach in some males of extant South American camelids.

Anterior stylids (protostylids and parastylids) on  $M_3$  are prominent, and the anterior fossette on  $M_1$  is still present, making this specimen correspond to the “wear stage 3” of Breyer’s classification (Breyer 1977). The molar lochs and lochids are rounded in occlusal view. “Ribs”

(labial styles) on the upper molars are well developed and  $P_4$  is triangular, with a fossetid on the distal lobe.

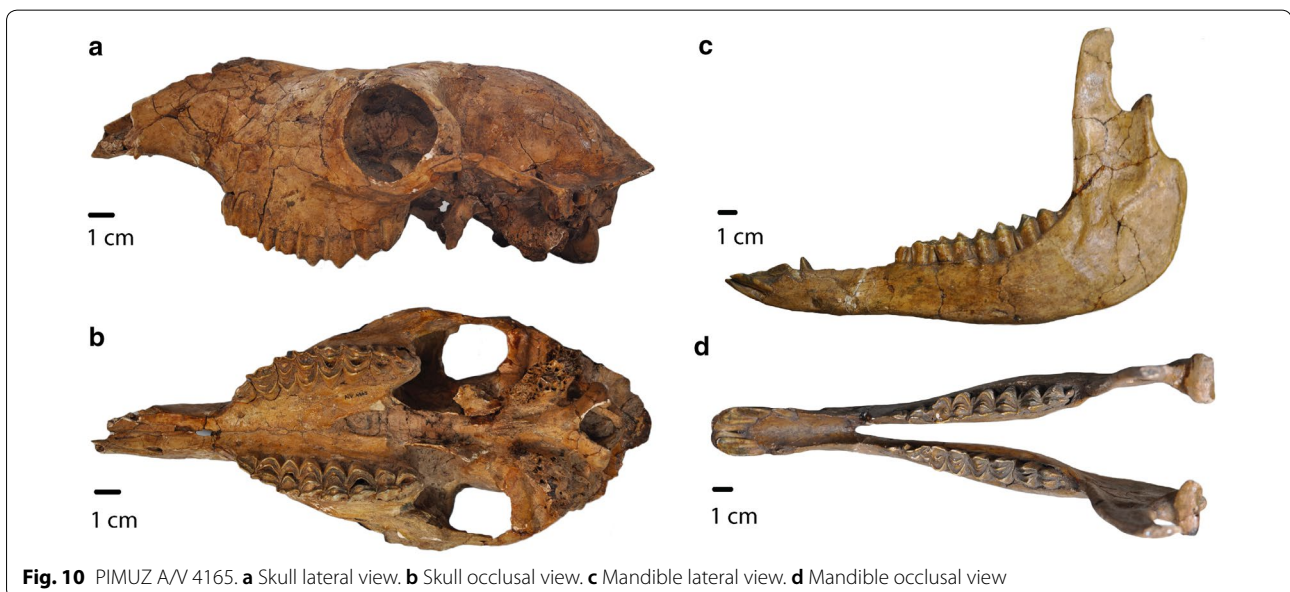
### Limbs

Compared to data on South American camelids from Scherer (2009, p. 163–166, figs. 18–19, p. 68–70), limb bones of this specimen are longer than in *Vicugna vicugna*. Most limb bones are also longer than *Lama guanicoe*, except its metacarpals which fall in the upper values of *L. guanicoe* (measurements of PIMUZ A/V 4165 in Additional file 1). Compared to *Palaeolama* and *Hemiauchenia*, metapodial lengths overlap with *Palaeolama* and are smaller than in *Hemiauchenia*. Tibiae and radio-ulnae lengths both overlap with *H. paradoxa* and *P. mirifica*, and, respectively, with *P. major* and *P. weddelli*. Stylopodials are shorter than in *Palaeolama* and *Hemiauchenia*.

Metacarpals are more gracile than in *Palaeolama*. Metatarsal gracility falls within the lower values of *Palaeolama major*. Metacarpals and metatarsals are more robust than in *Vicugna* and more in the range of *Lama*, *H. paradoxa*, and *H. macrocephala*.

Metacarpals and metatarsals are of comparable length, with the metacarpals slightly longer than the metatarsals. In *H. paradoxa*, *H. macrocephala*, *P. major*, *P. mirifica*, *L. castelnaudi*, *L. guanicoe*, and *V. provicugna* metacarpals are longer than metatarsals or of comparable length (char. 27 in Scherer (2013)). In *H. edensis*, *P. weddelli*, and *V. vicugna*, metacarpals are shorter than the metatarsals.

In the hind limbs, metatarsals are smaller than the tibiae and femora. Femora are shorter than the tibiae, as in *V. vicugna*, *H. paradoxa*, and *H. macrocephala* (char.



