Marvin R. Lamborg, Chairman Ralph W. F. Hardy, Co-Chairman E. A. Paul

6. Microbial Effects

Panel Members: G. Bethlenfalvey, Peter Dart, C.L. Luh, R.J. Luxmoore, Barbara Mazur, J.H. McBeath, Yaacov Okon, W.H. Patrick, Jr., W.J. Payne, G.A. Peters, T. St. John.

RESEARCH RECOMMENDATIONS AND SUMMARY

The recommendations for microbiological research needs are based on the following general scenario: a) increased atmospheric CO_2 will increase crop productivities by 10-40%, depending on crop and geographic area, which in turn will increase biomass and soil organic matter by 5-40%; b) additional root-derived materials and crop residue in the soil will increase soil microbial activities, producing a greater flux in most major cycles and possibly some changes in pool sizes of -10% to +30%; c) these effects will increase biological N₂ fixation, and the increased demand for N will place significant limitations on phosphorus and other mineral nutrients; d) no significant changes will occur in soil O₂ or CO_2 .

1) Quantification of the key soil components, processes and structure, as a function of increased atmospheric CO₂, is essential. Components and processes to be measured includes mycorrhiza, nitrogen fixation, biomass production, mineralization-immobilization, nitrification, denitrification, organic matter, available phosphorus and other nutrients, soil CO₂ and O₂, and soil structure. These determinations should be carried out in the key agricultural and forest ecosystems of the tropical and temperate zones. Information should also be collected on microbial mobilization of limiting nutrients and on the sensitivity of mobilization to atmospheric CO₂. Organisms of note are those involved in N₂ fixation, nitrification, and denitrification, as well as mycorrhiza. Rhizosphere changes will probably precede those in the root-free zone.

Some of these measurements will of necessity be long term, perhaps 5-10 years.

2) Soil limitations should be defined in terms of N, P, and other nutrients. The N_2 -fixing organisms and mycorrhiza are especially important to study. The latter can scavenge P and perhaps other limiting nutrients in unfertilized areas, and there is insufficient information on them. Development of inoculants composed of nitrogen-fixing organisms, and especially mycorrhiza, are essential.

3) Minimum tillage is suggested as a large-scale operating model resembling the conditions of soils which may occur under elevated CO_2 .

4) On the assumption that additonal P and N will be needed to obtain crop production increases from elevated atmospheric CO_2 , alternative economical sources of N and P should be developed with emphasis on biological systems.

5) Among the questions to be addressed are: Is there a significant increase in the amount of N_2O released to the atmosphere? Are soil-borne plant diseases increased or decreased? What is the effect of possible decreased litter quality?

6) The rapid rate of technological change, especially in molecular genetics, dictates the need to develop scenarios for the impact of increased CO₂ based on the use of futuristic rather than current technologies. The recently established molecular biology and recombinant DNA technologies should be applied to increase the beneficial interaction of soil microbes and plants. In this way, the postulated nitrogen, phosphorus, and nutrient limitations may be reduced, and the requirement for multiple interacting organisms may be eliminated.

Summary

The postulated doubling of atmospheric CO_2 is not likely to have a direct effect on soil microbial activity because during the growing season, the concentration of CO_2 in the soil atmosphere is already ten to fifty times higher than existing atmospheric CO_2 . Based on all available experimental information, it is estimated that a doubling of atmospheric CO_2 will cause an increase in primary productivity of ten to forty percent, depending on locale. The increase in biomass will, in turn, produce a limitation of available soil nutrients, especially nitrogen and phosphorus. Increased organic carbon together with nitrogen and/or phosphorus limitation will result in a preferential increase in nitrogen fixation and mycorrhizal activities as the expedient means for supplying required nutrients to sustain the predicted increase in primary productivity. Therefore, increased emphasis should be placed on fundamental research related to soil microbiology with special reference to nitrogen-fixing, nitrifying and denitrifying bacteria, and to the mycorrhizal fungi.

The biogeochemical cycle of carbon 1. Global Effects. which integrates aerial, soil, and aquatic activities constitutes the basic mechanism for the production of renewable resources such as food, fiber, and fuel, and for the removal of organic detritus through mineralization. Man is increasingly affecting the carbon cycle by burning fossil fuels, by intensifying agriculture and forestry, and by destroying segments of the earth's plant cover. Soil microorganisms are an important determinant in defining the extent to which plants can respond to the increased level of atmospheric CO2. Because the concentration of CO₂ in the soil is already an order of magnitude higher than the atmospheric concentration, soil microbes are unlikely to be affected by the anticipated atmospheric change. They will, however, be affected by the primary plant response, increased CO₂ fixation and increased biomass. Predicting the response of soil microbes to this secondary extent was the charge given to the "Microbial Inputs" group. The major conclusion of the group's discussions is that soil microorganisms will provide the major source of nutrients required by plants in response to the increased atmospheric CO2. The extent of the plant response will be dictated by the soil activities. Increased knowledge of the soil changes may enable beneficial management for improved plant productivity under increased atmospheric CO2. A summary of our knowledge and postulated changes in the various soil activities and recommendations for future research follows.

1.1 Microorganisms and Photosynthesis. At first glance, bacterial and cyanobacterial photosynthesis may not loom as large in cultivated and non-cultivated ecosystems as green plant photosynthesis, but the contributions of microbes to carbon fixation cannot be ignored (Karagouni and Slater, 1978). Microbial CO_2 fixation is reviewed in this chapter (see Section 3). Nitrogen fixation, the second most significant source of energy potential and nutrients for plants (Hardy and Havelka, 1975), is tied both directly and indirectly to photosynthesis. The photosynthetic prokaryotes which form the base of many food chains also

fix nitrogen. In addition, the photosynthate generated by legumes and many other plants contributes the energy and electrons used by the nodule-forming and associative heterotrophic bacteria that fix nitrogen.

Carbon dioxide, a product of both aerobic and anaerobic metabolism, is important not only because it completes the carbon cycle but also because of its direct influence on microbial growth. Chemoautotrophic and photoautotrophic microorganisms must have CO_2 , as it is their sole carbonaceous nutrient. CO_2 is stimulatory to, and often required by, many heterotrophs. Frequently growth will not proceed in its absence. Yet high levels can be toxic.

1.2 CO₂ Concentrations in Soil and Factors Affecting It. The CO₂ concentration in soil is much higher than that in the atmosphere because of the continued generation of CO₂ by biological processes. General soil levels under aerobic conditions usually approximate 3,000 ppm where active microbial or plant growth occurs. Under waterlogged conditions, soil CO₂ levels of 10,000-30,000 ppm are usually considered to be the average. All measurements of CO₂ are made on the air in the macropores between soil aggregates. The actual microsite could be much higher; however, because respiring roots and microorganisms are usually on the outside of aggregates in larger pores, the inside of aggregates probably do not generate large amounts of CO₂.

The movement of CO_2 from reactive sites follows the general laws of gaseous diffusion (Fick's law). The diffusion rates of gases are linearly related to the concentrations in the air, the soil depth, and the diffusion constant in soil. The solubility of gases in the water phase depends on the type of gas, the temperature, the salt concentration, and their partial pressure in the atmosphere. CO_2 has a diffusion constant in air similar to O_2 and N_2 (0.2 cm² sec⁻¹). The relative diffusion constant of all gases in water is lower by a factor of 10^{-4} . Because CO^2 can become ionized, it has a much higher solubility in water than O^2 (0.9 cm /liter vs. 0.031 cm³/liter for oxygen at 10° C and 760 mm of mercury).

As concluded from Fick's law, the diffusion rate across water films will control 0_2 and $C0_2$ content relative to the fluxes associated with 0_2 use and $C0_2$ production in soil. Calculations based on the diffusion coefficients and solubilities of $C0_2$ and 0_2 in water indicate that $C0_2$ should move away from the respiring root 23 times more quickly than 0_2 will move in (Greenwood, 1970). Thus, under normal circumstances 0_2 will become limiting before growth of plant roots and microorganisms is inhibited by an excess of CO₂ (Garrett, 1981). This could change, however, if a proliferation of microbial, fungal, and other plant material raises the below-ground concentration of CO₂ above 100,000 ppm by elevating HCO3⁻ concentrations to the range known to inhibit fungal metabolism (see Section 5.6) (Griffin, 1972).

The soil volume is approximately half solid and half pore space. The water content of the pore space controls the proportion of soil air, with an average medium textured soil having approximately 25% air space (one half of the pores are water filled). The proportion of pore space may increase with increased organic matter (Russell, 1973). Any associated increase in total porosity would be expected to increase the gas diffusion coefficient, favoring rapid gas exchange.

During the growing season, the CO2 concentration in soil is usually much higher than that of the atmosphere because of the generation of CO2 by biological processes. On the average, the CO2 concentration of the soil atmosphere varies from 10 to 50 times higher than the atmospheric CO2. The seasonal change is the most important variable. Temperature has a major influence on plant productivity and litter decomposition rates. Moisture, while having the same effect as temperature, also has a major influence, in that CO2 concentrations can become very high when major pore spaces are filled during or immediately after rainfall or irrigation. Table 1 shows the range of CO₂ concentrations in a North American grassland during the growing season. Diffusion during the winter had equalized soil CO2 levels at various depths. Initiation of plant growth increased CO2 at the surface, with two rainfall events in June and July resulting in CO2 concentrations above 10,000 ppm occurring in the surface layers. As the winter approached, surface layers had less CO2 due to decreased plant growth and microbial activity. The CO2 content at lower depths could have been due to slow diffusion rates or continued microbial activity in soils that were still relatively warm during this time of the vear.

The position of the water table is a major factor controlling CO_2 in the soil profile. In contrast to the data shown in Table 1, the highest CO_2 concentrations in Figure 1 occurred adjacent to the water table. The high gradient between 35 and 60 cm, illustrated in Figure 1 is

Soil depth (cm)	CO_2 ppm (x 10^2)							
	May 24	June 6	July 6	Aug. 5	Sept. 9	0ct. 10		
15	15	109	161	33	10	3		
30	12	41	123	73	9	4		
45	10	14	40	92	6	4		
90	10	13	30	23	14	33		
150	7	18	24	30	38	29		

Table 1. CO_2 concentrations in a native grassland during the growing season (from de Jong, <u>et al.</u>, 1974).

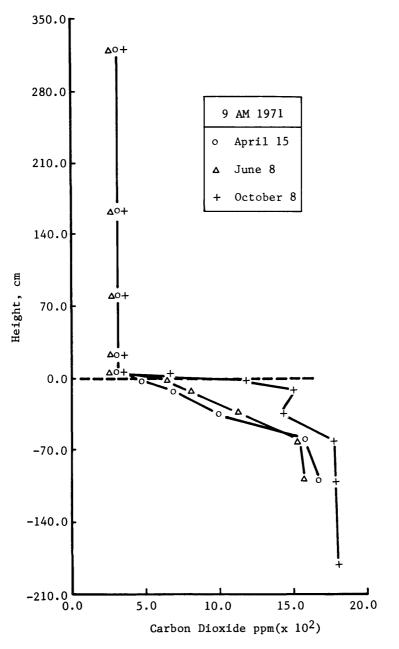


Figure 1. Representative profiles of carbon dioxide concentration above the ground-water table on three selected dates. From Schwartz and Bazzaz (1977).

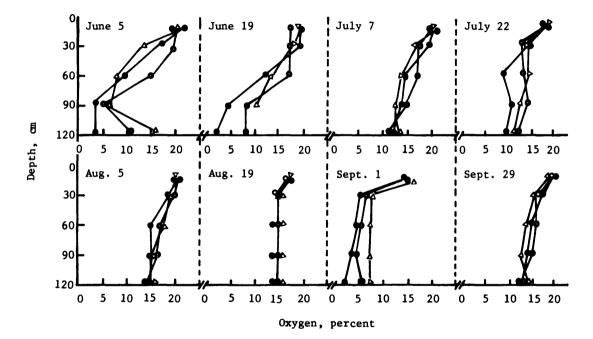


Figure 2. Oxygen content during the growing season in three replicated profiles in soil planted to cotton (Gossypium <u>hirsutum L.</u>). From Patrick (1977).

related to clay content and vegetative growth resulting from the development of root structures.

