1	Brain potentials evoked by intraepidermal electrical stimuli
2	reflect the central sensitization of nociceptive pathways
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9	Running head: IES brain potentials reflect secondary hyperalgesia
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29 Abstract

30 Central sensitization (CS), the increased sensitivity of the central nervous system to somatosensory inputs, accounts for secondary hyperalgesia, a typical sign of several painful 31 32 clinical conditions. Brain potentials elicited by mechanical punctate stimulation using flat-tip 33 probes can provide neural correlates of CS, but their signal-to-noise ratio is limited by poor synchronisation of the afferent nociceptive input. Additionally, mechanical punctate 34 35 stimulation does not activate nociceptors exclusively. In contrast, low-intensity intra-36 epidermal electrical stimulation (IES) allows selective activation of type-II A $\delta$  mechano-heat 37 nociceptors (II-AMHs), and elicits reproducible brain potentials. However, it is unclear 38 whether hyperalgesia from IES occurs and co-exists with secondary mechanical punctate 39 hyperalgesia, and whether the magnitude of the EEG responses evoked by IES within the hyperalgesic area is increased. To address these questions, we explored the modulation of 40 the psychophysical and EEG responses to IES by intra-epidermal injection of capsaicin in 41 42 healthy human subjects. We obtained three main results. First, the intensity of the 43 sensation elicited by IES was significantly increased in participants who developed robust 44 mechanical punctate hyperalgesia after capsaicin injection (i.e., responders), indicating that 45 hyperalgesia from IES co-exists with punctate mechanical hyperalgesia. Second, the N2 peak magnitude of the EEG responses elicited by IES were significantly increased after the intra-46 47 epidermal injection of capsaicin in responders only. Third, a receiver-operator 48 characteristics analysis showed that the N2 peak amplitude is clearly predictive of the presence of CS. These findings suggest that the EEG responses elicited by IES reflect 49 50 secondary hyperalgesia, and therefore represent an objective correlate of CS.

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# 52 New & Noteworthy

53 Secondary mechanical punctate hyperalgesia is a cardinal sign of central sensitization (CS), 54 an important mechanism for chronic pain. Our study demonstrates that hyperalgesia from 55 intra-epidermal electrical stimulation coexists with mechanical punctate hyperalgesia and 56 elicits electroencephalographic (EEG) potentials that predict the robust occurrence of 57 punctate hyperalgesia in a human experimental model of CS. These findings inform clinical 58 development of EEG-based biomarkers of CS.

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# 62 Keywords:

63 central sensitization; secondary hyperalgesia; mechanical punctate stimulation;
64 intraepidermal electrical stimulation; EEG.

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# 67 Introduction

68 Central sensitization (CS) refers to the increased sensitivity of the central nervous system to somatosensory inputs. CS accounts for the enhanced painful percepts elicited by nociceptive 69 70 stimulation of the skin surrounding a site of tissue injury (secondary hyperalgesia) 71 (Ringkamp et al. 2013), and it has been suggested to be an important contributor to several 72 chronic pain states (Ji et al. 2003; Latremoliere and Woolf 2009). A cardinal sign of CS is 73 secondary hyperalgesia to nociceptive punctate mechanical stimuli, also known as 74 secondary mechanical punctate hyperalgesia. Such punctate stimuli, when delivered using flat-tip probes, preferentially activate the free-nerve endings of type-I A& mechano-heat 75 76 nociceptors (I-AMH) (Magerl et al. 2001). CS is typically established by an intense activation 77 of C-fibre skin nociceptors: the resulting afferent barrage to the dorsal horn results in a 78 hetero-synaptic facilitation of I-AMH inputs, which substantiates secondary mechanical 79 punctate hyperalgesia (Geber et al. 2007; Ziegler et al. 1999).