**Fig. 10** PIMUZ A/V 4165. **a** Skull lateral view. **b** Skull occlusal view. **c** Mandible lateral view. **d** Mandible occlusal view

26 in Scherer (2013)). In the forelimbs, humeri are the shortest bones, and radio-ulnae are the longest. This is in contrast to some Lamini, including extant forms and *Palaeolama*, whose humeri are longer or of comparable length than their metacarpals (char. 28 in Scherer (2013)) (Figs. 11, 12).

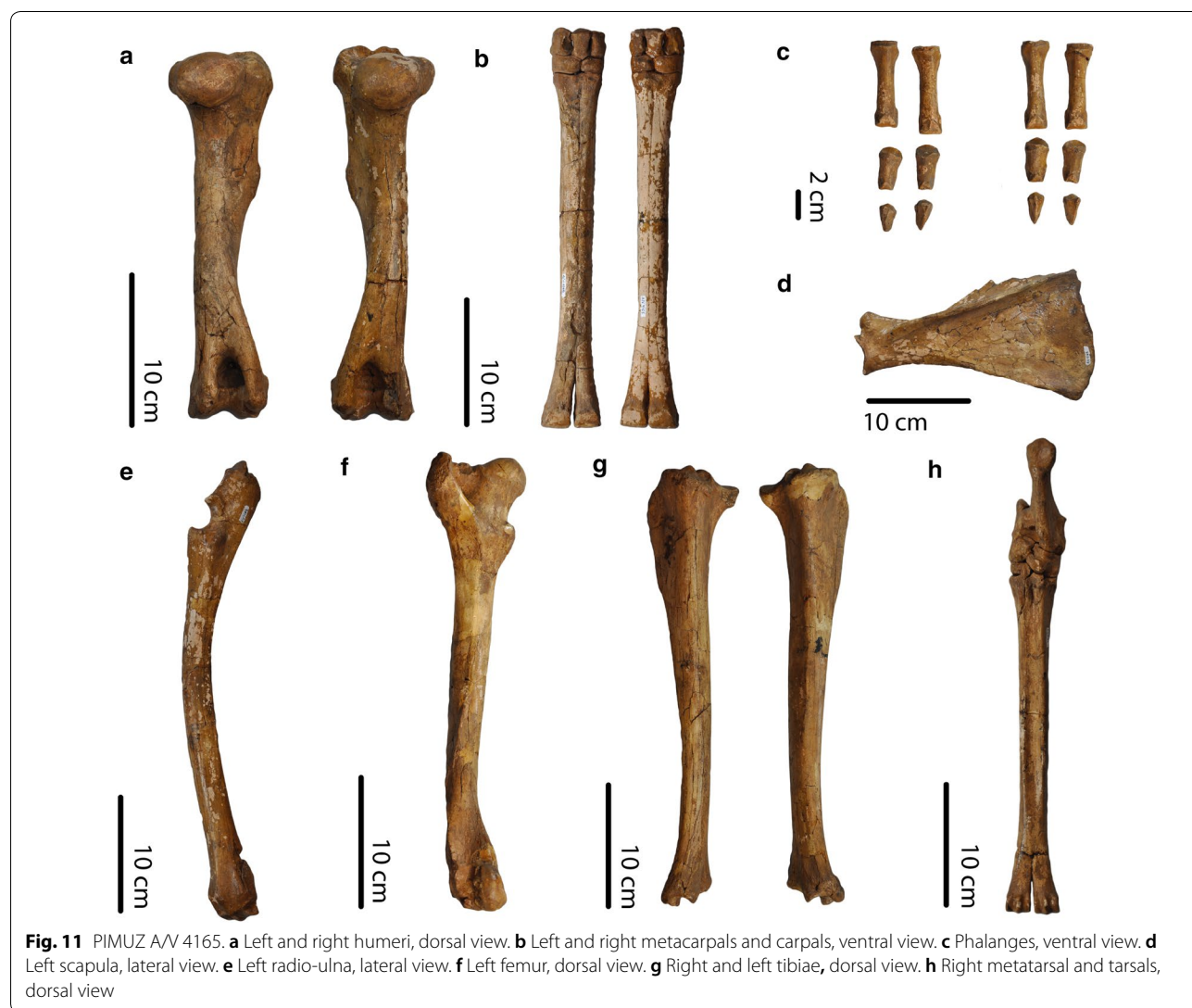
## Results

The cladistic analysis resulted in one most parsimonious tree (Fig. 12) with length 149 (CI=0.423, RI=0.675, RC=0.286). Synapomorphies are listed in Additional file 6. Both Bootstrap values and Bremer supports are low overall.