On a macro basis, 0_2 curves are generally reciprocal to the $C0_2$ curves, justifying the assumption that 0_2 is consumed in the production of $C0_2$. But the agreement is not absolute and the discrepancies are considerably beyond the limit of experimental error. Oxygen content of Louisiana soils were low in the sub-soil of silty clay loam growing cotton early in the spring, but increased as the season progressed. Heavy rains between the 19 August and 1 September samplings interfered with oxygen exchange in soil and caused a temporary decrease in oxygen content (Figure 2).

Roots are the major contributors to total soil respiration, with an average of 20 to 50% of the total soil respiration being attributed to roots and the closely associated microflora (Johnen, 1974; DeJong and Paul, 1979). Similar values are obtained for forests as for agricultural sites (Reichle et al., 1973; Coleman, 1973).

In summary, soil CO₂ under aerobic conditions approximate 3000 ppm where active microbial or plant growth is occurring. Under waterlogged conditions, CO₂ levels of 10,000 to 30,000 ppm are usually considered to be an average that plant roots and microorganisms are exposed to. No significant change in soil microbial activity is likely as a direct result of the projected increase in atmospheric CO2, since soil CO2 is always several orders of magnitude higher than ambient CO2. In wetland soils that are marginally aerated, increased plant residue decomposition resulting from higher plant production may further restrict aeration and cause a shift toward microbial activity which uses electron acceptors other than 0_2 . This should affect sensitive microbial reactions such as denitrification, and possibly result in the production of some inhibitory metabolites as well as decrease the decay of plant residues.

1.3 Effect of Increased Levels of CO_2 on Solution Equilibrium Reactions. The carbonate system is comprised of gaseous CO_2 , dissolved CO_2 , carbonic acid (H₂CO₃), bicarbonate (HCO₃⁻), carbonate (CO₃⁻), and carbonatecontaining materials. The equilibrium concentrations of minerals containing Ca, P, Fe, Mn, Cd, Cu, and Pb are affected by CO_2 partial pressures. The CaSO₄.2H₂O (gypsum)-CaCo₃ (calcite) equilibrium is operative in many dry land soils. The solubility of calcite is decreased as CO_2 pressures increase. At 340 ppm CO₂ in the atmos-

phere, calcium sulfate and calcium carbonate coexist at pH 7.8. Increasing the CO_2 concentration lowers the pH at which the two minerals can coexist. Similarly, the equilibrium between dolomite [MgCa(CO_3)₂] and calcite (CaCO₃) is shifted to dolomite by increasing CO_2 . It has recently been postulated that the formation of calcium carbonates could provide a sink for increased CO_2 because the inorganic carbonates are in equilibrium with atmospheric CO_2 , provided that exchangeable calcium and complexes such as gypsum are available in the soil. In flooded soils, the siderite (FeCO₃)-CO₂ equilibrium has a major buffering effect on soil CO_2 levels.

Because carbonate minerals tend to be relatively insoluble, an increased CO_2 concentration generally leads to decreases in the solute concentrations of cations such as iron and copper, and heavy metals such as lead and cadmium. The inorganic chemistry of phosphate, however, is complex; increased pH and increased CO_2 result in greater solubility of certain phosphate minerals. A 10-fold increase in CO_2 was said by Lindsay (1979) to shift the solubility line of octacalcium phosphate upward by 1.3 log units $H_2PO_4^=$.

Calculations show that CO_2 at 600 ppm in equilibrium with rain water would decrease the pH 0.15 unit. The buffering capacity of soil, and the great range in normal soil and sediment, however, are more important than the effects of increased CO_2 in the atmosphere. It has generally been supposed that roots excrete H⁺ to the soil system and render nutrients available. It is now believed that when plants utilize NO3⁻, the absorption of more anions than cations results in an excretion of HCO3⁻. This yields an increase in pH. Roots growing on NH4⁺ and N₂-fixing organisms should cause a pH drop surrounding the roots. There are, however, other effects, and higher HCO3⁻ concentrations are said to increase root hair density (Nye and Tinker, 1977).

1.4 CO_2 and Soil Organic Matter Levels. Erosion plays a major role in the amount of humus in both cultivated and forested soils. The majority of organic materials are usually deposited near their source. Sediments are slowly decomposed. Addition of nutrients to sediments, rivers and oceans can cause increases in eutrophication and a net sequestering of C. Eroded sites present new landscapes and soil organic matter formation occurs rapidly. This has been shown in the revegetation of mine spoils, where the general rise in soil organic matter averages approximately 400 kg/ha/yr over the first 50 years of revegetation. Climatic alterations due to a "greenhouse effect" could have major influences on soil organic matter levels. Peat lands, taiga forests and sub-alpine tundra exposed to warmer temperatures would have both greater productivity and greater diversity. The amount of potential agricultural land that could be exploited in the northern hemisphere is limited by terrain and by endogenous nutrients. It is known, however, that transformation of northern forested land to agricultural land, and formation of sod Podzolic soils will increase soil organic matter levels.

Soil organic matter comprises approximately 1700 x 10^{15} g of C on a global basis. This is three times the level found in the atmosphere or in the land biota. Movements of C in and out of this source, therefore, can have major impacts on the global CO_2 levels (Bolin et al., 1979). Soil organic matter also plays a major role in soil properties and nutrient dynamics and will be vital to the future response of plants and microorganisms to increased CO_2 levels.

Predictive models of man's influence on soil organic matter (Jenkinson and Rayner, 1977; van Veen and Paul, 1981) have been developed which mimic the effect of past management practices and predict possible future levels. This has been possible since soil humus is composed of a number of constituents which are acted on by a large but generally inactive microbial population. The general concept that soils of the tropics have lower humus levels than those of temperate areas has been disproved (Sanchez et al., 1982a,b). Chemical analyses have shown that all soils have similar components and carbon dating has demonstrated that half of the soil humus is comprised of recalcitrant fractions with mean turnover times of at least 1000 years (van Veen and Paul, 1981).

In aerobic environments, the level of plant C addition has a limited effect on soil humus contents at low plant C/N ratios. Organic matter is stabilized by the reaction of phenol and lignin with inorganic and organic N. The content of these aromatic fractions, their reaction with inorganic particles, and their aggregation leads to long-term stabilization. Green manuring with N-rich legume can have major impacts on the N economy of soil and can relieve much of the fossil fuel requirement for the production of nitrogen fertilizer. Green manures will not, however, lead to major increases in soil organic matter levels unless other management techniques are also incorporated.

	Upland Temperate Zone			Upland Tropics				Wetlands			
	Agri. Total	Culti- vated	Forest	Grass- land	Agri. Legume	C ₄ Plants	Grass- land	Forest	Paddy	Other*	Tundra
I. Plant matter	1.3	1.25	1.2	1.35	1.4	1.2	1.3	1.10	1.4	1.3	1.3
Soil organic matter	1.1	1.05	1.2	1.2	1.1	1.05	1.15	1.05	1.1	1.4	1.4
Soil nutrients remain	ing										
N	1.1	1.05	1.1	1.2	1.1	1.05	1.15	1.05	1.1	1.2	1.3
P	0.95		.95	.95			1.0	0.9	0.95	1.1	0.9
Micronutrients			. 95	.95			1.0	0.9	0.95	1.0	0.9
Microorganisms		1.05	1.5	1.5	1.1	1.05	1.1	1.05	1.1	1.5	1.00
II. Present soil-C (10 ¹⁵⁸)	73		210	147			197	90		313-30	0 30
Estimated new levels (10 ¹⁵ 8)	80		252	176			226	94		?	42

Table 2. Effect of Gradual Doubling of Atmospheric CO2 on Plant Residues, Soil Organic Matter, and Soil Biomass

Area and soil C levels from Bolin et al., 1979.

Values in Part T are ratios of predicted versus present values.

*Includes swamps, marshes and peat bogs.

Crop breeding has been selecting for plants with low C/N ratios, low root production, and low phenolic contents. However, in contrast to maize, cotton which has a high C/N ratio, can yield increased humus levels when grown over a 20-30 year period (Lal and Kang, 1982). The organic matter model of van Veen and Paul (1981) demonstrated that straw incorporation over a 50-year period could result in a stabilization of soil humus levels that were 5-10% higher than those where straw had not been incorporated.

2. Estimates of the Effect of Doubling Atmospheric CO2 on Soil Organic Matter and Biomass. The carbonaceous substrates drive soil microbiological processes ranging from mineralization-immobilization, through nitrogen fixation, to denitrification. An understanding of these effects has to be based on estimates of the changes in available carbon attributable to plant residue additions, to exudates and other root derivatives. Table 2 shows the estimates of some of these variables for a number of representative plant community types. Those estimates for productivity are generally lower than values given by Rogers et al. (1980), Idso (1981), and Idso and Kimball (1982). Idso and Kimball (1982) have recently reviewed the literature on the effects of CO₂ on the yield of crops. Of the 355 separate samples exposed to CO₂ for extended periods, the enrichment increased yields up to 28%. Goudriaan and Atjay (1979) concluded that, in the field, plant production is usually limited by shortages of water and nutrients. Mulching with plant residues has increased both plant yields (Goswami and Suri, 1982) and nitrogen fixation in soybeans (Shivashankar et al., 1976), although the extent to which this is due to CO₂ or to indirect effects on soil temperature and water evaporation is not known. Water and temperature were not considered as limiting factors in calculating productivity in Table 2. This can in part be rationalized by the fact that new moisture and temperature regimes will merely represent a displacement of these zones from their present locations. To simplify comparisons, values are given as a ratio of present levels. Thus, a value of 1.25 for plant matter represents a 25% increase in plant productivity from present 1982 levels.

This possible displacement of agricultural crops or ecosystems to different parts of the world would require slow adjustments in associated soil organic matter levels. The 100 year time frame involved probably means that this is possible for agricultural crops. However, forested areas and wetlands such as peats cannot as easily be transposed to other portions of the globe without producing major dislocations on the source-sink relationships. The

possible utilization of peat lands for agricultural crops after attendant drainage, would result in significant increases in atmospheric CO_2 and major mineralization of sulfur, nitrogen and phosphorus.

Soil organic matter levels are estimated to respond only slightly to increased productivity. This is in agreement with the results of long term nutrient application trials and in model predictions such as those of van Veen and Paul (1981). It also takes into account that green manures raise soil fertility, but not long term soil organic matter levels.

Phosphorus and nitrogen availability will be major limiting factors for both crop production and soil organic matter stabilization. The increases in productivity shown in Table 2 would require a major input of fertilizers for agricultural lands. In natural ecosystems where fertilization is not feasible, limitations of N, P and trace elements will limit plant adaptation to increased CO_2 .

The good response of legumes to CO_2 was considered in the estimates for cultivated and tropical dryland and for legume and grassland systems. Rice is also known to respond well. Temperate cultivation includes a mixture of legumes and less responsive C₄ crops which should have an intermediate increase in plant productivity. Tropical C₄ crops are predicted to respond poorly, since some of these grasses have low nutritive value and grow in soils which are both deficient in P and contain toxic levels of Al.

