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Lectin binding patterns and immunohistochemical antigen detection in placenta and lungs of *Brucella abortus*-bovine infected fetuses

María Andrea Fiorentino^{1,*}, Fernando Alberto Paolicchi¹, Carlos Manuel Campero¹ and Claudio G. Barbeito²

¹INTA, Centro Regional Buenos Aires Sur, Estación Experimental Agropecuaria Balcarce, 7620 Balcarce, Argentina

²Cátedra de Histología y Embriología e Instituto de Patología, Facultad de Ciencias Veterinarias Universidad Nacional de La Plata, La Plata, B1900 AVW, Argentina

Abstract

Lectin binding relies on the affinity of these substances for specific terminal sugars. The method facilitates the identification of complex structures to which the terminal sugar attaches and may reveal physiological or pathological changes in cells, intracellular interactions or extracellular transport pathways. This study was carried out to investigate the effect of infection with *Brucella abortus* on the pattern of lectin binding in bovine fetal lungs (n=6) and bovine placentas (n=5). Fetal lungs and placenta from heifers experimentally inoculated with *B. abortus*, strain 2308 were examined by histological, lectin-histochemical, immunohistochemical and cultural techniques. *B. abortus* antigens were immunohistochemically detected in fetal lungs and placenta. An increase in the labeling with UEA-1, DBA, PNA, RCA-1 and SBA was found in the lungs and an increase in the labeling with UEA-1, ConA, PNA, DBA was found in the placentas. The present lectin histochemical study revealed a distinctive pattern of oligosaccharide distribution in the lungs and placenta of *B. abortus*-infected fetuses.

Keywords: Bovine fetuses, *Brucella abortus*, Immunohistochemical, Lectin, Placenta.

Introduction

Brucella abortus is a Gram negative, facultative intracellular pathogen and the main causative agent of brucellosis in cattle. The disease in pregnant females can result in massive infection of the placenta, which in turn, leads to infertility and abortion in the late stages of gestation (Olsen and Palmer, 2014). In Argentina, cattle brucellosis prevalence of the 2.10 % was found in 2004, although a program of control and eradication was implemented (Aznar *et al.*, 2014).

Lectin binding relies on the affinity of these substances for specific terminal sugars and for α or β anomeric linkages to the penultimate residues. The lectin histochemistry method facilitates the identification of complex structures to which the terminal sugar attaches and may reveal physiological or pathological changes in cells, intracellular interactions or extracellular transport pathways (Gemeneir *et al.*, 2009). Lectin histochemistry has been used in the evaluation of glycoprotein expression in reproductive infections in several animal species, and changes in glycoproteins have been suggested in different studies where the patterns of lectin binding were modified (Pérez *et al.*, 1998).

This fact has been reported in bovine sexually-transmitted diseases as the uterine and tubarian epithelium of heifers infected with *Tritrichomonas foetus* and *Campylobacter fetus venerealis* (Cipolla *et*

al., 1998; Cobo *et al.*, 2004). These changes are related to the adhesion capacity and invasivity of the microorganisms. In relation with *B. abortus*, only two reports about the binding pattern of different bacterial strains have been published (Corbel *et al.*, 1983a,b). More recently, the presence of beta 1,2 glucooligosaccharides in *B. abortus* was demonstrated as these sugars were recognized by c-lectins of the immune system cells and were involved in the pathogenesis of the disease (Poester *et al.*, 2013; Zhang *et al.*, 2016).

However, no information has been found about the carbohydrate pattern changes related to *B. abortus* infection in bovine fetuses and placentas.

The purpose of this study was to demonstrate the effects of presence of *B. abortus* on the lectin binding pattern on fetal lungs and placenta with the isolation of the bacterium.

Materials and Methods

Sample selection

Samples of bovine fetuses or stillborn lungs (n=6) and placenta (n=5), with *B. abortus* isolation, were selected from a previous study of experimentally inoculated heifers at 5 months of gestation with *B. abortus* 2308 (Fiorentino *et al.*, 2008). Negative controls consisted of 3 fetuses and placenta with non-infectious abortion and with similar gestational age (> 7 months) than fetuses with *B. abortus* diagnosis.

