

Regulation of cochlear outer hair cells

Insights from mathematical modelling

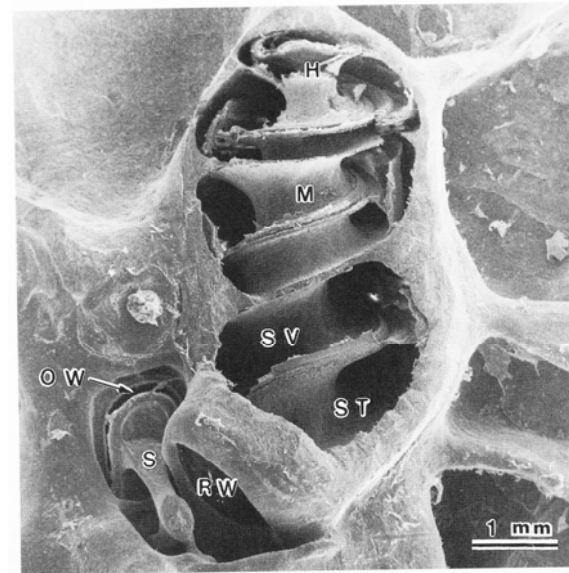
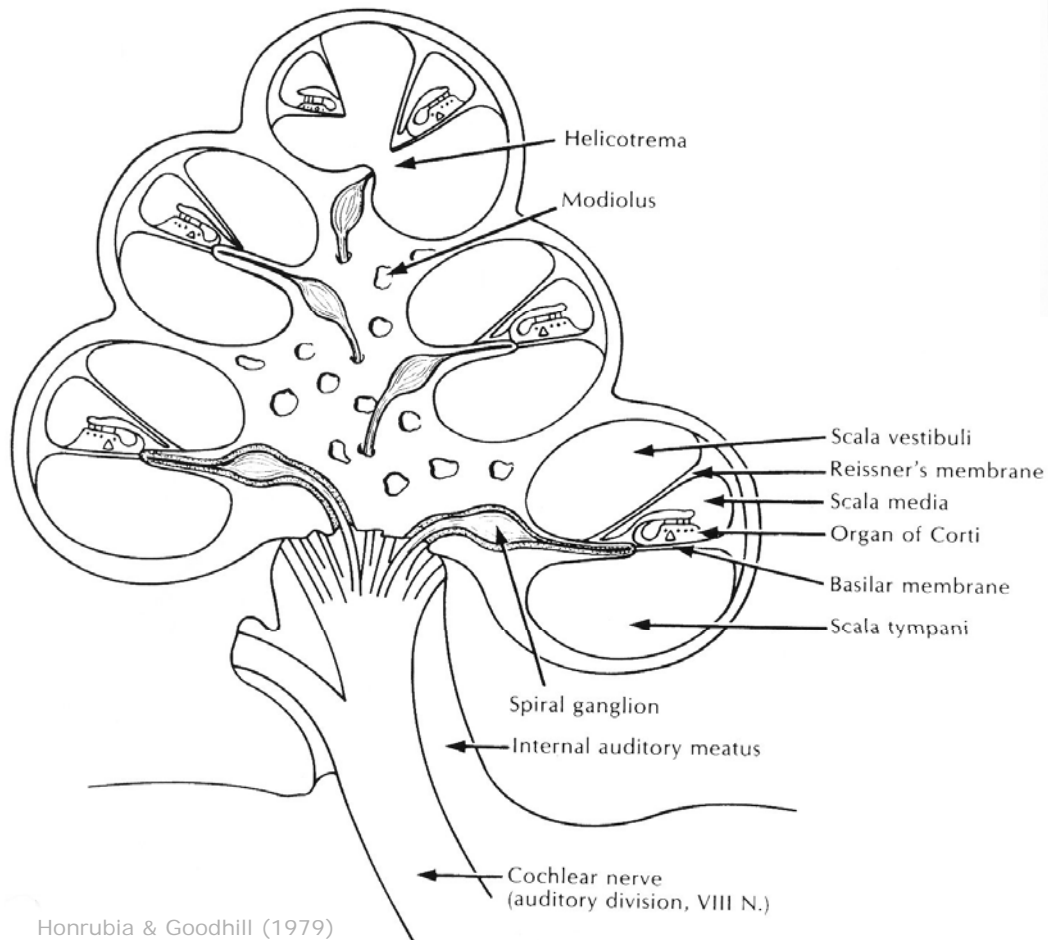
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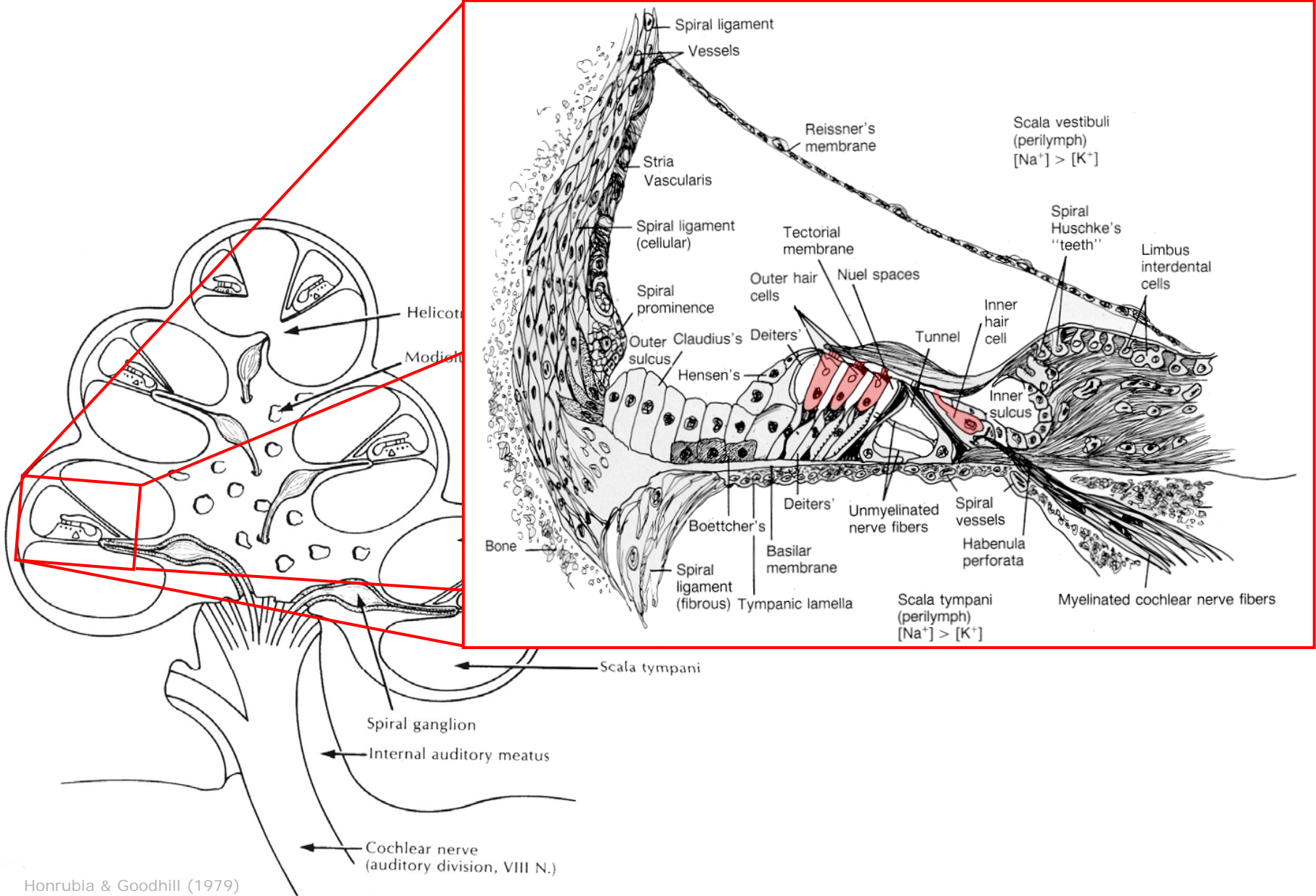
The Auditory Laboratory, Physiology
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Mid-modiolar cross-section of the cochlea



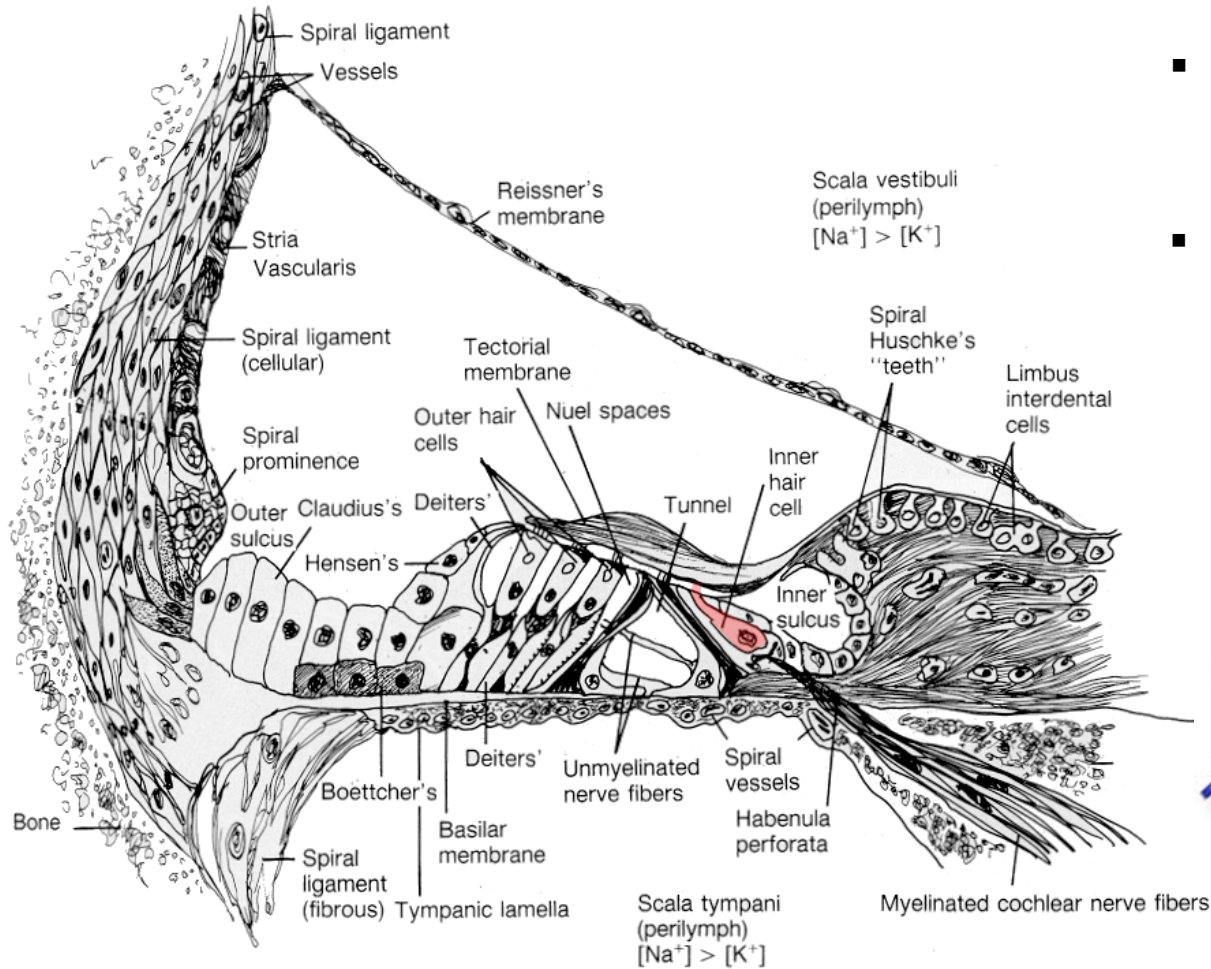
View of the cochlea after removal of the otic capsule

Mid-modiolar cross-section of the cochlea

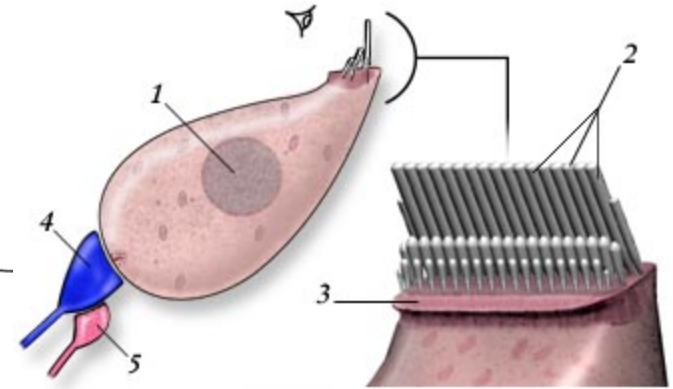


Honrubia & Goodhill (1979)