Tropical and temperate forests contain a large initial biomass of nonphotosynthetic tissue so that only a modest increase in biomass is predicted as the result of increased atmospheric CO₂. These areas sequester nutrients in biomass. Therefore, increased growth will result in major nutrient stress causing increased C:N ratios of the substrate and lowering of decomposition rates due to changes in plant type and composition. For example, beech leaves (Fagus grundifolia) with a lignin content of 24%, have a half-life in litter of nine years, while the halflife for ash leaves (Fraxinus americana) which have 12% lignin, is 1.4 years (Mellilo et al., 1982). Many temperate forests have a small proportion of nutrients tied up in the phytomass.

Kimmins et al. (1979) and Swank and Waide (1980) summarized studies of thirteen forest sites. The data show that 10.3% (std. dev. 2.9%) of the nitrogen, and for

seven sites 21.0% (std. dev. 19.2%) of the phosphorus was associated with the vegetation (roots included). The total system nutrient levels included the soil chemical content to a depth of 1 m or less. Sufficient nutrient capital exists within many temperate forests for a growth response to elevated CO₂ if mechanisms for increasing nutrient availability exist. One possible mechanism of increasing phosphorus supply to trees may result from increased oxalate excretion by mycorrhizal symbionts. Graustein et al. (1977) have demonstrated oxalate release by mycorrhizae, and have outlined feasible reactions by which oxalate could increase phosphate concentration in soil solution. The possibility of increased phosphorus, nitrogen (Section 5.5), and water supply (Section 5.6.4) led Luxmoore (1981) to hypothesize an increase in phytomass and nutrient retention of forests with an increase in atmospheric CO₂. This contrasts with other assessments (Goudriaan and Atjay, 1979; Kramer, 1981) which suggest that existing water and nutrient limits on growth may continue to limit plant response to increased CO2.

The projected yield increases and organic matter production will require major nutrient transfers from both increased N fixation and from mining of presently insoluble P forms. For example, temperate forests would require a conversion of one-third of their present inorganic P to biomass and soil organic matter for the above production figures to be attained. Mycorrhizal effects and biological N₂ fixation will have to be even more significant than at present; mycorrhizal activity will probably be the major limiting factor in nutrient mobilization.

These data show that microbial soil populations will be exposed to significantly more available substrate. Most of the newly produced organic matter is more easily mineralized than preexisting biomass after the initial immobilization has been overcome. This will result in bursts of microbial activity. Thus, the large initial size of the current soil biomass in all studied ecosystems, together with increased activity, precludes major changes in biomass. Pulses of microbial growth will cause larger short term fluctuations in biomass and nutrient turnover than the steady state levels indicated in Table 2. The estimates for increased soil organic matter levels include changes in management, such as zero tillage. Residue handling problems, toxic intermediates, and the use of more of the plant residue for biofuels could alter these calculations.

The estimated new equilibrium levels for soil organic

matter were obtained by multiplying estimates for present levels of soil carbon by the expected increase reached after the soils have attained new steady state levels. The estimates for carbon in peat and swamplands varies from 300 $x \ 10^{15}$ g (Atjay et al., 1979) to 900 x 10^{15} g (Bolin et al., 1979). This represents the largest carbon reservoir, which has been built up over many years. The method used for calculating new equilibrium levels cannot be applied to these soils. The present productivity of these wetlands is estimated at 4 x 10^{15} g C yr⁻¹ out of a total of 63 x 1 10^{15} g C yr⁻¹. The proportion that is stabilized in the ecosystem is difficult to estimate, but it is much higher than that for upland sites, indicating that these sites will continue to be sinks for fixed C as long as climatic changes do not materially dislocate the lowland areas from present sites.

3. CO₂ Production and Effects

3.1 Roots. Any increased root growth that may occur under elevated atmospheric CO₂ will increase the oxygen demand for root respiration. Root proliferation will be favored in zones with adequate aeration. However, at very high root densities the oxygen supply to roots may limit respiration. Generally, excess CO2 will not limit root respiration as much as 0₂ deficiency. Also, the apparent length of the diffusion path in the liquid phase surrounding roots is more often a factor limiting normal root respiration than is the gaseous composition of soil pores (Grable, 1966). This factor could become more significant at elevated CO_2 if soil water films tend to be thicker as a result of a slower rate of plant water use. In fine textured soils and marginally wet soil areas, root growth may be limited by oxygen supply, and responses to increased atmospheric CO_2 may not be expected in these areas.

<u>3.2</u> Fungi. Changes in the soil atmosphere affect fungal growth. Soil fungi are relatively insensitive to reductions in 0_2 concentrations to about 4% in the gas phase. At lower concentrations of 0_2 , there are some marked differences among species, and nearly all fungal growth rates decline at 1% 0_2 (Griffin, 1972). In the presence of 21% 0_2 , CO₂ has little effect on the growth rate of most fungi until CO₂ exceeds 10,000 ppm (Smith and Griffin, 1971). Within the soil, increased partial pressures of CO₂ are always associated with a reduced partial pressure of 0_2 . Therefore, enhanced fungal growth in gas mixtures containing more CO₂ and less 0_2 than the

atmosphere may not be unusual (Griffin, 1972). However, when the CO_2/O_2 ratio exceeds unity, significant changes in the mycoflora occur (Macauley and Griffin, 1969). A significant alteration of the soil atmosphere due to both increased root respiration and to degradation of more soil organic matter could therefore produce unforeseeable changes in the soil mycoflora.

Vesicular Arbuscular Mycorrhiza (VAM) Fungi. These organisms may represent up to 20% of the dry weight of the fungus-root association in some plants (Bethlenfalvay et al., 1982), and total respiration rates in mycorrhizal roots may be twice those in roots not colonized by VAM fungi. If these high respiration rates were to raise the concentration of bicarbonate ion in the soil solution, it could conceivably inhibit VAM fungal growth. Should this be the case, P limitation might lead to diminished symbiotic N₂ fixation. It could also indirectly diminish associative N₂ fixation by decreasing plant growth and root exudation (see Section 5).

3.3 Return of Crop Residues to Soil. With increased above-ground biomass production levels, there will be a greater return of crop residues to the soil, particularly in minimum tillage situations, where straw is not burned. Straw residues of temperate cereals such as wheat can influence development of subsequent crops adversely if there is insufficient time for the residues to be decomposed by microbes. Early in the breakdown process organic acids can accumulate in cool temperate climates, which can inhibit cereal seed germination (Lynch, 1978). To obviate this effect will require attention to such agronomic details as the time and depth of planting of the subsequent crop.

In tropical environments a product of sorghum residue breakdown can severely inhibit the growth of a subsequent crop. Polyphenolic compounds are implicated, and their toxic effect can be ameliorated by allowing a longer period for microbial breakdown of the residues before planting a subsequent crop. It is possible that inoculation of the soil with bacteria can hasten this process.

In semi-arid tropical environments, the growth of annual crops in soils with low nitrogen content may be enhanced by the addition of relatively small amounts (5 t/ha) of organic matter to the soil in the form of farmyard manure. The reason for the beneficial effect does not appear to be the extra supply of plant nutrients. Such

plant growth responses suggest that processes which increase the rate of return of plant residues to the soil can have beneficial effects on subsequent crop growth.

4. CO₂ Uptake

4.1 General Observations. A relatively comprehensive assessment of the effect of CO₂ concentration factors, though somewhat outdated and preceding recognition of C₄, is found in the chapter from "Photosynthesis, Vol. II, Part 1" by Rabinowitch (1951). Additional good discussions are found in "Prediction and Measurement of Photosynthetic Activity" (Denmead and Lemon, 1970). While no attempt will be made to address this aspect in detail, several points will be made. Given a doubling in atmospheric CO₂, plant biomass can be expected to increase, although for this to occur, more combined nitrogen may be required. It is also possible that more root exudation will occur. This could stimulate microbial growth in the root zone, especially in tropical and subtropical soils where carbon sources are likely to be limiting.

<u>4.2 Microbial Effects</u>. As the principal and ultimate degraders of carbonaceous plant and animal litter, detritus, and ejecta, heterotrophic bacteria and fungi would be affected indirectly by increased CO_2 by having more oxidizable or fermentable substrate to deal with and more CO_2 to assimilate. Further, if the mean temperature rises as a consequence of CO_2 accumulation, rates of heterotrophic respiratory and fermentative activity may be expected to rise significantly. It will be important to know whether successions of microbial species in various habitats change from indirect influences such as greater carbon availability and higher temperatures.

The K_M for cell free fixation of CO_2 by the autotrophic nitrifying bacteria <u>Nitrobacter</u> has been calculated to be 5.9 x 10^{-3} M with respect to bicarbonate ion. At pH 7.6 and 30°, a CO_2 of approximately 1 vol % is required to produce this HCO_3^- concentration (Kiesow et al., 1972). The intercellular CO_2 of <u>Nitrobacter</u> equilibrated with air was found by the above authors to be 1.2 vol %, indicating that the CO_2 within the cells is substantially higher than the partial pressure of CO_2 in the atmosphere. It is generally considered that the requirement for CO_2 of autotrophic bacteria such as nitrifiers is met when biologically produced CO_2 in the soil must diffuse through approximately one inch of soil. Soil CO_2 levels are normally not thought to have a direct effect on nitrification and, subsequently, denitrification.

The increased CO₂ resulting from the microbial degradation of soil organic matter of a rewetted rice soil has been reported to result in a flush of green algae and a depression of the cyanobacteria (Rogers and Kulasooria, 1980). Cyanobacteria, however, have a competitive advantage over green algae at the alkaline pH often found in flooded fields during high photosynthetic periods. The decreased light intensities and N levels associated with the latter periods of rice growth also favor cyanobacteria growth.

Nitrogen-fixing cyanobacteria (blue-green algae) appear to have a ubiquitous distribution on earth. In the free-living state, this distribution seems to be most influenced by the pH of the milieu in which the algae grow. Generally, they are inhibited by acid conditions where the pH is less than pH 5.5-6.0, but particular forms apparently have adapted to more acid conditions. Their considerable growth and nitrogen-fixing activity in estuarine, marine, and inland lake environments is dealt with in Chapter 6. They are also well adapted to growing on the soil surface over a very wide range of environments in forests, on beach and river sands, pastures, and arable fields in both temperate and tropical environments.

Cyanobacteria are often numerous enough to develop a soil crust, usually in association with green algae. The balance between green algae and cyanobacteria in these soil niches is influenced by light intensity with the cyanobacteria favored at the low-light intensity associated with greater plant canopy closure. Cyanobacteria are also favored by lower soil mineral nitrogen levels, and possibly by the CO₂ levels in the atmosphere surrounding the algae. Temperature and soil moisture conditions, and climatic factors indirectly influenced by atmospheric CO₂ concentrations, are also likely to influence the growth of cyanobacteria on the soil surface. Cyanobacteria seem to be adapted to higher soil temperatures and, because they produce resistant spores, to periodic wetting and drying phases. The latter is a situation likely to increase in some parts of the earth as part of the CO_2 -induced climatic changes. Thus cyanobacteria distribution is likely to be only modestly affected by increases in atmospheric CO2, although their occurrence on soils receiving little or no nitrogen fertilizer may increase.

The nitrogen-fixing activity of these cyanobacteria

can have a significant influence on the nitrogen economy of their habitat. In desert and tundra soils they appear to be a major current source of nitrogen input to soil. In agricultural lands, they occur as soil crusts under annual crops in both temperate and tropical regions, and in rice they float freely in the paddy water, as well as encrusting the basal parts of the rice stems. They also occur in pastures receiving little or no N fertilizer, but their extent is very much influenced by the canopy development of the associated plant species. In certain situations, such as tropical sorghum and grass fields (Dart and Wani, 1982), the nitrogen-fixing activity can be high, with estimates of up to 25 Kg N/ha/season. On the Broadbalk long-term wheat experiment started in 1843, this amount of N would be sufficient to provide all the nitrogen input necessary to produce the crop. In tropical soils, the cyanobacterial nitrogen fixation seems to be mainly limited by surface soil moisture, and can be reduced to negligible levels within three days of rain or irrigation because of soil drying.