80 Secondary mechanical punctate hyperalgesia has been quantified by measuring the brain activity using non-invasive functional neuroimaging techniques, like functional magnetic 81 82 resonance imaging (fMRI) (Lee et al. 2008) and magnetoencephalography (MEG) (Maihofner 83 et al. 2010). Given that secondary hyperalgesia is a well-established surrogate model for 84 centrally generated hyperalgesia in chronic pain patients, such neural correlates have potential clinical and pharmaceutical applications. However, fMRI and MEG are costly and 85 86 not readily available. In contrast, electroencephalography (EEG) is more affordable and 87 routinely used in clinical practice. Moreover, previous studies have shown that punctate 88 stimulation causing pin-prick-like pain can elicit EEG potentials, whose amplitudes reflect 89 subjective reports of secondary mechanical punctate hyperalgesia (Davies et al. 2010;

90 lannetti et al. 2013). However, there are technical and physiological constrains that may hamper clinical translation of pinprick-evoked potentials. First, the mechanical stimulus is 91 generated by hand-held probes. The use of hand-held probes is operator dependent, which 92 93 limits reproducibility of stimulus delivery. Second, given that the force exerted is driven 94 passively by a weighted cylinder (Magerl et al. 2001), the probe needs to be held 95 perpendicularly to both the skin and the ground, in order to ensure that a consistent force is 96 applied. This limits the number of body territories that can be effectively stimulated. Pneumatically driven (Kohlloffel et al. 1991) or solenoid-powered (Davies et al. 2010) 97 98 mechanical devices have also be described: they circumvent some of the difficulties 99 associated with the use of hand-held probes. However, any device that relies on mechanical stimulation to activate cutaneous nociceptors remains limited by a crucial factor, the 100 101 variability in skin compliance. This limits the synchronicity of nociceptor activation, 102 introduces high variability of spatial and temporal summation at central synapses, and thus makes the estimation of response latency and amplitude difficult. Third, when using 103 104 mechanical probes, the spatial location of the stimulated spot is typically changed between 105 trials, which further increases the variability of the afferent nociceptive input. Lastly, and 106 most importantly, mechanical punctate stimulation activates intra-epidermal nociceptive 107 nerve endings preferentially, but not selectively. Indeed, at higher stimulus intensities the 108 dermis and subcutaneous tissues are more likely to become temporarily deformed, which 109 may result in a certain degree of activation of deeper A $\beta$  afferents (Treede et al. 2002).

110 A possible alternative to punctate stimulation is the selective activation of A $\delta$  nociceptors by 111 simple and affordable concentric electrodes that are designed to deliver currents exclusively 112 to the epidermal skin layers, where the free nerve-endings of nociceptors ramify (Inui and

113 Kakigi 2012; Inui et al. 2002). Psychophysical, behavioural and electrophysiological data 114 indicate that, when used at low-intensity of current, intra-epidermal electrical stimulation 115 (IES) activates A $\delta$  nociceptors *selectively*, i.e. without coactivating A $\beta$  afferents (Mouraux et 116 al. 2010). Still, it remains to be determined whether the psychophysical and EEG responses 117 evoked by IES are increased in the presence of secondary mechanical punctate hyperalgesia. 118 This question is physiologically pertinent: given the evidence that IES predominantly activate 119 type-II AMHs (Mouraux et al. 2010; Treede and Magerl 2000), the observation that EEG 120 responses to IES are increased would imply that hyperalgesia from IES is also mediated by 121 this class of nociceptive afferents.

122 Here, we explored whether IES evoked potentials hold promise as an objective neural 123 correlate of secondary hyperalgesia. We intra-epidermally injected capsaicin in right hand of 124 healthy subjects to induce a state of CS. Participants were classified in responders and non-125 responders based on whether or not they developed robust secondary mechanical punctate 126 hyperalgesia. We then tested (1) whether subjects who developed secondary mechanical 127 hyperalgesia also developed secondary hyperalgesia from nociceptive-specific IES. We also 128 (2) explored whether the magnitude of the EEG responses to nociceptive IES delivered to 129 the secondary hyperalgesic area was significantly increased and (3) quantified the sensitivity and specificity of the EEG responses elicited by IES for detecting the presence of secondary 130 131 hyperalgesia in our study cohort.

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# 133 Materials and Methods

#### 134 Participants

135 Fourteen healthy right-handed volunteers participated in this study. All participants were 136 pain-free, not taking any medication and did not have any history of severe allergic reactions to chilli peppers at the time of testing. They all gave signed written informed 137 138 consent, and the experimental procedures were approved by the UCL Research Ethics 139 Committee. Before the electrophysiological recording, the experimental setup and the 140 psychophysical rating task were clearly explained to the participants, who were also 141 familiarized with the sensation elicited by IES. Data from two participants were discarded 142 because no clear event-related potential (ERP) could be identified, and the data from the 143 remaining twelve participants (22-39 years, 7 female) were analysed.