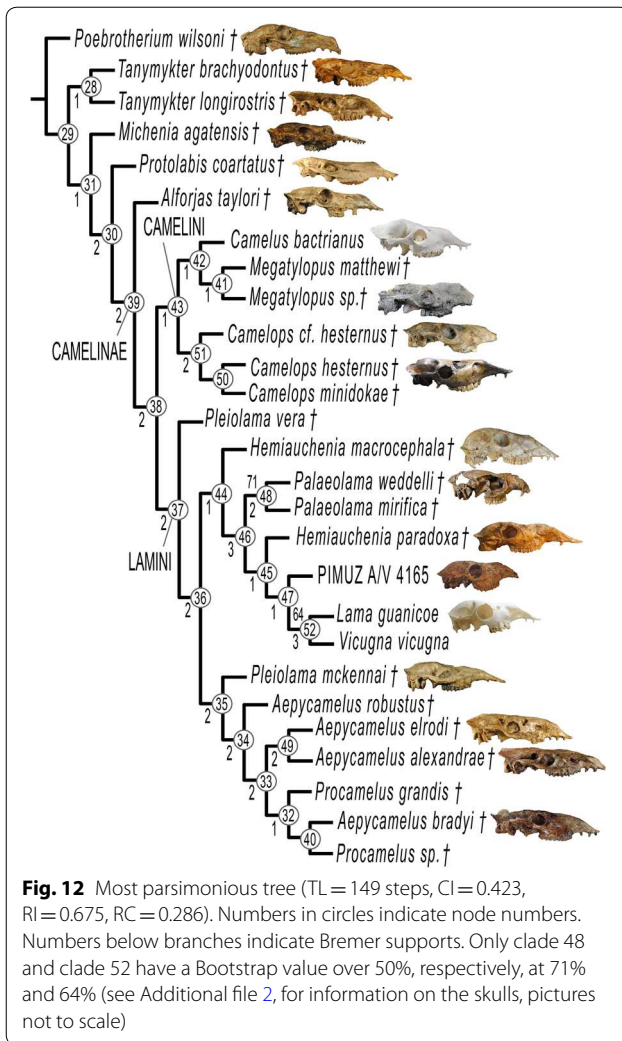
PIMUZ A/V 4165 is positioned within South American Lamini and is the sister taxon to the clade formed by *Lama guanicoe* and *Vicugna vicugna* (node 52). The synapomorphy that holds these three taxa together

at node 47 is a small postglenoid process (char. 35). PIMUZ A/V 4165 has a single autapomorphy: a lower canine that is close to  $I_3$  (<1 cm) (char. 7).

Well-developed protostylids and parastylids (“llama buttresses”) (char. 15) are the single synapomorphy of the clade formed by *H. paradoxa*, PIMUZ A/V 4165, *L. guanicoe*, and *V. vicugna* (node 45). *Hemiauchenia* appears as polyphyletic. *Palaeolama* (node 48) is monophyletic; its synapomorphies are a quadrangular  $P_4$  with fossetids in the mesial and distal lobes (char. 14), and triangular labial lophids (char. 16). The clade formed by *L. guanicoe* and *V. vicugna* (node 52) is held together by the absence of  $P^3$  and  $P_3$  (chars. 12 and 13), and the foramen ovale and median lacerate foramen are completely separated by a thin spine of the alisphenoid (char. 33). The confluence of these foramina is an autapomorphy of *L. guanicoe*.



**Fig. 11** PIMUZ A/V 4165. **a** Left and right humeri, dorsal view. **b** Left and right metacarpals and carpals, ventral view. **c** Phalanges, ventral view. **d** Left scapula, lateral view. **e** Left radio-ulna, lateral view. **f** Left femur, dorsal view. **g** Right and left tibiae, dorsal view. **h** Right metatarsal and tarsals, dorsal view



Two synapomorphies support the monophyly of Lamini (node 37): the anterior end of the nasals arched in the transverse section (char. 23) and the anterior edge of the choanae positioned at the level of  $M^2$  or between  $M^2$  and  $M^1$  (char. 29). An important split within the Lamini (node 36) separates two clades: one formed by North American taxa from the Miocene (node 35), and the other formed mostly by South American taxa and two North American ones (*H. macrocephala* and *P. mirifica*) from the Pliocene–Pleistocene to the present (node 44).