Cyanobacteria also occur in symbiotic association with higher plants, such as mosses, ferns, cycads, and angiosperms, where their nitrogen fixation contributes significantly to plant growth. One particular association, that between the floating fern <u>Azolla</u>, and the cyanobacterium, <u>Anabaena azollae</u>, has particular significance in wet land rice cultivation, and in fresh water ponds, in both tropical and temperate environments (Moore, 1969; Peters and Calvert, 1982).

Considering surface soil microbes, specifically free-living photosynthetic prokaryotes and lichens, it is necessary to appreciate that CO₂ concentration at the soil-atmosphere interface is influenced to some extent by the biota itself (see Section 1 and 2). It should also be noted that a number of cyanobacteria and unicellular green algae (eukaryotes) which exhibit C3 photosynthesis, exhibit very low CO₂ compensation points and/or no O₂ inhibition of photosynthesis (Bidwell, 1977; Lloyd et al, 1977) because they possess a CO₂ concentrating mechanism when grown under current ambient CO₂ (Berry et al., 1976). This mechanism is not present at elevated pCO2 partial pressures (Kaplin et al, 1982) and it is probable that in organisms possessing it there will be little effect from increasing CO2. Higher CO2 levels may favor those cyanobacteria and other algae which lack such a mechanism. Photoheterotrophic growth and/or heterotrophic growth of such organisms might be stimulated in nature through an increase in soil organic matter.

5. Plant Microbial Associations

5.1 Introduction. Microorganism populations are stimulated many fold in the vicinity of plant roots as a result of the release of root derived substances as exudates and as a result of the decay of root cap and cortical cells. Bacteria live within root tissues and in specialized root structures such as nodules. This microbial population has a direct effect on plant growth through its production of plant growth promoters and inhibitors. through their competition for plant nutrients (immobilization), and through their release of nutrients by breakdown of dead cells (mineralization). Certain bacteria, such as Azospirillum, can enhance nutrient uptake by crop plants by a mechanism that is currently not clear. The activity of rhizosphere microorganisms will be enhanced by the larger photosynthetic activity of plants at higher CO2 which will release a greater supply of microbial substrates through the roots. Current evidence suggests that the microbial population will be qualitatively similar, and that the results of their enhanced activity will be, in general, beneficial for plant growth.

Although more CO₂ is likely to initially increase immobilization of nutrients into microbial biomass, their subsequent turnover through mineralization should favor enhanced nutrient supply under maintenance fertilization regimes. If nutrients are not supplied as fertilizer, then immobilization of nutrients is likely to reduce plant growth, even though plant adaptation features, such as reduced concentration of nutrients per unit of dry matter, will work towards maintaining plant growth rates. Increased plant growth will lead to an increased demand for plant nutrients such as P and S. The reduced levels of combined nitrogen in the rhizosphere will favor nitrogen-fixing bacteria.

Root associated bacteria can also modify the pH of the immediate root environment, thereby affecting the solubility of certain plant nutrients. For pastures in South-East Australia dominated by annual clover plants, one of the consequences of nitrogen fixation by the root nodules has been a gradual lowering of soil pH to values which may limit continued reinfection and nodulation by <u>Rhizobium</u>. With enhanced N₂ fixation rates predicted as atmospheric CO_2 rises, this problem could be exacerbated, requiring new management techniques to maintain the pasture

5.2 Organic Mineralization-Immobilization Reactions. The mineralization of nutrients in the geocycle of elements can

occur by specific enzymes elaborated during periods of nutrient deficiencies. It is known that phosphatases and sulfatases are produced under such conditions. Amidases are known to occur, but it is generally considered that N accumulates in the $\rm NH_4+$ form only when microorganisms mineralize more $\rm NH_4+$ as a byproduct than is required for microbial growth.

The current mineralization-immobilization rates of nutrients such as N, S and P occur at two to three times the net rates measured by crop uptake or nutrient accumulation under fallow conditions. The microbial biomass accounts for 2-5% of the soil carbon and 3-8% of the N of surface soils (Jenkinson and Ladd, 1980). Microbial products with fairly rapid turnover rates comprise an equal fraction. Management of the biomass and their metabolites through residue management and timing of cultivation and of fertilizer application could result in a major decrease in the nitrogen that is lost from most soil systems. Immobilization should occur during periods of maximum crop growth. Even a 10% reduction in the high levels of nitrogen loss, usually associated with crop growth, would have a major impact on fertilizer requirements and on nutrient availability.

Increased plant productivity will increase the amounts of carbon entering the organic pool, first as live biomass and later as soil organic matter. We have estimated earlier the soil organic matter build-up from the input of more live biomass. The large anticipated increase in total biomass will produce a considerably lower increase in organic matter.

The increased mineralization of plant material will increase heterotrophic microbial activity. An increased demand for the mineral nutrients needed for microbial decomposition that are not contained in plant material will further stress the soil resources of nitrogen, phosphorus, potassium and other essential minerals. The increased requirement for nitrogen can be partially met by increased fixation of atmospheric N₂, but the increased requirement for other nutrients will necessarily be met by either soil resources, in an unfertilized ecosystem, or by both soil and fertilizer sources in agricultural systems. Table 2 predicts some depletion in the soil reservoir of these nutrients. The decreased availability of these plant nutrients will exert a limiting effect on plant production.

The higher rate of biomass production should stimulate biological nitrogen fixation, since a widening of the C:N

ratio of the soil stimulates N_2 fixation by both free-living organisms and N_2 -fixing organisms. The widening ratios may also encourage increased migration of legumes into grassland. It is unlikely, however, that enough nitrogen will be fixed to maintain the soil C:N ratio and some slight, but permanent, increase in the soil C:N ratio can be predicted to accompany increasing biomass.

For wetlands and poorly aerated uplands, the higher demand for oxygen resulting from increased plant production will cause increased accumulation of plant material, as peat in the wetlands or as increased soil organic matter in the poorly aerated uplands. This organic matter will have a high C:N ratio because of its limited decomposition. The absolute amounts of carbon, nitrogen, sulfur, and other mineral nutrients tied up in this organic material will significantly increase.

A similar effect will occur in those upland areas where the additional plant biomass is difficult to decompose because of its content of inhibitory compounds such as lignins and tannins. This condition will exist in some forests and will result in a higher organic matter accumulation and a wider C:N ratio.

Incomplete oxidation of plant material, especially under conditions of limited soil aeration, will produce organic acids, such as acetate, propionate and butyrate, which are toxic or inhibitory to some biological processes. It is safe to predict an increased concentration of these compounds in oxygen deficient systems. Under current atmospheric CO_2 , residues of crop plants such as sorghums and millets produce high amounts of such compounds and the crops must therefore be grown in rotation to avoid such inhibitory effects (Dart, personal communication).

Under seriously limited aeration, the mineralization of increased amounts of biomass produces reduced sulfur compounds such as hydrogen sulfide (H_2S), dimethyl sulfide, and methyl mercaptans which are toxic to both microorganisms and higher plants. Ecosystems that are marginally aerated may be adversely affected by an increased oxygen demand.

Less sulfur and phosphorus will be available under conditions of increased biomass production and the consequent increase in the organic matter pool dilutes what may be an already limited supply. Except for the input of additional sulfur from atmospheric fallout and sulfur and phosphorus fertilizers, plants are limited to the existing soil supplies.

5.3 Nitrification. With available moisture and oxygen, and the simultaneously increased availability of CO₂ and NH₃ from mineralization, nitrification by autotrophic bacteria in the surface layer of soil will be stimulated. Nitrite and nitrate, the intermediate and the final products of nitrification, may still serve as plant nutrients, but the nitrogen in these oxidized forms is vulnerable to loss through leaching and/or denitrification by subsurface bacteria. In addition, if oxygen becomes limiting during nitrification so that only the ammoniaoxidizing bacteria function, toxic levels of nitrite may result. Finally, an adventitious result of nitrification is the release of as yet unpredictable amounts of nitric and nitrous oxide (Lipschultz et al., 1981). More NO and N₂O is liberated by denitrifying than by nitrifying organisms. Nitrous oxide moves into the earth's atmosphere and may erode the stratospheric ozone layer. There is currently an equilibrium between loss and replacement of ozone. Increased rate of nitrogen oxide release might decrease the protection afforded by ozone, and increase the penetration of ultraviolet radiation.

5.4 Denitrification. Denitrifying bacteria are widely distributed in the soil. In an anaerobic environment, these organisms convert nitrate and nitrite to NO, N₂O and N₂. Populations are greatest in the zone reaching from just below the surface to a depth of 15 to 30 cm. Penetration of this zone by nitrate and nitrite added as fertilizer or generated at or near the surface, coupled with increased availability of soil organic matter, will result in increased rates of release of N₂ and a loss of a significant plant nutrient. Smaller but significant quantities of NO and N₂O relative to N₂ will also be released (McKenney et al., 1982) and added as the major source to the stratospheric load described in the previous section.

5.5 Nitrogen Uptake. Of the essential elements limiting plant growth, N is frequently in notoriously short supply (Subba Rao, 1977). Thus, if CO_2 availability were removed as a limiting factor, increased reliance on N_2 fixation as a source of N would occur. A stimulation of N_2 fixation as a result of atmospheric CO_2 fertilization has been observed (Havelka and Hardy, 1976). This would shift the floristic composition of the vegetation toward legumes and nonlegumes capable of symbiotic N_2 fixation (Newton et al., 1977). Plants and bacteria mutually benefiting from associative N_2 fixation (Vose and Ruschel, 1981) would also flourish.

5.5.1 Symbiotic N₂ Fixation

5.5.1.1 Effects of Higher Atmospheric CO₂ Levels. The major effects of CO2 enrichment are probably via indirect effects on the symbiotic organisms. Nitrogen-fixing organisms such as Rhizobium and Frankia have major requirements for energy provided as plant photosynthate. They in turn are the major organisms contributing N₂ to agricultural and natural ecosystems, respectively. Highly productive systems cannot be achieved or maintained until a large stock of organic matter is built up during a number of years of N₂ fixation. Early experiments with low light levels and with defoliation indicated that these treatments greatly affected the root-associated symbionts. Quantitative data concerning the effects of CO2 concentrations are limited. Mulder and van Veen (1960) observed an increase in N2 fixation when hydroponic culture solution was aerated with 40 x 10^3 ppm CO₂.

Hardy and Havelka's field experiment (1975) showed that over a nine-week period plants grown with supplemental CO₂ reduced five times as much C_2H_2 and had a greater mass of root nodules than untreated controls. Senescence of the N₂-enriched plants had significantly more nodules than untreated controls. Senescence of the N2-fixing system was delayed. The work by Phillips et al. (1976) indicated that CO₂-enriched plants had significantly more nodules than the controls. However, CO2 enrichment had no effect on the number of nodules which developed per unit dry weight of a pea plant and a linear regression existed between the number of root nodules and the dry weight of pea plants. They concluded that CO₂ enrichment resulted in an integrated growth of the entire plant with no special promotion of nodule growth, while Hardy and Havelka (1975) and Rogers et al. (1980) showed preferential fixation of N_2 vs. use of fixed N by CO₂ enrichment.

5.5.1.2 Partitioning of Photosynthates. Nitrogen fixation is a highly energy-intensive process and represents a significant sink for photosynthate (Phillips, 1980). Minchin and Pate (1973), in their classic study of C partitioning in nodulated legumes, found that approximately 10% of the fresh assimilate is used as a substrate for nitrogenase. This figure was later confirmed by Kucey and Paul (1982) in mycorrhizal hosts. Increased N input into the symbiotic association by the microsymbiont in turn stimulates plant growth and photosynthesis. The resulting

autocatalytic cycle (Bethlenfalvay et al., 1978) may finally be limited by other factors.

