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Abstract: Ectomycorrhizas of Scots pine (*Pinus sylvestris* L.) and beech (*Fagus sylvatica* L.) were sampled in a mature forest ecosystem exposed for more than 40 years to moderate levels of gaseous and dust pollutants. Soil of the forest site was characterised by low pH and accumulation of heavy metals (Pb, Mn, Zn, Cu, Cd, Fe). Mycorrhizal vitality and enzyme activity of the root-surface and soil acid phosphatase (AcPase) were studied at 17 measurement points (0–5 cm soil depth) in relation to the content of inorganic phosphate (Pi) and aluminium ions (Al^{3+}) in the soil. Anatomy of Scots pine and beech mycorrhizas taken from different measurement points was observed. The concentration of essential nutrients (C, N, P, Ca, Mg) and the ratios Ca/Al, Mg/Al and N/P were analysed in fine roots. High concentrations of Al^{3+} in the soil (40–118 meq kg^{-1}) and low levels of Pi (12–44 mg P_2O_5 kg^{-1}) were accompanied by high activity of the root-surface AcPase of pine and beech mycorrhizas (25–67 and 33–86 μmol pNP g^{-1} fresh weight h^{-1}, respectively) and soil AcPase (6.8–22.4 μmol pNP g^{-1} dry weight h^{-1}). The results indicate that fine tree roots are undoubtedly under stress as evidenced by a disturbance in P uptake and accumulation. However, the high vitality of mycorrhizas and the high Ca/Al ratio in fine roots suggest that the defence mechanisms of mycorrhizas and the rhizosphere are still able to ameliorate the influence of anthropogenic pollution.

Additional key words: acid phosphatase, fluorescein diacetate, soil, *Fagus sylvatica*, *Pinus sylvestris*, root anatomy

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Introduction

It is well established that anthropogenic stresses are important factors in forest decline (Schütt and Cowling 1985). Many forest ecosystems in Poland have been exposed to heavy industrial pollution, mainly sulphur dioxide (SO_2), nitrogen oxides (NO_x), hydrogen fluoride (HF), acid rain and heavy metals for about half a century (Białobok 1984; Godzik 1990). In recent years the phytotoxic effects of increased levels of tropospheric ozone (O_3) has been emphasised (Bytnerowicz et al. 1993; Godzik 1997). Although during the recent years the emissions of SO_2 to the atmosphere in Poland significantly decreased, toxic gases such as NO_x, HF, NH_3 and O_3 are important compounds influencing forest ecosystems and the area of injured forests is still increasing (Vancura et al. 2000).

In the early eighties attention has been paid to the response of tree root systems to anthropogenic pollutants. It has been suggested that forest decline is a consequence of damage to fine tree roots (Ulrich 1983; Meyer 1985) and that the sensitivity of roots to environmental pollution could be used as a marker for physiological damage to trees prior to visible symptoms of decline (Richards 1989). Forests of temperate regions usually grow on acidic soils, with a high accumulation of organic matter at the soil surface and limited availability of nutrients. An addi-

tional input of acid deposition may cause a decline in the decomposition rate of organic matter and chemical changes resulting in an increased availability of soluble aluminium, manganese and heavy metals and an accelerated leaching of base cations (Ca^{2+}, Mg^{2+}, K^+, Na^+) down the soil profile (Ulrich 1983; Grodziński et al. 1990).

Most tree species in temperate forest ecosystems are ectomycorrhizal. Ectomycorrhizas (fungal mantle and extramatrical mycelium) are known to improve the uptake of water and nutrients (particularly nitrogen and phosphorus) by plants from the soil solution. The influx of Pi in mycorrhizal roots can be 3–5 times higher than in nonmycorrhizal roots (Smith and Read 1997). A major part of the phosphorus (up to 80% or more) contained in the upper horizons of forest soils, where most ectomycorrhizas are localised, is present in organic forms (e.g. Dalal 1977). Ectomycorrhizas are able to acquire large amounts of phosphorus from organic sources that are not available directly to the plant. One of the mechanisms by which mycorrhizal fungi and mycorrhizas may increase the uptake of phosphorus from the soil is the production of surface-accessible acid phosphatase (acid phosphomonoesterase = AcPase, EC 3.1.3.2) (Harley 1989; Smith and Read 1997). In ectomycorrhizal root tips the activity of AcPase has been detected mainly in the matrix of the fungal mantle and along the plasma membranes of the Hartig net (Dexheimer et al. 1986; Turnau and Dexheimer 1995). The activity of AcPase has been shown to be more active in ectomycorrhizal than in nonmycorrhizal roots of forest trees (Barlett and Lewis 1973; Williamson and Alexander 1975; MacFall et al. 1991; Kieliszewska-Rokicka 1992; Pasqualini et al. 1992). The enzyme activity can vary between ectomycorrhizal morphotypes (Antibus and Linkins 1992, Kieliszewska-Rokicka 1999), and exhibit seasonal variation with a maximum activity in autumn (Rejšek 1991; Antibus and Linkins 1992).

The main function of mycorrhizas – nutrient transfer between the fungus and the plant, take place at the plant-fungus interface in living mycorrhizas. From a functional standpoint in vital ectomycorrhizas, both the fungus hyphae and the plant cells are vital. Initiation, ageing and death of ectomycorrhizas of forest trees in temperate regions are dynamic processes dependent on the seasonal cycle, weather conditions and the plant and the fungus species (Smith and Read 1997). Anthropogenic stresses can decrease the life-span of individual mycorrhizas, the frequency of vital mycorrhizas in the pool studied (Ritter et al. 1989; Kottke et al. 1993; Münzenberger et al. 1995), and nutrient balance in fine roots (Joslin et al. 1988; Persson and Majdi 1995).

Forest soils contain extracellular acid phosphatase (AcPase) released by plant roots, mycorrhizas and soil microorganisms (fungi, bacteria, algae). The soil AcPase may exist temporarily in complexes enzyme-substrate, adsorbed on clay minerals or associated with humic colloids (Burns 1982), takes part in the hydrolysis of polyphosphates and organic phosphates in the soil (Boutin and Roux 1974) and in Pi absorption by the plant (Bieleski 1973). Inverse relations have been shown between Pi supply and AcPase activity, both in soil (Spiers and McGill 1979) and in ectomycorrhizas of forest trees (Alexander and Hardy 1981; MacFall et al. 1992; Kieliszewska-Rokicka 1992; Pasqualini et al. 1992). Decreased activity of the root-surface AcPase has been reported from forest sites exposed to anthropogenic pollutants such as sulphur (Rejšek 1991) and the combined effects of toxic gases and heavy metals (Kieliszewska-Rokicka 1999). Activity of extracellular soil acid phosphatase can be depressed by increased concentrations of heavy metals in the soil (Tyler 1974; Juma and Tabatabai 1977; Kuperman and Carreiro 1997; Nowak et al. 1999).

It is considered that some parameters of fine roots and mycorrhizas, such as root biomass and turnover, the degree of mycorrhizal colonisation, anatomical changes of fine roots, nutrients and trace elements ratios and physiological activity, can be used as early indicators of environmental changes (Vogt et al. 1993). Extracellular soil AcPase can be a useful and sensitive estimate of metabolic response of soils to environmental stress (Tyler 1974; Nannipieri et al. 1990; Nannipieri 1994).

This study is a part of three-seasons of observation of the mycorrhizal status of Scots pine and beech in a small forest catchment of Ratanica stream located in southern Poland (49°51′N, 20°02′E) 40 km south of Cracow, in the Carpathian Foothills, in the vicinity of a big water reservoir supplying drinking water for the city of Cracow. The catchment has been exposed to moderate levels of gaseous and dust pollutants transported to this area by winds from distant industrial agglomerations (Upper Silesia, Cracow, Ostrava region in Czech Republic) (Tomaszewska and Walczewski 1992) and by local emissions from surrounding villages (Manecki and Tarkowski 1993). The Ratanica catchment belongs to a network of study plots of the International Co-operative Programme on Integrated Monitoring of Air Pollution Effects on Ecosystems in Europe. Numerous studies on air, soil, water and vegetation quality have been conducted in this forest ecosystem (see: Grodzińska and Weiner 1994; Grodzińska and Szarek 1995; Grodzińska and Laskowski 1996; Kieliszewska-Rokicka et al. 1997).