Inner hair cells

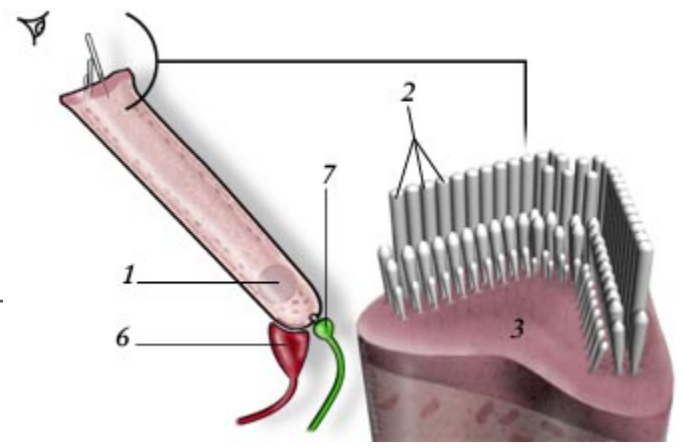
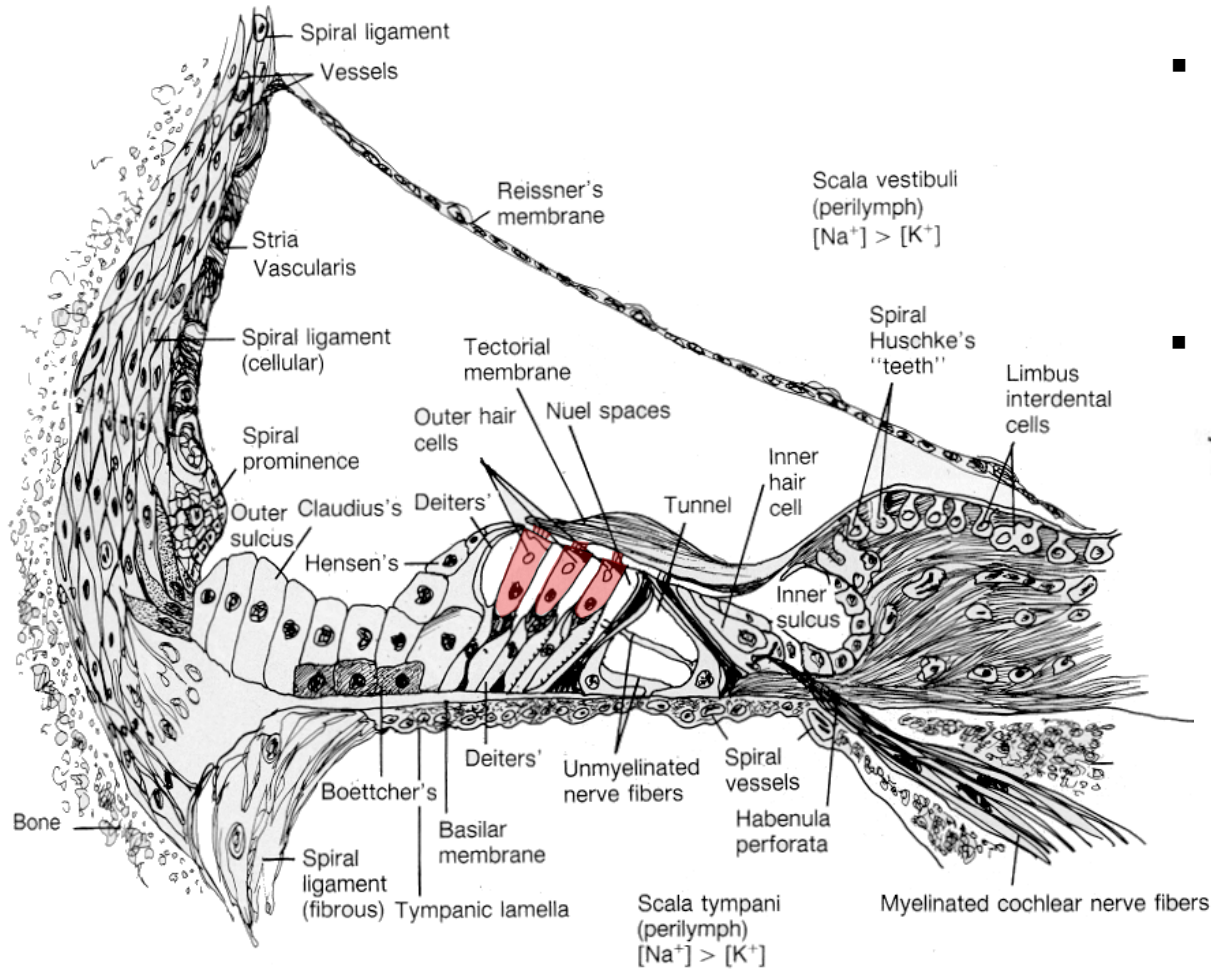


- Inner hair cells (IHCs) carry out **mechanoelectrical transduction** (or MET)
- Stereociliary deflection results in depolarisation and release of neurotransmitter

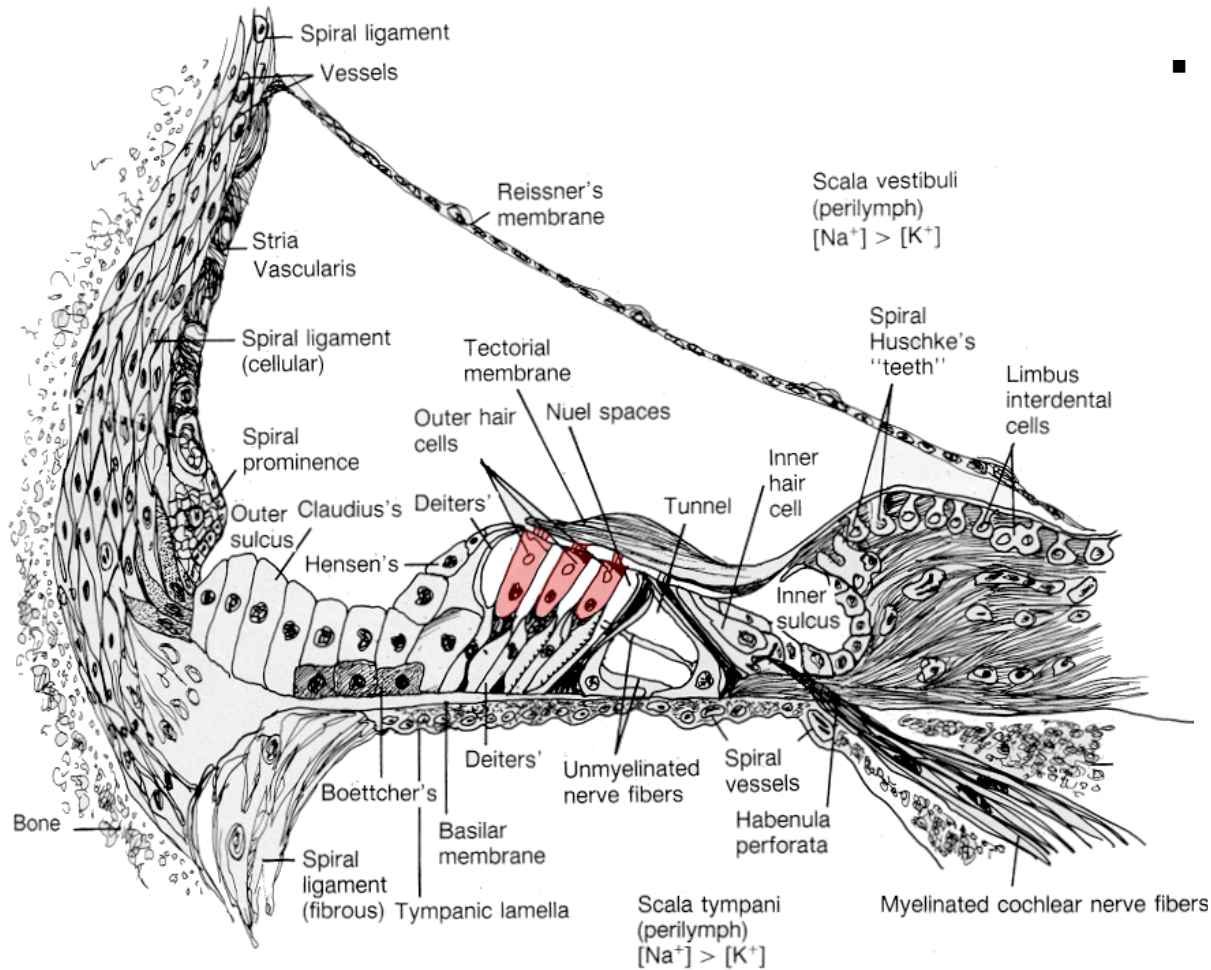


Outer hair cells

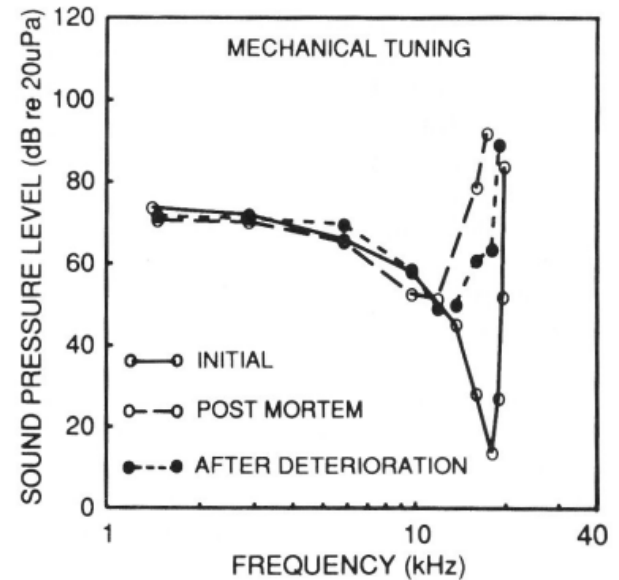
- Outer hair cells (OHCs) perform mechano-electrical transduction, but also carry out **electromechanical transduction (EMT)**
- That is, they are **motile**.



Outer hair cells



- Outer hair cells (OHCs) perform mechano-electrical transduction, but also carry out **electromechanical transduction (EMT)**

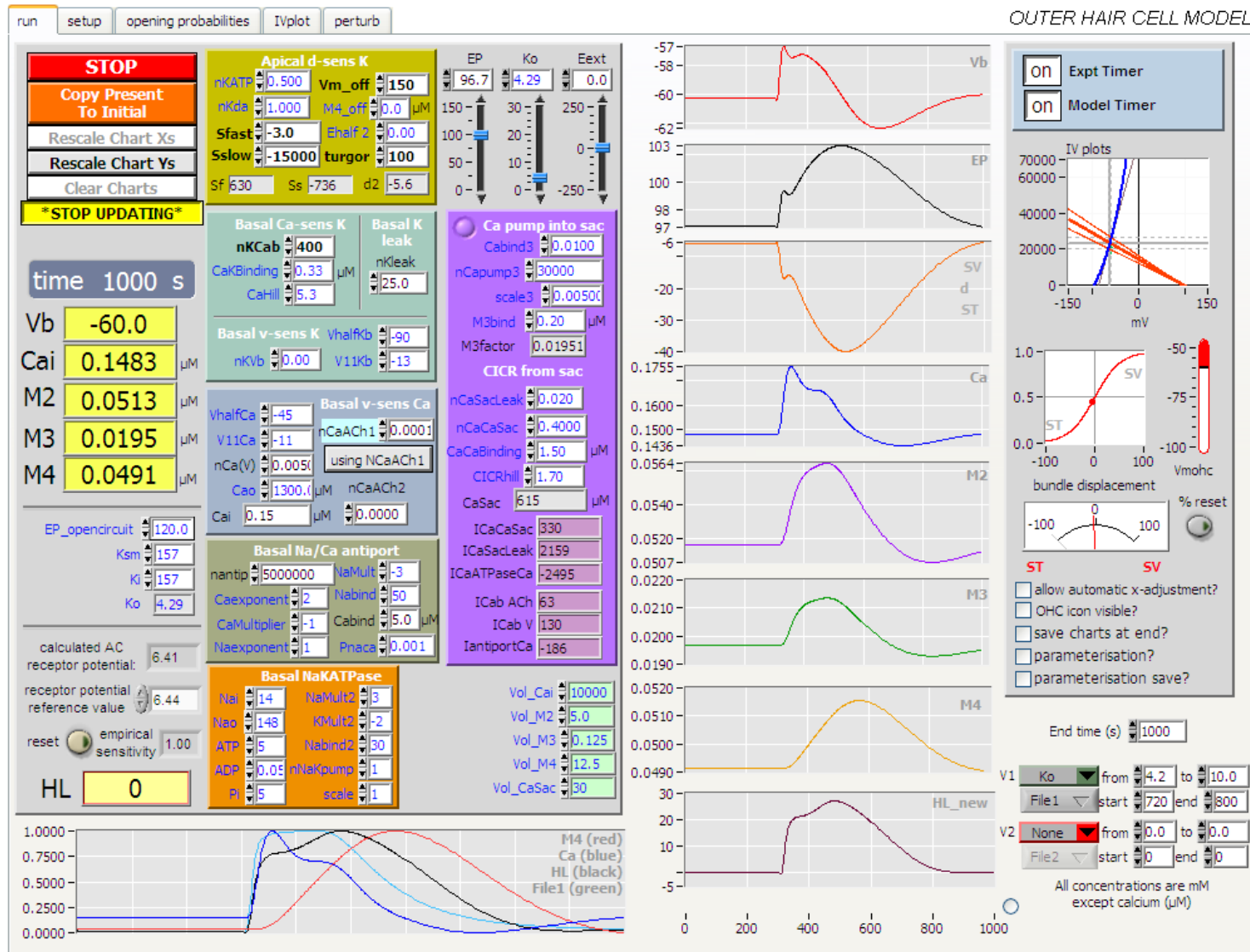


Patuzzi, 1992

- The "active process" increases basilar membrane vibration x1000 (or 60 dB)
- Allow sharp mechanical tuning and frequency resolution
- Compressive nonlinearity allows a million-fold range of sound pressures

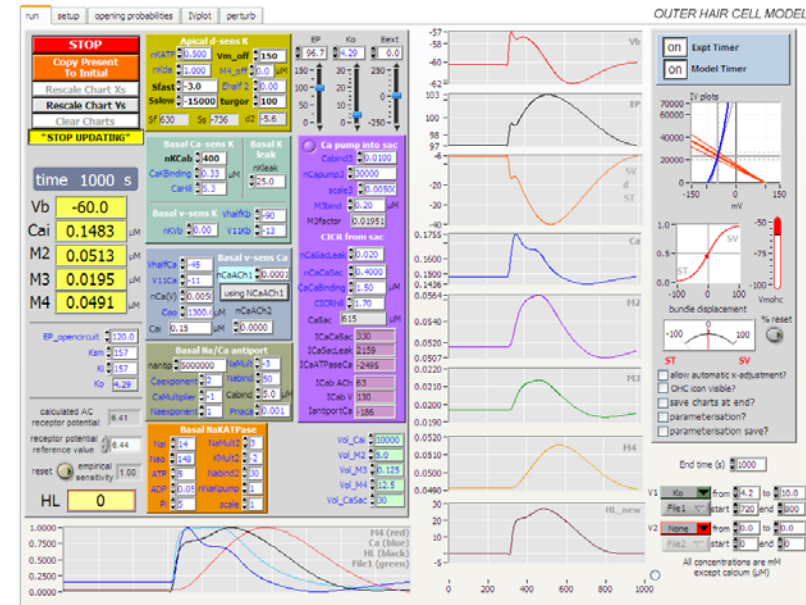
Outer hair cell mathematical model

O'Beirne, G.A. and Patuzzi, R.B. (2007). Mathematical model of outer hair cell regulation including ion transport and cell motility. *Hearing Research* 234:29-51.