*Pleiolama* is not monophyletic, appearing at two different positions within Lamini: *P. vera* is the basal-most Lamini taxon (node 37), while *P. mckennai* is the most basal taxon in the clade formed mostly by North American Lamini from the Miocene (node 35). The genera *Aepycamelus* and *Procamelus* are, respectively, polyphyletic and paraphyletic. *A. elrodi* and *A. alexandrae* are held together by four synapomorphies (node 49); *A. bradyi* forms a clade with *Procamelus sp.* (node 40) and

*A. robustus* is more basal (node 34). *Procamelus* also appears in two different places, but close to each other: nodes 32 and 40.

*Camelops* (node 51) is monophyletic and placed within the Camelini (node 43). This genus is united by the absence of  $P^1$  (char. 8), anterior edge of the choanae U-shaped (char. 30), ethmoid foramen positioned in the fossa formed by a division of the lateral pterygoid muscle (char. 32), and the ovale and median lacerate foramina are completely separated by a thin spine of the alisphenoid (char. 33).

Camelinae (node 39) are monophyletic with three synapomorphies supporting the group: caniniform  $P^1$  and  $P_1$  (char. 8 and 9), and absence of  $P^2$  (char. 10). *Alforjas taylori* is a basal Camelinae, sister taxon to the clade formed by the ‘tribes’ Camelini and Lamini. Our definitions of Camelini and Lamini differ from those of Honey et al. (1998). We interpret the first major division within the Camelinae (node 38) as the split between these two clades.

The three synapomorphies uniting the Camelini (node 43) are the absence of  $P_3$  (char. 13), well-developed upper molar labial styles (“ribs”) (char. 17), and a lacrimal vacuity bordered by four bones, with the lacrimal border greatly reduced and the frontal border greatly enlarged (char. 25). The lacrimal vacuity bordered by two bones, frontal and maxillary, is an autapomorphy of *Camelus bactrianus*. *Megatylopus* and *Camelus bactrianus* (node 42) are united by a single synapomorphy: the absence of the premaxillary notch (char. 22).

The Protolabinae (composed of *Tanymyktar*, *Michenia*, and *Protolabis*) are paraphyletic in our results. The genus *Tanymyktar* (node 28) is monophyletic and supported by two synapomorphies.

## Discussion

Most of the terminal taxa were scored on the skull of a single specimen; therefore, our results could be altered by a larger sampling for each species and the addition of postcranial characters.

PIMUZ A/V 4165 appears most closely related to *L. guanicoe* and *V. vicugna* (node 47), to the exclusion of *H. paradoxa*. We found the postglenoid process (char. 35) of *H. paradoxa* to be large, whereas a small postglenoid process unites PIMUZ A/V 4165 with *L. guanicoe* and *V. vicugna*. A lower canine closer to  $I_3$  (< 1 cm) is the autapomorphy for PIMUZ A/V 4165. In *H. paradoxa*, *L. guanicoe*, and *V. vicugna*, the lower canine is positioned far from  $I_3$  ( $\geq 1$  cm) (char. 7).

We found important differences between the postcranials of PIMUZ A/V 4165 and those of *L. guanicoe* and *V. vicugna*. Among these, PIMUZ A/V 4165 has longer limb bones than *V. vicugna* and longer or among the

largest sizes of *L. guanicoe* (Scherer 2009). In *V. vicugna*, metacarpals are shorter than metatarsals (Scherer 2013), whereas in PIMUZ A/V 4165, metacarpals are slightly longer than metatarsals. Metacarpals are longer than the humeri in PIMUZ A/V 4165, in contrast to *L. guanicoe* whose metacarpals are shorter than the humeri (Scherer 2009, p. 201), and *V. vicugna* whose metacarpals and humeri lengths are comparable (Scherer 2009, p. 232).

The postcranial characters observable on PIMUZ A/V 4165 are most similar to *H. paradoxa* in Scherer's (2013) matrix (see Additional file 7). For example, as in PIMUZ A/V 4165, metacarpals are longer than humeri in *H. paradoxa*. However, an analysis using Scherer's (2013) matrix results in a polytomy with PIMUZ A/V 4165 within the Lamini (see Additional file 7). Furthermore, compared to data from Scherer (2009), PIMUZ A/V 4165 has shorter metatarsals and humeri than *H. paradoxa*. Although the parameters of our study cannot confirm it, the fact that the morphology of PIMUZ A/V 4165 is not consistent with any single species could indicate that it belongs to a new species.