The aim of the present study was to determine whether the long-term pressure of moderate anthropogenic pollution had an influence on the physiological mechanisms of phosphate mobilisation and uptake.

Materials and method

Study site

The entire Ratanica catchment covers 241.5 ha. The upper part of the catchment (88 ha) is covered by mixed forest with a predominance of *Pinus sylvestris* (L.) (63%) and *Fagus sylvatica* (L.) (21%), while the lower part is occupied by fields and meadows (Raimer et al. 1990). The highest point of the catchment is a hill of 427 m above sea level and the lowest one lies at 270 m a.s.l. More than half of the area is situated from 330 to 390 m a.s.l. The soil of the Ratanica catchment is an acidic podsol with a layer of mor-type humus. The soil solution is strongly acidic, especially in the upper part of the soil profile (Grodzińska and Laskowski 1996). The average concentration of SO_2 in the period 1991–1994 was 21.1 mg m^{-3} with SO_2 as high as 100 mg m^{-3} and NO_2 – 8.7 mg m^{-3} with high values up to 30 mg m^{-3} (Szarek 1995). The average concentration of tropospheric O_3 between June 1995 and October 1995 was 36.2 ppb (16.6–55.8) (Grodzińska and Laskowski 1996). The annual dust fall in the area of Ratanica forest was reported as 35–48 ton km^{-2} (Manecki and Tarkowski 1993). The concentrations of heavy metals (Pb, Mn, Zn, Cu, Cd, Fe) in the upper soil layers (particularly in the organic layer) were higher than at unpolluted sites in Poland (Grodzińska and Laskowski 1996), however only the concentration of Pb exceeded the range accepted as normal (Kabata-Pendias and Pendias 1979). The deposition of nitrogen (mainly as NH_4^+) in the Ratanica forest ecosystem was estimated as 24 kg ha^{-1} year^{-1}. The mean annual precipitation in the period 1991–1994 was 700 mm (Grodzińska and Laskowski 1996). The concentrations of sulphur in needles and Fe and Pb in bark of pines grown in Ratanica forest were significantly higher than in relatively unpolluted areas in Poland (Molski and Dmuchowski 1990; Szarek et al. 1993).

Most of the chemical analyses mentioned above were carried out at 29 points, located at three transects traced across the forested part of the catchment and on a permanent plot (PP), of 0.25 ha, located in the upper part of the catchment at an elevation of 370–380 m a.s.l. (Grodzińska and Laskowski 1996) (Fig. 1).

In the present studies soil cores were taken at every second transect point and at the permanent plot (17 measurement sites). The vitality of mycorrhizas was observed at 8 measurement sites.
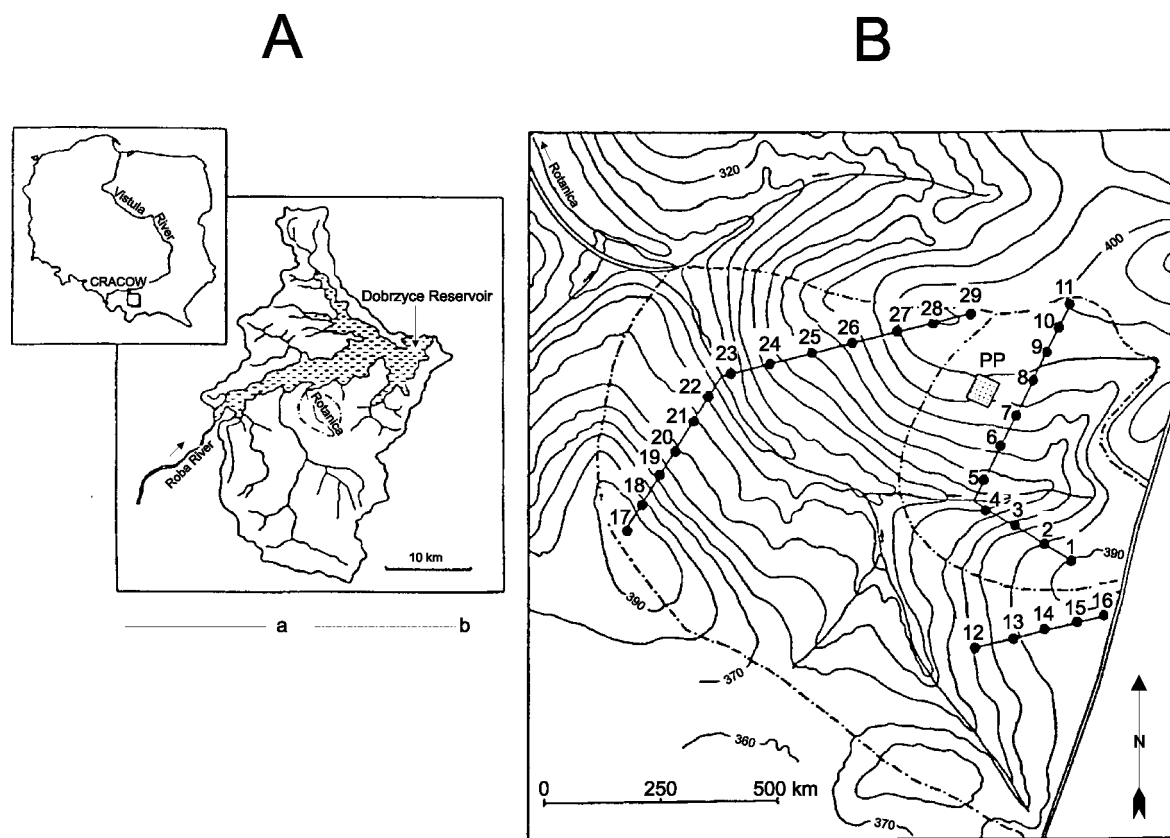

Fig. 1. A – Location of the Ratanica catchment: a – watershed of Dobczyce Reservoir basin, b – Ratanica catchment; B – Location of the measurement points in the Ratanica catchment (according to: Grodzińska 1994)

Sampling and sample preparation

Soil core samples were collected in October 1997 with a 5 cm dia soil corer to a depth 5 cm after removal of the litter layer. All cores were taken at 0.7–1 m from tree bases. Intact soil cores were sealed in plastic bags and kept cool until used, within one week of collection. Soil samples for AcPase activity determination were sieved at 2 mm sieve to remove roots. Fine roots for vitality estimation were separated from soil and organic matter on a sieve under a stream of cold water, excised from the main root and cleaned with running water to remove adhering organic matter. Final separation of roots was conducted under a stereomicroscope. All visibly dead and injured roots, as well as roots of annual plants, were removed.

Microscopic studies

To estimate the development of the fungal mantle and the Hartig net, fresh ectomycorrhizas were cut into transverse sections using a razor blade. The sections were placed in a drop of glycerol and examined under a light microscope (Nikon, Optiphot-2) with a camera attachment (Nikon, Microflex UFX-DX). At each measurement point 50 transverse sections prepared from different mycorrhizal morphotypes were analysed.

FDA (fluorescein diacetate) vital staining of ectomycorrhizas

Fresh mycorrhizas (60–80 from one measurement point) were sectioned longitudinally using a razor blade. The fresh sections were stained immediately with the FDA-stain according to Ritter et al. (1986) for 5–15 minutes, washed with distilled water and observed under a fluorescence microscope (Olympus Provis). Vitality of mycorrhizas, characterised by the light green FDA-fluorescence was documented with a microscope camera using Fuji film, 400 ASA. Five stages of ectomycorrhizal vitality were distinguished according to Ritter et al. (1989): 1) entirely vital – all regions of mycorrhiza are vital (the fungal mantle, the cortex including the Hartig net, the central cylinder, the meristem), 2) largely vital – most of the mycorrhizal compartments are vital (the cortex with the Hartig net, the central cylinder, the meristem), 3) reduced vital – only the central cylinder and meristem are vital, 4) dying mycorrhiza – only the central cylinder is vital, 5) dead mycorrhiza – all root compartments are dead.