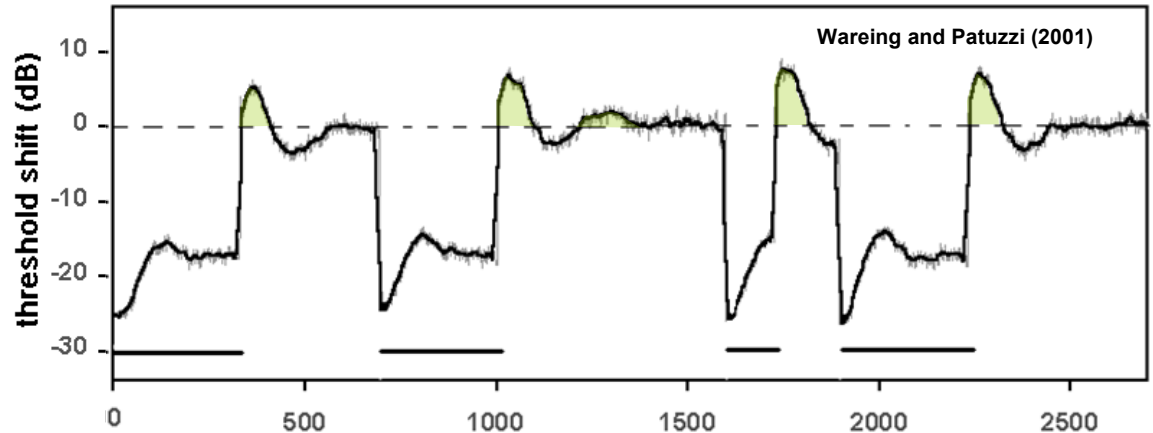
Outer hair cell mathematical model

- incorporates known OHC electrophysiology
- includes effect of hair cell motility on the conductance of the apical membrane
- simulates perturbations by systematic and timed variation of model parameters
- predicts changes in active gain due to OHCs, and therefore hearing loss
- provides a plausible and consistent explanation for **slow oscillatory behaviour** observed in the cochlea

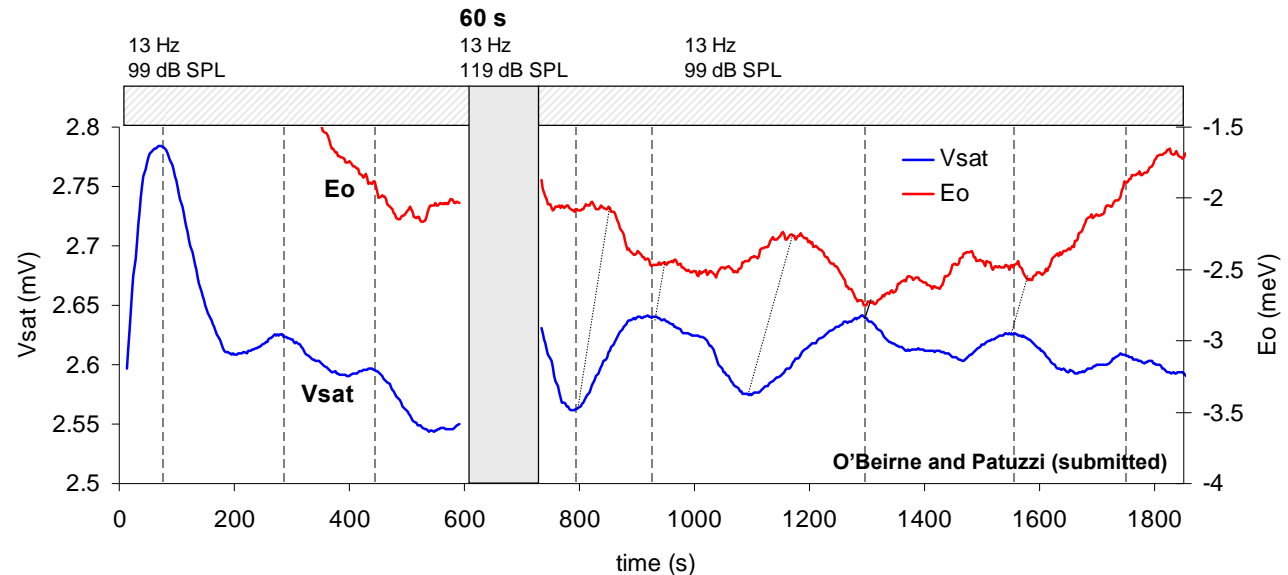


The low-frequency “bounce phenomenon”

- Oscillation of psychophysical thresholds in human subjects (Hirsch and Ward, 1952; Zwicker and Hesse, 1984; Wareing and Patuzzi, 2001)



- Mechanical in origin (Kemp, 1982, 1986; Kirk and Patuzzi, 1997; Kirk et al., 1997)



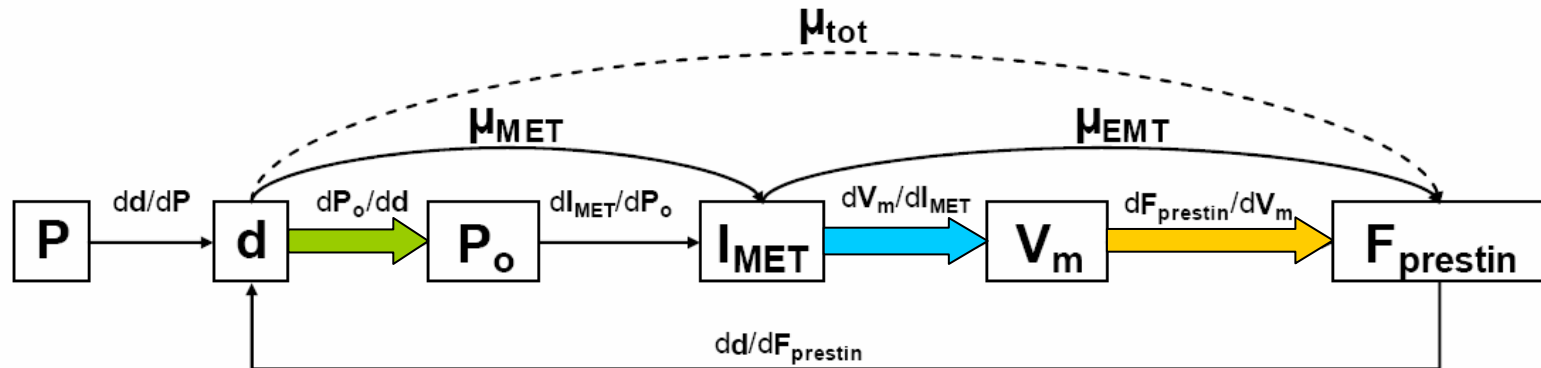
- Oscillations in measures of hair cell function (OAEs, LF CM) (Kirk and Patuzzi, 1997; Kirk et al., 1997; O'Beirne and Patuzzi, submitted.)

How? Why? What?

- How does the presentation of a low-frequency tone cause oscillations in hair cell performance?
- How do the hair cells perform *better than normal*?
- Why aren't they that good all the time?
- What is the underlying mechanism for the bounce?

What do OHCs need for maximum amplification?

- The OHC active process depends on the efficiency of each step from sound-induced movement to hair-cell produced movement.

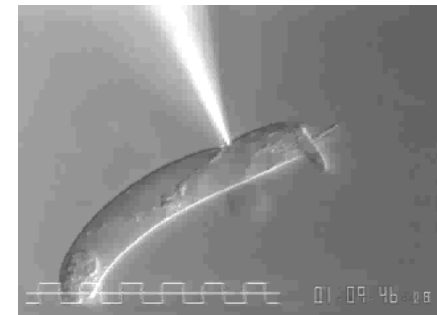
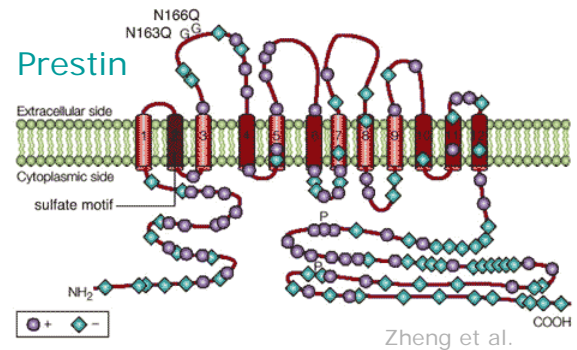
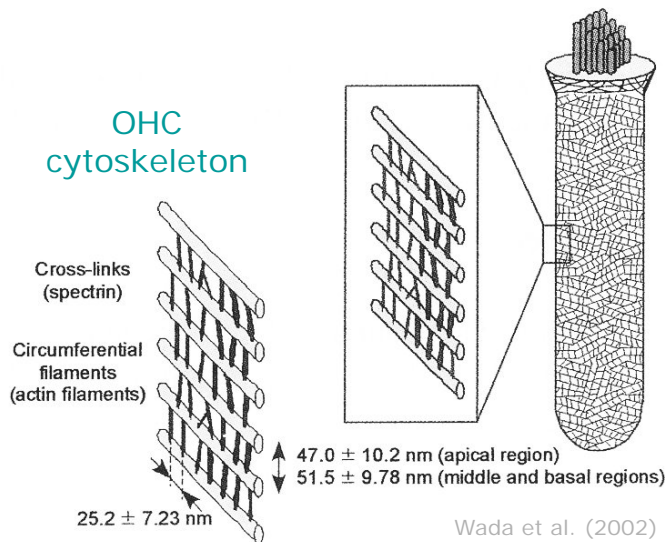


O'Beirne (2005)

- In terms of normal regulation, the most critical factors are:
 - that the hair bundles are in their **most sensitive position** (*i.e. maximal receptor current for a given displacement*)
 - that the **resistance** of the OHC basolateral wall is high (*i.e. maximal receptor potential for a given receptor current*)
 - that V_m is in the **most sensitive** region for motor proteins (*i.e. maximal force production for given receptor potential*)

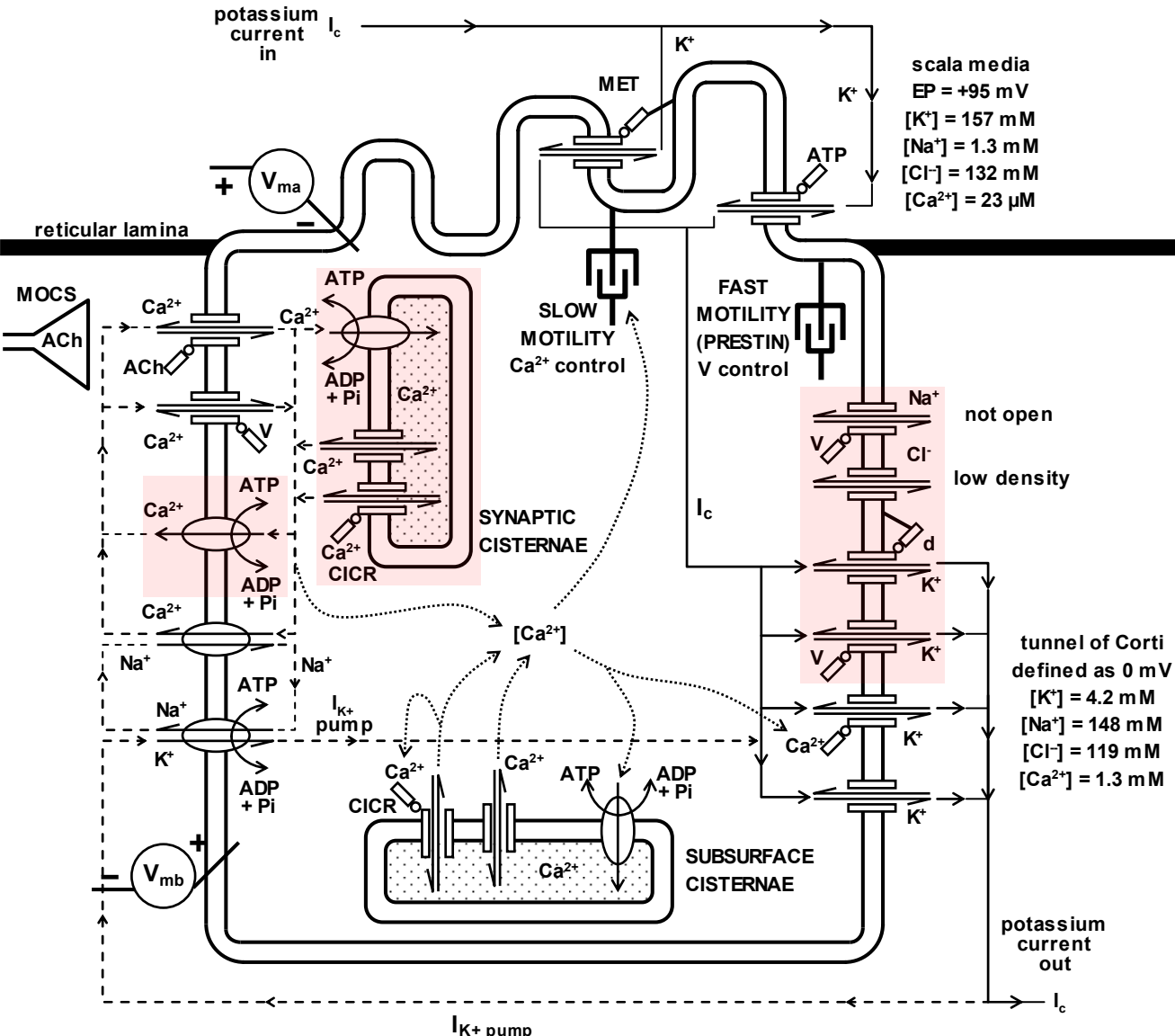
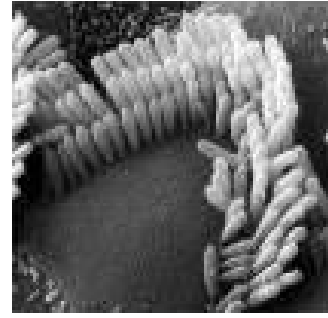
How does the OHC maintain hair bundle angle?

- Motility
 - Contractions or elongations of the hair cell alter the resting hair bundle angle (operating point)
 - Two types of motility:
 - slow (calcium-based)
 - fast (electromotility)



Frolenkov et al. (1998)

Schematic diagram of OHC cellular components



Apex

- d-sensitive K^+ channel
- ATP-sensitive cation channels

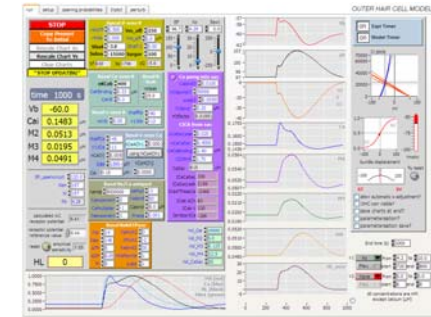
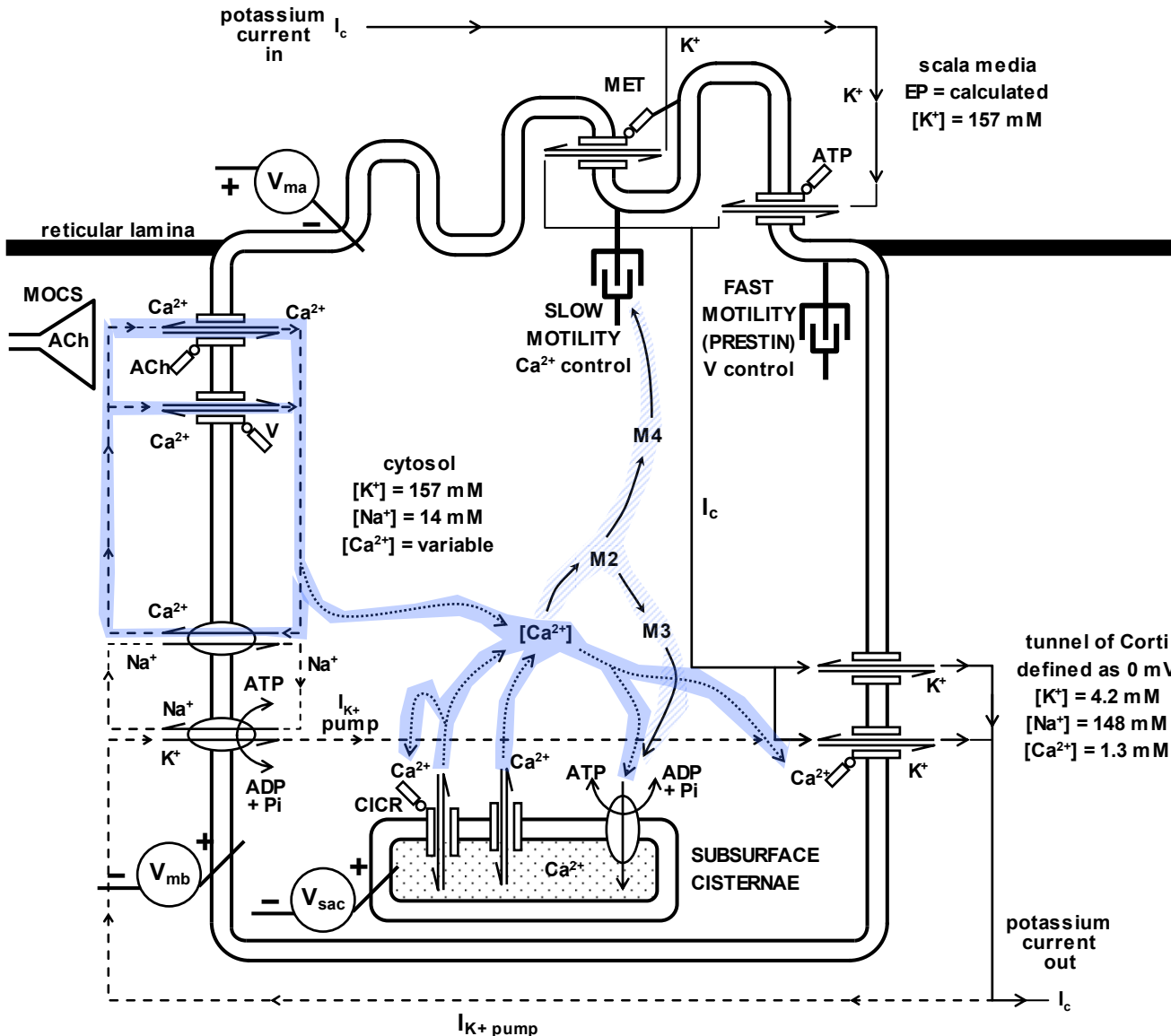
Base