*Corresponding Author: María Andrea Fiorentino. INTA, Centro Regional Buenos Aires Sur, Estación Experimental Agropecuaria Balcarce, 7620 Balcarce, Argentina. Email: fiorentino.maria@inta.gob.ar

Histology

For histological examination, tissues obtained from the lungs and placenta were fixed in neutral-buffered 10% formalin, routinely embedded in paraffin wax, sectioned at 5 μ m, and stained with hematoxylin and eosin (H&E). Histopathological changes were identified and lesions were recorded and classified (Campero *et al.*, 2003).

Immunohistochemistry

A commercial avidin-biotin complex kit (Vectastain ABC Elite, Rabbit IgG; Vector Laboratories, Burlingame, CA, USA) was used (Cobo *et al.*, 2004; Campero *et al.*, 2005). Briefly, paraffin-embedded tissue sections (5 μ m) were mounted on 3-aminopropyltriethoxy-silane coated slides (P8920; Sigma Diagnostics, St Louis, MO, USA) and dewaxed with xylene. To quench endogenous peroxidase, the sections were incubated in H₂O₂ 3% in methanol for 10 min at room temperature, and then passed through a graded alcohol series before rinsing three times in deionized water and phosphate-buffered saline (PBS; pH 7.2, 0.01M). Afterwards, 0.1% protease enzyme (P1547; Sigma) in Tris-buffered saline (pH 7.5; Dako, Carpinteria, CA, USA) was added for antigen retrieval. The slides were then rinsed twice for 3-min in PBS and treated with blocking solution (ABC kit) for 30 min at room temperature to block non-specific sites. Primary *B. abortus* antibody (*B. abortus* antiserum, DIFCO LABS, Detroit, Michigan, USA), diluted 1/200 in PBS, was applied for 45 min at 37°C. After three rinses in PBS, sections were incubated for 30 min at 37°C with secondary biotinylated rabbit anti-bovine IgG (Vector). After three additional rinses in PBS, peroxidase-labelled avidin was applied at 37°C for 30 min. Sections were rinsed again in PBS, and the enzyme activity was detected by treatment for 10 min with 3% 3-amino-9-ethylcarbazole in N,N-dimethylformamide (AEC Substrate-Chromogen System; Dako). After counterstaining with Mayer's hematoxylin, the slides were dehydrated and mounted (Crystal Mount; Biomed Corporation, Foster City, CA, USA) for examination. *B. abortus*-natural infected fetal lung tissue was used as a positive control and nonimmune normal rabbit serum was substituted for primary antibody as a negative control.

Lectin Histochemistry

Paraffin wax-embedded sections (5 mm) were used. After dewaxing, sections were treated with 0.3% hydrogen peroxide in methanol for 30 min at room temperature (to inhibit the endogenous peroxidase), rinsed several times in 0.01M PBS (pH 7.2), and treated with 0.1% bovine serum albumin in PBS for 15 min. The sections were then incubated for 1 h at room temperature with biotinylated lectins. Seven lectins (Lectin Kit BK 1000; Vector) with different specificity were used (Table 1).

Table 1. Lectins used in the histochemical studies and their major specificities.

Lectin	Acronym	Carbohydrate specificity ^{a,b}
<u>Concanavalia</u> <u>ensiformis</u>	ConA	α -D-Man; α -D-Glc
Glycine max	SBA	α -D-GalNAc; α and β -Gal
Dolichos biflorus	DBA	α -D-GalNAc
Ulex europaeus-I	UEA-1	α -L-Fuc
Triticum vulgare	WGA	α -D-GlcNAc>>NeuNAc
<u>Arachis</u> <u>hypogaea</u>	PNA	β -D-Gal(1-3)GalNA
Ricinus communis-I	RCA-1	β -D-Gal > α -D-Gal

^aGolstein and Hayes (1978); ^bFuc=Fucose; Gal=Galactose; GalNAc=N-acetyl-galactosamine; Glc=Glucose; GlcNAc=N-acetyl-glucosamine; Man=Mannose; NeuNAc=N-acetyl-neuraminic acid.

Optimal lectin concentration was 30 mg/mL in PBS for all lectins, except for PNA, which was applied at a concentration of 10 mg/mL. The slides were incubated with an avidin-biotin-peroxidase complex (ABC) (Vector) for 45 min. The horseradish peroxidase was activated by incubation for 1–2 min with a diaminobenzidine commercial kit (Dako). Specimens were rinsed in distilled water, dehydrated with graded ethanol solutions, cleared in xylene and mounted in Permount (Fisher Scientific International, Hampton, NH, USA). Controls for lectin staining included: exposure to horseradish-peroxidase and substrate medium without lectin and blocking by incubation with the appropriate blocking sugars (0.1 to 0.2M in PBS) for 1 h at room temperature before applying to the sections. The intensity of lectin binding was subjectively scored from (-) (none) to +++ (strongly positive) following the system used previously (Fernández *et al.*, 2014; Díaz *et al.*, 2017). The structures studied in the lungs were glycocalyx and cytoplasm of the epithelium of bronchi, bronchioli and alveoli and vascular endothelium. Endothelium, mesenchyme, mononuclear throphoblast and binucleate giant cells were observed in placentas.