Activity of root surface acid phosphatase

AcPase activity associated with excised ectomycorrhizal root tips was measured using a modified method of Bartlett and Lewis (1973). The incubation medium contained 1.8 ml of 0.1 M Na-acetate buffer, pH 5.0, 0.2 ml 0.05 M p-nitrophenyl phosphate (pNPP) and 10–20 mg fresh weight of root tips. In the reference samples roots were omitted. After 1h in a shaking bath at 30°C, the roots were removed and 1ml of reaction mixture was added to 9 ml of 0.5 M NaOH. The liberated p-nitrophenol (pNP) was determined at 400 nm. Enzyme activity was expressed as μmol pNP g^{-1} fresh weight h^{-1}.

Activity of soil acid phosphatase

Two ml of 0.2 M phosphate buffer (pH 6.5), 0.5 ml of 0.115 M p-nitrophenyl phosphate (pNPP) and 0.1 ml of toluene were added to 0.5 g of air-dried soil and incubated at 37°C for 1h. Adding 2 ml of 0.5 M NaOH and 0.5 ml of 0.5 M $CaCl_2$ stopped the reaction. After filtration the solution was diluted 10 times and the liberated p-nitrophenol determined at 400 nm (Tabatabai and Bremner 1969). Each sample was accompanied by a reference sample, where p-nitrophenol was omitted.

Chemical analyses of soil and fine roots

The pH of the soil layer from which roots were removed was measured with a glass electrode in soil-water and soil-salt (0.01 M $CaCl_2$) suspensions (Anonymous 1994). The concentration of P_2O_5 in soil was measured by the method of Egner-Riehm, the concentration of extractable aluminium and hydrogen (Al^{3+}, H^+) were estimated by the method of Sokołow (Ostrowska et al. 1991). The phosphorus concentration (PO_4-P) in fine roots was measured using a standard colorimetric method (Allen 1989). Carbon and nitrogen content were estimated by isotope ratio mass spectrometry (IRMS) Europa 20–20. The concentration of Ca in the plant material was measured by atomic emission spectrometry (Flapho 4) and concentrations of Mg and Al by atomic flame absorption spectrophotometry (Varian 20 BQ) (Grodzińska and Laskowski 1996).

Statistics

Relationships between AcPase activities and Pi content in soil were evaluated using correlation and regression analyses (p<0.05).

Results

Phosphate and aluminium content in soil

The concentrations of extractable Pi, measured as P_2O_5, aluminium ions (Al^{3+}), hydrogen ions (H^+) and pH values of the upper soil layer (0–5 cm) in the experimental forest are given in Table 1. All soil samples were generally strongly acidic, however variability among the 17 measurement sites was found. Available phosphorus concentration varied among the measurements sites within a range from 12 to 44 mg P_2O_5 kg^{-1} of soil and the concentration of Al^{3+} from 40.6 to 118.4 meq kg^{-1} of soil.

Table 1. Soil acidity and concentrations of extractable Pi and Al in the upper soil layer (0–5 cm) at 17 measurement sites in the Ratanica forest catchment. The measurement sites were located along three transects traced across the whole catchment (see: Fig. 1)

Measurement sites	pH_{water}	pH_{salt}	H^+ (meq· kg^{-1} of soil)	Pi (mg P_2O_5· kg^{-1} of soil)	Al^{3+} (meq·kg^{-1} of soil)
1	4.53	3.42	0.8	22	40.6
3	4.14	3.11	1.6	34	41.4
5	4.26	3.22	0.6	15	40.2
7	4.43	3.27	0.5	12	52.3
PP	4.05	3.47	1.5	37	60.7
9	4.35	3.25	1.3	22	77.8
11	4.30	3.57	1.4	29	93.8
12	4.15	3.10	1.6	28	80.8
14	4.16	3.36	1.3	26	72.6
16	4.18	3.53	1.0	24	64.0
17	4.22	3.43	2.0	37	79.6
18	4.14	3.10	1.0	41	47.6
20	3.92	3.86	1.0	42	81.5
22	3.78	3.08	1.3	43	78.8
23	3.82	3.16	1.2	42	74.5
25	3.86	3.12	1.0	17	74.0
27	3.80	3.47	4.0	44	118.4
Mean	4.12	3.32	1.3	30.3	69.3
± SD	± 0.22	± 0.22	± 0.08	± 10.6	± 20.8

Average Ca/Al (mol/mol) = 0.04*; Mg/Al (mol/mol) = 0.58*
*The concentrations of Ca and Mg in soil were taken from Grodzińska and Laskowski (1996)

Anatomy of ectomycorrhizas

The microscope observations of the transverse and the longitudinal sections of mycorrhizas revealed the presence of different types of fungal mantle on the surface of the fine roots and the Hartig net in the intercellular spaces of the root cortex. In beech mycorrhizas the Hartig net penetrated the cortex to a depth of 1–3 cortical cell layers and in Scots pine mycorrhizas reached the endodermis (Fig. 2).

Vitality of ectomycorrhizas

Different stages of mycorrhizal vitality that reflect phases in the ageing process of ectomycorrhizas are presented in Fig. 3. The vitality of Scots pine and beech ectomycorrhizas was estimated at 8 measurement sites (1, 5, PP, 11, 14, 17, 22, 27). Frequency of mycorrhizas with vital fungal hyphae (stages 1 + 2) varied slightly between the measurement points and was from 40 to 53% for Scots pine and from 42 to 57% for beech. The frequency of dead mycorrhizas was from 10 to 19% for Scots pine and from 11 to 16% for beech (Fig. 4). The differences in mycorrhizal vitality did not differ significantly between the measurement sites.

Root-surface and soil acid phosphatase

The activity of the surface AcPase of Scots pine mycorrhizas ranged between 25.3 and 66.7 μmol pNP g^{-1} fresh weight h^{-1} and the enzyme activity of beech mycorrhizas between 33 to 86,4　mol pNP g^{-1} fresh weight h^{-1}, depending on the measurement site. The highest AcPase activities were found at measurement sites No 5, 9 and 25 (Table 2).

The activity of extracellular AcPase in the soil horizon from which the mycorrhizas were taken varied between 6.8 at measurement site No 17 and 22.4 at measurement site No 5 (Table 2). An inverse significant correlation (p<0.05) between the concentration of extractable inorganic phosphorus in the soil layer 0–5 cm and the extracellular soil AcPase activity and the root-surface AcPase of ectomycorrhizas of Scots pine and beech was found (Fig. 5).