- K^+ leakage channel
- Ca^{2+} -sensitive K^+ channel
- Na^+/K^+ ATPase
- Ca^{2+}/Na^+ antiport
- v-sensitive Ca^{2+} channel
- ACh-sensitive Ca^{2+} channel

Subsurface/Synaptic Cisternae

- Ca^{2+} induced Ca^{2+} release
- Ca^{2+} leakage
- Ca^{2+} ATPase

Simplifications/alterations required for modelling



Apex

- d-sensitive K^+ channel
- ATP-sensitive cation channels

Base

- K^+ leakage channel
- Ca^{2+} -sensitive K^+ channel
- $Na^+/K^+/ATPase$
- Ca^{2+}/Na^+ antiport
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Subsurface/Synaptic Cisternae

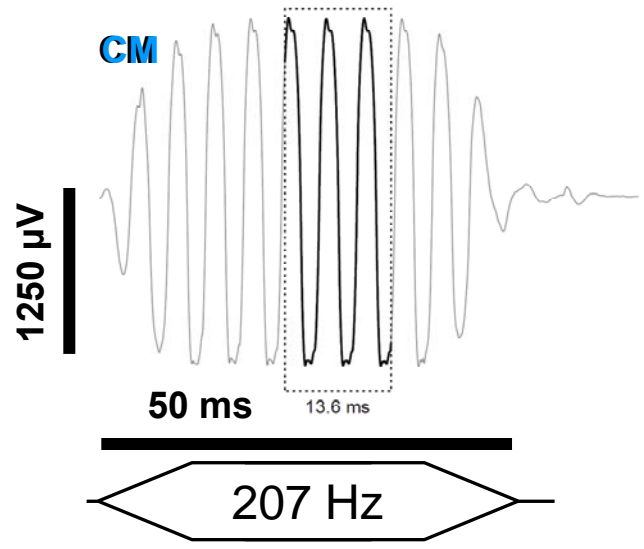
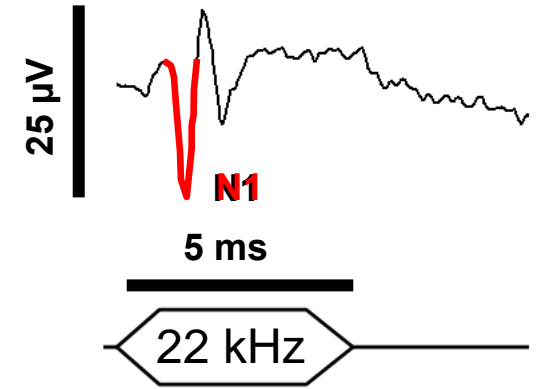
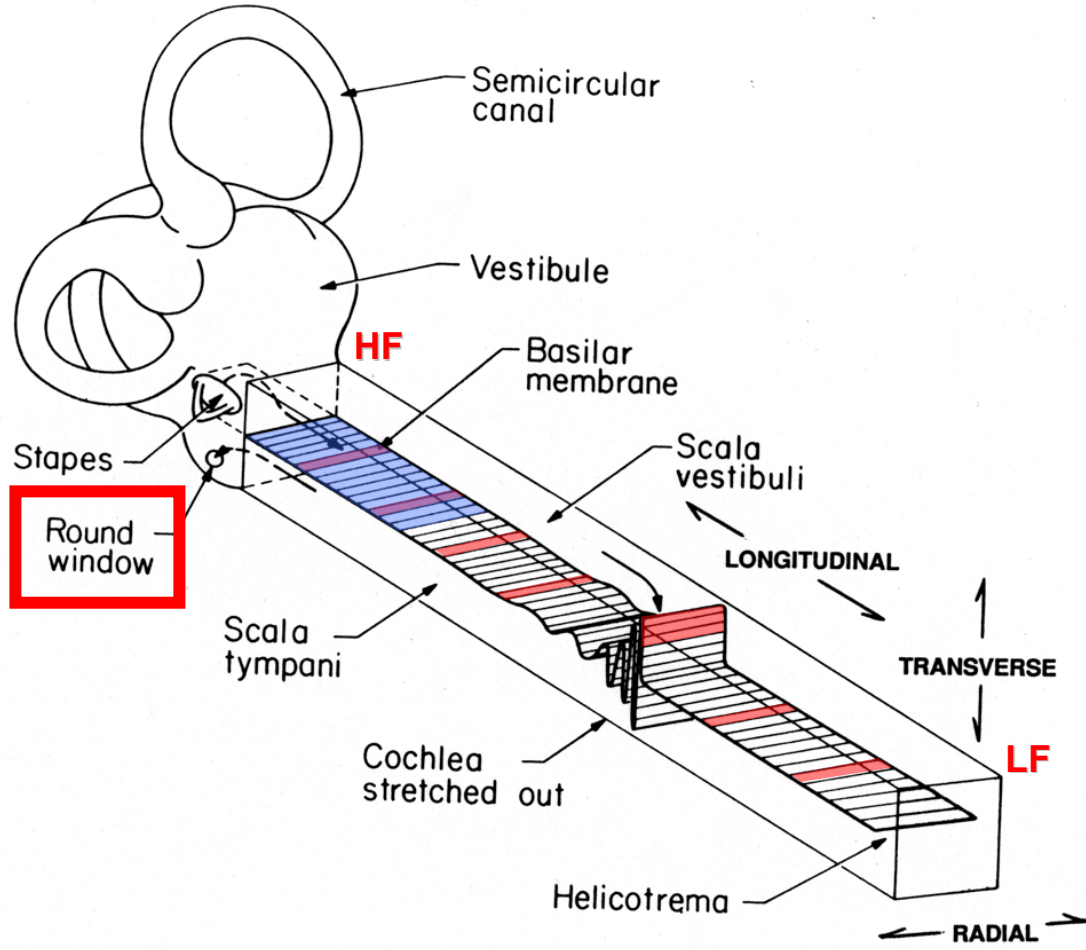
- Ca^{2+} induced Ca^{2+} release
- Ca^{2+} leakage
- Ca^{2+} ATPase

Selection of model parameters based on experimental data

1. Electrophysiological experiments in guinea pigs
 - perturb the cochlea in some way (e.g. low-frequency tones, perfusions of artificial perilymph, current injection, mechanical bias)
 - measure just about everything simultaneously
 - analyse interactions between measured cochlear parameters
2. Simulate the experiment in the mathematical model and fine-tune parameters to match the experimental data
3. Repeat.

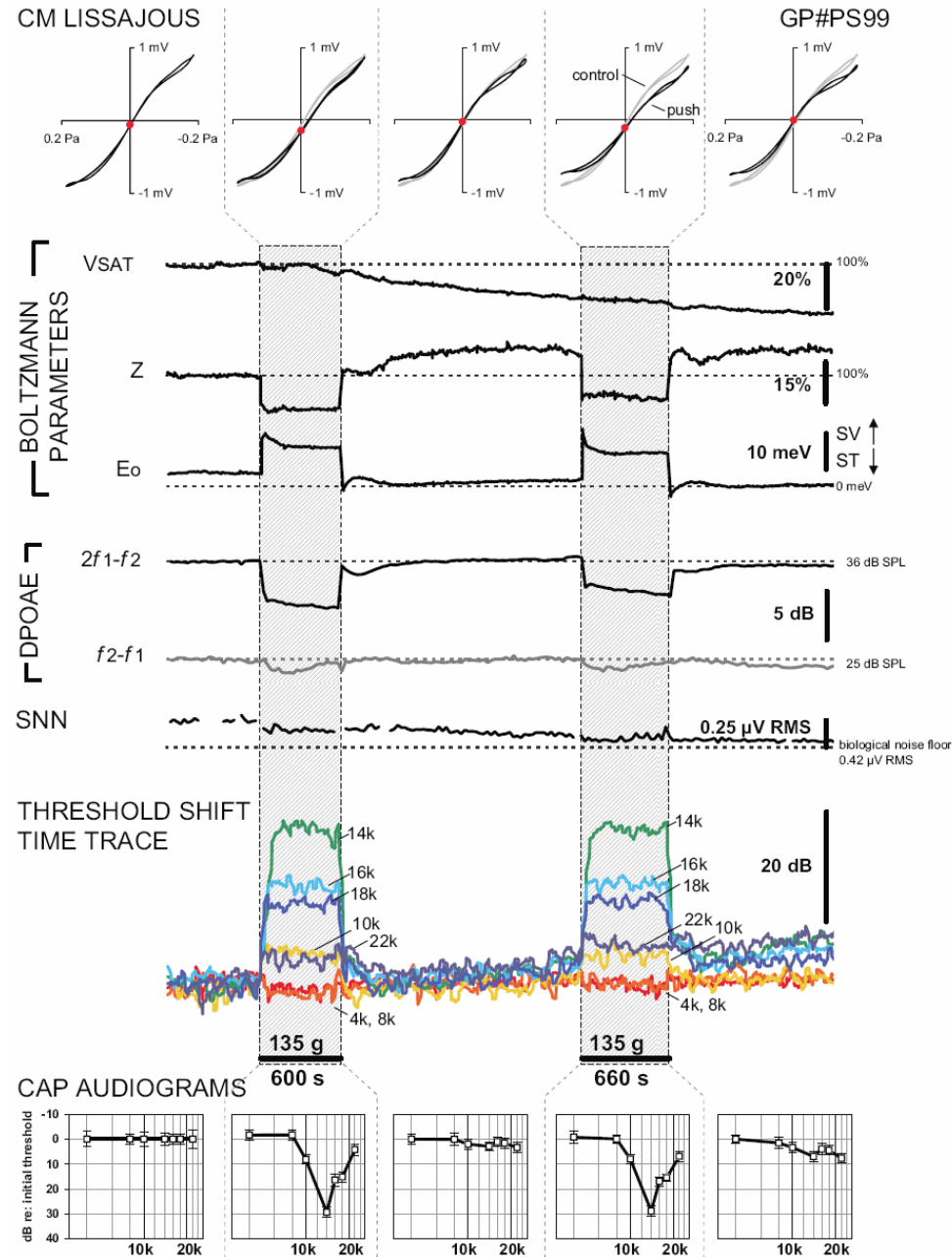
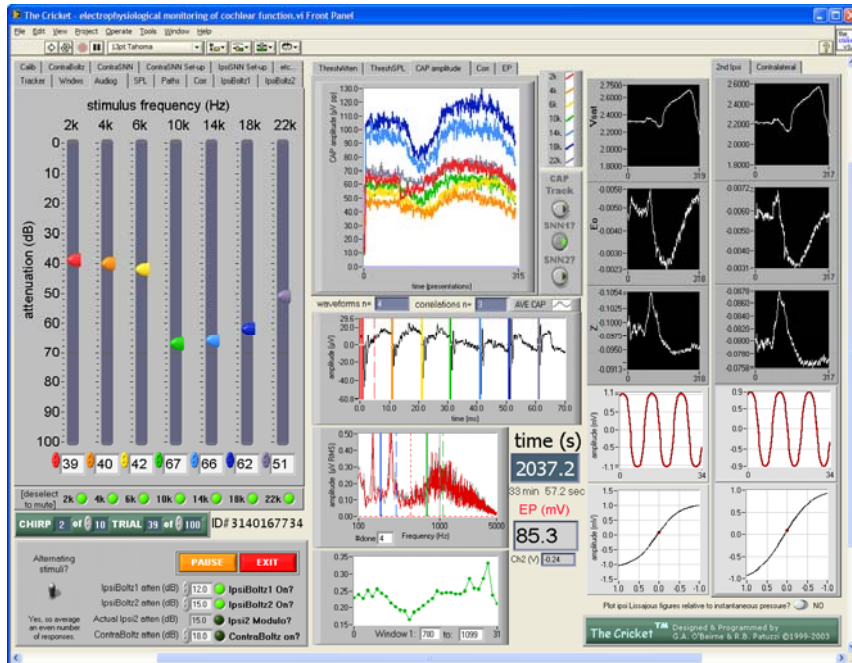
Electrophysiological monitoring of cochlear function

- Round-window electrocochleography



Electrophysiological monitoring of cochlear function

- Custom-written software allowed near-simultaneous measurement of:
 - CAP thresholds and waveforms at seven different frequencies
 - Boltzmann analysis of 200 Hz CM
 - Distortion-product OAEs
 - Endocochlear potential (EP)
 - Spectrum of neural noise (SNN)