5.5.1.3 Productivity. Two major effects on root and soil processes will be produced by increased plant productivity. Water and minerals will become even more limiting to plants. This is especially true for nitrogen, because it is frequently the most limiting nutrient, and nitrogen from the mineralization of soil organic nitrogen will not increase as rapidly as demand for crop production. Thus, the need for fertilizer will increase more rapidly than increases in productivity. This will also be expected for nitrogen-fixing crops such as soybeans.

Thus, nitrogen fixation may increase faster than productivity, due to the limited ability of the soil to supply additional nitrogen. Associative and free-living nitrogen fixers may be stimulated in the same way, but the effect may be less since they are more dependent on soil nitrogen concentration rather than plant demand. Mulching, and increased soil organic matter content, expected to occur under minimum tillage practices, may also have a significant effect on N_2 fixation.

5.5.1.4 CO₂ Concentration in the Rhizosphere. The effects of below-ground CO₂ may be different for host and endophyte. Low concentration of soil CO₂ may be necessary for optimal root growth, and higher concentrations can be inhibitory, with peas being inhibited at CO₂ concentrations as low as 15,000 ppm. At 65,000 ppm of CO₂, beans and sunflowers are strongly inhibited, while barley and oats are not (Stolwijk and Thimann, 1957). Under somewhat different experimental conditions, 80,000 ppm CO₂ was moderately inhibitory to both peas and barley (Geisler, 1967).

Removal of CO_2 from air is inhibitory to both the nodulation of legumes and the growth of rhizobia in culture (Mulder and van Veen, 1960; Lowe and Evans, 1962). The effect of elevated CO_2 concentration is less clear. It has been reported that nitrogenase activity, measured by acetylene reduction, is not measurably different at 0 and 30,000 ppm CO_2 in pea and soybean nodules (Mahon, 1979; Coker and Schubert, 1981). But in alders, 30,000 ppm CO_2 inhibits nitrogenase activity 20% (Winship and Tjepkema, 1982), while no consistent effect is found in soybean nodules (Coker and Schubert, 1981). In peas, the respiration of nodulated root systems is substantially inhibited by 30,000 ppm CO_2 , but this is apparently due to inhibition of root respiration rather than to nodule respiration (Mahon, 1979).

Increased CO_2 could lead to more productive rhizobia/ legume associations. In legumes, high concentrations of CO_2 can increase root and nodule growth due to PEP-carboxylase catalyzed carbon fixation. The fixed carbon serves as a carbon skeleton for nitrogen fixation, and as an energyproducing metabolic intermediate. CO_2 at sufficiently high concentrations can be toxic to plant roots, but good growth occurs even at 240,000 ppm CO_2 , and thus increased atmospheric CO_2 should not be a significant factor.

5.5.2 Nodulated Non-Legumes. Frankia, an actinomycete, nodulates a relatively diverse group of wood dicotyledonous plants including Alnus, Myrica gale, Shepherdia and Casuarina. These N-fixing symbiotic associations are a significant source of combined N in various ecosystems. Since the plants are C₃, it is anticipated that an increase in atmospheric CO₂ will increase rates of photosynthesis and photosynthate for the nodule endophyte. This may enhance total N₂ fixation and N input from these associations.

<u>5.5.3 Nitrogen-fixing associations</u>. Nitrogen-fixing bacteria occur in the rhizosphere of many plants, particularly in the tropics (Dart and Day, 1975; Neyra and Dobereiner, 1977). Nitrogen balance studies for soil-grown plants in pots containing measured amounts of combined nitrogen, show that such bacteria may supply nitrogen for plant growth. Studies with $^{15}N_2$ for sorghum, sugar cane, <u>Paspalum notatum</u>, <u>Digitaria decumbans and Setaria</u> <u>italica confirmed that biologically fixed nitrogen was</u> transferred to the plant tops (DePolli et al., unpublished; Wani, Day and Dart, unpublished).

More than 50 plant species from both temperate and tropical environments, including agriculturally important crops such as sorghum, pearl millet, setaria, maize, rice, and wheat are known to stimulate nitrogenase activity in their rhizospheres as measured by acetylene reduction assays. For rice, sorghum, millet, setaria and <u>Paspalum</u> <u>notatum</u>, there appears to be genotypic differences in activity between lines (Dart and Wani, 1982).

The Broadbalk wilderness experiment at Rothamsted, started in 1888, suggests that nitrogen fixation associated with the roots of several weed species resulted in the build up of soil organic matter, calculated to be 40 kg N/ha/yr from fixation (Jenkinson and Rayner, 1977).

Other N balance field experiments in Brazil and India suggest that biological nitrogen fixation contributes significantly to the plant nitrogen economy. In India a cross between <u>Pennisetum purpureum</u> and <u>P. americanum</u> produced 136 tonnes/ha above ground dry matter in a low fertility alfisol soil. The crop removed 1185 kg N/ha in 30 months, without addition of N fertilizer.

This nitrogen-fixing activity also plays an important part in the nitrogen economy of estuarine communities and for <u>Spartina alterniflora</u> in salt marshes, appears to provide most of their nitrogen requirement (Buresh et al., 1980). Nitrogen-fixing activity is generally stimulated by high photosynthetic activity and by high soil moisture (Dart and Wani, 1982). Both of these are likely to be enhanced by higher atmospheric CO_2 .

If the C to N ratio of the extra plant tissue derived from increased photosynthetic activity does not change, then this will create a greater demand for nitrogen, which must be supplied either by fertilizer or by increased nitrogen fixation.

5.5.3.1 Inoculation responses. Many different types of nitrogen-fixing bacteria have been isolated from the rhizosphere. Some, such as Azotobacter and Azospirillum, can also influence plant growth by other means (Okon, 1982), perhaps by production of plant growth promoters and by enrichment of mineral uptake. It is, however, difficult to quantify the contribution of each process following inoculation of plants with N₂-fixing bacteria.

Inoculation with N_2 -fixing bacteria has increased the yield of nitrogen and dry matter of a variety of cereal crops in both pot and field experiements in Israel, India, and Brazil (Okon, 1982; Dart and Wani, 1982; Boddey and Dobereiner, 1982). Plants inoculated with Azospirillum form associations with roots of cereals and forage grasses which significantly increase mineral uptake (N, P and K) by the roots and also contribute fixed nitrogen to the plant, mainly during flowering (Okon, 1982). Significant increases in yields were obtained in Azospirillum inoculated fields that received an intermediate level of nitrogen fertilization at planting. Effects of this type (N₂ fixation, enhancement of mineral uptake) associated with rhizosphere bacteria such as Azospirillum, will be further enhanced by the higher rate of root surface activity resulting from higher photosynthate supply to the roots. Furthermore, if the root respiration rates increase, there may be a selection for bacteria that adapt

and are more efficient under microaerobic conditions, such as Azospirillum.

It is not clear if rhizosphere bacteria are specially active with C4 plants compared with C3 plants. Some promising Azospirillum isolates have been obtained from the roots of weeds. It will be important to know if high CO_2 will enhance weed growth more than that of crops. To exploit the potential of root associations with bacteria in agricultural systems will require careful management of any fertilizer additions to obtain maximum benefit under 600 ppm CO_2 .

5.5.4 Azolla. Azolla is a floating aquatic fern which contains a cyanobacterium, Anabaena azolla, as a symbiont. Both partners are photosynthetic, and the cyanobacterium can provide the association with its total N requirement. Azolla is a C3 plant with Calvin cycle intermediates, an 0_2 inhibition of photosynthesis and an 0_2 dependent $C0_2$ compensation point. Under optimized laboratory conditions these associations have been shown to double their biomass in 1.6 to 2.0 days and to contain 5-6% N on a dry weight basis. Under good field conditions, doubling times are on the order of 3-5 days with 4-5% N on a dry weight basis. These associations provide an alternative N source for rice. While both rice and Azolla may be expected to respond to CO₂ enrichment of the atmosphere, rice will require additional combined nitrogen if quality is maintained and yields enhanced. Azolla-Anabaena associations may play a significant role in meeting this additional N requirement for rice, and perhaps for other aquatic crops, such as taro.

5.6 Mycorrhiza. Mycorrhizae are symbiotic associations between the host-plant root and a mycorrhizal fungus. The fungi are generally referred to as ecto- or endomycorrhiza. These fungi belong to different classes of the Eumycota. Ectomycorrhizae (ECM) generally colonize woody plant species and are important in forest ecology. Endomycorrhizal fungi are mostly categorized as "vesiculararbuscular" because of their distinctive morphological characteristics. This latter category colonizes practically all native and agricultural plant species except species belonging to the orders Caryophyllales, Polygonales, and Capparales (Gerdemann, 1968). Ectomycorrhizae are generally biotrophic, but often can be grown in pure culture. Vesicular-arbuscular mycorrhizal (VAM) fungi are obligate biotrophs. They lack host specificity, and some species are world-wide in distribution.

5.6.1 Inoculum. Vesicular-arbuscular mycorrhizal fungi have not yet been cultured under axenic conditions in the absence of a host plant, in spite of considerable research efforts in several laboratories. Efforts are being made to produce VAM fungi for large-scale inoculation by soil culture (J. Menge, personal communication) and by a nutrient-film technique (Rothamsted Report, 1981).

Some data are available which indicate that certain host-endophyte combinations are more effective than others, in spite of a general lack of host specificity. The effectiveness of VAM fungi appears to be more affected by soil characteristics than by compatibility with any specific host plant (Mosse, 1975).

Inoculation of field crops with VAM fungi may be successful when native soil populations are sparse or consist of relatively ineffective species (Hayman, 1974). Yield or plant growth increases resulting from field inoculations have been reported by Black and Tinker (1977). Inoculation of cowpea with <u>Rhizobium</u> and chopped roots of a mycorrhizal native grass was carried out in a Brazilian oxisol by S.M. LaTorraca and T.V. St. John (unpublished data). Yield of VAM plants in five fertilization treatments were up to twice those of controls that received sterilized grass roots and live Rhizobium.

Field inoculations of ectomycorrhizal fungi already play an important role in establishment of exotic forest species and reclamation of mine spoils and other unfavorable sites (Marx and Krupa, 1978). Thus, techniques already exist that would allow new ectomycorrhizal fungi to be introduced as needed to deal with changing conditions brought about by increasing atmospheric CO₂.

5.6.2 Development. Effects of CO₂ on aerial portions of the plant (increased photosynthesis) can be expected to favorably influence the mycorrhizal symbiosis (Hayman, 1974). Competition for carbohydrates by the symbiosis under limiting light intensity was shown to result in growth depression of the host plant (Bethlenfalvay et al., 1982b). The development of host and VAM endophyte appear to be interrelated ontogenetically with source-sink relationships as a controlling factor (Bethlenfalvay et al., 1982a, 1982b). A possible increase in world mean temperature might influence fungal physiology (Furlan and Fortin, 1973; Hayman, 1974). Spore germination, infection, proliferation and sporulation of mycorrhizal fungi all depend on favorable environmental conditions. Some environmental factors to which VAM fungi are particularly

sensitive are pH, temperature, and excess water (Mosse et al., 1981).

The presence of decomposing soil organic particles stimulates the proliferation of VAM extra-matrical mycelium (St. John et al., submitted). Since soil hyphae are able to initiate new VAM infection points (Powell, 1976), the presence of increased soil organic matter may encourage more rapid spread of VAM. The direct effects of CO₂ on mycorrhizae have not been studied.

Saif (1981) examined the effect of oxygen concentration on mycorrhizal infection and plant growth response. He found that plants responded better to VAM at CO_2 concentrations of 12 and 16% than at 21%. His artificial atmosphere did not contain even the normal ambient amount of CO_2 . His techniques could be used to study CO_2 effects, and work at Rothamsted Experiment Station has apparently shown VAM infection to respond favorably to small increases in CO_2 concentration (B. Mosse, personal communication).