Nutrient content in fine roots

The concentrations of C, N, Pi, Ca, Mg and Al analysed in fine roots (0<1 mm) of Scots pine taken from the upper soil layer (0–5 cm) are presented in Table 3. The percentage of C in pine fine roots was similar at all sites (38–45,4%) and the percentage of N varied between 1,20 and 1,91%. The concentration of Pi in
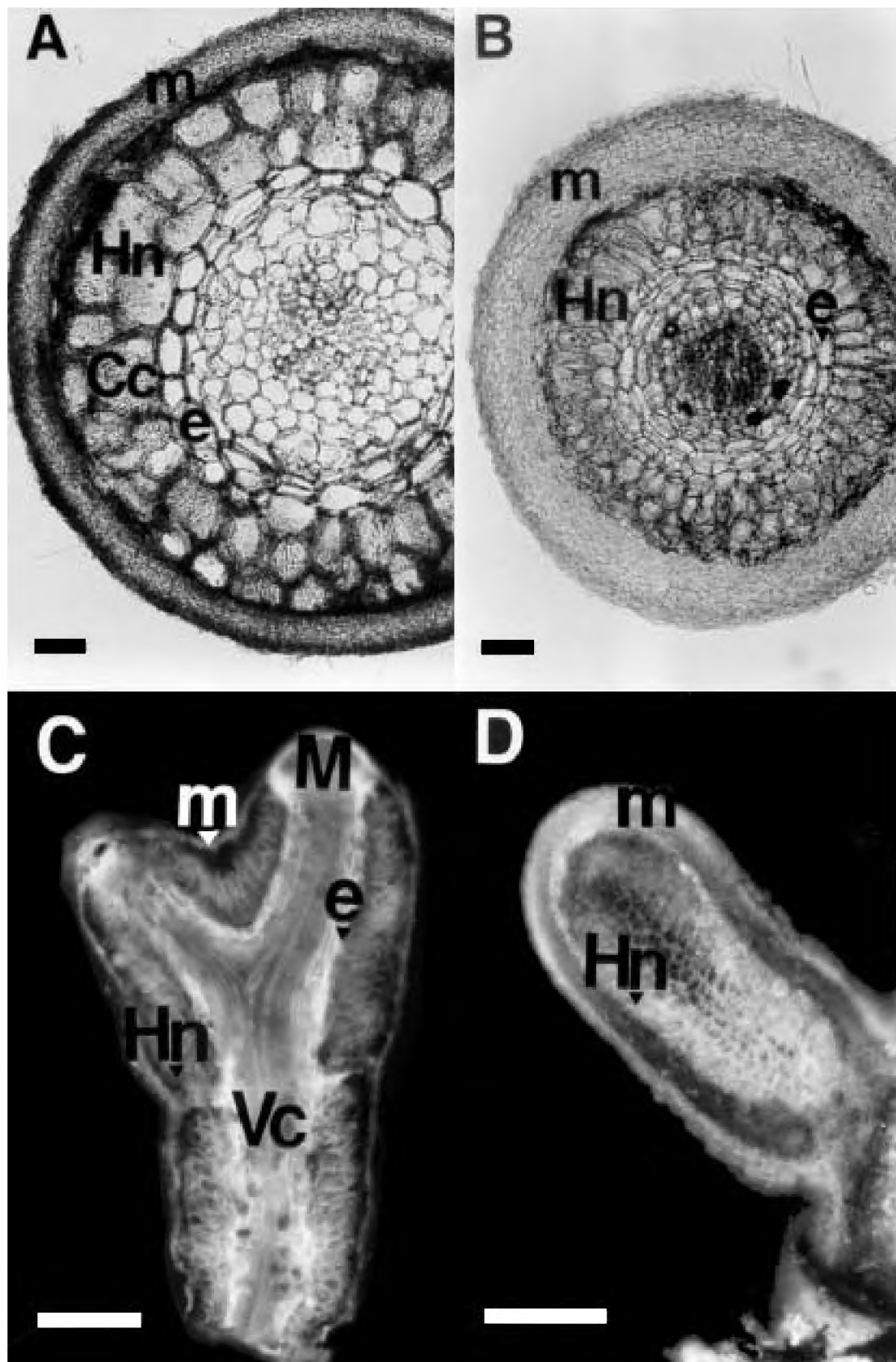
Fig. 2. Microscope observations of ectomycorrhizas. A – crossection of Scots pine mycorrhiza (scale mark=50 m); B – crossection of beech mycorrhiza (scale mark=50 m); C – longitudinal section of Scots pine mycorrhiza (scale mark= 200 m); D – longitudinal section of beech mycorrhiza (scale mark=100 m); m = fungal mantle, Hn = Hartig net, e = endodermis; M = meristem, Vc – vascular cylinder

pine fine roots was generally low (0.05–0.15%). The proportions of nitrogen in relation to C and Pi in fine roots are presented in Table 3.
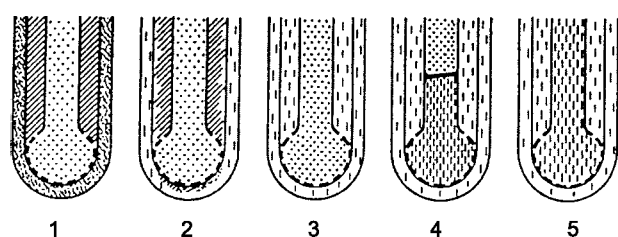
Fine roots of Scots pine and beech accumulated high concentrations of extractable aluminium (Al^{3+}) and also high amounts of Ca and Mg. The Ca/Al and Mg/Al (mol/mol) ratios were calculated (Table 3).

Discussion

This study demonstrated the relations between soil parameters in a moderately polluted forest and some aspects of physiological activity of ectomycorrhizas of Scots pine and beech. We have focused on the concentration of available inorganic phosphorus in the soil and the efficiency of mycorrhizas in phosphorus mobilisation and accumulation of Pi in fine roots. The average concentration of Pi in the Ratanica catchment was relatively low (30.3 mg P_2O_5 kg^{-1} of soil), as compared with other forest sites in Poland, however some differences in Pi content among the 17 measurement sites in the Ratanica forest were found (Table 1). Similar or lower levels of Pi in the upper soil layer have been found at severely polluted sites, such as Świerklaniec in Upper Silesia (5–8 mg P_2O_5 kg^{-1} of soil), Szklarska Poręba in the Sudeten Mountains (9–12 mg P_2O_5 kg^{-1} of soil) (Walendzik et al.

1995a,b) and in the vicinity of the Warsaw Steelworks (12–44 mg P_2O_5 kg^{-1} of soil) (Kieliszewska-Rokicka 1999), whereas soil in relatively unpolluted areas (Białowieża National Park and Kampinos National Park) contained higher concentrations of Pi (30–50 and 83 mg P_2O_5 kg^{-1} of soil, respectively) (Walendzik et al. 1995a; Kieliszewska-Rokicka 1999). Soil in the Ratanica forest was further characterised by low soil pH, high concentrations of Al^{3+} (Table 1) and accumulation of heavy metals in the upper soil layers (Niklińska et al. 1995; Grodzińska and Laskowski 1996).

The average concentration of aluminium ions (Al^{3+}) in the upper soil layer in the Ratanica forest was relatively high (69.6 meq kg^{-1} of soil) as compared with forest sites characterised by strongly acidic soils, such as Szklarska Poręba (55 meq kg^{-1} of soil) and Świerklaniec and Białowieża National Park (10–20 meq kg^{-1} of soil) (Walendzik et al. 1995a,b). High levels of Al^{3+} resulted from the dissolution of inorganic Al at low soil pH is considered as an important stress factor for tree roots in forest ecosystems,


A   Scots pine mycorrhizas

B   Beech mycorrhizas

Fig. 3. Stages of ectomycorrhizal vitality, determined by FDA vital staining of hand cut medial longitudinal sections: 1. entirely vital – all regions of mycorrhiza are vital (the fungal mantle, the cortex including the Hartig net, the central cylinder, the meristem, 2. largely vital – most of the mycorrhizal compartments are vital (the cortex with the Hartig net, the central cylinder, the meristem), 3. reduced vital – only the central cylinder and meristem are vital, 4. dying mycorrhiza – only the central cylinder is vital, 5. dead mycorrhiza – all root compartments are dead (according to: Ritter et al. 1989)

– living ectomycorrhizal mantle

– living Hartig net

– living vascular and meristematic tissues

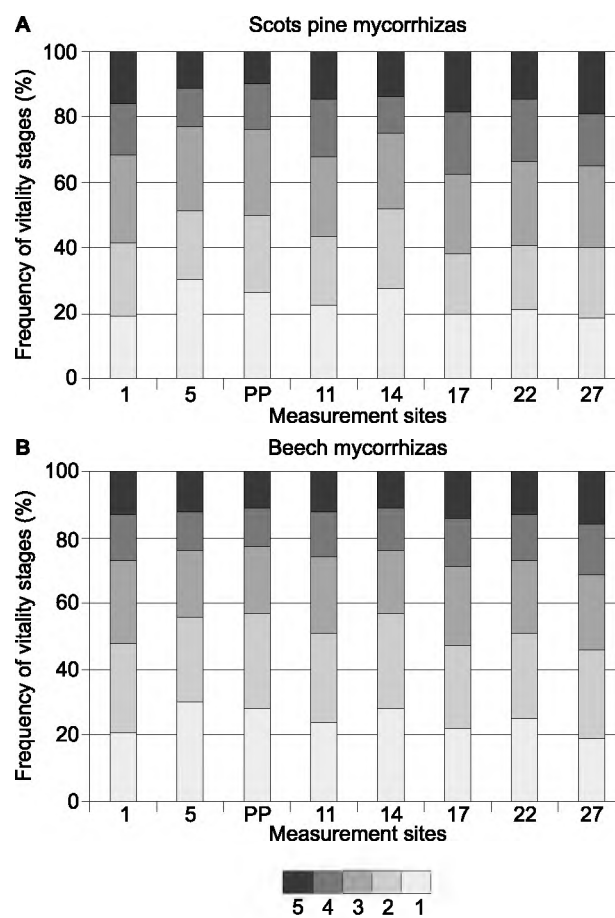– dead parts of mycorrhiza

– – – – metacutilization layer

Fig. 4. Frequency of vitality stages of ectomycorrhizas of Scots pine (*Pinus sylvestris* L.) and beech (*Fagus sylvatica* L.) in 8 measurement sites of the Ratanica catchment (see: Fig. 1). Sixty to eighty mycorrhizas taken from 0–5 cm soil depth were classified according to 5 vitality stages (see: Fig. 3)

