# Hope College Digital Commons

# **Faculty Publications**

9-5-2014

# Intergeniculate Leaflet Lesions Result in Differential Activation of Brain Regions Following the Presentation of Photic Stimuli in Nile Grass Rats

Andrew J. Gall Michigan State University, gall@hope.edu

Lily Yan Michigan State University

Laura Smale Michigan State University

Antonio A. Nunez Michigan State University

Follow this and additional works at: https://digitalcommons.hope.edu/faculty\_publications

Part of the Behavioral Neurobiology Commons, and the Biological Psychology Commons

# **Recommended Citation**

**Repository citation**: Gall, Andrew J.; Yan, Lily; Smale, Laura; and Nunez, Antonio A., "Intergeniculate Leaflet Lesions Result in Differential Activation of Brain Regions Following the Presentation of Photic Stimuli in Nile Grass Rats" (2014). *Faculty Publications.* Paper 1502.

https://digitalcommons.hope.edu/faculty\_publications/1502

Published in: *Neuroscience Letters*, Volume 579, September 5, 2014, pages 101-105. Copyright © 2014 Elsevier.

This Article is brought to you for free and open access by Hope College Digital Commons. It has been accepted for inclusion in Faculty Publications by an authorized administrator of Hope College Digital Commons. For more information, please contact digitalcommons@hope.edu.



# NIH Public Access

**Author Manuscript** 

Neurosci Lett. Author manuscript; available in PMC 2015 September 05.

#### Published in final edited form as:

Neurosci Lett. 2014 September 5; 579: 101–105. doi:10.1016/j.neulet.2014.07.016.

# Intergeniculate leaflet lesions result in differential activation of brain regions following the presentation of photic stimuli in nile grass rats

Andrew J. Gall, Lily Yan, Laura Smale, and Antonio A. Nunez

Department of Psychology and Neuroscience Program Michigan State University East Lansing, MI USA

# Abstract

The intergeniculate leaflet (IGL) plays an important role in the entrainment of circadian rhythms and the mediation of acute behavioral responses to light (i.e., masking). Recently, we reported that IGL lesions in diurnal grass rats result in a reversal in masking responses to light as compared to controls. Here, we used Fos as a marker of neural activation to examine the mechanisms by which the IGL may influence this masking effect of light in grass rats. Specifically, we examined the patterns of Fos activation in retinorecipient areas and in brain regions that receive IGL inputs following 1-h light pulses given during the early night in IGL-lesioned and sham-operated grass rats. Three patterns emerged: (1) IGL lesions had no effect on the Fos response to light, (2) IGL lesions resulted in a reversal in Fos responses to light, and (3) IGL lesions resulted in a lack of a Fos response to light. Of specific interest were the suprachiasmatic nucleus (SCN) and the olivary pretectal nucleus (OPT), both of which are retinorecipient and connect reciprocally with the IGL. Light-induced Fos expression in the SCN was unaffected by IGL lesions, whereas the OPT exhibited a significant reduction in Fos expression following a light pulse in animals with IGL lesions. Altogether, our results suggest that the OPT, but not the SCN, exhibits a reversal in Fos responses to light following IGL lesions that reverse masking responses in diurnal grass rats. Our results suggest that interconnections between the IGL and downstream brain areas (e.g., OPT) may play a role in determining the direction of the behavioral response to light.

#### Keywords

intergeniculate leaflet; Fos; masking; grass rat; diurnality

The authors declare no competing financial interests.

<sup>© 2014</sup> Elsevier Ireland Ltd. All rights reserved

Corresponding author: Andrew J. Gall, Ph.D. Department of Psychology Michigan State University East Lansing, MI 48824 Phone: 517.355.6873 Fax: 517.432.2744 gall@msu.edu.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

#### Introduction

Light and darkness modulate behavior and physiology not only by entraining circadian rhythms, but also by acutely inhibiting or stimulating activity, a process called masking [1]. In diurnal animals, light generally stimulates activity, whereas light generally inhibits activity in nocturnal animals [2,3]. The neural substrates involved in masking responses to light are relatively unknown, especially in diurnal animals.