Examples of experimental perturbations and model results

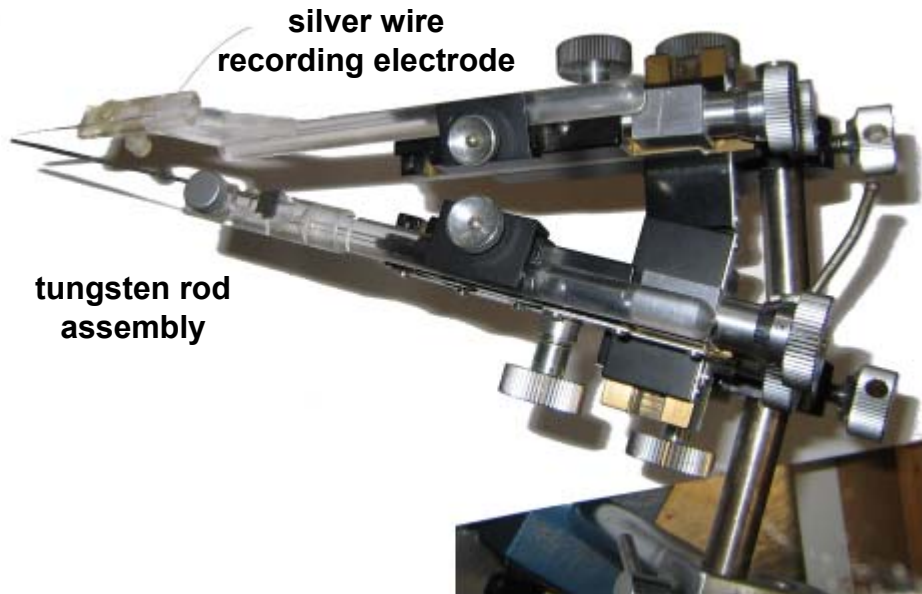
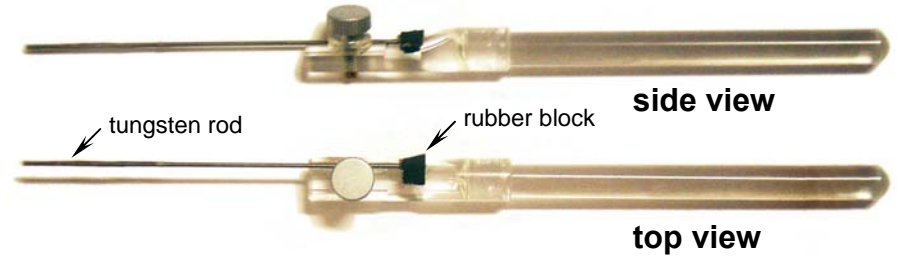
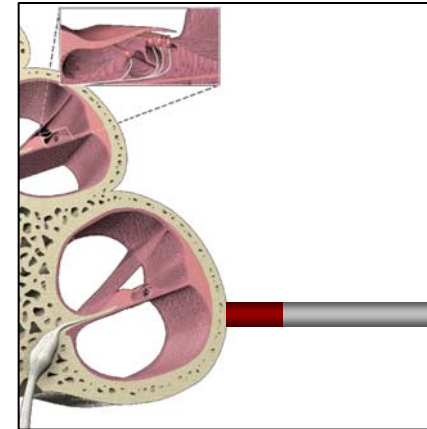
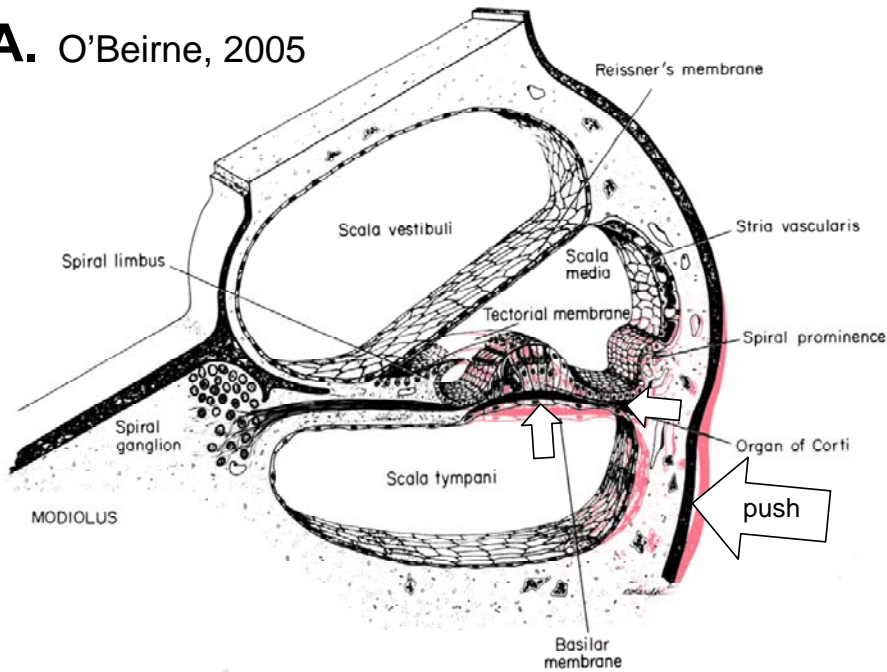
- Model tested and refined using data from experiments conducted in the guinea pig

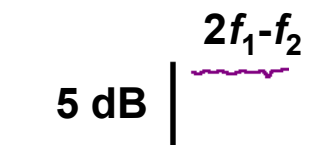
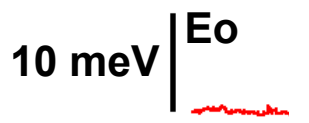
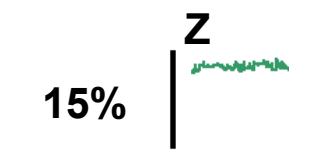
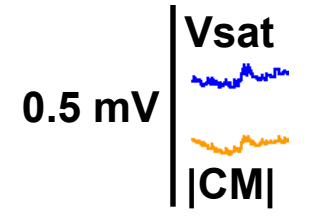
- **Examples:**
 - Cochlear perfusions
 - Systematic variation of model parameters
 - Application of force to the cochlear wall
 - Generation of hair cell regulation curves
 - The bounce
 - Accumulation and removal of intracellular calcium
 - Effect on basolateral permeability

Application of force to the cochlear wall

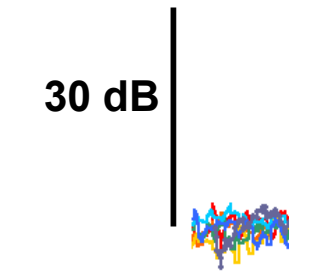
- Force applied to the cochlear wall using a 1 mm diameter insulated thoriated-tungsten rod.
- Static displacement of basilar membrane.

A. O'Beirne, 2005



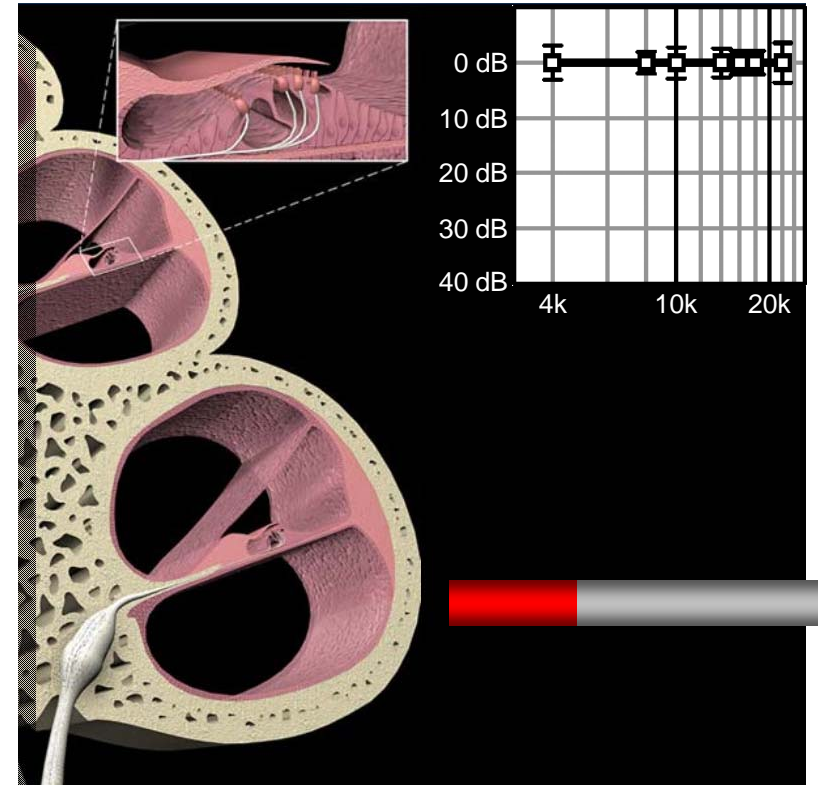
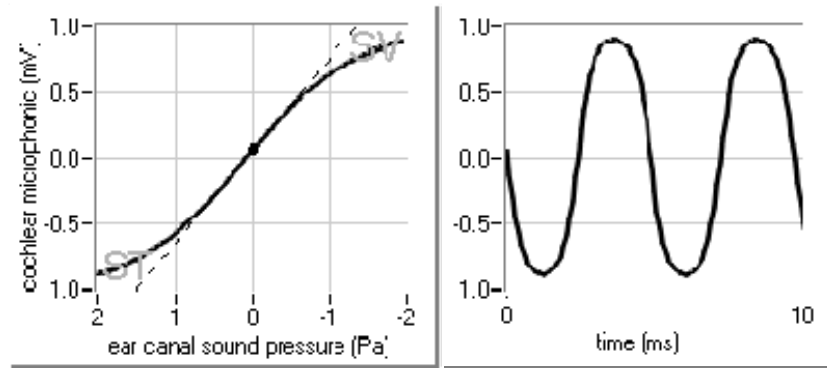


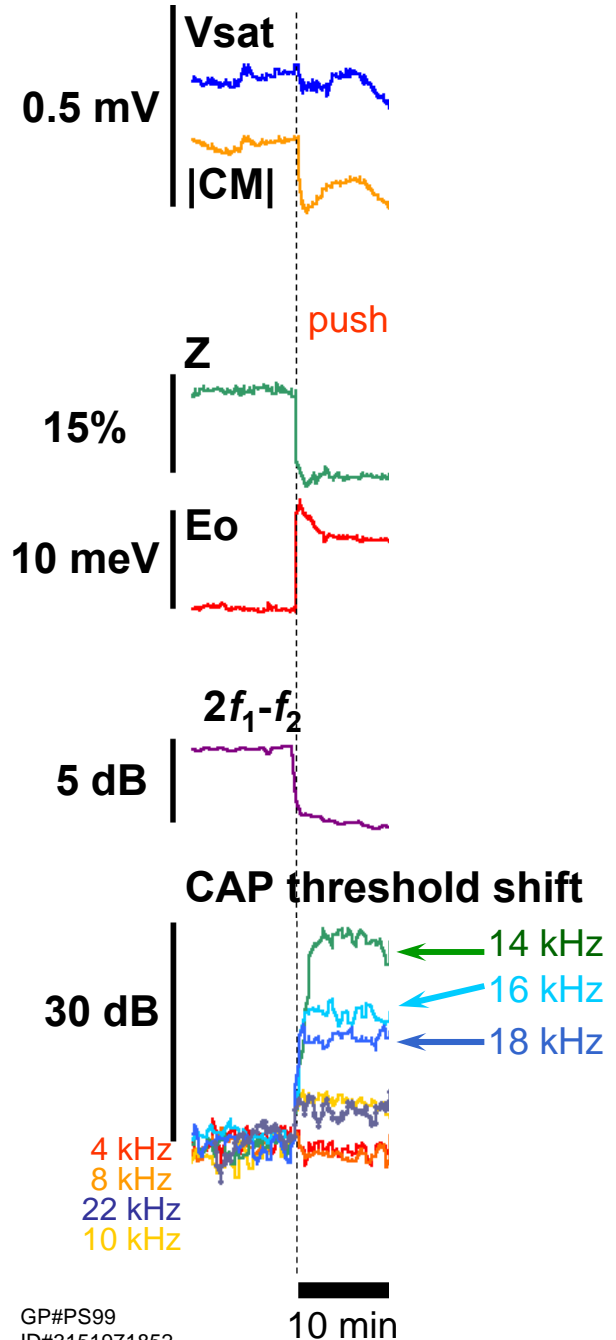
CAP threshold shift



SV ↑
ST ↓

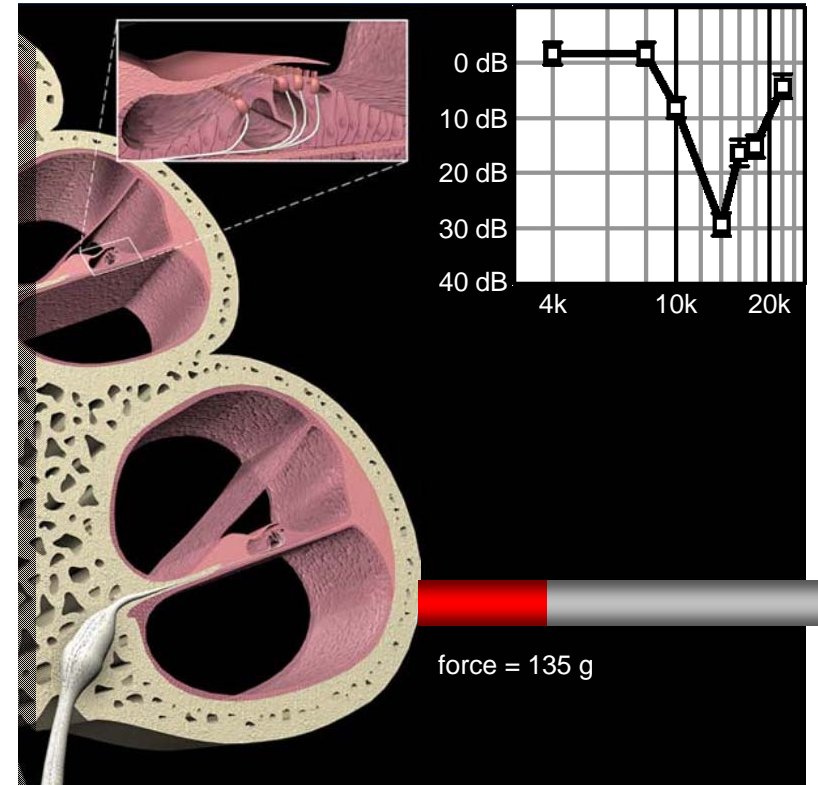
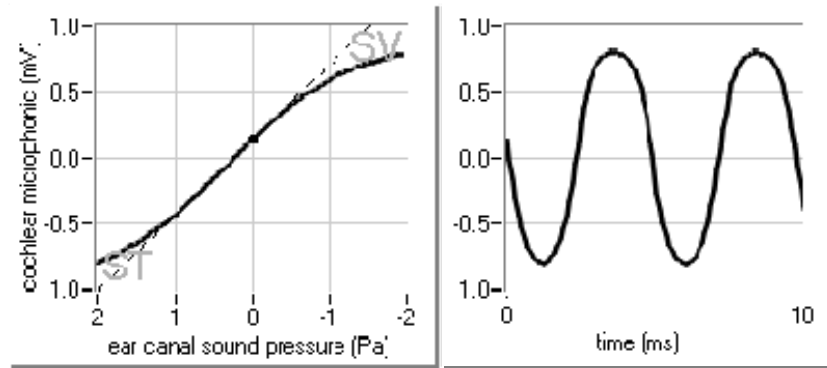
application of force to the cochlear wall