Table 2. Enzyme activity of surface acid phosphatase (AcPase) of excised ectomycorrhizas of Scots pine (*Pinus sylvestris* L.) and beech (*Fagus sylvatica* L.) and of soil at 17 measurement sites in Ratanica catchment (see: Fig. 1). Soil and mycorrhizal root tips were taken from a 0–5 cm depth. AcPase activities of pine and beech ectomycorrhizas are the means of 7–11 analyses ± SD and AcPase activities of soil are the mean of 6 analyses ± SD

Measurement sites	AcPase activity of ectomycorrhizas (mol pNP· g⁻¹fresh weight· h⁻¹)		AcPase activity of soil (mol pNP · g⁻¹dry weight· h⁻¹)
	pine mycorrhizas	beech mycorrhizas	
1	45.5 ± 3.1	44.7 ± 6.2	8.2 ± 1.7
3	46.5 ± 8.4	–	7.5 ± 1.3
5	66.7 ± 7.1	86.4 ± 16.6	22.4 ± 3.5
7	59.5 ± 18.0	66.6 ± 7.7	14.9 ± 2.3
PP	56.5 ± 5.3	64.6 ± 5.5	9.8 ± 1.1
9	61.5 ± 10.0	71.3 ± 7.6	13.1 ± 2.2
11	52.7 ± 6.2	55.1 ± 4.8	13.6 ± 1.8
12	43.3 ± 4.8	47.0 ± 4.4	11.3 ± 1.6
14	37.3 ± 4.4	45.5 ± 5.5	8.9 ± 1.1
16	45.8 ± 6.5	53.4 ± 5.9	12.0 ± 1.6
17	25.3 ± 5.2	33.0 ± 3.8	6.8 ± 1.1
18	29.5 ± 6.2	38.5 ± 5.8	12.1 ± 1.2
20	40.4 ± 11.9	47.9 ± 10.6	12.8 ± 2.7
22	34.0 ± 6.4	38.0 ± 4.3	12.9 ± 3.4
23	34.2 ± 4.4	–	12.0 ± 2.6
25	65.7 ± 9.8	70.0 ± 7.7	21.6 ± 3.9
27	35.5 ± 7.7	42.7 ± 4.9	7.6 ± 0.9
Mean	45.8 ± 14.4	53.0 ± 15.8	12.2 ± 4.4

A Scots pine mycorrhizas

$y = -0,9341x + 74,104$
$R^2 = 0,5933$

AcPase activity (mol pNp g⁻¹ fresh weight h⁻¹)

Soil Pi (mg P_2O_5 kg⁻¹)

B Beech mycorrhizas

$y = -0,9974x + 82,817$
$R^2 = 0,5059$

AcPase activity (mol pNP g⁻¹ fresh weight h⁻¹)

Soil Pi (mg P_2O_5 kg⁻¹)

C Soil (0-5 cm)

$y = -0,2311x + 19,319$
$R^2 = 0,3091$

AcPase activity (mol pNP g⁻¹ dry weight h⁻¹)
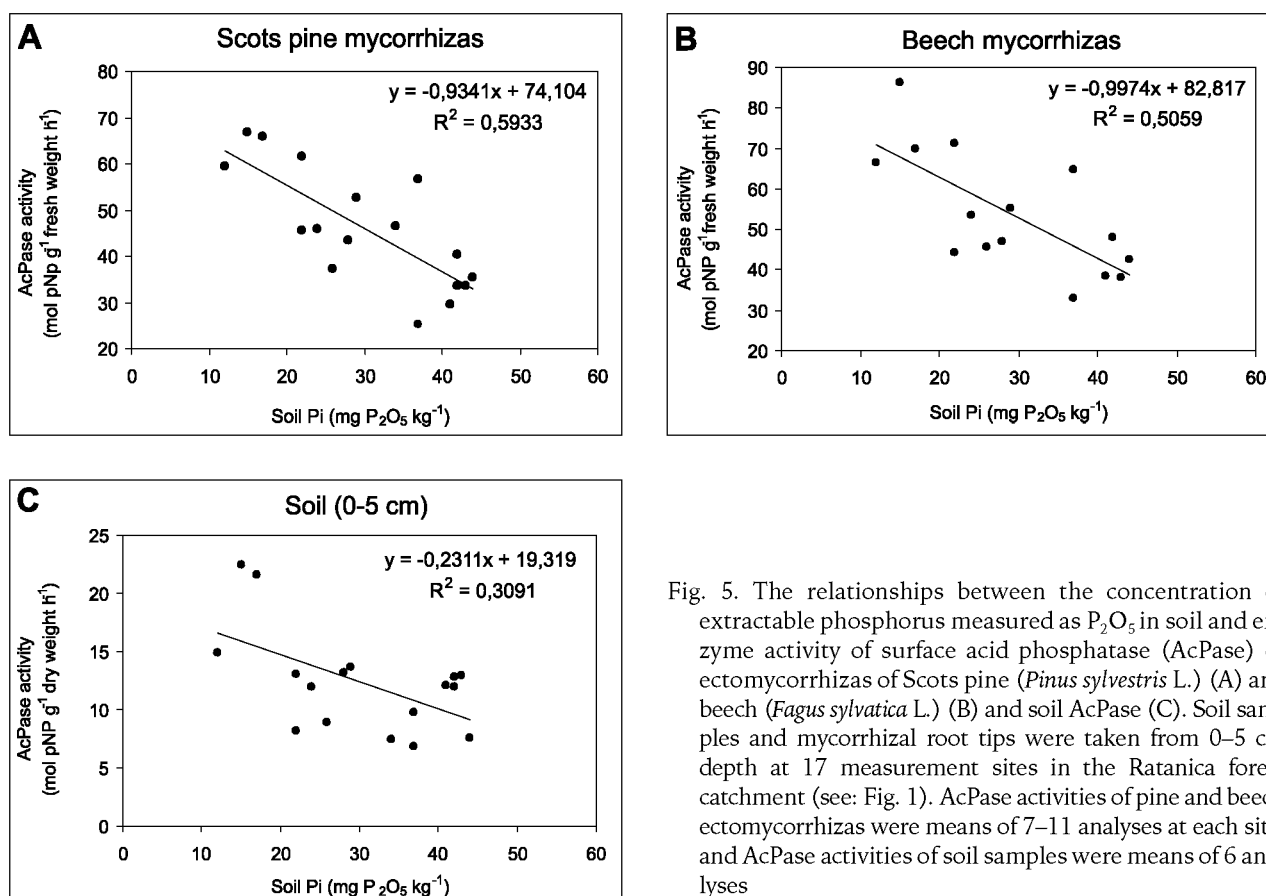
Soil Pi (mg P_2O_5 kg⁻¹)

Fig. 5. The relationships between the concentration of extractable phosphorus measured as P_2O_5 in soil and enzyme activity of surface acid phosphatase (AcPase) of ectomycorrhizas of Scots pine (*Pinus sylvestris* L.) (A) and beech (*Fagus sylvatica* L.) (B) and soil AcPase (C). Soil samples and mycorrhizal root tips were taken from 0–5 cm depth at 17 measurement sites in the Ratanica forest catchment (see: Fig. 1). AcPase activities of pine and beech ectomycorrhizas were means of 7–11 analyses at each site, and AcPase activities of soil samples were means of 6 analyses

Table 3. Concentrations of nutrients in fine roots ($\varnothing$<2 mm) of Scots pine and beech in the Ratanica forest catchment. Data represent means and ranges of the nutrient concentrations and proportions in root samples taken from 0–5 cm depth at 12 measurement sites