The intergeniculate leaflet (IGL), which receives direct retinal input [4], has been implicated in the mediation of masking responses in nocturnal rodents [5]. We recently showed that light pulses result in a significant reduction in activity levels following IGL lesions in diurnal grass rats (*Arvicanthis niloticus*), whereas control grass rats exhibit a significant increase in activity levels [6]. Therefore, destroying the IGL in these diurnal animals alters their masking patterns to resemble those of nocturnal animals. The mechanisms by which this reversal in behavior occurs following IGL lesions have not yet been examined.

Here, we used Fos expression to examine whether the IGL influences responses to light in retinorecipient regions in the brain, or in regions that are involved in sleep and arousal. We focused particularly on the suprachiasmatic nucleus (SCN) and olivary pretectal nucleus (OPT) since both areas are retinorecipient [7,8], connect reciprocally with the IGL [4,9], play a functional role in sleep-wake processes [10,11], and have been linked to masking [12,13], although the role of the SCN in the mediation of masking responses to light remains controversial [14]. Further, since light is capable of inducing sleep in nocturnal animals [2] and arousal in diurnal ones [15], we also measured Fos expression in brain regions that receive inputs from the IGL and are involved in the control of sleep or wakefulness, including the ventrolateral preoptic nucleus (VLPO), ventral subparaventricular zone (vSPVZ), dorsomedial hypothalamus (DMH), locus coeruleus (LC), and dorsal raphe (DR) [16].

Based on our earlier observations [6], we predicted that the SCN's response to light would be unaffected by the IGL lesions, since the effects of light on the period of activity rhythms was unaffected by IGL lesions. In addition, we predicted that one or more brain areas outside the SCN would exhibit a reversal in Fos expression following light pulses given to IGL-lesioned animals. Such a finding would suggest a mechanism by which the IGL might promote a diurnal pattern of masking behavior in grass rats.

### Materials and Methods

All experiments were carried out in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals (NIH Publication No. 80-23) and were approved by the Institutional Animal Care and Use Committee of Michigan State University. All efforts were made to minimize the number of animals used.

#### Subjects

Twenty-eight singly housed adult female grass rats (n = 20 complete IGL-lesioned subjects, n = 8 shams) were used. These 28 animals were the same used in a previous report, which

describes the behavioral effects of IGL lesions that also extended beyond the IGL (see [6] for surgical and histological details). Female grass rats do not exhibit estrous cycles in the laboratory [17], and do not differ from males in masking responses to light [3].

#### **Experimental Procedures**

**Light treatment procedure**—As described previously [6], at least 10 weeks after surgery, following the placement of animals in constant dark (DD) and constant light (LL), animals were re-entrained to a 12:12 LD cycle, and then subjected to a series of dark and light pulses given in 12:12 LD (lights on = ZT 0) while behavior was monitored. Finally, approximately half of the animals (n = 9 lesions, n = 4 shams) received a 1-h light pulse (300 lux of white light) at ZT14 during the dark phase of a 12:12 LD cycle, and were sacrificed at ZT15. The other half (n = 11 lesions, n = 4 shams) were also sacrificed at ZT15, but without receiving a light pulse (LP).

#### c-Fos Immunohistochemistry

Following transcardial perfusion, brains were removed and sectioned as described previously [6]. Labeling of Fos-immunoreactive (Fos-ir) cells followed protocols previously established in the grass rat brain [18]. Sections were incubated in Fos antibody raised in rabbit (1:25,000, Santa Cruz Biotechnology, Santa Cruz, CA, USA) and processed with avidin-biotin-immunoperoxidase using DAB (3,3'-diaminobenzidine) as the chromogen enhanced with nickel sulfate. Sections were mounted on gelatin-coated glass slides, dehydrated, and coverslipped with dibutyl phthalate xylene (DPX; Sigma-Aldrich, St. Louis, MO, USA).

#### Cell counting

For quantifying Fos expression, observers blind to experimental condition selected two sections containing each brain region of interest, including the SCN, VLPO, OPT, vSPVZ, DMH, LC, and DR. Sections were examined under a light microscope (Leitz, Laborlux S, Wetzlar, Germany) equipped with a drawing tube to produce bilateral maps of Fos positive cells. Counting boxes were used to delineate the VLPO (190  $\mu$ m × 190  $\mu$ m; [19]), vSPVZ (215  $\mu$ m × 160  $\mu$ m; [20]), LC (400  $\mu$ m × 700  $\mu$ m; [18]), and DR (150  $\mu$ m × 650  $\mu$ m; [18]). The SCN, OPT, and DMH were outlined using thionin counterstained tissue. For the OPT, Fos-positive cells were counted separately for the shell and core, but since the counts were not significantly different between the two areas, data for the OPT are reported as the total of shell plus core. The number of Fos-positive cells for each region were counted bilaterally and divided by 2 to obtain an average of unilateral Fos-ir counts. The Fos-ir counts from the OPT of two brains were excluded from the analysis due to the region being partially lesioned bilaterally.