ECM fungi have been studied only slightly more than VAM fungi. Telson et al. (1980) reported that the difference in growth of mycorrhizal and non-mycorrhizal <u>Pinus sylvestris</u> was greater under conditions of higher CO_2 concentration. The growth of the mycorrhizal fungus <u>Pisolithus tinctorius</u> in pure culture was improved by higher than normal CO_2 concentrations.

Read and Armstrong (1972) showed that formation of the mantle of ECM in certain conifers is dependent on transport of oxygen through roots. They noted that oxygen can be very deficient in coniferous humus layers and suggested that mycorrhizal function in the humus environment is dependent on diffusion of O_2 from host roots. If they are correct, ECM establishment and function may be essentially independent of ambient CO_2 levels in nature.

Destruction of native vegetation, the main consumer of CO_2 , sometimes has the additional effect of the modification or destruction of mycorrhizal inocula (Reeves et al., 1979). Successional vegetation or agricultural and ornamental species subsequently occupying the site may be strongly influenced by the lack of inocula (Janos, 1980). One consequence is likely to be reduced production on that site, and therefore O_2 production and CO_2 consumption.

5.6.3 Effect on Plant Growth. Most of the P present in mineral soils is precipitated as the aluminum or iron

salt in acid soils and as the calcium salt in alkaline soils (Focht and Martin, 1979). Only a small fraction of the total P present in a soil may be available to plants. As a result, P is one of the major factors which limit plant growth under natural and cultural conditons. Mycorrhizal fungi may significantly increase the uptake of P and other relatively immobile nutrients, such as Zn and Cu, by their host plants (Hayman, 1978). The mechanism for this phenomenon is a thorough permeation of the soil volume availabe to the host by the extra-matrical fungal mycelium, which can reach soil microsites not exploited by the host alone (Tinker, 1978). In addition, mycorrhizae release oxylate which, in turn, causes the release of some additional nutrients from soil minerals (Graustein et al., 1977).

5.6.4 Effect on Water Uptake. The effect of mycorrhizal fungi on plants when water is the major limiting factor may be significant, but is not well known (Trappe, 1981). While few crop plants will survive when soil water potential is below -15×10^5 Pa (Hall et al., 1979), many fungi and bacteria will thrive at values far below this (Focht and Martin, 1979). Of particular interest in this respect are VAM, which appear to be common in desert plants (Trappe, 1981; Bethlenfalvay, unpublished data). Modification of plant water status by VAM may result in increased rates of photosynthesis under intermittent drought conditions, thus providing a better sink for CO_2 . The mechanism of enhanced water uptake by mycorrhiza is controversial. Higher rates of P uptake and the improved nutritional status of mycorrhizal plants have been implicated as the causal factors (Safir et al., 1972). More recent work, some of which was reviewed by Hardie and Leyton (1981), suggests that VAM may help plants acquire soil water not available to uninfected plants.

5.6.5 Respiratory Cost. Mycorrhizae growth could be limited by high soil CO_2 concentrations. VAM may represent up to 20% of the dry weight of the fungus-root association (Bethlenfalvay et al., 1982d). As respiration by fungi is 2 to 4 times higher than that of vascular plant tissue, and as CO_2 evolution by mycorrhizae is probably enhanced by infection respiration (Cooke, 1977), total respiration rates in mycorrhizae may be up to twice as high as in roots not colonized by VAM fungi. Increases in above-ground biomass may therefore be translated into below-ground respiration rates considerably higher than those found in experimentation with nonmycorrhizal plants under controlled conditions. 5.6.6 Effect on N_2 fixation. There are close linkages between organic C and N accumulation in mature soils and P contents in the original parent materials. Microbial growth processes are the principal arena for the adjustment of N supply to the supply of P (Cole and Heil, 1981). The availability of P is crucial to N₂ fixation, a process which has a many-fold higher requirement of P than does the host plant (Mosse, 1976). Thus the availability of P is a factor which may determine the rate at which N_2 fixation will be able to accommodate the increased N requirements of plants growing at a higher CO₂ concentration. Increased uptake of P by mycorrhizal fungi has been shown to enhance symbiotic N₂ fixation (Bethlenfalvay and Yoder, 1981) and associative N₂ fixation (Bagyaraj and Menge, 1978). These processes may therefore occupy a key position in modifying and controlling the flux of P and N to plants experiencing enhanced growth due to CO₂ fertilization.

5.7 Effects on Plant Pathogens

5.7.1 Fungus. Elevation of CO_2 concentration in the atmosphere and in soil may affect fungal development. Fungal species such as <u>Sclerotinia minor</u> and <u>Phytophthora</u> <u>infestans</u> are quite sensitive to CO_2 . Species like <u>Penicillium nigricans</u>, <u>Cochliobolus sativus</u> and many species of <u>Fusarium</u> are quite tolerant of high CO_2 . A rapid increase in CO_2 may have an adverse effect on some of the fungi, and changes in the compostion of the fungal community would be expected. This phenomenon might be observed in soil pores and microsites which contain a high concentration of entrapped CO_2 gas. However, given the adaptative capabilities of microorganisms, a gradual increase of CO_2 concentration over a long period of time may cause little overall change to the species present, and/or their growth.

Extremely high CO_2 concentrations (200,000 ppm or more) inhibit the hyphal growth of fungi, but sporulation and development of fruiting bodies (morphogenesis) seems to be much more sensitive to CO_2 . In mushrooms, for instance, production can be improved by manipulating CO_2 . At a high CO_2 , the pathogenicity of <u>Rhizoctonia solani</u> and <u>Sclerotium rolfsii</u> is reduced. High CO_2 can also reduce losses by inhibiting the growth of fruit-rotting fungi on fruits and vegetables.

5.7.2 Bacteria and other plant pathogens. Effects of doubling the CO_2 in the atmosphere on the growth and community composition of plant pathogenic bacteria may be

important because bacteria can survive from season to season by living in decaying plant tissues. Hence, the return of extra plant residues to soil may increase the population of pathogens as well as useful organisms.

For other plant pathogens -- virus, mycoplasma, rickettsia, etc. -- that are closely associated with the host system, little direct effect can be expected from increased CO₂ in the atmosphere.

6. Management Options

Direct effects of increased CO_2 levels are not considered to be of major significance on microbial soil activity. This conclusion is based on the assumption that soil CO_2 will not be significantly increased by a doubling of atmospheric CO_2 . Photosynthetic microorganisms on the soil surface will be unaffected by increased CO_2 because they already possess a CO_2 pumping system.

Indirect effects of increased atmospheric CO_2 as the result of increased plant productivity will strongly affect microbial soil activity which in turn will control the extent of the crop productivity that is realized. Management of the soil system including the application of existing and yet to be developed technologies will be key to the degree of crop productivity that is realized. Furthermore, some of these technologies, such as minimum tillage, provide models to study the impact of increased crop production.

Techniques such as minimum tillage greatly reduce erosion and the requirements of energy for cultivation. They increase the potential for crop growth and for stabilization of soil organic matter (Phillips et al., The plant cover and absence of physical disturbance 1980). produces a 2°-6°C drop in soil temperature, while soil moisture and aggregate stability are maintained. Minimum tillage in the U.S. is projected to increase from 2.2 Mha in 1974 to 62 Mha, or 45% of the total U.S. cropland, by 2000. At Sydney, Montana, minimum tillage maintained soil organic matter at the level of a virgin grassland site, while conventional tillage reduced organic matter content by 20%. The nitrogen requirement, at least in the short term, is increased, possibly due to increased immobilization and/or losses due to denitrification. Phosphate requirements in minimum tillage may parallel those of nitrogen.

The 1,600 - 2,000 Mha of potentially available land in

the tropics that is subject to rapid decomposition rates and erosion may benefit from minimum tillage. At present, 60% of this area is devoted to shifting cultivation. Management and maintenance of soil organic matter levels are essential for erosion control, water penetration, buffering against acidification, blocking P absorbing sites, and as a nutrient reservoir. An organic matter increase of 625 kg/ha for annual pastures and 1150 for perennial pastures has been observed in Australia. Bush fallow produces similar increases. Residue mulch - minimum tillage has maintained tropical soil and temperate organic matter levels 20% above those under conventional tillage.

6.2 Intercropping, Multiple Cropping, Green Manures. The use of legumes for intercropping, multiple cropping or as green manures is a management option for nitrogen provision. Combination of hairy vetch in minimum tillage maize production is an example. Hairy vetch grown during the winter periods in Kentucky and other southern states not only decreases soil erosion and weed growth, but has the potential for supplying 100-200 kg N/ha to the following corn crop. The use of a herbicide at the time of maize growth and the perennial nature of hairy vetch enables the cycle vetch-maize-vetch.

6.3 Microorganism Addition. Phosphorus limitation and aluminum toxicity inhibit plant growth in a vast area of tropical savanna such as the cerrado of Brazil (Sanchez et al., 1982a). Mycorrhizal fungi, as discussed earlier, have the potential to cause major increases in P uptake under tropical conditions of both upland and lowland crops (Islam et al., 1980; Tinker, 1982). Mycorrhizal inoculation coupled with their increased activity from increased crop production will impact positively on these large areas. Systems for production and inoculation with preferred mycorrhizal strains are needed. Mycorrhizal research is needed to provide management options for P input.

After phosphorus, nitrogen is probably the most important nutrient for increased crop productivity by increased CO_2 . Several management options exist. In the case of legumes, inoculation with the most effective N-fixing microbe will continue to be most important. The criteria for strain selection are ability to successfully compete with ineffective indigenous rhizobia, ability to nodulate the host plant, and ability to efficiently convert N₂ to ammonia. Similar approaches are being used with Frankia for the non-legume N₂-fixing trees. Other options exist for other crops. The Azolla-Anabaena association is

being evaluated as an intercrop with paddy-grown rice. Some management changes may be needed to optimize nitrogen input by this system, since in the presence of elevated CO_2 the canopy may close earlier than it does now. The associative N₂-fixing systems such as between cerial grains and <u>Azospirillum</u> have been shown to definitely increase crop yield in at least a few locations (see Section 5.5.3). <u>Azospirillum</u> inoculation may evolve as a management option for increasing nitrogen input.

<u>6.4 New Technology Inputs</u>. Manipulation of genetic materials of both host and microbial symbionts has great potential to provide new options for increased productivity. The rate of scientific advances in molecular genetics has been great relative to the modest rate of increase in atmospheric CO_2 . It is thus reasonably safe to assume that this evolving technology will obviate many of the limitations of existing technologies.

A discussion of possible relevant molecular genetic inputs follows. The time required for most of these technological advances is longer than many have suggested, but the advances within the next 25 years should be great. Molecular genetic manipulations of the N₂ fixation system pose some of the most difficult challenges in agriculture because of the high multiplicity of the genes and because of the sensitivity of nitrogenase to O_2 .

With the above qualifications, the potentials for genetic manipulation in N_2 fixation and denitrification will be considerable. Genetic studies have defined many of the nitrogen fixation genes in blue-green algae, in freeliving organisms, and in <u>Rhizobium</u>, and the genes responsible for nodulation are similarly being characterized. Improved versions of these genes will be developed and transferred among strains. <u>Rhizobium</u> which most efficiently fix nitrogen may be transformed to become the more effective nodulators, and, reciprocally, strains which dominate in the field may be the recipients of improved nitrogen fixation genes. Nodulation genes could also be transferred among <u>Rhizobium</u> strains in order to alter their host specificity. Alternatively, nitrogen fixation genes could be moved into mycorrhiza or perhaps even plants.