Nutrients	Scots pine	Beech
C (%)	42.3 ± 2.4 (39.3–44.8)	–
N (%)	1.58 ± 0.22 (1.20–1.91)	–
$PO_4 - P$ (%)	0.093 ± 0.029 (0.05–0.15)	–
C/N (weight ratio)	26.8 ± 3.95 (21.2–34.7)	–
N : P (weight ratio)	100 : 5.8 (100 : 4.1–100 : 7. 9)	–
Ca (mg g⁻¹ dry weight)	4.04 ± 0.75 (3.26–4.75)	3.73 ± 0.59 (3.15–4.33)
Mg (mg g⁻¹ dry weight)	1.77 ± 0.41 (1.40–2.2)	1.55 ± 0.28 (1.27–1.83)
Al (mg g⁻¹ dry weight)	1.69 ± 0.58 (1.14–2.30)	1.36 ± 0.62 (0.67–1.90)
Ca/Al (mol/mol)	1.79 ± 0.75 (0.96–2.44)	2.39 ± 1.74 (1.13–4.36)
Mg/Al (mol/mol)	1.28 ± 0.53 (0.67–1.67)	1.89 ± 1.0 (0.75–3.03)

which inhibits the uptake and transport of essential cations (Ca^{2+}, Mg^{2+}) and important anions (PO_4^{3-}, SO_4^{2-}) (Foy et al. 1978). It is accepted that aluminium toxicity is related to a decreased Ca/Al ratio (<1.0) than to Al^{3+} concentration alone (Ulrich 1983; Rost-Siebert 1983). The mobile Al^{3+} and heavy metal ions in the soil may reduce Pi concentration by immobilisation in metal-polyphosphate complexes as confirmed by Turnau et al. (1993) who found a high content of Al, Pb, Cd, Zn in polyphosphate granules localised in the fungal vacuoles of *Paxillus involutus/Pinus sylvestris* mycorrhizas taken from a heavily polluted forest. Scots pine seedlings grown in the presence of increased levels of Al^{3+} in the soil substrate had decreased concentrations of Pi in the root and shoot of mycorrhizal seedlings, as compared with the control (Kieliszewska-Rokicka et al. 1998; Rudawska et al. 2000). Moreover, acid deposition and high concentrations of heavy metals may be toxic to soil microorganisms involved in the decomposition of organic matter and nutrient cycling (Bååth 1989; Zwoliński 1995) and can inhibit enzymatic systems including acid phosphatase (Tyler 1974; Tyler et al. 1989; Kuperman and Carreiro 1997). One of mechanisms by which mycorrhizal fungi and mycorrhizas may increase the uptake of phosphorus from soils under Pi-limiting conditions is the production of a root-surface AcPase (Harley 1989; Smith and Read 1997).

Our results revealed an inverse correlation between the concentration of Pi in the soil and the activities of the root-surface and soil extracellular AcPase. This indicates that the functioning of a feedback mechanism which induces the AcPase activity in response to a deficiency of plant and soil Pi as shown for ectomycorrhizal fungi and ectomycorrhizas (i.e. Alexander and Hardy 1981; MacFall et al. 1991; Kieliszewska-Rokicka 1992; Pasqualini et al. 1992) and for forest soil (i.e. Spiers and McGill 1979). Increased activities of the root-surface AcPase in Ratanica forest (Table 2, Fig. 5) suggest a high physiological activity of ectomycorrhizas as confirmed by FDA vital staining (Figs. 2, 4). The frequency of vital mycorrhizas, containing vital fungal hyphae (stages 1, 2) ranged between 20 and 40%, whereas the percentage of dead mycorrhizas was relatively low (between 10 and 18%). Similar results have been reported for unpolluted and moderately polluted forest sites in Poland (Józefaciukowa et al. 1995) and in Germany (Kottke et al. 1993; Münzenberger et al. 1995).

In trees, the concentration of elements in the fine roots is regarded as a better indicator of the nutritional conditions in the forest soil than the foliar concentration (Joslin et al. 1988; Person and Ahlström 1991). In spite of the increased phosphatase activity, the concentrations of Pi and the weight proportions

N/P in pine fine roots were below the range considered as optimal. According to Ingestad (1979), the optimum weight proportion of nitrogen to phosphorus for Scots pine is 100 N : 15 P. In this study Pi concentration was 2–3 times below the optimum resulting in a 100 N : 5.8 P average (Table 3).

A low Ca/Al (<0.1 mol/mol) in fine roots is considered as an indicator of the toxic effect of aluminium (Meiwes et al.1986). Although the Ca/Al ratio in the soil of the Ratanica forest was much below the level believed as safe for plants (Table 2), the Ca/Al in fine tree roots was relatively high (0.96–2.44 in Scots pine and 1.13–4.34 in beech) (Table 3). The average Ca/Al ratio in fine roots in the Ratanica forest was higher than that found by Józefaciukowa et al. (1995) in fine roots (<2 mm) of Scots pine from 11 forest sites in Poland (Ca/Al=0.1–0.7) and by Persson and Majdi (1995) in fine roots (<2 mm) of mature Scots pine (Ca/Al=0.11–0.22) and Norway spruce (0.05–0.57) in various Swedish forest stands. Similarly, no deficiency of Mg was found in the fine roots of pine and beech from the Ratanica catchment (Table 3). Keltjens and van Loenen (1989), in hydroponic experiments, have found a higher accumulation of Al in roots of coniferous evergreens (Scots pine and Douglas-fir) than in deciduous broad-leaved species (oak and birch) and intermediate Al levels in larch. The differences in Al concentrations in fine roots of Scots pine and beech from Ratanica forest were not significant.

These results led us to the conclusion that the long-term influence of air pollution that induced significant changes in soil chemistry in the Ratanica forest catchment caused a moderately disadvantageous effect on the functioning of ectomycorrhizas of Scots pine and beech, as demonstrated by some nutrient imbalance in the fine roots of these trees. The anatomical structure of ectomycorrhizas, the relatively high frequency of vital ectomycorrhizas and the high activity of the enzyme involved in the nutritive metabolism (AcPase) showed that the physiological activity was not inhibited by the anthropogenic stress factors, however it was insufficient with respect to phosphate mobilisation and uptake. The high buffer capacity of the soil in Ratanica forest, especially in the organic soil layer, reported by Grodzińska and Laskowski (1996), could alleviate the potentially harmful effects of increased levels of aluminium and heavy metals ions in the soil.

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References

Allen S. (ed.) 1989. Chemical Analysis of Ecological Materials. Blackwell Scientific Publications.

Alexander I.J., Hardy K. 1981. Surface phosphatase activity of Sitka spruce mycorrhizas from a Serpentine site. Soil Biology and Biochemistry 13: 301–305.

Anonymous 1994. Mannual on methods and criteria for harmonized sampling assessment, monitoring and analysis of the effects of air pollution on forests. Programme Coordinating centres, Hamburg and Prague.

Antibus R.K., Linkins A.E. 1992. Effect of liming a red pine forest floor on mycorrhizal numbers and mycorrhizal and soil acid phosphatase activities. Soil Biology and Biochemistry 24: 479–487.

Barlett E.M., Lewis D.H. 1973. Surface phosphatase activity of mycorrhizal roots of beech. Soil Biology and Biochemistry 5: 249–257.

Bååth E. 1989. Effects of heavy metals in soil on microbial processes and populations (a review). Water, Air and Soil Pollution 47: 335–379.

Białobok S. (ed.). 1989. Życie drzew w skażonym środowisku. PWN, Warszawa–Poznań, 501 pp.

Bieleski R.L. 1973. Phosphate pools, phosphate transport and phosphate availability. Annual Review of Plant Physiology 24: 225–257.

Boutin J.P., Roux   L. 1984. L'hydrolyse des polyphosphates par les racines des plantes. I. Role des racineset des microorganismes. Agrochimica 18: 424–431.

Burns R.G. 1982. Enzyme activity in soil: location and possible role in microbial ecology. Soil Biology and Biochemistry 14: 423–427.

Bytnerowicz A., Manning W.J., Grosjean D., Chmielewski W., Dmuchowski W., Grodzińska K., Godzik B. 1993. Detecting ozone and demonstrating its phytotoxicity in forested areas of Poland: a pilot study. Environmental Pollution 80: 301–305.

Dalal R.C. 1977. Soil organic phosphorus. Advances in Agronomy 29: 83–117.