#### Statistical analysis

A two-way ANOVA was used to analyze the data with a  $2 \times 2$  factorial design [surgical condition (sham vs. IGL lesion) × lighting condition (darkness vs. light pulse)]. Significant interactions were followed by evaluation of simple main effects using independent sample t-

tests. For all analyses, differences were significant when p < 0.05. All means are presented with their standard errors.

### Results

#### IGL lesions did not affect Fos responses to light in the SCN or VLPO

For the SCN (Figure 1), a two-way ANOVA found a significant main effect of lighting condition ( $F_{1,24} = 63.4$ , p < .0001), but not of surgical condition ( $F_{1,24} = .04$ , p = .837), and no interaction between the two variables ( $F_{1,24} = .003$ , p = .959). For the VLPO (Table 1), a two-way ANOVA found a significant main effect of lighting condition ( $F_{1,24} = 45.1$ , p < .0001), and of surgical condition ( $F_{1,24} = 4.5$ , p = .045), but no interaction between the two variables ( $F_{1,24} = 1.5$ , p = .229).

#### IGL lesions resulted in a reversal in Fos responses to light in the OPT

For the OPT (Figure 2), a two-way ANOVA found a significant interaction between lighting and surgical condition ( $F_{1,22} = 79.0, p < .0001$ ). Analysis of simple main effects of lighting condition revealed that Fos-ir was significantly increased following a light pulse in shams ( $t_6 = 8.0, p < .0001$ ), but significantly decreased in lesioned animals ( $t_{16} = -3.3, p = .004$ ). Analysis of simple main effects of surgical condition showed that following a light pulse, IGL-lesioned animals exhibited significantly decreased Fos-ir as compared to shams ( $t_{10} = 5.8, p < .0001$ ). For the control night, IGL-lesioned animals exhibited significantly increased Fos-ir as compared to shams ( $t_{12} = -6.8, p < .0001$ ).

#### IGL lesions resulted in a lack of Fos responses to light in the vSPVZ, DMH, LC, and DR

In the vSPVZ, DMH, LC, and DR (Table 1), two-way ANOVAs found significant interactions between lighting and surgical condition ( $F_{1,24}s > 4.3$ , ps < .05). Analysis of simple main effects of lighting condition revealed that Fos-ir was significantly increased following a light pulse in all four areas for shams ( $t_6s > 4.3$ , ps < .005), but not for lesioned animals ( $t_{18}s < 1.7$ , ps > .096). In the vSPVZ and LC, the simple main effects of surgical condition on Fos-ir did not reach statistical significance following a light pulse ( $t_{11}s < 1.9$ , ps > .089) or for the control night ( $t_{13}s < 1.9$ , ps > .084). However, in the DMH and DR, the simple main effect of surgical condition was significant for the light pulse condition; IGLlesioned animals exhibited significantly less Fos-ir as compared to shams ( $t_{11}s > 2.6$ , ps < .024). For the control night, no significant differences were found in Fos-ir for IGL-lesioned animals as compared to shams ( $t_{13}s < 1.3$ , ps > .218).

### Discussion

IGL lesions in grass rats result in a redistribution of activity across the circadian cycle and a striking reversal in the direction of behavioral masking responses to light pulses presented at ZT14 as compared to controls [6]. The current study used Fos expression to investigate how selected brain regions change their responses to light after IGL lesions, and to use that information to identify possible pathways through which the IGL might influence masking effects of light in grass rats. Our results demonstrate that the OPT, vSPVZ, DMH, DR, and LC, but not the SCN or VLPO, change their responses to light following IGL lesions.

Interestingly, the OPT exhibited a reversal in Fos induction following light pulses in IGLlesioned grass rats, raising the possibility that interconnections between the IGL and OPT play a role in determining the direction of the behavioral response to light.