Results and Discussion

Lectins are glycoproteins or proteins of non-immune origin that are obtained from plants and animals. They have been used in many areas of diagnostic research, mainly those related to changes in the expression of membrane and cytoplasmic glycoconjugates (Walker, 1989; Pilobello and Mahal, 2007). In the present work, the effect of *B. abortus* on lectin-histochemical patterns in infected lungs and placentas in comparison to uninfected controls was evaluated.

As expected, no histopathological lesions were found in the non-infected fetuses and placentas. Table 2 shows the histopathological and immunohistochemical findings in fetuses/stillbirth lungs and placentas with *B. abortus* isolation.

Table 2. Histopathological and immunohistochemical findings in fetuses/stillbirth lungs and placentas with *Brucella abortus* isolation.

Sample N°	Fetuses/stillbirth age	Culture	Histopathological lesions ¹		Immunohistochemical ²	
			Lungs	Placenta	Lungs	Placenta
99	1 day	+	+++	nc	+++	nc
100	3 days	+	+++	+	+	+++
161	8 months	+	+++	nc	+++	nc
160	8 months	+	+++	nc	++	nc
135	8 months	+	+++	nc	++	nc
141	8 months	+	nc*	+	nc*	+++
142	8 months	+	nc*	++	nc*	+++
97	7 months	+	+++	+	+	++
150	8 months	+	nc*	++	nc*	+++
Controls	> 7 months	(-)	(-)	(-)	(-)	(-)

(1): Lesions: (-) no lesions observed; +: mild; ++: moderate; +++: severe. (2): Amount of antigen: (-): negative; +: low; ++: moderate; +++: abundant. nc: not collected sample. nc*: alive calf.

B. abortus-positive aborted fetuses developed histopathological changes similar to those described by other authors in experimental and natural infections (López et al., 1984; Sözmen et al., 2004). A multifocal distribution of severe bronchopneumonia was the most common change that was found in the lungs of the examined fetuses. The predominant inflammatory infiltrate was composed of macrophages, lymphocytes and neutrophils. This infiltrate was located in the interalveolar septa and was less abundant in lumina of alveoli and bronchi. In agreement with that previously reported, the fibrinous pleuritis was a lesion found only in some infected fetuses (Xavier et al., 2009). Placentitis was found in all the placentas examined, but the intensity of the lesion varied from slight to moderate. Mononuclear infiltration was found more frequently than polymorphonuclear infiltration as was previously reported in ovines by Yazicioglu and Hazaroglu (2000). Bacterial colonies were found in the necrotic areas of villi of one sample. Necrotic vasculitis with necrosis and mononuclear cell infiltration in the vessel walls was observed in two placentas. The necrotic placentitis affecting the depth of the villus was similar to that previously described in bovines and ovines (Yazicioglu and Haziroglu, 2000; Xavier et al., 2009). However, differently from that observed in previous reports, the polymorphonuclear neutrophils were found only in two placentas (Pérez et al., 1998). This observation is in agreement with the lung results, where only one fetus showed neutrophilic infiltrate, but not in agreement with the results of other works (Pérez et al., 1998). These differences could be originated by some differences in the experimental conditions, as Xavier et al. (2009) inoculated the animals at a later age while Perez et al. (1998) obtained the lungs only by natural infection abortion.