Dexheimer J., Aubert-Dufresne M.P., Gerard J., LeTacon F., Mousain D. 1986. Etude de la localization ultrastructurale des activites phosphasiques acides dans deux types d'Ectomycorhizas: *Pinus nigra nigricans/ Hebeloma crustuliniforme* et *Pinus pinaster/ Pisolithus tinctorius*. Bulletin Société Botanique de France 133: 343–352.

Foy C.D., Chaney R.C., White M.C. 1978. The physiology of metal toxicity in plants. Annual Review of Plant Physiology 29: 511–566.

Freer-Smith P. H. 1996. Forest growth and decline: What is the role of air pollutants? In: Plant Response to Air Pollution. Yunus M., Iqbal M. (eds.). John Wiley & Sons, Chichester, England. pp. 437–447.

Godzik B. 1997. Ground level ozone concentrations in the Kraków region, southern Poland. Environmental Pollution 98: 273–280.

Godzik S., Sienkiewicz J. 1990. Air pollution and forest health in Central Europe: Poland, Czechoslovakia and the German Democratic Republic. In: Ecological Risks-Perspectives from Poland and the United States. Grodziński W., Cowling W.B., Breymeyer A.I. (eds.). National Academy Press, New Haven, Connecticut, pp. 22–74.

Grodzińska K. 1994. Biogeochemical studies in the Ratanica forest catchment (Wieliczka Foothills, s. Poland). Geographia Polonica 62: 99–104.

Grodzińska K., Laskowski R. (eds.). 1996. Ocena stanu środowiska i procesów zachodzących w lasach zlewni potoku Ratanica (Pogórze Wielickie, Polska południowa). Państwowa Inspekcja Ochrony Środowiska, Warszawa, 143 pp.

Grodzińska K., Szarek G. 1995. Ecological studies in the Ratanica catchment (Carpathian Foothills, S. Poland) – An overview. Water, Air and Soil Pollution 85: 1635–1640.

Grodzińska K., Weiner J. (eds.). 1993. Watershed processes and vegetation in the region of chronic atmospheric pollution (Carpathian Foothilld, s. Poland). Ekologia Polska. Polish Journal of Ecology 41(3–4): 285–450.

Grodziński W., Greszta J., Laskowski R., Maryański M., Rożen A. 1990. Effect of the chemical composition of industrial dusts on forest floor organic matter accumulation. Water, Air and Soil Pollution 53: 169–178.

Harley J.L. 1989. The significance of mycorrhiza. Mycological Research 92: 129–139.

Ingestad T. 1979. Mineral nutrient requirements of *Pinus sylvestris* and *Picea abies* seedlings. Physiologia Plantarum 45: 373–380.

Joslin J.D., Kelly J.M., Wolfe M.H., Rustad L.E. 1988. Elemental patterns in roots and foliage of mature spruce across a gradient of soil aluminium. Water, Air and Soil Pollution 40: 375–390.

Józefaciukowa W., Dobrowolska D., Farfał D. 1995. Ilościowa i jakościowa ocena korzeni drobnych sosny zwyczajnej w gradiencie skażenia środowiska leśnego. Prace Instytutu Badawczego Leśnictwa (Poland), seria B, 24: 47–61.

Juma N.G., Tabatabai M.A. 1977. Effects of trace elements on phosphatase activity in soil. Soil Science American Journal 41: 343–346.

Kabata-Pendias A., Pendias H. 1979. Pierwiastki śladowe w środowisku biologicznym. Wydawnictwo Geologiczne, Warszawa, 300 pp.

Keltjens W.G., van Loenen E. 1989. Effects of aluminium and mineral nutrition on growth and chemical composition of hydroponically grown seedlings of five different forest tree species. Plant and Soil 119: 39–50.

Kieliszewska-Rokicka B. 1992. Acid phosphatase activity in mycorrhizal and non-mycorrhizal Scots pine seedlings in relation to nitrogen and phosphorus nutrition. Acta Societatis Botanicorum Poloniae 61(2): 253–264.

Kieliszewska-Rokicka B. 1999. Phosphate status and acid phosphatase activity in soil and ectomycorrhizas in two mature stands of Scots pine (*Pinus sylvestris* L.) exposed to different levels of anthropogenic pollution. Acta Societatis Botanicorum Poloniae 68(4): 311–317.

Kieliszewska-Rokicka B., Rudawska M., Leski T. 1997. Ectomycorrhizae of young and mature Scots pine trees in industrial regions in Poland. Environmental Pollution 98(3): 315–324.

Kottke I., Weber R., Ritter T., Oberwinkler F. 1993. Vitality of mycorrhizas and health status of trees in diverse forest stands in West Germany. In: Forest Decline in the Atlantic and Pacific Region, Hüttl R.F., Mueller-Dombois D. (eds.). Springer-Verlag, Berlin, Heidelberg, pp. 189–201.

Kuperman R.G., Carreiro M. 1997. Soil heavy metal concentrations, microbial biomass and enzyme activities in a contaminated grassland ecosystem. Soil Biology and Biochemistry 29(2): 179–190.

MacFall J., Slack S.A., Iyer J. 1991. Effect of *Hebeloma arenosa* and phosphorus fertility on root acid phosphatase activity of red pine (*Pinus resinosa*) seedlings. Canadian Journal of Botany 69: 380–383.

Manecki A., Tarkowski J. 1993. Mineralogical-chemical characteristics of atmospheric dust pollution in the surroundings of Dobczyce reservoir supplying drinking water for the city Cracow. Ekologia Polska (Polish Journal of Ecology) 41: 289–306.

Meiwes K.J., Koenig N. 1986. H-Ionen Deposition in Waldekosystemen in Norddeutschland. GSF München, BPT-Bericht 8: 25–35.

Meyer F.H. 1985. Einfluss des Stickstoff-Faktors auf den Mycorrhizabesatz von Fichtensämlingen im Humus einer Waldschadensfläche. Allgemeine Forstzeitschrift 40: 208–219.

Molski B.A., Dmuchowski W. 1990. Distribution and movement of selected elements in Poland using Pine needles analysis. In: Ecological Risks. Perspectives from Poland and the United States. Grodziński W., Cowling E.B., Breymeyer A.I. (eds.). National Academy Press, Washington DC, pp. 215–231.

Münzenberger B., Schmincke B., Strubelt F., Hüttl R.F. 1995. Reaction of mycorrhizal and

non-mycorrhizal pine roots along a deposition gradient of air pollutants in eastern Germany. Water, Air and Soil Pollution 85: 1191–1196.

Nannipieri P. 1994. The potential use of soil enzymes as indicators of productivity, sustainability and pollution. In: Soil Biota: management in sustainable farming systems. Pankhurst C.E., Doube B.M., Gupta V.V.S.R., Grace P.R. (eds.). CSIRO Australia, pp. 238–245.

Nannipieri P., Ceccanti B., Grego S. 1990. Ecological significance of the biological activity in soil. In: Soil Biochemistry, vol. 6. Bollag J.-M., Stotzky G. (eds.). Marcel Dekker, New York, pp. 293–355.

Niklińska M., Maryański M., Szarek G., Laskowski R. 1995. Chemical input/output for a moderately polluted forest catchment in southern Poland. Water, Air and Soil Pollution 85: 1771–1776.

Nowak J., Niedźwiecki E., Dziel M. 1999. Wpływ metali ciężkich na zmiany aktywności enzymatycznej gleby. Roczniki Gleboznawcze 50(1/2): 61–68.

Ostrowska A., Gawliński S., Strzubiałka Ż., 1991. Metody analizy i oceny właściwości gleb i roślin. Instytut Ochrony Środowiska (ed.), Warszawa, 67–77.

Pasqualini S., Panara F., Antonelli M. 1992. Acid phosphatase activity in *Pinus pinea – Tuber albidum* ectomycorrhizal association. Canadian Journal of Botany 70: 1377–1383.

Persson H., Majdi H., 1995. Effects of acid deposition on tree roots in Swedish forest stands. Water, Air and Soil Pollution 85: 1287–1292.

Raimer J., Rutkowska L., Grabczyńska S., Orzeł S., Rieger R. 1990. Ocena biomasy i produktywności drzewostanów kompleksu leśnego „Ratanica" na Pogórzu Wielickim. Acta Agraria et Silvestria, Series Silvestris 29: 89–103.