Considering the key role that the SCN plays in the circadian regulation of behavior, and given its possible, but controversial, involvement in mediating behavioral masking responses to light [12,14], it may seem surprising that Fos-ir in the SCN was unaffected by light pulses given to grass rats with IGL lesions, even though circadian activity is significantly affected by these lesions. However, as mentioned above, IGL lesions in the same animals did not affect how constant light and darkness influenced the period of free running activity rhythms in grass rats [6], thus suggesting normal SCN responses to light after IGL lesions. Further, there is no consensus about the necessity for a functional SCN for the mediation of masking responses, since following complete SCN lesions, hamsters can show masking responses to light [14]. Taken together, our past [6] and current data indicate that IGL lesions can have profound effects on masking responses and on the distribution of daily activity without apparent changes in how the SCN responds to light.

Although our results show that Fos expression in the SCN in response to light is unaffected by IGL lesions, we cannot rule out that other aspects of light transduction by the SCN were not affected by the lesions. Importantly, light stimulation during the subjective day increases the number of action potentials recorded from SCN neurons in nocturnal rats *in vivo* [21] and induces clock gene expression in the adrenal gland [22], without inducing Fos or clock-gene expression in the SCN [22,23]. Therefore, the SCN may be capable of transducing photic input in a phase-independent manner without altering gene expression or the phase and period of its oscillator. Thus, the lack of change in light-induced Fos expression observed in grass rats following IGL lesions is not sufficient to conclude that IGL lesions are incapable of affecting photic transduction by SCN neurons.

The OPT is retinorecipient [8], reciprocally connected to the IGL [4,9], and exhibits a Fos response to light [13,24]. The OPT mediates pupillary reflexes to light and is involved in the generation of eye movements [25], but may also be involved in sleep and circadian behavior [10,26,27]. Here, the phase-dependent increase in activity at night, along with the reversal in the direction of masking responses of grass rats after IGL lesions [6], was matched by a reversal in Fos expression in the OPT of these animals, thus suggesting that normal behavioral and neural responses to light by this diurnal species require an intact IGL-OPT circuit.

Other observations also point to the OPT as a candidate for mediating responses to light in a chronotype-specific fashion. For example, for grass rats that become night-active when given a running wheel, the OPT does not exhibit light-induced Fos at ZT14, whereas it does so in day-active wheel-runners [24]. Also, light induces Fos in the OPT of intact grass rats, but decreases it in intact mice [28]; this result stands in contrast to findings in laboratory rats [13], in which 2-h light pulses given at ZT19 induce increased Fos-ir in the OPT. This difference in lab rats could be due to a difference in the time of day in which animals were light pulsed, or could indicate a species difference. Taken together, data obtained from night-active grass rats, nocturnal mice, and IGL-lesioned grass rats with enhanced nocturnal

activity, show that the induction of Fos-ir by light within the OPT at ZT14 is either reversed or absent in night-active compared to day-active animals. Therefore, the OPT is a possible candidate for mediating the differences observed in masking between night-active and day-active phenotypes at ZT14, both within and between species.

The VLPO is a forebrain structure that is retinorecipient [7], but its role in masking is virtually unknown. We reported recently that light presented at ZT14 results in a significant increase in Fos-ir in the VLPO in grass rats [28]. This appears to be a paradoxical response, given that the VLPO is a sleep-active region in both diurnal and nocturnal species [19,29]. Galanin-containing cells of the VLPO appear to be responsible for inducing sleep in nocturnal animals [30]. One untested possibility is that light stimulates a different subset of cells within the VLPO in grass rats, perhaps a population of local inhibitory neurons that synapses on the galanin-containing cells. As shown here, lesions of the IGL do not affect light-induced Fos responsiveness in the VLPO, even though the masking response is reversed in these animals. Since IGL-lesioned grass rats behave more like nocturnal animals in terms of masking and circadian behavior [6], we predict that the same cells that respond to light in nocturnal species respond to light in IGL- lesioned grass rats.