The monophyly of the Protolabinae recovered by Honey and Taylor (1978), Harrison (1985), Honey et al. (1998) and Scherer (2013) is not supported here. The species previously assigned to this group display much morphological disparity. For example, *Michenia agatenensis* and *Tanymycter* retain  $I^1$  and  $I^2$  (chars. 1 and 2), while *Protolabis coartatus* has lost these teeth. We found these absences (chars. 1 and 2) to be synapomorphies uniting *P. coartatus* to the Camelinae (node 30), whereas in Scherer (2013), these were synapomorphies in her definition of the Camelinae. Here, the caniniform  $P_1$  and  $P^1$  (chars. 8 and 9) and the absence of  $P^2$  (char. 10) are also synapomorphies of the Camelinae (node 39). Of these, the synapomorphies of characters 8 and 9 are in agreement with Honey et al. (1998), and the synapomorphy of character 10 is in agreement with Harrison (1979).

Our phylogeny places *Alforjas taylori* in the most basal position among the Camelinae—this is in significant contrast to previous phylogenies. Harrison (1979, 1985) and Scherer (2013) placed this species in a more derived position within Lamini. In Scherer (2013), this was partly due to postcranial characters. Here and in Scherer (2013), the presence of the protostylids and parastylids (“llama butresses”) (char. 15) on the lower molars is not a synapomorphy for the Lamini (node 37). In contrast, Harrison (1979, 1985) found this feature as a synapomorphy for the Lamini.

In Harrison (1979, 1985) and Honey et al. (1998), *Alforjas* and *Camelops* were united partly by a similar degree of hypsodonty. In these previous works and in Scherer (2013), *Alforjas* and *Camelops* were placed within Lamini and shared with them dorsally arched

nasal bones in the transverse section (char. 23). We agree with this being a synapomorphy for Lamini (node 37), but we were unable to score it on *A. taylori* due to the poor preservation of the nasal bones. In *C. hesternus* and *C. cf. hesternus*, we found this feature to be dorso-ventrally flattened.

In our results, *Camelops* (node 51) is included in the Camelini (node 43). This is in agreement with recent proteomic and genetic studies which defined *Camelops* as more closely related to Camelini than to the Lamini (Buckley et al., 2019; Heintzman et al., 2015).

Harrison (1979, 1985) and Scherer (2013) found rounded canines (char. 5) to be a synapomorphy of the Camelini, whereas we found it to be a synapomorphy of the clade composed of *Aepycamelus* and *Procamelus* (node 34). *Procamelus* appeared as a basal Camelinae in Scherer (2013) and Harrison (1979). Honey et al. (1998) placed it within the Camelini. Here, as in Harrison (1985), *Procamelus* is more derived and placed within the Lamini. *Aepycamelus* is also positioned within the Lamini, as in Scherer (2013) and Honey et al. (1998). However, these genera are not monophyletic in our results, which indicates that further taxonomic revisions are necessary both taxa. We agree with observations by Honey et al. (1998) on the ambiguous status of *Aepycamelus*. We found important morphological differences between the different species of *Aepycamelus*. For instance,  $P^2$  (char. 10) is absent in *A. robustus*, whereas it is present in *A. alexandrae*, *A. bradyi*, and *A. elrodi*.

Scherer (2013) did not find *Hemiauchenia* to be monophyletic and we agree on this point. However, we also find a polyphyletic separation between *H. paradoxa* and *H. macrocephala*. The monophyly of *Palaeolama* (node 48) was also recovered in Scherer (2013), and in both studies, a quadrangular  $P_4$ , with fossetids in the mesial and distal lobes (char. 14), is a synapomorphy for this genus. Here, *Palaeolama* is closely related to the genera *Hemiauchenia*, *Vicugna*, and *Lama*, whereas in Scherer (2013), *Palaeolama* was in a more basal position.

#### Dental characters

When present,  $P^1$ ,  $P_1$ , and  $P_2$  (chars. 8, 9, and 11) are either premolariform or caniniform. We ordered these characters, assuming a linear evolution from a plesiomorphic premolariform state towards a caniniform state, with eventual loss in more derived taxa. At multiple points in our results, the states for these characters did not follow this linear path. For example, one of the synapomorphies at node 34 is a change from absent to a caniniform  $P_2$  (char. 11). Similarly, on node 33,  $P_2$  changes from caniniform to premolariform.



### Implications on the evolution of the Camelinae

The oldest Camelinae fossils in our analysis are those of *Aepycamelus alexandrae*, *Aepycamelus elrodi*, and *Aepycamelus robustus*, and are geologically aged to the early Barstovian, approximately (age unknown for the specimen OMNH 016,560, *A. robustus*). They all form part of the Lamini. This would imply that the appearance of the Camelinae, the divergence of Lamini from Camelini (node 38), and the main split among Lamini (node 36), all occurred before 16 Mya. Such a finding would suggest multiple ghost lineages across most of our trees. A more parsimonious conclusion would be a continuing unstable position for *Aepycamelus*, with a possible earlier divergence time for the Lamini and Camelini clades (node 38) than that estimated by Honey et al. (1998).