Rejšek K. 1991. Acid phosphomonoesterase activity of ectomycorrhizal roots in Norway spruce pure stands exposed to pollution. Soil Biology and Biochemistry 23: 667–671.

Richards J.H. 1989. Evaluation of root functioning of trees exposed to air pollutants In: Biologic Markers of Air Pollution Stress and Damage in Forests (ed. By Committee on Biologic Markers of Air-Pollution Damage in Trees). National Academic Press, Washington, DC.

Ritter T. 1990. Fluoreszenzmikroskopische Untersuchungen zur Vitalität der Mykorrhizen von Fichten (*Picea abies* (L.) Karst.) und Tannen (*Abies alba* Mill.) unterschiedlich geschädigter Bestädigter Bestände im Schwarzwald. Dissertation, Tübingen.

Ritter T., Kottke I., Oberwinkler F. 1986. Nachweis der Vitalität von Mykorrhizen durch FDA-Vitalfluorochromierung. Biologie in unserer Zeit 16(6): 179–185.

Ritter T., Kottke I., Oberwinkler F. 1989. Vitality and ageing of the ectomycorrhizae of damaged and undamaged trees. Agriculture, Ecosystems and Environment 28: 415–419.

Rost-Siebert K. 1983. Aluminium Toxicität und Toleranz an Keimpflanzen von Fichte (*Picea abies* Karst.) und Buche (*Fagus sylvatica* L.). Allgemeine Forstzeitung 38: 686–689.

Rudawska M., Kieliszewska-Rokicka B., Leski T. 2000. Effect of aluminium on *Pinus sylvestris* seedlings mycorrhizal with aluminium-tolerant and aluminium-sensitive strains of *Suillus luteus*. Dendrobiology 45: 93–100.

Schütt P., Cowling E.B. 1985. Waldsterben, a general decline of forests in central Europe. Symptoms, development and possible causes. Plant Disease 69(7): 548–558.

Smith S.E., Read D.J. 1997. Mycorrhizal Symbiosis. Second edition. Academic Press, London, 605 pp.

Spiers G.A., McGill W.B. 1979. Effects of phosphorus addition and energy supply on acid phosphatase production and activity in soils. Soil Biology and Biochemistry 11: 3–8.

Szarek G. 1995. Sulphur and nitrogen input to the Ratanica forested catchment (Carpathian Foothills, southern Poland). Water, Air and Soil Pollution 85: 1765–1770.

Szarek G., Braniewski S., Chrzanowska E., Rutkowska L. 1993. Nutrients and pollutants in forest vegetation in the Ratanica watershed (Carpathian Foothills, southern Poland). Ekologia Polska (Polish Journal of Ecology) 41: 375–392.

Tabatabai M.A., Bremner J.M. 1969. Use of p-nitrophenyl phosphate for assay of soil phosphatase activity. Soil Biology and Biochemistry 1: 301–307.

Tomaszewska M., Walczewski J. 1992. Zasięg smug dymowych widocznych na zdjęciach satelitarnych, a stan równowagi atmosfery. Zeszyty Naukowe AGH 1440, Sozologia, Sozotechnika 35: 35–59.

Turnau K., Dexheimer J. 1995. Acid phosphatase activity in *Pisolithus arrhizus* mycelium treated with cadmium dust. Mycorrhiza 5: 205–211.

Turnau K., Kottke I., Oberwinkler F. 1993. *Paxillus involutus/Pinus sylvestris* mycorrhizae from heavily polluted forest. I. Element localization using electron energy loss spectroscopy and imaging. New Phytologist 123: 313–324.

Tyler G. 1974. Heavy metal pollution and soil enzymatic activity. Plant and Soil 41: 303–311.

Tyler G., Pahlsson A., Bengtsson G., Bååth E., Tranvik L. 1989. Heavy-metal ecology of terrestrial plants, microorganisms and invertebrates. Water, Air and Soil Pollution 47: 189–215.

Ulrich B. 1983. Soil acidity and its relation to acid deposition. In: Effects of Accumulation of Air Pollu-

tion in Forest Ecosystems, Ulrich B., Pankrath J. (eds.), Reidel, Dordrecht, Boston, London, pp. 127–146).

Walendzik R.J., Szołtyk G., Olejarski I. 1995a. The factors that differentiate properties of the forest soils in contaminated and uncontaminated areas. In: Proceedings of International Symposium: Estimation and Condition of Natural Environment in Poland and other Countries. Jachranka – Poland, 10–13.10.1995., H. Lorenc (ed.). Institute of Meteorology and Water Management, Warsaw, pp. 157–159.

Walendzik R.J., Szołtyk G., Olejarski I., Przyborowska I. 1995b. Wpływ wapnowania na zdegradowane gleby w Sudetach. Prace Instytutu Badawczego Leśnictwa, ser. B, 24: 117–129.

Williamson B., Alexander I.J. 1975. Acid phosphatase localised in the sheath of beech mycorrhiza. Soil Biology and Biochemistry 7: 195–198.

Vancura K., Raben G., Gorzelak A., Mikulowski M., Caboun V., Oleksyn J. 2000. Impact of air pollution on the forests of central and eastern Europe. In: Forest Dynamics in Heavily Polluted Regions. Innes J.L., Oleksyn J. (eds.), CAB International, pp. 121–146.

Vogt, K.A., Publicover D.A., Bloomfield D.J., Perez J.M., Vogt D.J., Silver W.L. 1993. Belowground responses as indicators of environmental change. Environmental Experimental Botany 33: 189–205.

Zwoliński J. 1995. Wpływ emisji zakładów przemysłu metali nieżelaznych na środowisko leśne – rola metali ciężkich w degradacji lasów. (Effects of emissions from non-ferrous metal works on forest environment – the role of heavy metals in forest degradation.) Prace Instytutu Badawczego Leśnictwa, seria A, 809: 1–86.

Aktywność fizjologiczna ektomikoryz w średnio zanieczyszczonym lesie (zlewnia potoku Ratanica, południowa Polska)

Streszczenie

Badania ektomikoryz sosny (*Pinus sylvestris* L.) i buka (*Fagus sylvatica* L.) prowadzono na Pogórzu Karpackim w dorosłym drzewostanie, który przez ponad 40 lat znajdował się pod wpływem gazowych i pyłowych zanieczyszczeń powietrza o średnich stężeniach. Gleby badanego ekosystemu leśnego cechowało niskie pH oraz znaczna akumulacja metali ciężkich (Pb, Mn, Zn, Cu, Cd, Fe). Badano zależność między stężeniem nieorganicznego fosforanu w glebie, a aktywnością enzymatyczną kwaśnej fosfatazy mikoryz i gleby. Analizowano także stężenia pierwiastków odżywczych (C, N, P, Ca, Mg) oraz jonów glinu (Al^{3+}) w korzeniach drobnych. Mikoryzy sosny i buka były analizowane pod względem budowy anatomicznej oraz żywotności. Stwierdzono stosunkowo wysokie stężenie jonów Al^{3+} w glebie (40–118 meq kg^{-1}) i niski poziom nieorganicznego fosforanu (Pi)

(12–44 mg P_2O_5 kg^{-1}). Jednocześnie zarejestrowano wysoką aktywność powierzchniowej kwaśnej fosfatazy mikoryz sosny i buka (odpowiednio, 25–67 i 33–86 μmoli pNP g^{-1} świeżej masy h^{-1}) oraz kwaśnej fosfatazy gleby (6,8–22,4 μmoli pNP g^{-1} suchej masy h^{-1}). Stężenia fosforu w korzeniach drobnych (poniżej stężeń uważanych za optymalne) oraz wysoki stosunek N/P pokazują, że pobieranie i akumulację fosforu przez systemy korzeni drobnych sosny i buka jest niedostateczne, mimo wysokiej aktywności kwaśnej fosfatazy. Budowa anatomiczna mikoryz, stosunkowo duży udział żywych mikoryz w systemie korzeni drobnych oraz wysoki stosunek Ca/Al w korzeniach sosny i buka (odpowiednio, 1,79 i 2,38) sugerują, że mechanizmy obronne mikoryz i mikroorganizmów ryzosfery mają zdolność łagodzenia wpływu zanieczyszczeń antropogenicznych.