The vSPVZ [23,31,32], DMH [31], LC [33], and DR [27,34] exhibit a Fos response to light, with orexin being necessary for this response in the DR of grass rats [34]. As shown here, these areas responded to light in control grass rats, but not in animals with IGL lesions. Thus, connections between the IGL and these areas may be vital to their ability to respond to light. It is possible that these sites receive information about light directly from the IGL or via pathways affected by IGL lesions (e.g., orexinergic pathways). Different IGL neurons express neuropeptide Y (NPY) or enkephalin (Enk) [35]; we are currently testing the hypothesis that one or both of these cell types project to downstream areas (e.g., OPT) to modulate neural and behavioral responses to light.

# Conclusions

Grass rats with complete IGL lesions behave more like nocturnal animals in terms of masking and circadian behavior [6]. Concurrent with a reversal in behavior are changes in Fos expression following light pulses within the vSPVZ, DMH, LC, DR, and OPT, but not within the SCN or VLPO. Importantly, only the OPT exhibited a reversal in its response to light following IGL lesions. Altogether, interconnections between the IGL and downstream brain areas such as the OPT may play a role in determining the direction of the behavioral response to light that differs across day-active and night-active individuals.

#### Acknowledgments

We thank Dorela Shuboni, Jennifer Langel, Ken Bennett, Megan Luck, and Thomas Groves for technical assistance. Supported by a National Science Foundation (NSF) grant (IOS-1051919, to L.S., A.A.N., and L.Y.) and A.J.G. was supported by a National Institutes of Health (NIH) Ruth L. Kirschstein National Research Service Award (NRSA) from the National Institute of Neurological Disorders and Stroke (NINDS) (F32 NS083360-01).

# Abbreviations

DR	dorsal raphe		
DMH	dorsomedial hypothalamus		
IGL	intergeniculate leaflet		
LP	light pulse		
LC	locus coeruleus		
OPT	olivary pretectal nucleus		
SCN	suprachiasmatic nucleus		
vSPVZ	ventral subparaventricular zone		
VLPO	ventrolateral preoptic nucleus		
ZT	Zeitgeber time		

#### References

- Redlin U. Neural basis and biological function of masking by light in mammals: suppression of melatonin and locomotor activity. Chronobiol Int. 2001; 18:737–758. [PubMed: 11763983]
- Morin LP. Nocturnal light and nocturnal rodents: similar regulation of disparate functions? J Biol Rhythms. 2013; 28:95–106. [PubMed: 23606609]
- Shuboni DD, Cramm S, Yan L, Nunez AA, Smale L. Acute behavioral responses to light and darkness in nocturnal Mus musculus and diurnal Arvicanthis niloticus. J Biol Rhythms. 2012; 27:299–307. [PubMed: 22855574]
- 4. Moore RY, Card JP. Intergeniculate leaflet: an anatomically and functionally distinct subdivision of the lateral geniculate complex. J Comp Neurol. 1994; 344:403–430. [PubMed: 8063960]
- Redlin U, Vrang N, Mrosovsky N. Enhanced masking response to light in hamsters with IGL lesions. J Comp Physiol A. 1999; 184:449–456. [PubMed: 10377979]
- Gall AJ, Smale L, Yan L, Nunez AA. Lesions of the Intergeniculate Leaflet Lead to a Reorganization in Circadian Regulation and a Reversal in Masking Responses to Photic Stimuli in the Nile Grass Rat. PLoS One. 2013; 8:e67387. [PubMed: 23840688]
- Gaillard F, Karten HJ, Sauve Y. Retinorecipient areas in the diurnal murine rodent Arvicanthis niloticus: A disproportionally large superior colliculus. J Comp Neurol. 2013; 521 Spc1.
- Morin LP, Blanchard JH. Neuropeptide Y and enkephalin immunoreactivity in retinorecipient nuclei of the hamster pretectum and thalamus. Vis Neurosci. 1997; 14:765–777. [PubMed: 9279004]
- 9. Morin LP. The circadian visual system. Brain Res Brain Res Rev. 1994; 19:102–127. [PubMed: 7909471]
- Miller AM, Miller RB, Obermeyer WH, Behan M, Benca RM. The pretectum mediates rapid eye movement sleep regulation by light. Behav Neurosci. 1999; 113:755–765. [PubMed: 10495083]
- Ibuka N, Inouye SI, Kawamura H. Analysis of sleep-wakefulness rhythms in male rats after suprachiasmatic nucleus lesions and ocular enucleation. Brain Res. 1977; 122:33–47. [PubMed: 837222]
- Li X, Gilbert J, Davis FC. Disruption of masking by hypothalamic lesions in Syrian hamsters. J Comp Physiol A Neuroethol Sens Neural Behav Physiol. 2005; 191:23–30. [PubMed: 15449094]
- Prichard JR, Stoffel RT, Quimby DL, Obermeyer WH, Benca RM, et al. Fos immunoreactivity in rat subcortical visual shell in response to illuminance changes. Neuroscience. 2002; 114:781–793. [PubMed: 12220578]
- Redlin U, Mrosovsky N. Masking by light in hamsters with SCN lesions. J Comp Physiol A. 1999; 184:439–448. [PubMed: 10377978]