Following parallel lines of experimentation, nodulation and association genes could be transferred to associative bacteria. Identification of the plant genes necessary for these interactions will allow new plant hosts to be developed through genetic transfer of the plant symbiosis genes. Of particular interest in this regard would be both transferring symbiosis genes to crop plants such as corn and wheat, and transferring stem nodulation genes from peanuts, <u>Sesbania</u>, or <u>Aeschynemene</u> to new hosts. Stem nodules might be less susceptible to soil nitrate/ammonia inhibition and might be more able to take advantage of more CO_2 .

The problem of denitrification may similarly yield to genetic solutions. The genes that produce N_20 uptake enzymes could be altered to increase their affinities for N_20 . They could then scavenge N_20 and reduce it before its escape to the atmosphere.

Finally, opportunities will arise to create strains with novel synthetic enzymes. Hybrid genes are now being created by splicing segments of different genes together, and we can expect that novel nitrogenase genes will also be formed through gene synthesis and gene splicing. Such constructions should yield nitrogen-fixing enzymes that bear little resemblance to the present enzymes. One purpose for generating such modified enzymes would be to fix nitrogen to products other than ammonia. Many more such opportunities to create novel enzymes will undoubtedly present themselves. These technologies will greatly expand the management options in the coming decades.

9. Literature Cited

- At jay, G.L., P. Ketner and P. Duvigneaud. 1979. Terrestrial primary production and phytomass. In <u>Scope 13, The</u> <u>Global Carbon Cycle</u>, B. Bolin, E.T. Degens, S. Kempe and P. Ketner, Eds., John Wiley and Sons, New York, pp. 129-181.
- Bagyaraj, D.J. and J.A. Menge. 1978. Interaction between a VA mycorrhiza and <u>Azotobacter</u> and their effects on rhizosphere microflora and plant growth. <u>New Phytol</u>. 80, 567-573.
- Bauer, A. and A.L. Black. 1981. Soil carbon, nitrogen and bulk density comparisons in two crop plant tillage systems after 25 years and in virgin grasslands. <u>Soil</u> Sci. Sco. Am. J. 45, 1166-1170.
- Berry, J., J. Boynton, A. Kaplan, and M. Badger. 1976. Growth and photosynthesis of <u>Chlamydomonas reinhardii</u> as a function of CO₂ concentration. <u>Carnegie Institute</u> Year Book. 75, 423-432

Bethlenfalvay, G.J. S.S. Abu-Shakra, and D.A.

Phillips, 1978. Interdependence of nitrogen and photosynthesis in <u>Pisum Sativum</u> L. II. Host plant response to nitrogen fixation by Rhizobium strains. Plant Physiol. 62, 131-133.

- Bethlenfalvay, G.J. and J.F. Yoder. 1981. The <u>Glycine-</u> <u>Glomus-Rhizobium</u> symbiosis. <u>Physiol. Plant</u>. 52, <u>141-145</u>.
- Bethlenfalvay, G.J., M.S. Brown, and R.S. Pacovsky. 1982a. Relationships between host and endophyte development in mycorrhizal soybeans. New Phytol. 90, 537-543.
- Bethlenfalvay, G.J., R.S. Pacovsky, M.S. Brown, and G. Fuller. 1982b. Mycotrophic growth and mutualistic development of host plant and fungal endophyte in an endomycorrhizal symbiosis. Plant Soil (in press).
- Bethlenfalvay, G.J., R.S. Pacovsky, H.M. Bayne, and A. Stafford. 1982c. Interactions between N₂ fixation mycorrhizal colonization and host plant growth in the <u>Phaseolus-Rhizobium-Glomus</u> symbiosis. <u>Plant Physiol.</u> (in press).
- Bethlenfalvay, G.J., R.S. Pacovsky, and M.S. Brown. 1982d. Parasitic and mutualistic associations between a mycorrhizal fungus and soybean: development of the endophyte. Phytopathology. 72, 894-897.
- Bidwell, R.G.S. 1977. Photosynthesis and light and dark respiration in fresh water algae. <u>Can. J. Bot.</u> 55, 809-818.
- Black, R.L.B. and P.B. Tinker. 1977. Interaction between effects of vesicular-arbuscular mycorrhiza and fertilizer phosphorus on yields of potatoes in the field. Nature 267, 510-511.
- Boddey, R.M. and G. Dobereiner. 1982. Association of <u>Azospirillum</u>: Other diazotrophs with tropical Gramineae. <u>12th Intl. Congress of Soil Science. New</u> <u>Delhi, India, Feb. 8-16, 1982. Symposia Papers</u> pp. 23-47.
- Bolin, B., E.T. Degen, S. Kempe and P. Ketner, Eds. 1979. <u>The Global Carbon Cycle</u>. John Wiley and Sons, New York.

Buresch, P.J., M.E Casselman and W.H. Patrick, Jr. 1980.

Nitrogen fixation in flooded soil systems, a review. Adv. Agron. 33, 149-192.

- Coker, G.T. and K. Schubert. 1981. Carbon dioxide fixation in soybean roots and nodules. <u>Plant Physiol.</u> 67, 691-696.
- Cole, C. V. and R. D. Heil. 1981. Phosphorus effect on terrestrial nitrogen cycling. In F. E. Clark and T. Rosswell, Eds, <u>Terrestrial N-Cycles Ecological</u> Bulletin. Vol. 3. Stockholm, pp. 363-374.
- Coleman, D.C. 1973. Compartmental analysis of "total soil respiration": an exploratory study. Oikos 24, 361-366.
- Cooke, R. 1977. <u>The Biology of Symbiotic Fungi</u>. John Wiley and Sons, London.
- Dart, P.J. and J.M. Day. 1975. In <u>Non-symbiotic Nitrogen</u> <u>Fixation</u>, N. Walker, Ed. John Wiley and Sons, New York, pp. 225-252.
- Dart, P.J. and S.D. Wani. 1982. Nonsymbiotic nitrogen fixation and soil fertility. In <u>The Tropics Trans.</u> 12th Intl. Congress of Soil Science 1, 3-27.
- de Jong, E. and E.A. Paul. 1979. The composition of the soil atomsphere and its relationship to soil aeration and respiration. In <u>Encyclopedia of Earth Science</u> <u>Series. Vol. VI. Soil Science and Applied Geology.</u> O.W. Finkle, Jr., Ed. Reinhold Book Corp., New York.
- de Jong, E., H.J.V. Schappert and K.B. MacDonald. 1974. Carbon dioxide evolution from virgin and cultivated soil as affected by management practices and climate. Canadian J. Soil Sci. 54, 299-307.
- Denmead, O.T. and E.R. Lemon. 1970. <u>Prediction and</u> <u>measurement of photosynthetic activity</u>. Wageningen Center for Agricultural Publishing and Documentation, pp. 149-164 and 199-205.
- De-Polli, H., M. Eiichi and J. Dobereiner. 1977. Confirmation of nitrogen fixation in two tropical grasses by molecular nitrogen 15 incorporation. <u>Soil Biol.</u> Biochem. 9, 119-123.
- Dreyfus, B.L. and J.R. Dommergue. 1981. Nitrogen fixing nodules induced by <u>Rhizobium</u> on the stem of tropical legume Sesbania rostrata. FEMS Lett. 10, 313-317.

- Focht, D.D. and J.P. Martin. 1979. Microbiological and biochemical aspects of semi-arid agricultural soils. In <u>Agriculture in Semi-Arid Environments</u>, A.E. Hall, G.H. Cannel and H.W. Lawton, Eds. Springer-Verlag, Berlin, pp. 119-147.
- Furlan, V. and J.A. Fortin. 1973. Formation of endomycorrhizae by Endogone calospora on Allium cepa under three temperature regimes. Nat. Can. 100, 467-477.
- Garrett, S.D. 1981. Soil Fungi and Soil Fertility. Pergamon Press, Oxford.
- Geisler, G. 1967. Interactive effects of CO₂ and O₂ in soil on root and top growth of barley and peas. <u>Plant</u> Physiol. 42, 305-307.
- Gerdemann, J.W. 1968. Vesicular-arbuscular mycorrhiza and plant growth. Ann. Rev. Phytopathol. 6, 397-418.
- Goswami, K. and V.K. Suri. 1982. Carbon fertilization: Influence of simulated field soil respiration on soybean crop. Proc. 12th ISSS Congress, Delhi.
- Goudriaan, J. and G.L. Ajtay. 1979. The possible effects of increased CO₂ on photosynthesis. In <u>The</u> <u>Global Carbon Cycle</u>, B. Bolin, E.T. Degens, S. Kempe and P. Ketner, Eds. John Wiley and Sons, New York, pp. 237-250.
- Grable, A.R. 1966. Soil aeration and plant growth. Adv. Agron. 18, 57-106.
- Graustein, W.C., K. Cromack and P. Sollins. 1977. Calcium oxalate: Occurrence in soils and effect on nutrient and geochemical cycles. Science. 198, 1252-1254.
- Greenwood, D.J. 1970. Distribution of carbon dioxide in the aqueous phase of aerobic soils. J. Soil Sci. 21, 314-329.
- Griffin, D.M. 1972. Ecology of Soil Fungi. Syracuse Univ. Press., Syracuse, N.Y.
- Hall, A.E., K.W. Foster and J.G. Waines. 1979. Crop adaptation to semi-arid environments. In <u>Agriculture in</u> <u>Semi-Arid Environments</u>, A.E. Hall, G.H. Cannell and H.W. Lawton, Eds. Springer-Verlag, Berlin, pp. 148-179.

- Hardie, K. and L. Leyton. 1981. The influence of vesicular-arbuscular mycorrhiza on growth and water relations of red clover. I. In phosphate deficient soil. New Phytol. 89, 599-608.
- Hardy, R.W.F. and U.D. Havelka. 1975. Photosynthate as a major factor limiting N₂ fixation by field grown legumes with emphasis on soybeans. In <u>Symbiotic</u> <u>Nitrogen Fixation in Plants</u>, R.S. Nutman, Ed., <u>Cambridge University Press</u>, London, pp. 421-439.
- Havelka, U.D. and R.W.F. Hardy. 1976. Legume N₂ fixation as a problem in carbon nutrition. In <u>Nitrogen</u> <u>Fixation</u>, W.E. Newton and C.J. Nyman, Eds., Vol. 2., Washington State Univ. Press, pp. 456-475.
- Hayman, D.S. 1974. Plant growth responses to vesiculararbuscular <u>mycorrhiza</u>. VI. Effect of light and temperature. New Phytol. 73, 71-80.
- Idso, S.B. 1981. Carbon dioxide an alternative view. <u>New</u> Scientist 92, 444-446.
- Idso, S.B. and B.A. Kimball. 1982. Man, carbon dioxide, climate and food: A global perspective. <u>Symposium on</u> Plant Production Under Drought Conditions. Tulsa, OK.
- Islam, R., A. Ayanaba and F.E. Sander. 1980. Response of cowpea (<u>Vigna unguiculata</u>) to inoculation with VA mycorrhizal fungi and to rock phosphate fertilization in some unsterilized Nigerian soils. <u>Plant and Soil</u> 54, 107-117.
- Janos, D.P. 1980. <u>Mycorrhizae</u> influence tropical succession. Biotropica 12 (Supplement), 56-64.
- Jenkinson, D.S. and J.H. Rayner. 1977. The turnover of soil organic matter in some of the Rothamsted classical experiments. Soil Sci. 123, 298.
- Jenkinson, D.S. and J.W. Ladd. 1980. In <u>Soil Biochemistry</u>, Vol. 5, E.A. Paul and J.W. Ladd, Eds., Marcel Dekker, New York.
- Johnen, B.G. 1974. <u>Bildung, Menge und Umsetzung von</u> <u>Pflanzenwurzein im Boden</u>. Agrikulturchemischen Institut der Rheinishchen Friedrich-Wilhelms-Universitat, Bonn.