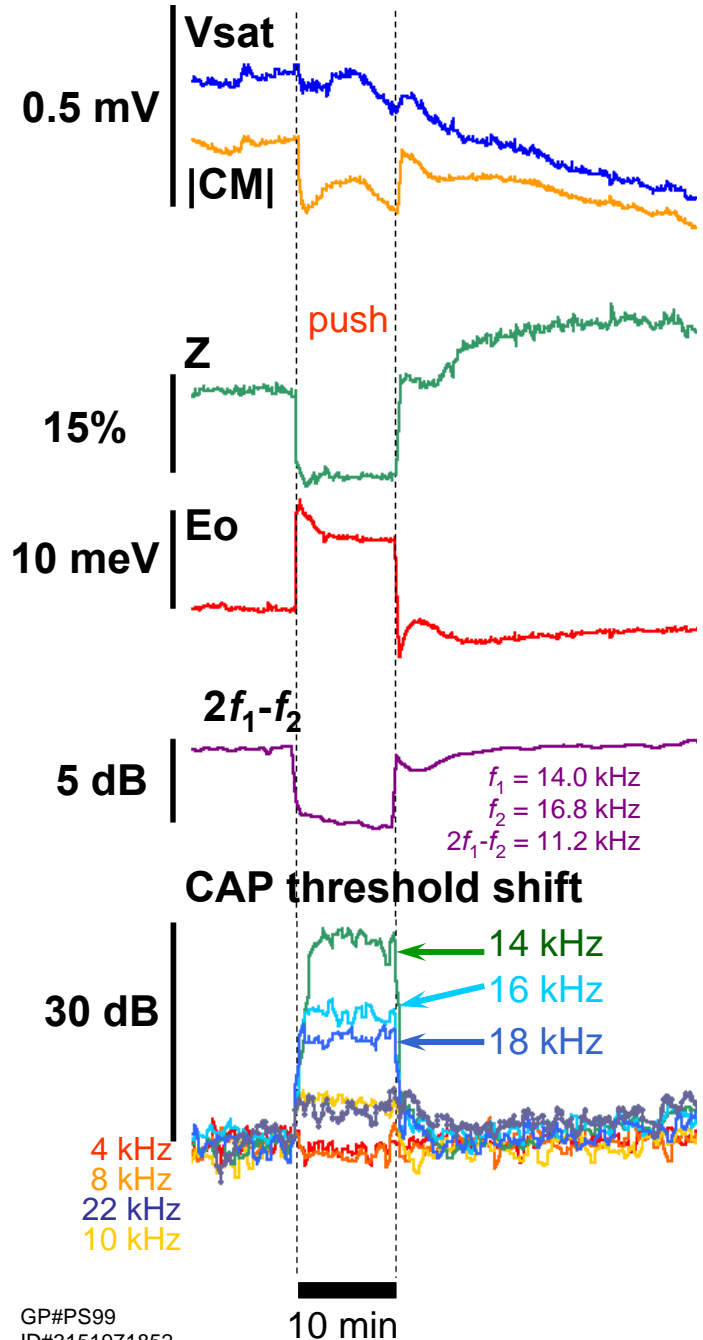




SV ↑
ST ↓

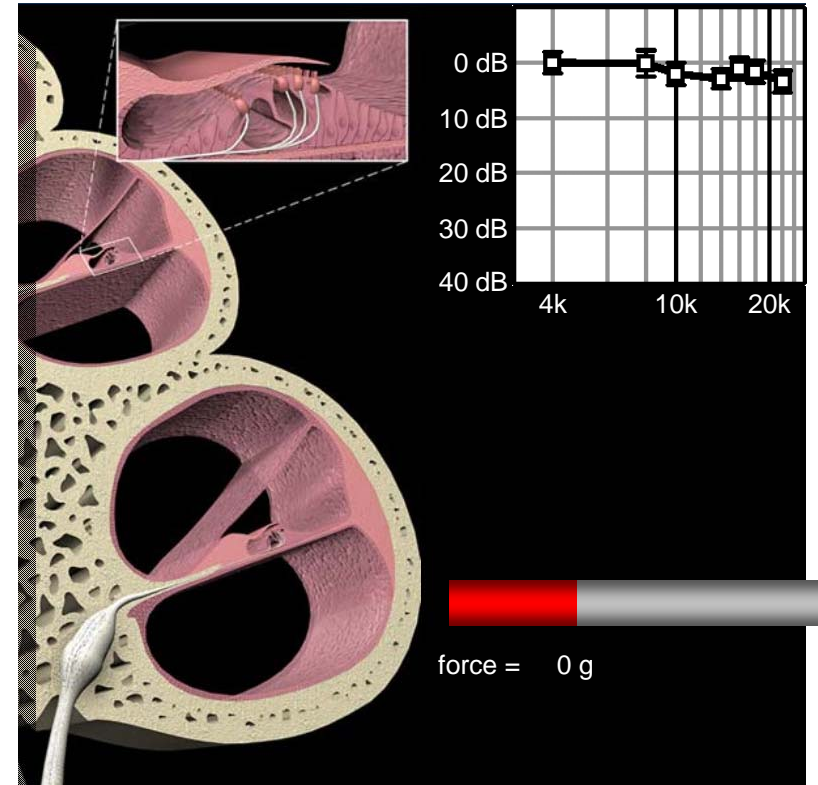
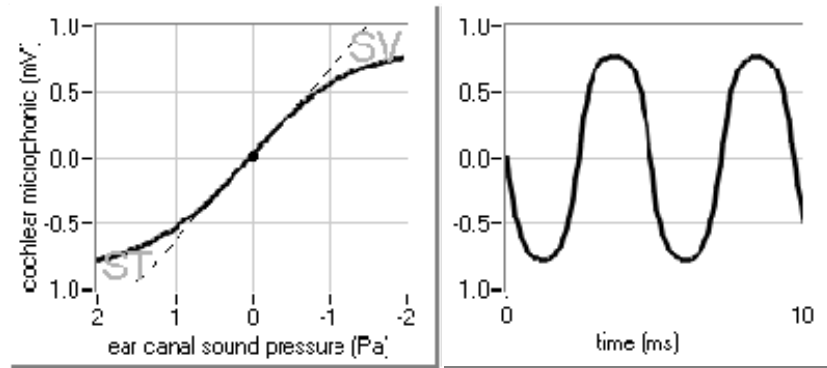
application of force to the cochlear wall

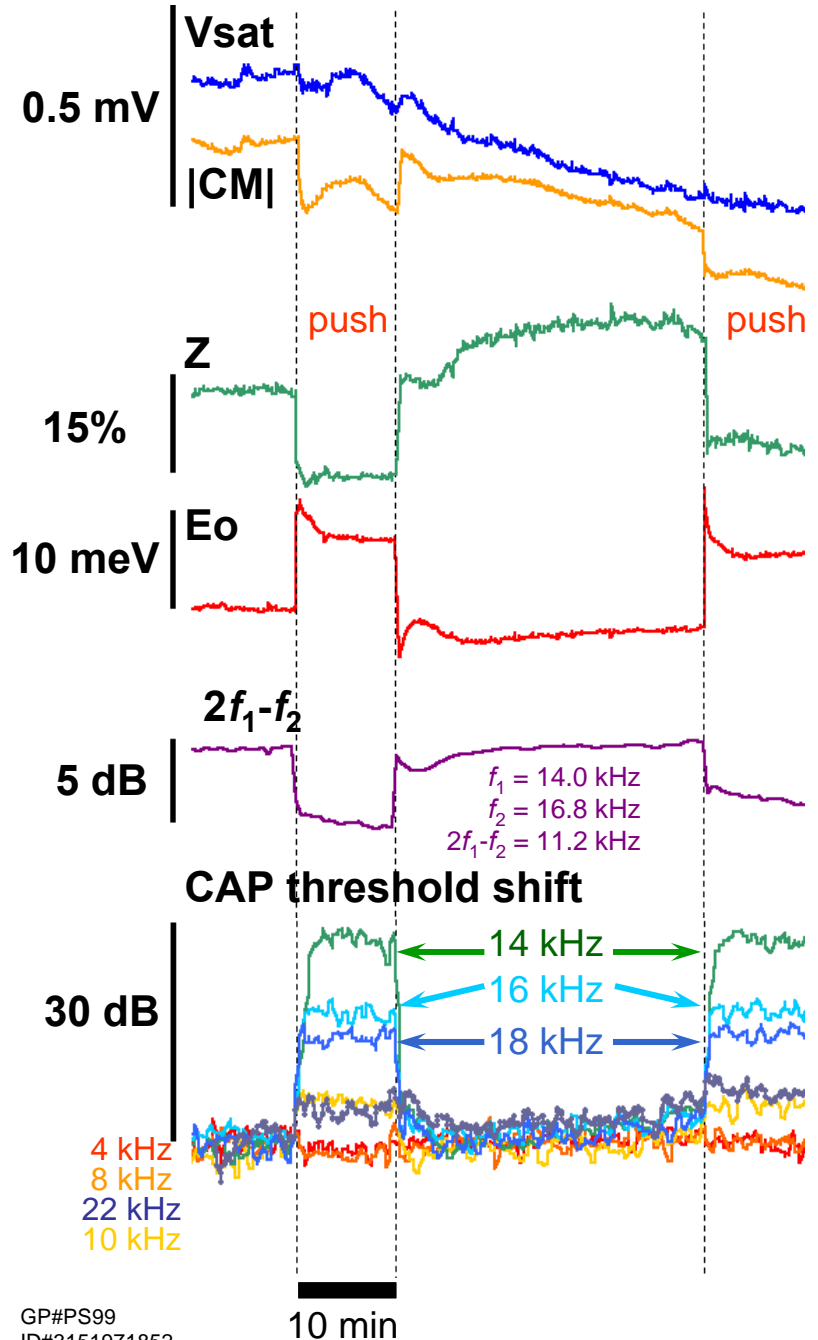




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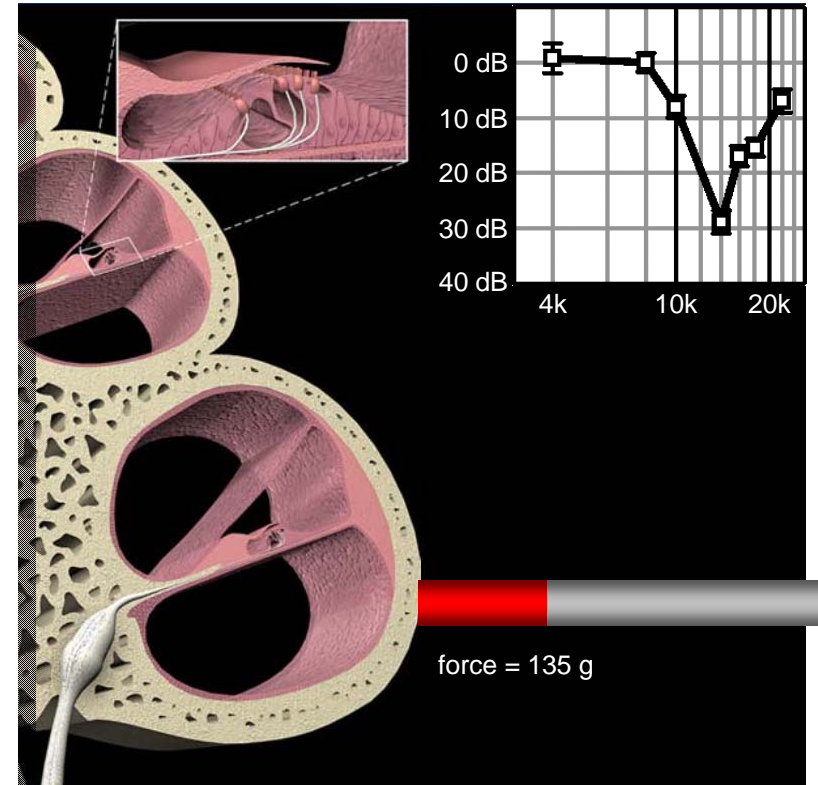
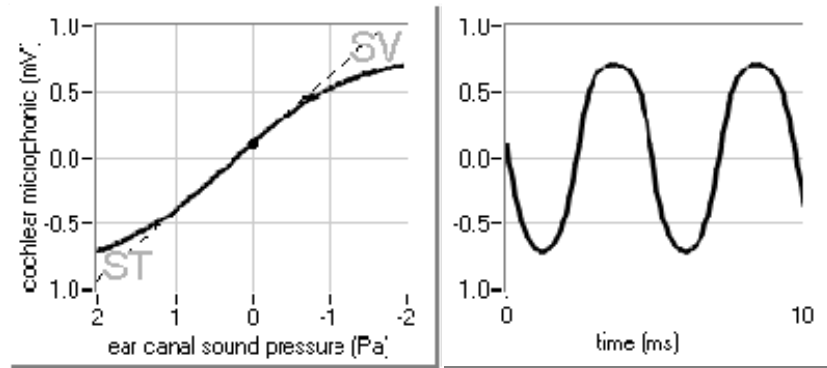
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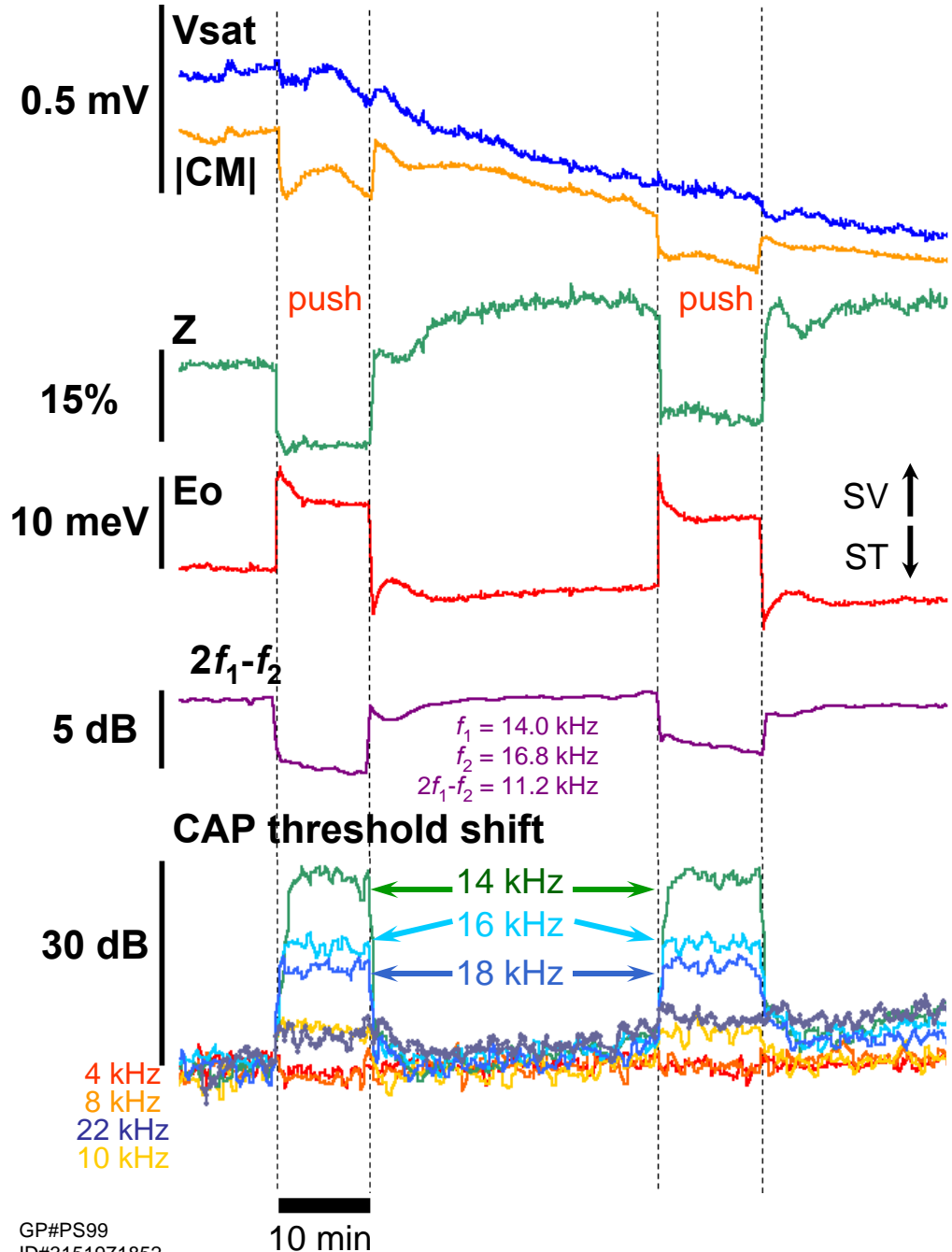