### Conclusion

Based on craniomandibular and dental characters, PIMUZ A/V 4165 appears more closely related to *L. guanicoe* and *V. vicugna*. However, there are important differences between the postcranials of *L. guanicoe*, *V. vicugna*, and this specimen. Although observable postcranial characters on PIMUZ A/V 4165 coincide mostly with *H. paradoxa* (Scherer 2013), their affinity was not supported in our analysis based on that matrix. We found differences in the length of some limb bones of PIMUZ A/V 4165 and *H. paradoxa*. Therefore, we suggest that PIMUZ A/V 4165 could be considered as a new species. Future systematic studies of this group with a larger sample will be able to test our results.

We propose several new craniomandibular and dental characters and restructure some of the relationships among Camelinae. Our results do not support the monophyly of the *Protolabinae*, *Aepycamelus*, *Procamelus*, *Pleiolama*, or *Hemiauchenia*. The monophyly of the *Camelinae*, *Camelops*, and *Palaeolama* are supported. We hypothesize that *Aepycamelus* and *Procamelus* form part of the Lamini and that *Camelops* is a member of the Camelini.

Several synapomorphies presented in earlier phylogenies were also recovered here, including the arched nasals (char. 23) for the Lamini (Harrison 1979, 1985; Honey et al. 1998; Scherer 2013), and the quadrangular P<sub>4</sub> with fossetids in the mesial and distal lobes (char. 14) for *Palaeolama* (Scherer 2013). We disagree, however, with the “llama buttresses” (char. 15) as a synapomorphy for the Lamini (Harrison 1979, 1985). Instead, we find this feature only in a more derived subgroup of this clade (node 45). We also propose new synapomorphies for several clades. Among these, we found the absence of the premaxillary notch (char. 22) to be a synapomorphy for the clade joining *Megatylopus* and *Camelus bactrianus* (node 42).

Our results superficially indicate a potential divergence between Camelini and Lamini clades before 16 Mya. This would imply that the Lamini and Camelini taxa were already present during the fourth radiation of the camelids (late Hemingfordian-to-early Barstovian) when Honey et al. (1998) date the appearance of Camelinae. More likely, the placement of the geologically older *Aepycamelus* taxa in a derived position is an indication of their continued ambiguous phylogenetic placement.

### Supplementary information

**Supplementary information** accompanies this paper at <https://doi.org/10.1186/s13358-020-00208-6>

**Additional file 1.** Postcranial measurements of PIMUZ A/V 4165.

**Additional file 2.** List of specimens studied and their dental wear stage.

**Additional file 3.** Figures of measurements for ratio characters (chars. 44–49).

**Additional file 4.** Measurements for ratio characters.

**Additional file 5.** Character matrix.

**Additional file 6.** Synapomorphies.

**Additional file 7.** Phylogeny based on matrix by Scherer (2013) with PIMUZ A/V 4165.

### Abbreviations

UF: University of Florida; UCMP: University of California Museum of Paleontology; AMNH: American Museum of Natural History; OMNH: Oklahoma Museum of Natural History; PIMUZ: Paläontologisches Museum der Universität Zürich; ZM: Zoologisches Museum der Universität Zürich; CM: Carnegie Museum of Natural History; MACN PV: Colección de Paleovertebrados, Museo Argentino de Ciencias Naturales “Bernardino Rivadavia”; MHNH: Muséum National d’Histoire Naturelle, Paris.

### Acknowledgements

We thank Richard C. Hulbert Jr. (UF), Pat Holroyd (UCMP), Judith Galkin (AMNH), Jennifer Larsen (OMNH), Amy Henrici (CM), Sergio Lucero (MACN PV), and Martina Schenkel (UZH) for access to collections and material under their care; Guillaume Billet (MHNH) for bringing the specimen PUN1 to Zürich; Eric Holt (UCMP) for providing us with pictures of the specimen UCMP 20040; Gabriel Aguirre Fernandez for his advice; anonymous reviewers for their helpful comments; Financial support SNF grant no. 31003A-169395.

### Authors’ contributions

SL collected all data and wrote this paper. AMB advised on phylogenetics, character formulation, and revised paper. MSV advised on phylogenetic methods, and revised and approved paper. All authors read and approved the final manuscript.

### Funding

SNF Grant No. 31003A-169395.

### Availability of data and materials

Data for the comparison of limb lengths, gracility and proportions used in the description of PIMUZ A/V 4165 can be found in Scherer (2009). The matrix used for the phylogeny, list of characters, measurements for characters, and the list of specimens studied are included in the Supplementary information files.

### Competing interests

The authors declare that they have no competing interests.