The results obtained by immunohistochemistry are similar in bovines, goats, ovines and mice infected with *Brucella* spp (Meador et al., 1986; Pérez et al., 1998; Yazicioglu and Haziroglu, 2000; Sözmen et al., 2004). Labelled *B. abortus* was seen in all evaluated tissues from infected fetuses and placentas but not in controls. It was demonstrated that *B. abortus* is able to replicate and persist within mouse and human lung epithelial cells (Kahl-McDonagh et al., 2007; Ferrero et al., 2009). However, in the present work, immunohistochemistry positivity to *B. abortus* was not found in epithelial lung cells. In the placentas, bacteria were detected extracellular in the necrotic areas and in the macrophages and mononuclear trophoblasts, but not in the cytoplasm of binucleated giant cells. In the lungs, immunolabeling was detected in macrophages and extracellular in the exudate found in the alveoli and bronchial lumen (Fig. 1). These localizations are in accordance with the characteristics of *Brucella* infections. The members of the *Brucella* genus are facultative intracellular pathogens that enter phagocytic cells such as neutrophils, macrophages and trophoblast cells and inhibit the killing mechanism of these professional phagocytic cells (Gorvel and Moreno, 2002). Furthermore, *Brucella* has the ability to interfere with intracellular trafficking, preventing fusion of the *Brucella*-containing vacuole with lysosome markers, and directing the vacuole towards a compartment that has rough endoplasmic reticulum, which is highly permissive to intracellular replication of *Brucella* (Pizarro-Cerdá et al., 2000). However, some non-phagocytic cells may also be invaded by *B. abortus* in cell lines *in vitro* (Pizarro-Cerdá et al., 1998). Lectin binding patterns for the seven lectins in lungs and placentas of the infected and non-infected animals are summarized in Tables 3 and 4.

Table 3. Lectin staining pattern in placentas of cows infected with *Brucella abortus* (Brc) and uninfected controls (C).

Placenta	UEA1		WGA		RCA1		ConA		PNA		SBA		DBA	
	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc
Trofoblast														
Glycocalix	(-)	+	+++	++	+++	+++	+	++	(-)	++	+++	+++	(-)	+++
Apical cytoplasm	(-)	+	+	+	++	++	(-)	+	(-)	+	+	++	(-)	++
Endothelium	(-)	(-)	+	+	++	+	+	+	(-)	(-)	+	+	(-)	(-)
Mesenchyme	(-)	+	++	+	+	+	+	+	(-)	(-)	(-)	+	(-)	(-)
Binucleate giant cells	(-)	(-)	++	+	+	+	(-)	+	(-)	+	(-)	(-)	(-)	(-)

Table 4. Lectin staining pattern in fetal bovine lungs infected with *Brucella abortus* (Brc) and uninfected controls (C).

Lung	UEA1		WGA		RCA1		ConA		PNA		SBA		DBA	
	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc	C	Brc
Bronchi														
Glycocalix	(-)	++	++	++	+	+++	++	+++	(-)	++	++	+++	(-)	+++
Apical cytoplasm	(-)	++	+	+	(-)	++	+	++	(-)	+	+	++	(-)	++
Bronchiole														
Glycocalix	(-)	++	++	+++	+	+++	++	+++	(-)	++	++	+++	(-)	+
Apical cytoplasm	(-)	+	+	++	(-)	++	+	++	(-)	++	+	++	(-)	(-)
Alveoli														
Glycocalix	(-)	+	+++	++	++	+++	+	++	(-)	++	++	+++	(-)	+
Apical cytoplasm	(-)	(-)	+	++	(-)	++	(-)	++	(-)	+	+	++	(-)	(-)
Endothelium	(-)	(-)	(-)	++	++	++	(-)	++	(-)	(-)	++	+	(-)	+

An increase in the binding to the majority of the lectins was observed in the lungs. The changes were most evident for UEA-1, RCA-1, PNA and DBA (Fig. 2), thus demonstrating an increase in the fucose, galactose and N-acetyl-galactosamine residues (Golstein and Hayes, 1978). DBA binding was found as a finely granular intracytoplasmic staining in some macrophages.

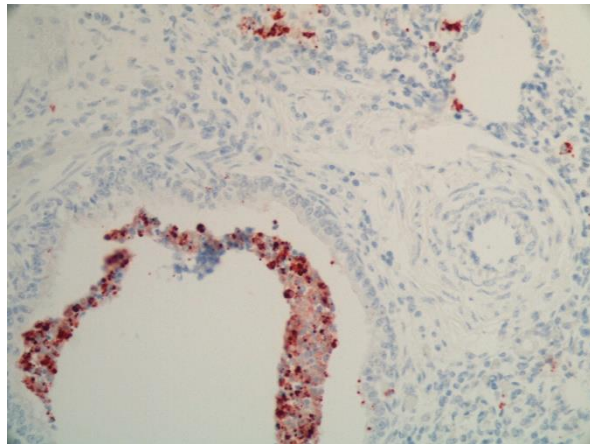


Fig. 1. *Brucella abortus* in the bronchial lumen of a bovine fetus with bronchopneumonia. IHQ 40x.