Gall et al.

15. Chellappa SL, Steiner R, Blattner P, Oelhafen P, Gotz T, et al. Non-visual effects of light on melatonin, alertness and cognitive performance: can blue-enriched light keep us alert? PLoS One. 2011; 6:e16429. [PubMed: 21298068]

Page 8

- 16. Morin LP. Neuroanatomy of the extended circadian rhythm system. Exp Neurol. 2012
- 17. McElhinny, T. Masters Dissertation. Michigan State University; East Lansing: 1996. Reproductive Biology and Biological Rhythms in Arvicanthis niloticus.
- Castillo-Ruiz A, Nunez AA. Fos expression in arousal and reward areas of the brain in grass rats following induced wakefulness. Physiol Behav. 2011; 103:384–392. [PubMed: 21402088]
- Novak CM, Smale L, Nunez AA. Fos expression in the sleep-active cell group of the ventrolateral preoptic area in the diurnal murid rodent, Arvicanthis niloticus. Brain Res. 1999; 818:375–382. [PubMed: 10082823]
- Schwartz MD, Nunez AA, Smale L. Differences in the suprachiasmatic nucleus and lower subparaventricular zone of diurnal and nocturnal rodents. Neuroscience. 2004; 127:13–23. [PubMed: 15219664]
- Meijer JH, Watanabe K, Schaap J, Albus H, Detari L. Light responsiveness of the suprachiasmatic nucleus: long-term multiunit and single-unit recordings in freely moving rats. J Neurosci. 1998; 18:9078–9087. [PubMed: 9787011]
- 22. Kiessling S, Sollars PJ, Pickard GE. Light stimulates the mouse adrenal through a retinohypothalamic pathway independent of an effect on the clock in the suprachiasmatic nucleus. PLoS One. 2014; 9:e92959. [PubMed: 24658072]
- Mahoney M, Bult A, Smale L. Phase response curve and light-induced fos expression in the suprachiasmatic nucleus and adjacent hypothalamus of Arvicanthis niloticus. J Biol Rhythms. 2001; 16:149–162. [PubMed: 11302557]
- 24. Langel J, Yan L, Nunez AA, Smale L. Behavioral masking, cFos responses, and time of day in day and night active grass rats. Journal of Biological Rhythms. 2014 In press.
- 25. Sefton, AJ.; Dreher, B. Visual System. In: Paxinos, G., editor. The Rat Nervous System. 2nd ed.. Academic Press; New York: 1995. p. 833-880.
- Miller AM, Obermeyer WH, Behan M, Benca RM. The superior colliculus-pretectum mediates the direct effects of light on sleep. Proc Natl Acad Sci U S A. 1998; 95:8957–8962. [PubMed: 9671786]
- 27. Marchant EG, Morin LP. The hamster circadian rhythm system includes nuclei of the subcortical visual shell. J Neurosci. 1999; 19:10482–10493. [PubMed: 10575044]
- Shuboni, DD.; Cramm, S.; Yan, L.; Nunez, AA.; Smale, L. Society for Research in Biological Rhythms. Sandestin, FL: 2012. Masking responses and light-induced changes in Fos expression in nocturnal and diurnal rodents.
- Sherin JE, Shiromani PJ, McCarley RW, Saper CB. Activation of ventrolateral preoptic neurons during sleep. Science. 1996; 271:216–219. [PubMed: 8539624]
- Gaus SE, Strecker RE, Tate BA, Parker RA, Saper CB. Ventrolateral preoptic nucleus contains sleep-active, galaninergic neurons in multiple mammalian species. Neuroscience. 2002; 115:285– 294. [PubMed: 12401341]
- Brooks E, Waters E, Farrington L, Canal MM. Differential hypothalamic tyrosine hydroxylase distribution and activation by light in adult mice reared under different light conditions during the suckling period. Brain Struct Funct. 2011; 216:357–370. [PubMed: 21509614]
- 32. Todd WD, Gall AJ, Weiner JA, Blumberg MS. Distinct retinohypothalamic innervation patterns predict the developmental emergence of species-typical circadian phase preference in nocturnal Norway rats and diurnal nile grass rats. J Comp Neurol. 2012; 520:3277–3292. [PubMed: 22431036]
- 33. Pompeiano M, d'Ascanio P, Centini C, Pompeiano O, Balaban E. Short-term (Fos) and long-term (FRA) protein expression in rat locus coeruleus neurons during the neurolab mission: contribution of altered gravitational fields, stress, and other factors. Neuroscience. 2002; 115:111–123. [PubMed: 12401326]
- 34. Adidharma W, Leach G, Yan L. Orexinergic signaling mediates light-induced neuronal activation in the dorsal raphe nucleus. Neuroscience. 2012; 220:201–207. [PubMed: 22710065]