Received: 17 April 2020 Accepted: 18 August 2020  
Published online: 07 October 2020

## References

- Baskin, J., & Thomas, R. (2016). A review of *Camelops* (Mammalia, Artiodactyla, Camelidae), a giant llama from the Middle and Late Pleistocene (Irvingtonian and Rancholabrean) of North America. *Historical Biology*, 28, 120–127. <https://doi.org/10.1080/08912963.2015.1020800>.
- Bond, M. (1999). *Santiago Roth*. *Museo*, 2(13), 33–37.
- Bravo-Cuevas, V. M., & Jiménez-Hidalgo, E. (2015). First reported occurrence of *Palaeolama mirifica* (Camelidae, Lamini) from the Late Pleistocene (Rancholabrean) of Puebla, central Mexico. *Boletín de la Sociedad Geológica Mexicana*, 67, 13–20.
- Breyer, J. (1977). Intra- and interspecific variation in the lower jaw of *Hemiauchenia*. *Journal of Paleontology*, 51, 527–535.
- Buckley, M., Lawless, C., & Rybczynski, N. (2019). Collagen sequence analysis of fossil camels, *Camelops* and c.f. *Paracamelus*, from the Arctic and sub-Arctic of Plio-Pleistocene North America. *Journal of Proteomics*, 194, 218–225. <https://doi.org/10.1016/j.jprot.2018.11.014>.
- Cione AL, Tonni EP (2001) Correlation of Pliocene to Holocene southern South American and European vertebrate-bearing units. In: Rook L, Torre D (eds) *Bollettino della Società Paleontologica Italiana* 40(2):167–173 (Neogene and Quaternary continental stratigraphy and mammal evolution)
- Cione AL, Tonni EP (2005) Bioestratigrafía basada en mamíferos del Cenozoico superior de la provincia de Buenos Aires, Argentina. In: de Barrio RE, Etchegerry RO, Caballé MF, Llambías E (eds) *Geología y Recursos Minerales de la Provincia de Buenos Aires*. Relatorio del XVI Congreso Geológico Argentino, vol 11, pp 183–200
- Cione, A. L., Gasparini, G. M., Soibelzon, E., Soibelzon, L. H., & Tonni, E. P. (2015). Continental Relationships, Chronostratigraphy, Climates, and Mammalian Biogeography of Southern South America Since Late Miocene. *The Great American Biotic Interchange* (pp. 9–69). Netherlands, Dordrecht: Springer.
- Cui, P., Ji, R., Ding, F., et al. (2007). A complete mitochondrial genome sequence of the wild two-humped camel (*Camelus bactrianus ferus*): an evolutionary history of camelidae. *BMC Genomics*, 8, 241. <https://doi.org/10.1186/1471-2164-8-241>.
- El Allali, K., Achaâban, M., & Ouassat, M. (2017). Anatomy of the dromedary head skeleton revisited. *J Morphol Sci*, 34, 073–088. <https://doi.org/10.4322/jms.100916>.
- Goloboff, P. A., & Catalano, S. A. (2016). TNT version 1.5, including a full implementation of phylogenetic morphometrics. *Cladistics*, 32, 221–238. <https://doi.org/10.1111/cla.12160>.
- Harrison, J. A. (1979). Revision of the Camelinae (Artiodactyla, Tylopoda) and description of the new genus *Alforjas*. *University of Kansas Paleontological Contributions*, 95, 1–20.
- Harrison, J. A. (1985). Giant Camels from the Cenozoic of North America. *Smithsonian Contributions to Paleobiology*, 57, 1–29. <https://doi.org/10.5479/si.00810266.57.1>.
- Heintzman, P. D., Zazula, G. D., Cahill, J. A., et al. (2015). Genomic Data from Extinct North American *Camelops* Revise Camel Evolutionary History. *Molecular Biology and Evolution*, 32, 2433–2440. <https://doi.org/10.1093/molbev/msv128>.
- Hershkovitz, P. (1982). Neotropical deer (Cervidae), part I, Pudus, genus *Pudu* Gray. *Fieldiana Zoology*, 11, 1–86.
- Honey JG (2007) Family Camelidae. In: The Evolution of Artiodactyls. Johns Hopkins University Press, pp 177–188
- Honey, J. G., Harrison, J., Prothero, D., & Stevens, M. (1998). Camelidae. *Evolution of Tertiary Mammals of North America* (pp. 439–462). Scott KM, Jacobs LL, New York: Cambridge University Press. Janis CM.
- Honey, J. G., & Taylor, B. (1978). A generic revision of the Protolabidini (Mammalia, Camelidae), with a description of two new Protolabidines. *Bulletin of the American Museum of Natural History*, 161, 367–426.