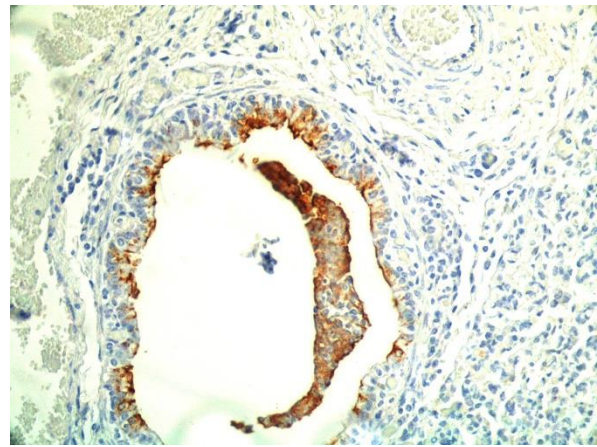


Fig 2. Pattern of DBA found in bovine fetal lung with isolation of *B. abortus*. 40x.

We have not found previous reports on lectin histochemistry in fetal bovine lungs infected with *B. abortus*. In normal human fetus lungs, Barkhordari *et al.* (2004) found a similar lectin binding pattern in the endothelium but not in bronchial or alveolar cells, thus demonstrating the variability between the species for the epithelium patterns of glycoconjugates. On the

other hand, the changes observed in fetuses infected with *Campylobacter fetus* were less marked than those found in the present study (Morrell *et al.*, 2010). Therefore, the comparison of these lectinohistochemical results demonstrates that the glyconjugate modifications in the lung epithelium are specific for each infecting microorganism. In our study, the lectin binding pattern in mononuclear normal trophoblasts was similar to that reported by Munson *et al.* (1989), shows the scarce binding by DBA and UEA-1 lectin in the villus.

In the binucleate giant cells, the binding to some lectins, such as UEA-1 and WGA, was similar to that found in previous reports in bovine near-term placenta (Jones *et al.*, 1994). However, the affinity for DBA and SBA in the near-term placenta binucleate giant cells was higher than the one found in the present study (Jones *et al.*, 1994).

This difference could be attributed to changes in the secretion activity of these cells during pregnancy (Woodwing and Burton, 2008). In the mononuclear trophoblast of the placentas of infected animals, an increase in the binding to PNA, DBA and UEA-1 was found. However, in the other analyzed structures like blood vessels, mesenchyme and binucleate giant cells, the changes were less relevant. This result is in agreement with the presence of *B. abortus* in the cytoplasm of these cells evidenced by immunolabeling. The changes in the lectin binding pattern could be a response to the penetration of the bacterium in the mononuclear trophoblast.

In a recent work, Carvalho Neta *et al.* (2008) reported changes in the expression of 62 genes (25 increased and 37 downregulated expression) in explants of chorioallantoic bovine membrane infected with *B. abortus*. The lectin pattern is an indirect result of the gene expression (Gimeno and Barbeito, 2004). In the present work, an increase in the binding of UEA-1, DBA and PNA in the trophoblasts of infected animals was found.

These changes in the labeling pattern evidenced a modification in the expression of lectins in cells infected with *Brucella*. In uninfected binucleated giant cells, there were no significant changes in the lectin pattern.

Notably, phagocytic cells as trophoblasts mononuclear and macrophages showed an intracellular labeling to DBA, similar to that observed in immunohistochemistry. As mentioned above, there are no previous reports on the affinity of lectins on animal tissues infected with *B. abortus*. However, in a study on *Brucella* cultures, the microorganism showed affinity for the Con-A lectin, demonstrating the presence of glucose and mannose residues, although they were negative for lectins that bind to N-acetylgalactosamine (Corbel *et al.*, 1983a,b).

In our study, *Brucella* labeling for the Con-A lectin was not observed but labeling was observed for DBA lectin in phagocytic cells. These differences could be due to changes in the bacterial surface generated by eukaryotic cell endocytic systems.

Conclusion

The present lectin histochemical study, supported by bacteriological cultures, immunohistochemistry and histopathology, revealed a distinctive pattern of oligosaccharide distribution in the lungs and placenta from *B. abortus*-infected fetuses and the positive DBA binding of this bacterium.

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Conflict of interest

The authors declare that there is no conflict of interests.

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