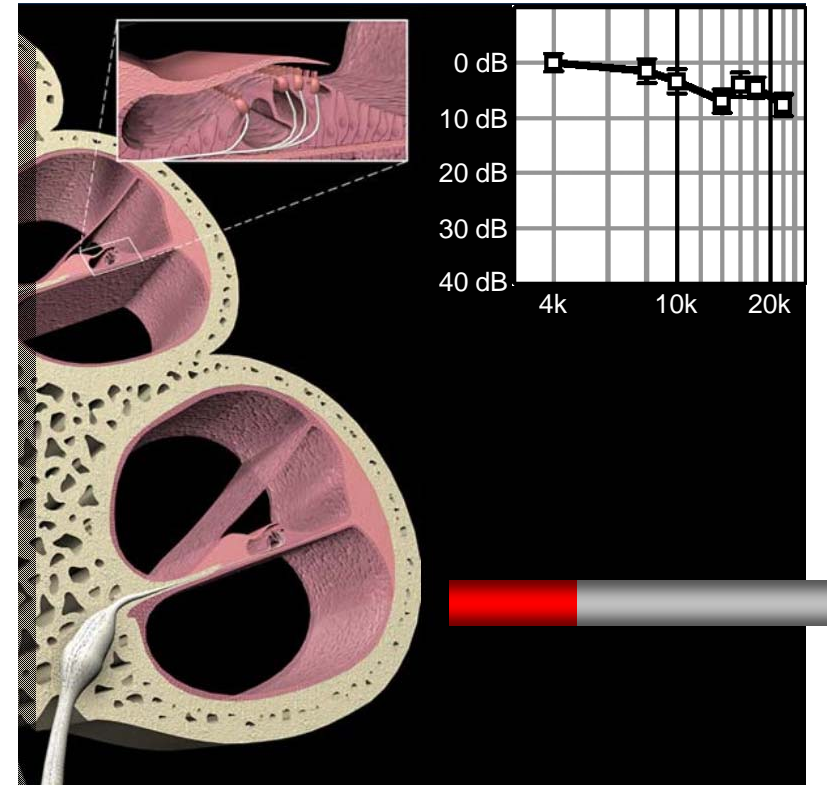
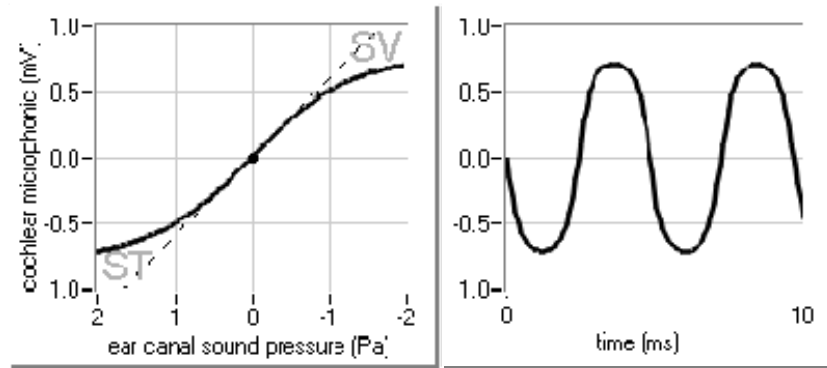
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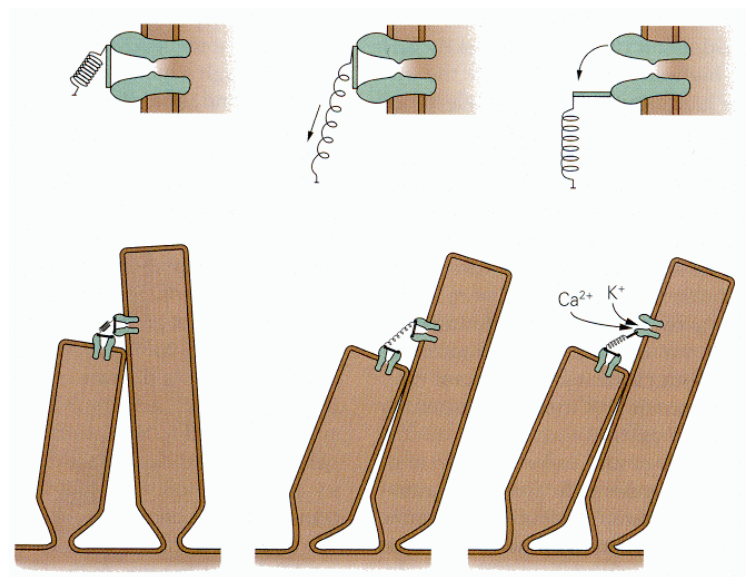
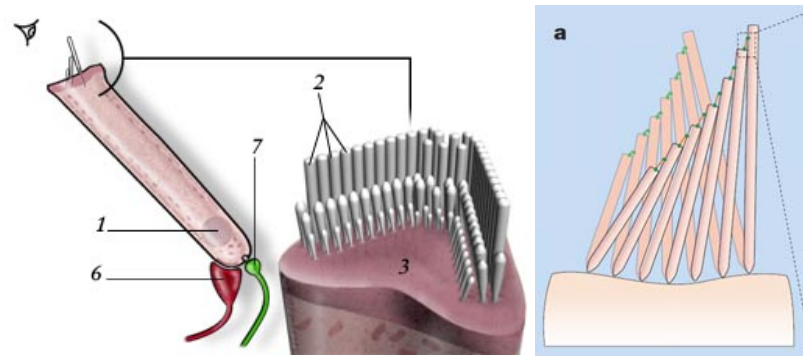
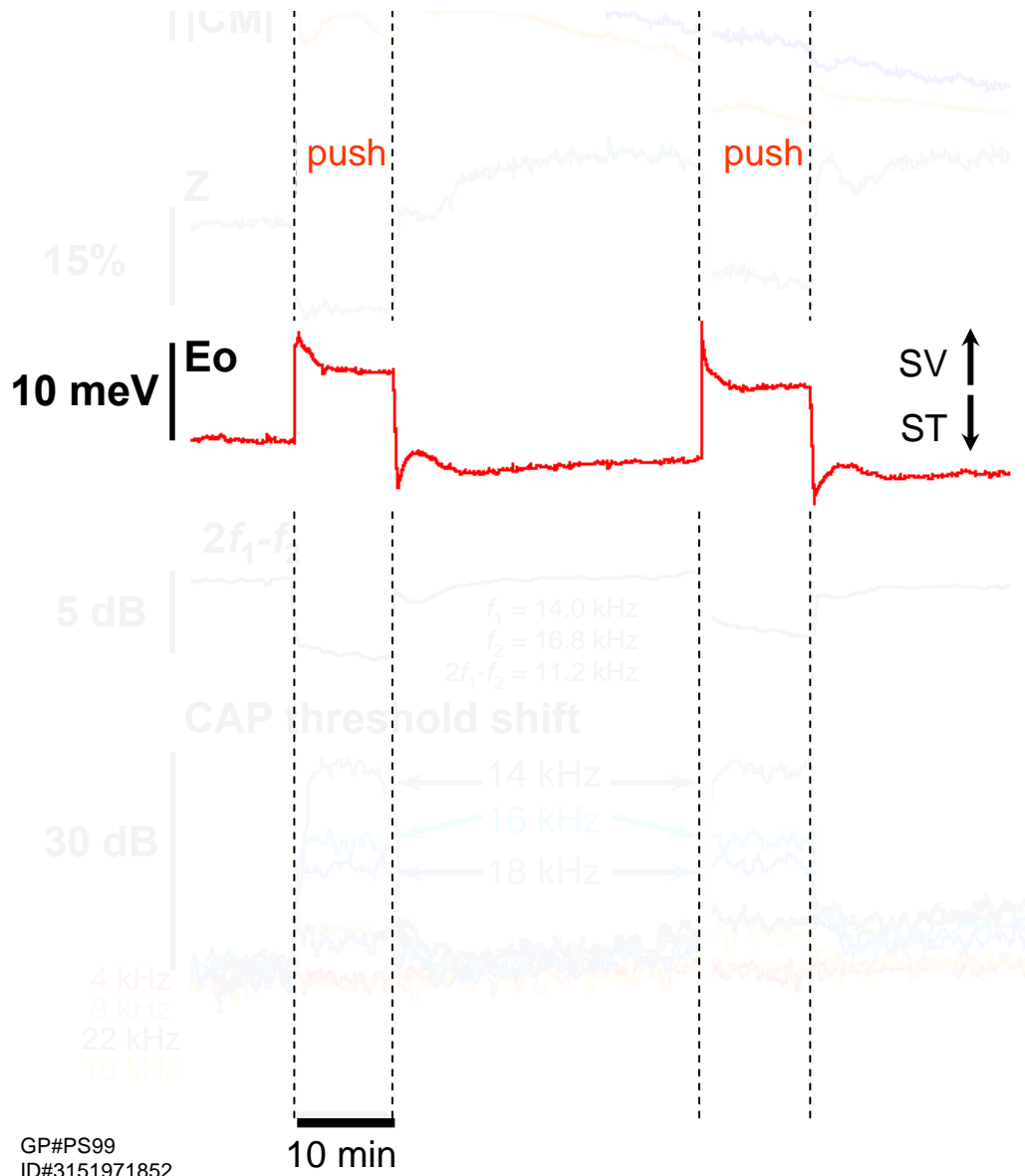


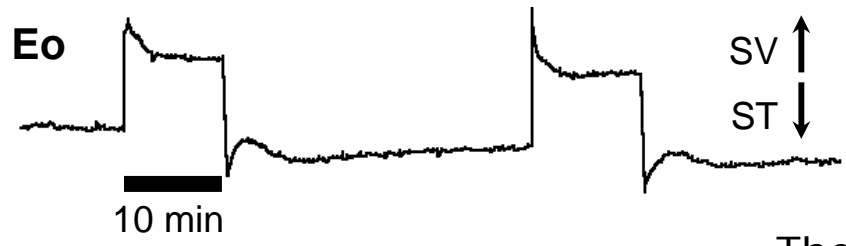


application of force to the cochlear wall



application of force to the cochlear wall



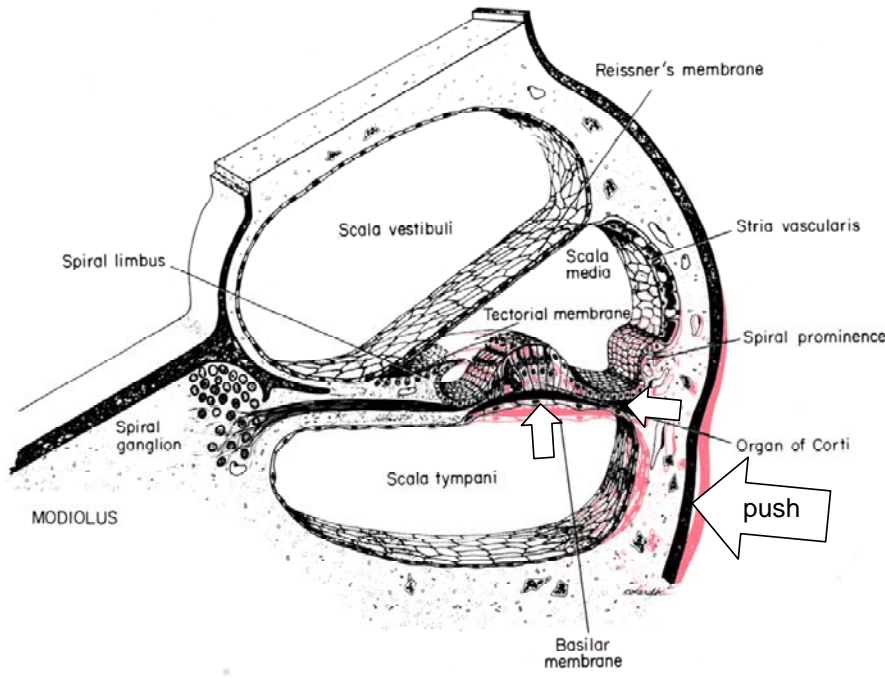
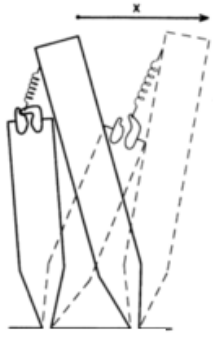
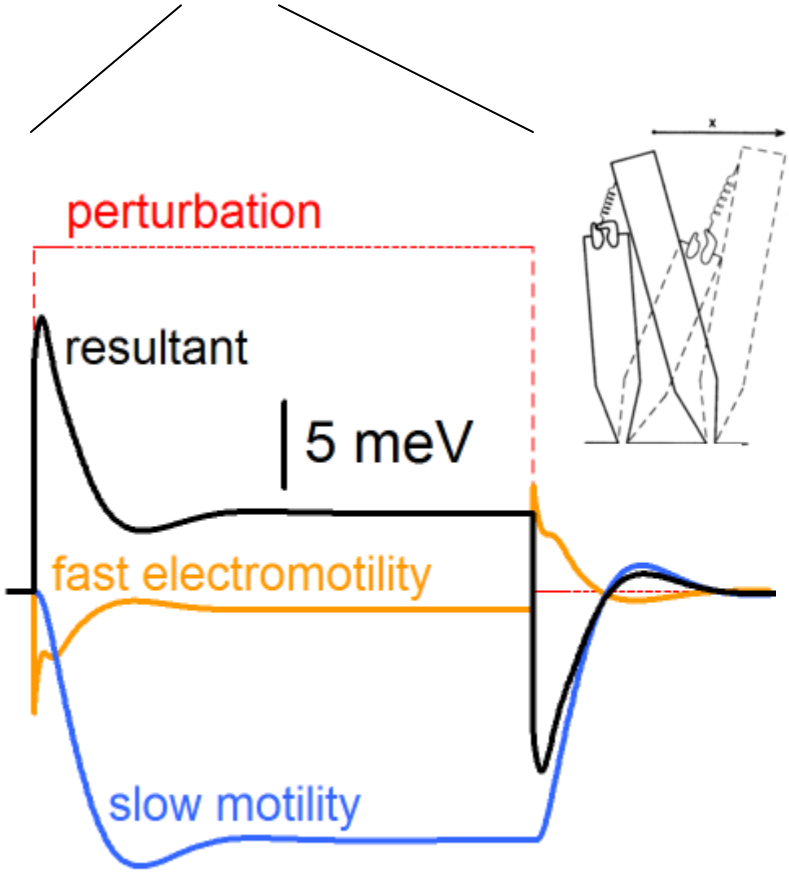


application of force to the cochlear wall

- Results from model -

The resultant operating point is sum of:

- a) Perturbation (SV direction)
 - i) Movement of the basilar membrane towards SV
 - ii) Deflection of stereocilia
 - iii) Opening of MET channels



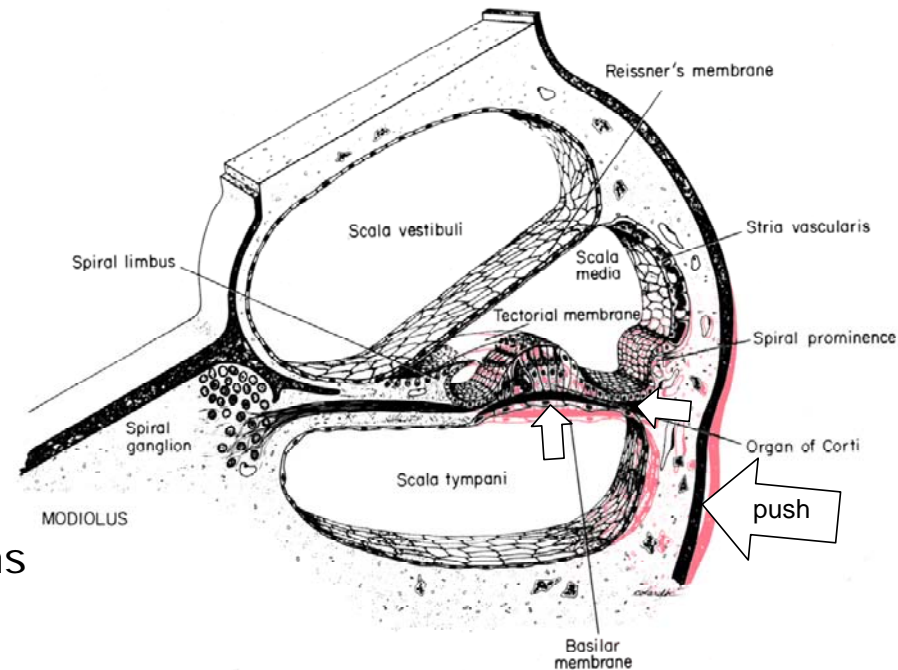
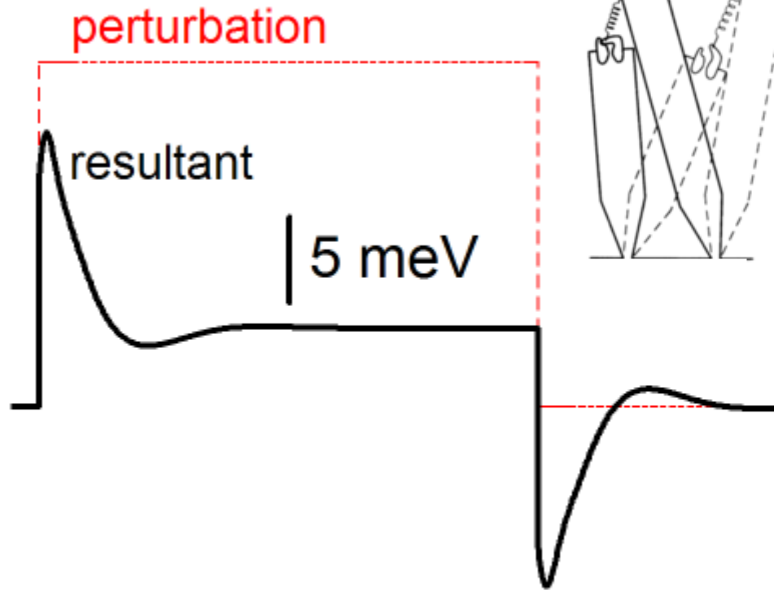


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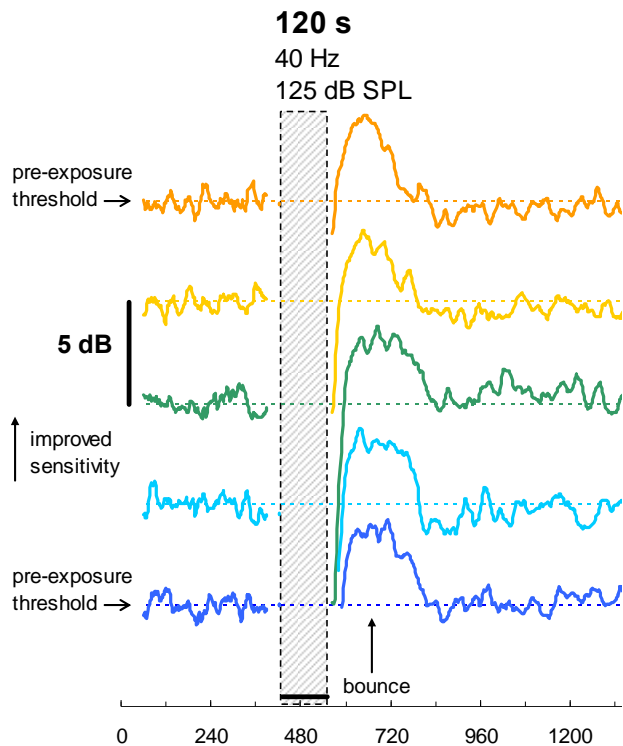
- Calcium mechanism dominates.
- OHC homeostatic regulation mechanisms are able to compensate for much of the perturbation, but not all of it.

Finally...

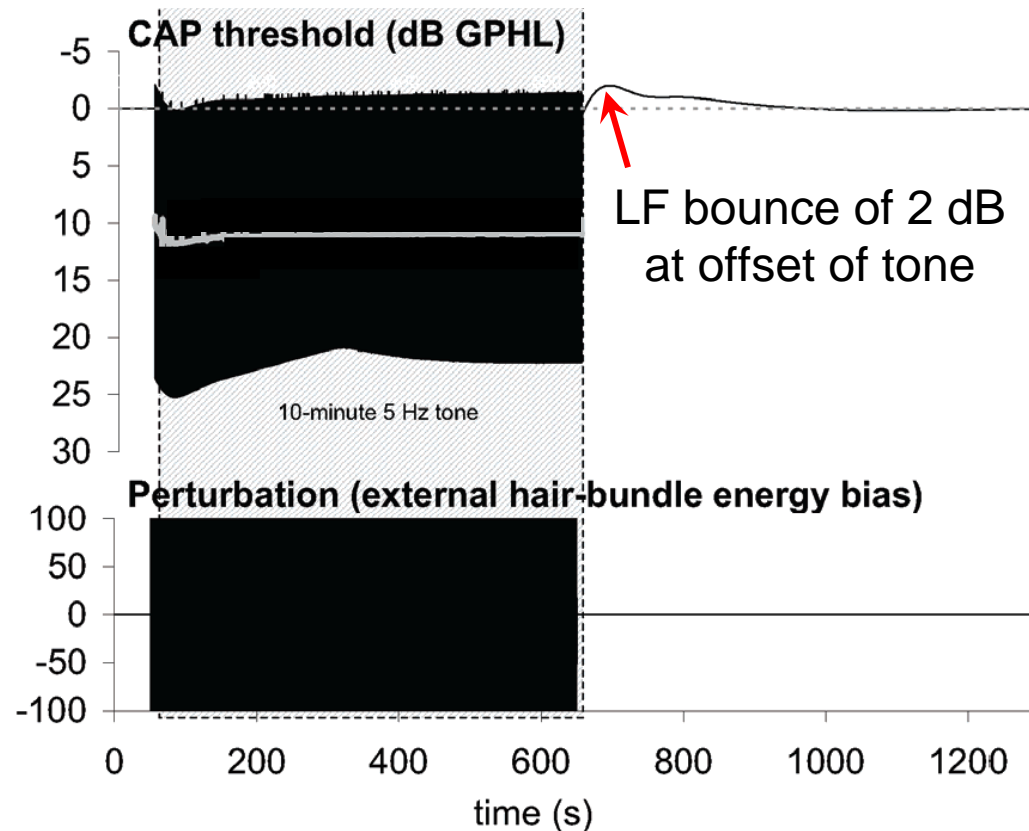
- What is the underlying mechanism for the bounce?

Experimental data:

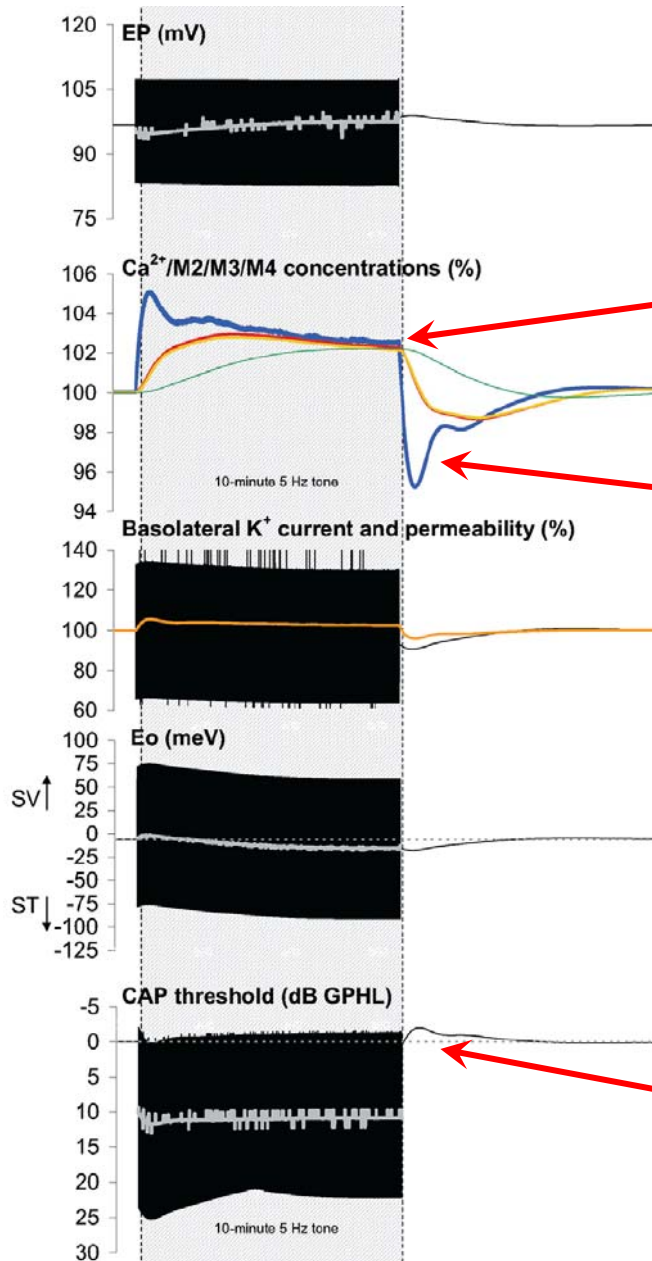
(O'Beirne & Patuzzi, submitted)



Model data:



Modelling of exposure to LF tones (the bounce)



Presence of LF tone causes cytosolic Ca²⁺ to rise, which causes a slow increase in messengers which accelerate Ca²⁺ removal

Ca²⁺ entry stops but its removal continues, causing Ca²⁺ levels to fall to 4.5% below control levels

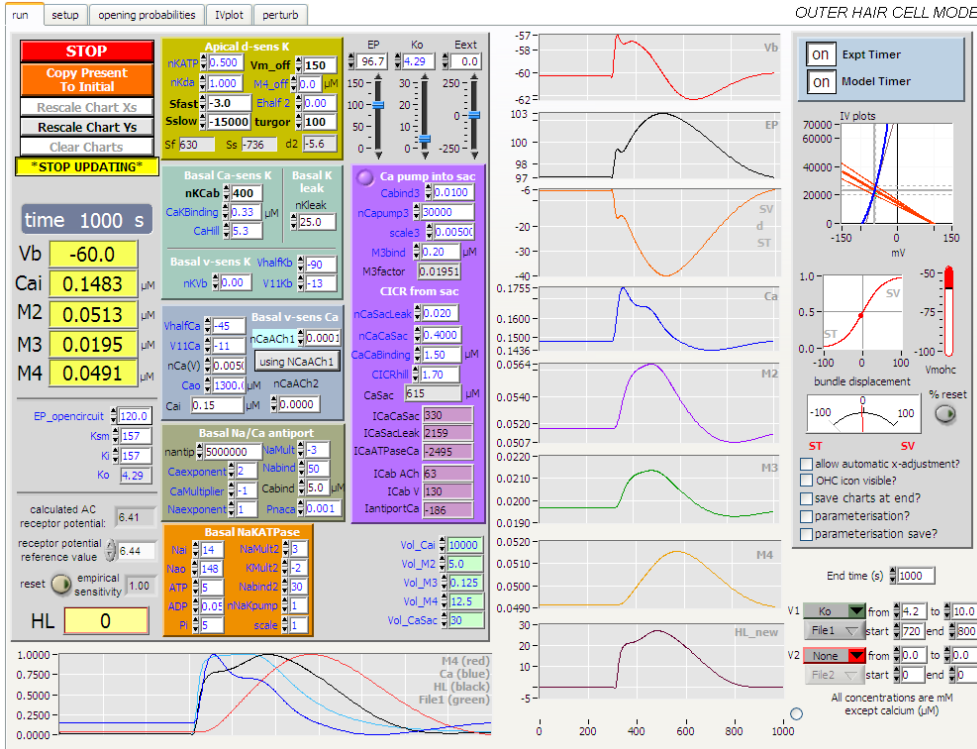
The reduced levels of calcium causes Ca²⁺-sensitive channels (SK type) in the OHC membrane to close, increasing basolateral resistance, and increasing the receptor potential that drives the active process ($V=IR$)

Increased sensitivity at offset of tone as Ca²⁺ falls below resting levels

Summary

- Known outer hair cell mechanisms are capable of regulating both membrane potential and MET operating point with great accuracy.
- First model to incorporate both the ion transport and mechanical properties of OHCs into a single framework
- Valuable step in understanding how perturbations of the ionic and electrical environment of the cochlea produce hearing loss and tinnitus.
- “Standard set” of model parameters capable of reproducing a number of key features of experimental data, but several key results the model cannot replicate.
- Modelling continuing using two functional pools of cytosolic Ca^{2+} , which decouples hair bundle angle and basolateral permeability, providing a better fit to these data.

Acknowledgements



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