 Smale L, Boverhof J. The suprachiasmatic nucleus and intergeniculate leaflet of Arvicanthis niloticus, a diurnal murid rodent from East Africa. J Comp Neurol. 1999; 403:190–208. [PubMed: 9886043]

# Highlights

• Light-induced Fos expression in the SCN was unaffected by IGL lesions.

- IGL lesions resulted in a reversal in Fos responses to light in the OPT.
- Interconnections between the IGL and OPT may mediate masking in grass rats.

Gall et al.



#### Figure 1. IGL lesions did not affect the Fos response to light in the SCN

(A) Mean number of Fos positive cells in the SCN in animals sacrificed on a control night (dark) as compared to those sacrificed following a LP. \* Significantly different from control night. (B) Representative photomicrographs of Fos induction in the SCN in shams and IGL-lesioned grass rats on a control night vs. following a LP. Sampling areas are outlined in the top left photomicrograph. Scale bar =  $100\mu m$ . Abbreviations: IGL: intergeniculate leaflet; SCN: suprachiasmatic nucleus; VLPO: ventrolateral preoptic nucleus; LP: light pulse.

Gall et al.



Figure 2. IGL lesions resulted in a reversal in Fos responses to light in the OPT

(A) Mean number of Fos positive cells in the OPT in animals sacrificed on a control night (dark) vs. those sacrificed following a LP. \* Significantly different from control night. <sup>a</sup> Significant difference between sham and lesion Fos-ir values on the control night. <sup>b</sup> Significant difference between sham and lesion Fos-ir values following a LP. (B) Representative photomicrographs of Fos induction in the OPT in shams and IGL-lesioned grass rats on a control night vs. following a LP. Sampling areas are outlined in the top left photomicrograph. Scale bar = 100µm. Abbreviations: IGL: intergeniculate leaflet; OPT: olivary pretectal nucleus; LP: light pulse.

#### Table 1

Mean number of Fos positive cells in the VLPO, vSPVZ, DMH, LC, and DR.

	Sham		Lesion	
	Control Night	Light Pulse	Control Night	Light Pulse
VLPO	11.0(3.6)	29.6 (5.3)	9.0 (0.9)	21.8 (1.6)
vSPVZ	23.1 (4.7)	57.2 (3.6)	41.1 (5.4)	41.8 (8.5)
DMH	95.3 (26.8)	<b>264.6</b> (29.4) <sup>a</sup>	142.8 (19.7)	167.9 (20.9) <sup>a</sup>
LC	20.6 (4.4)	67.1 (3.8)	39.8 (7.8)	45.7 (7.3)
DR	27.1 (4.3)	<b>58.3</b> ( <b>3.0</b> ) <sup><i>a</i></sup>	39.5 (7.3)	24.1 (3.5) <sup><i>a</i></sup>

Bold values indicate significant difference from control night.

Abbreviations: VLPO: ventrolateral preoptic nucleus; vSPVZ: ventral subparaventricular zone; DMH: dorsomedial hypothalamus; LC: locus coeruleus; DR: dorsal raphe.

 $^a\mathrm{Significant}$  difference between sham and lesion Fos-ir values following a LP.