- Kurtén, B., & Anderson, E. (1980). *Pleistocene mammals of North America*. New York: Columbia University Press.
- Pacheco Torres, V. R., Altamiro, A. J., & Guerra Porras, E. S. (1986). *The osteology of South American camelids*. Institute of Archaeology: University of California, Los Angeles.
- Pagnac, D. C. (2005). New camels (Mammalia: Artiodactyla) from the Barstow Formation (middle Miocene), San Bernardino County, California. *Paleobios*, 25, 19–31.
- Peterson, O. A. (1911). A new camel from the Miocene of western Nebraska. *Annals of the Carnegie Museum*, 7, 260–266.
- Prothero, D. R. (2005). Camelidae. In R. Emry (Ed.), *The Terrestrial Eocene-Oligocene Transition in North America* (pp. 609–651). Cambridge: Cambridge University Press.
- Roth, S. (1888). Beobachtungen über Entstehung und Alter der Pampasformation in Argentinien. *Zeitschrift der deutschen geologischen Gesellschaft*, 40, 375–464.
- Roth S (1889) Fossiles de la Pampa, Amérique du Sud (Catalogue No.5)
- Scherer, C. S. (2013). The Camelidae (Mammalia, Artiodactyla) from the Quaternary of South America: Cladistic and Biogeographic Hypotheses. *Journal of Mammalian Evolution*, 20, 45–56. <https://doi.org/10.1007/s10914-012-9203-4>.
- Scherer CS (2009) Os Camelidae Lamini (Mammalia, Artiodactyla) do Pleistoceno da América do Sul : aspectos taxonômicos e filogenéticos. (PhD thesis, unpubl.), Universidade Federal do Rio Grande do Sul. Instituto de Geociências, Programa de Pós-Graduação em Geociências, Porto Alegre, Brasil. <https://www.lume.ufrgs.br/handle/10183/49733>
- Scherer, C. S., Ferigolo, J., Ribeiro, A. M., & Guerra, C. C. (2007). Contribution to the knowledge of *Hemiauchenia paradoxa* (Artiodactyla, Camelidae) from the Pleistocene of southern Brazil. *Revista brasileira de paleontologia*, 10, 35–52.
- Schulthess, B. (1920). Beiträge zur Kenntnis der Xenarthra auf Grund der Santiago Roth'schen Sammlung des Zoologischen Museums der Universität Zürich. *Imprimerie Albert Kundig*, 44, 58–59.
- Shockey, B. J. (1997). Two new notoungulates (Family Notohippidae) from the Salla Beds of Bolivia (Deseadan: late Oligocene): systematics and functional morphology. *Journal of Vertebrate Paleontology*, 17, 584–599. <https://doi.org/10.1080/02724634.1997.10011005>.
- Soibelzon, E., Tonni, E. P., & Bidegain, J. C. (2008). Cronología, magnetoestratigrafía y caracterización bioestratigráfica del Ensenadense (Pleistoceno inferior-medio) en la ciudad de Buenos Aires. *Revista de la Asociación Geológica Argentina*, 63(3), 421–429.
- Vogolino D (2008) Las barrancas del río Paraná en la provincia de Buenos Aires. Un escalón en la llanura. In: CSIGA (ed) Sitios de Interés Geológico de la República Argentina. Instituto de Geología y Recursos Minerales, Servicio Geológico Minero Argentino, Buenos Aires, annals 46, II, pp 461–476
- Verzi, D., Deschamps, C., & Tonni, E. P. (2004). Biostratigraphic and palaeoclimatic meaning of the Middle Pleistocene South American rodent *Ctenomys kraglievichi* (Caviomorpha, Octodontidae). *Palaeogeography, Palaeoclimatology, Palaeoecology*, 212, 315–329. [https://doi.org/10.1016/S0031-0182\(04\)00328-1](https://doi.org/10.1016/S0031-0182(04)00328-1).
- Webb, S. D. (1965). The osteology of *Camelops*. *Bulletin of the Los Angeles County Museum*, 1, 1–54.
- Webb, S. D., & Meachen, J. (2004). On the origin of lamine Camelidae including a new genus from the Late Miocene of the high plains. *Bulletin of Carnegie Museum of Natural History*, 36, 349–362. [https://doi.org/10.2992/0145-9058\(2004\)36\[349:OTOOLC\]2.0.CO;2](https://doi.org/10.2992/0145-9058(2004)36[349:OTOOLC]2.0.CO;2).
- Woodburne, M. O. (2004). *Late Cretaceous and Cenozoic Mammals of North America: Biostratigraphy and Geochronology*. Columbia: Columbia University Press.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.