#### 144 Experimental design

145 The experimental design is summarised in Figure 1 (upper panel). Experiments were 146 conducted in a silent and temperature-controlled room. Throughout the experiment 147 participants sat on a comfortable chair with the hands resting on a table in front of them. 148 Participants were instructed to keep their gaze fixed on a black cross  $(2 \times 2 \text{ cm})$  placed centrally in front of them, at a distance of 1.5 m,  $\sim 20^{\circ}$  below eye level. To induce CS, 149 150 capsaicin was injected intra-epidermally on the right hand dorsum (Ziegler et al. 1999). IES were delivered in two separate blocks, one before ('pre-capsaicin') and one after capsaicin 151 injection ('post-capsaicin'). In the post-capsaicin block, IES were delivered only after 152 153 capsaicin-induced spontaneous pain had resolved. In each block we delivered 20 stimuli on 154 the left hand dorsum and 20 stimuli on the right hand dorsum, in pseudo-random order, with an inter-stimulus interval (ISI) of 8-12 s (rectangular distribution). Therefore, there 155 were four conditions: (1) Pre-capsaicin, right hand (PreRH); (2) Pre-capsaicin, left-hand 156 157 (PreLH); (3) Post-capsaicin, right-hand (PostRH); and (4) Post-capsaicin, left-hand (PostLH).

Three seconds after the stimulus onset, subjects were asked to state whether the stimulus was delivered on the right or the left hand, and to provide ratings of the perceived intensity of pinprick pain, using a numerical scale ranging from 0 (no pinprick sensation) to 100 (the most intense pinprick sensation imaginable).

#### 162 Intra-epidermal electrical stimulation (IES)

163 IES consisted of two constant-current square-wave pulses delivered in rapid succession, as 164 described in (Inui et al. 2002; Mouraux et al. 2010). Each pulse lasted 500 µs, and the inter-165 pulse interval was 10 ms (DS7, Digitimer, UK). Stimuli were delivered using a stainless steel 166 concentric bipolar needle electrode, consisting of a needle cathode (length: 0.1 mm,  $\emptyset$ : 0.2 mm) surrounded by a cylindrical anode ( $\emptyset$ : 1.4 mm) (Inui et al. 2002; Mouraux et al. 2010). 167 168 By gently pressing the device against the skin, the needle electrode was inserted into the 169 epidermis. Two electrodes were applied, one on the dorsum of each hand. Once the 170 electrodes were fixed, the thresholds for stimulus perception were determined for each hand and each subject, using an adaptive staircase procedure. The final intensity of the IES 171 172 for the experiment was set to twice the perceptual threshold, to ensure selective 173 stimulation of skin nociceptors (Mouraux et al. 2010).

After the thresholding procedure, we delivered a few stimuli at the intensity determined above, to familiarize the participant with the elicited sensation. If the participant reported a different perceived intensity on two hands, the location of the electrodes were adjusted on each participant until the reported intensities on both hands were similar, and then the thresholding procedure was repeated and the new stimulus intensity was determined.

#### 179 Capsaicin injection

To induce CS, we injected intra-epidermally a 10 mM solution of capsaicin (40 μg in a 12.5 μl volume of normal saline containing 0.16% Tween 80; for details, see (LaMotte et al. 1991). The capsaicin solution was injected at an angle of approximately 15° to the skin surface, using a 27-gauge disposable needle. The injection site was ~1.5 cm away from the IES electrode on the right hand dorsum. Therefore, IES was delivered on the skin area of secondary hyperalgesia away from the injection site where the skin would have been numbed by the local neurotoxic effects of capsaicin (LaMotte et al. 1992).

#### 187 Capsaicin-induced spontaneous pain and secondary hyperalgesia assessment

Spontaneous pain intensity after capsaicin injection was recorded using a numerical rating scale ranging between 0 (no pain) and 100 (worst pain imaginable). Participants were required to rate verbally the intensity of spontaneous pain every 10 s during the first 3 minutes and then every 30 s until the pain intensity ratings were less than 5 out of 100.