Kaplan, A., D. Zenvirth, L. Reinhold and J.A. Berry. 1982.

Involvement of a primary electrogenic pump in the mechanism for HCO₃⁻ uptake by the cyanobacterium Anabaena variablis. Plant Physiol. 69, 978-982.

- Karagouni, A.D. and J.H. Slater. 1978. Growth of the blue-green alga <u>Anacystis nidulans</u> during washout from light- and carbon dioxide-limited chemostats. <u>FEMS</u> Microbiol. Lett. 4, 295-299.
- Kimmins, J.P., J. de Catanzaro and D. Binkley. 1979. Tabular summary of data from the literature on the biogeochemistry of temperate forest ecoystems. <u>ENFOR</u> <u>Project P-8 Report</u>, Faculty of Forestry, Univ. of British Columbia, Vancouver, British Columbia, p. 104.
- Kramer, P.J. 1981. Carbon dioxide concentration, photosynthesis, and dry matter production. <u>BioScience</u> 31, 29-33.
- Kucey, R.M.N. and E.A. Paul. 1982. Carbon flow, photosynthesis and N₂ fixation in mycorrhizal and nodulated faba beans (<u>Vicea faba</u> L.). <u>Soil Biol</u>. Biochem., in press.
- Lal, R. and B.T. Kang. 1982. Management of organic matter in soils of the tropics and subtropics. Proc. 12th ISSS Congress, Delhi. 1, 152.
- Lindsay, W.L. 1979. <u>Chemical Equilibria in Soil</u>. John Wiley and Sons, New York.
- Lipschultz, F., O.C. Zafiriou, S.C. Wofsy, M.B. McElroy, F.W. Valoris and S.W. Watson. 1981. Production of NO and N₂O by soil nitrifying bacteria. <u>Nature</u> 294, 641-643.
- Lloyd, N.D.H., D.J. Canvin and D.A. Culver. 1977. Photosynthesis and photorespiration in algae. <u>Plant</u> Physiol. 59, 936-940.

Lowe, R.H. and H.J. Evans. 1962. Soil Science 94, 351-356.

Luxmoore, R.J. 1981. CO2 and phytomass. BioScience 31, 626.

- Lynch, J.N. 1978. Productivity and phytotoxicity of acetic acid in anaerobic soil and plant residues. <u>Soil Biol</u>. Biochem. 10, 131-135.
- Macauley, B.J. and D.M. Griffin. 1969. Effects of carbon dioxide and oxygen on the activity of some soil fungi. Trans. Brit. Mycol. Soc. 53, 53-62.

Mahon, J.D. 1979. Plant Physiol. 63, 892-897.

- Marx, D.H. and S.V. Krupa. 1978. Mycorrhizae. A. Ectomycorrhizae. In <u>Interactions between Non-pathogenic Soil</u> <u>Microorganisms and Plants</u>. Y.R. Dommergues and S.V. Krupa, Eds. Elsivier Scientific Publishing Co., Amsterdam, pp. 373-400.
- McKenney, D.J., K.F. Shuttleworth, J.R. Vriesacker and W.I. Findlay. 1982. Production and loss of nitric oxide from denitrification in anaerobic Brookston clay. Appl. Environ. Microbiol. 43, 534-541.
- Melillo, J.N., J.D. Aber and J.F. Muratore. 1982. Nitrogen and lignin control of hardwood leaf litter decomposition dynamics ecology (in press).
- Minchin, F.R. and J.S. Pate. 1973. The carbon balance of a legume and the functional economy of its nodules. J. Exp. Bot. 24, 259-271.
- Moore, A.W. 1969. Azolla: biology and agronomic significance. Bot. Rev. 35, 17-34.
- Mosse, B. 1975. Specificity in VA mycorrhizas. In Endomycorrhizas, F.E. Sanders, B. Mosse and P.B. Tinker, Eds., Academic Press, London. pp. 469-484.
- Mosse, B. 1976. The role of mycorrhiza in legume nutrition on marginal soils. In <u>Exploiting the Legume-</u> <u>Rhizobium Symbiosis in Tropical Agriculture</u>, J.M. Vincent, A.S. Whitney and J. Vose, Eds., <u>Coll. Trop.</u> Agri. Misc. Publ. 145. Univ. Hawaii, pp. 275-292.
- Mosse, B., D.P. Stribley and F. Le Tacon. 1981. Ecology of mycorrhizae and mycorrhizal fungi. <u>Advances in</u> <u>Microbial Ecology 5, 137-210.</u>
- Mulder, E.G. and W.L. van Veen. 1960. The influence of carbon dioxide on symbiotic nitrogen fixation. <u>Plant</u> and Soil 13, 265-278.
- Newton, W., J.R. Postgate, and C. Rodriguez-Barrueco. 1977. <u>Recent Developments in Nitrogen Fixation</u>. Academic Press, London.
- Neyra, C.A. and J. Dobreiner. 1977. Nitrogen fixation in grasses. Adv. Agron. 29, 1028.
- Nye, P.H. and P.B. Tinker. 1977. Solute movement in the

soil-root system. University of California Press, Berkeley.

- Okon, Y. 1982. Azospirillum: Physiological properties, mode of association with roots and its application for the benefit of cereal and forage grass crops. Special volume on Nitrogen Fixation. Israel Journal of Botany, in press.
- Patrick, W.H. 1977. Oxygen content of soil air by a field method. Soil Sci. Soc. Am. J. 41, 651-652.
- Payne, W.J. 1981. <u>Denitrification</u>. Wiley-Interscience, New York.
- Peters, G.A. and H.E. Calvert. 1982. The <u>Azolla-Anabaena</u> Symbiosis, Chapter in <u>Algal Symbiosis: A Continuum</u> of Interaction Strategies. Sponsored by the Phytological Society of America, Cambridge University Press (in press).
- Phillips, D.A., K.D. Newell, S.A. Hassell, and C.E. Felling. 1976. The effect of CO₂ enrichment on root nodule development and symbiotic N₂ reduction in Pisum sativan L. Amer. J. Bot. 63, 356-362.
- Phillips, R.E., R.E. Blevins, G.W. Thomas, W.W. Frye and S.G. Phillips. 1980. No tillage agriculture. <u>Science</u> 208, 1108-1113.
- Phillips, D.A. 1980. Efficiency of nitrogen fixation in legumes. Ann. Rev. Plant Physiol. 31, 29-49.
- Powell, C.L.I. 1976. Development of mycorrhizal infections from Endogone spores and infected root segments. Trans. Brit. Mycol. Soc. 66, 439-445.
- Rabinowitch, E.I. 1951. <u>Photosynthesis</u>, Vol. II, Part I. Interscience Publishers Inc., New York, p. 886.
- Read, D.J. and W. Armstrong. 1972. A relationship between oxygen transport and the formation of the ectotrophic mycorrhizal sheath in conifer seedlings. <u>New Phytol</u>. 71, 49-53.
- Reeves, F.B., D. Wagner, T. Moorman and J. Kiel. 1979. The role of endomycorrhizae in revegetation practices in the semi-arid West. I. A comparison of incidence of mycorrhizae in severely disturbed vs. natural environments. Am. J. Bot. 66, 6-13.

- Reichle, D.E., B.E. Dinger, N.T. Edwards, W.F. Harris and P. Sollins. 1973. Carbon flow and storage in a forest ecosystem. In G.M. Woodwell and E.V. Pecan Eds., <u>Proc. of the Symposium in Biology</u>, Upton, New York.
- Rogers, P.A. and S.A. Kulasooriya. 1980. <u>Blue-green algae</u> and rice. The International Rice Research Institute, Los Banos, Laguna, Phillipines.
- Rogers, H.H., G.E. Bingham, J.D. Cure, W.W. Heck, A.S. Heagle, D.W. Israel, J.M. Smith, K.A. Surano, and J.F. Thomas. 1980. <u>Response of vegetation to carbon</u> <u>dioxide</u>. U.S. Department of Energy and U.S. Department of Agriculture Joint Publication.
- Russell, E.W. 1973. Soil conditions and plant growth. 10th ed. Longmans, London, Chapter 18.
- Safir, G., J.S. Boyer and J.W. Gerdemann. 1972. Nutrient status and mycorrhizal enhancement of water transport in soybean. Plant Physiol. 49, 700-703.
- Saif, S.R. 1981. The influence of soil aeration on the efficiency of vesicular-arbuscular mycorrhizae.
 I. Effect of soil oxygen on the growth and mineral uptake of <u>Eupatorim odoratum</u> L. innoculated with Glomus macrocarpus. New Phytol. 88, 649-659.
- Sanchez, P.A., D.E. Bandy, J.F. Villachica and J.J. Nicholaides. 1982a. Amazon basin soils: Management for continuous crop production. Science 216, 821-827.
- Sanchez, P.A., M.P. Gichuru and L.B. Katz. 1982b. Organic matter in major soils of the tropical and temperate regions. Proc. 12th ISSS Congress, Delhi. 1, 99.
- Schwartz, D.M. and F.A. Bazzaz. 1973. In situ measurements of carbon dioxide gradients in a soil-plantatmosphere system. Oecologia 12, 161-167.
- Shivashankar, K., K. Vlassak and J. Livens. 1976. A comparison of the effect of straw incorporation and carbon dioxide enrichment on the growth, nitrogen fixation and yield of soybeans. J. Agric. Sci. 87, 81-85.
- Smith, A.M. and D.M. Griffin. 1971. Oxygen and the ecology of <u>Armillariella elegans</u> Heim. <u>Austr. J. Biol. Sci.</u> 24, 231-262.

- Stewart, W.D.P. 1975. <u>Nitrogen fixation by free-living</u> organisms. Cambridge Univ. Press.
- Stolwijk, J.A. and K.V. Thimann. 1957. On the uptake of carbon dioxide and bicarbonate by roots, and its influence on growth. Plant Physiol. 32, 513-520.
- Subba Rao, N.S. 1977. Nitrogen deficiency as a world-wide problem. In <u>A Treatise on Nitrogen Fixation</u>, R.W.F. Hardy and A.H. Gibson, Eds. John Wiley and Sons, New York, pp. 3-32.
- Swank, W.T. and J.B. Waide. 1980. Interpretations of nutrient cycling research in a management context: Evaluating potential effects of alternative management strategies on site productivity. In Forests: Fresh Perspectives from Ecosystem Analysis, R.W. Waring, Ed. Oregon State University Press, Corvallis, pp. 137-158.
- Telson, M., I.A. Leone and F.B. Flower. 1980. The role of an ectomycorrhizal fungus <u>Pisolthus tinctorius</u> in the survival and growth of scots pine subjected to landfill conditions. (Abstr.) Phytopathology 70, 470.
- Tinker, P.B. 1978. Effects of vesicular-arbuscular mycorrhizae on plant nutrition and plant growth. Physiol. Veg. 16, 743-751.
- Tinker, P.B. 1982. Mycorrhizas: The present position. In <u>Whither Soil Research.</u> Trans. 12th Intl Congress of Soil Science. New Delhi. 5, 140-166.
- Trappe, J.M. 1981. Mycorrhizae and productivity of arid and semi-arid rangelands. In <u>Advances in Food</u> <u>Producing Systems for Arid and Semi-Arid Lands</u>. Acad. Press, London, pp. 581-599.
- van Veen, J.A. and E.A. Paul. 1981. Organic carbon dynamics in grassland soils. I. Background information and computer simulation. <u>Can. J. Soil</u> Sci. 61, 185-201.
- Vose, P.B. and A.P. Ruschel. 1981. Associative N2 fixation, Vols. 1 and 2. CRC Press, Boca Raton.
- Winship, L.J. and J.D. Tjepkema. 1982. <u>Plant Physiol</u>. (accepted).