192 The development of mechanical hyperalgesia in the skin area surrounding the injection site 193 was confirmed by punctate mechanical stimulation of the skin adjacent (within 1 cm) to the 194 external circumference of the concentric IES electrode, using a flat-tip punctate probe (256 195 mN). This probe comprises a stainless steel wire tip ( $\phi$ : 0.25 mm) attached to a mounted 196 weight (256 mN) that glides smoothly within a hollow handheld cylindrical tube. When 197 applied perpendicularly to the skin, the weight of the probe rested entirely on the wire tip, 198 thus exerting a constant force of 256 mN. More details and a depiction of the punctate 199 probe can be found in (lannetti et al. 2013), as well as in the manufacturer website (MRC 200 Systems GmbH; http://www.mrc-systems.de/en/products/pinprick). The same mechanical 201 stimulus was applied to the corresponding position of the left hand, to obtain a baseline for 202 quantifying the effect of secondary hyperalgesia, as follows. Participants were asked to

203 report the intensity of punctate stimulation of the right hand (capsaicin-injected) and of the left hand (control), using a numerical rating scale that ranged between 0 (no pinprick 204 205 sensation) and 100 (the most intense pinprick sensation imaginable). For each hand, 206 punctate stimuli were applied three times with an inter-stimulus interval of approximately 5 207 sec, after the spontaneous pain induced by the capsaicin injection in the right hand had 208 decreased to less than 5 out of 100 (Figure 2). For every individual, the mean ratings of the 209 sensations elicited by the three stimuli was obtained for each hand and condition. The 210 intensity of secondary hyperalgesia was quantified as the ratio of the subjective ratings of 211 the pinprick sensation elicited by mechanical stimulation of the right and the left hands 212 (Right/Left). Participants were considered to have developed robust secondary hyperalgesia 213 from punctate stimuli if the ratio was  $\geq 2$ , and were thus classified as responders. All other 214 participants were classified as non-responders. This ratio was chosen based on a previous 215 EEG study, which showed that an approximately two-fold increase (+93%) in pinprick 216 sensation elicited by punctate stimulation after capsaicin sensitisation was associated with 217 significant increases in the evoked EEG response (Iannetti et al. 2013).

#### 218 **EEG recording**

The EEG was recorded using 31 Ag-AgCl electrodes placed on the scalp according to the International 10-20 system, and referenced to the nose. Ocular movements and eye blinks were recorded using two surface electrodes, one placed over the right lower eyelid, the other placed approximately 1 cm lateral to the lateral corner of the right orbit. Signals were amplified and digitized using a sampling rate of 1,024 Hz (SD32, Micromed, Italy).

# 224 Behavioural data analysis

Single trial ratings of the sensation elicited by IES were first normalized between 0 and 100, for each participant (the minimum value was set to 0 and the maximum value was set to 100). This procedure mitigates the differences in the range of values on the numerical rating scale with which individuals reported the intensity of pinprick pain elicited by IES (Huang et al. 2013). Normalized stimulus intensity ratings were subsequently averaged across trials for each condition, resulting in four average values for each participant (PreRH, PreLH, PostRH, PostLH).

To test whether capsaicin injection had an effect on the perceived IES intensity, we performed a three-way ANOVA, with the following experimental factors: Group (two levels: Responders, Non-Responders), Session (two levels: Pre-capsaicin, Post-capsaicin), and Hand (two levels: Injected [Right], Control [Left]).

Where effects were significant, post-hoc analyses were performed to define their direction and possible interactions. Two-way repeated-measures ANOVA for the main and interaction effects of Session and Hand were performed to define the effects of capsaicin injection on the intensity of the sensation elicited by IES, within each group. The statistical threshold of the post hoc analyses was Bonferroni corrected accounting for the number of comparisons (P = 0.05/2 = 0.025).

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# 243 **EEG data analyses**

EEG data analyses were performed using Letswave (<u>www.nocions.org</u>) (Mouraux and lannetti 2008) and Matlab (The MathWorks, Natick, MA). Continuous EEG recordings were segmented into epochs using a time window of 2 s (-0.5 to 1.5 s relative to the stimulus 247 onset). Each epoch was baseline corrected (baseline interval ranging from -0.2 to 0 s), and band-pass filtered (1–30 Hz). Artefacts produced by eye blinks or eye movements were 248 249 subtracted using a validated method based on independent component analysis (Jung et al. 250 2000). In all data sets, independent components related to eye movements had a large 251 electrooculogram channel contribution and a frontal scalp distribution. In addition, epochs 252 with amplitude values exceeding  $\pm 100 \,\mu$ V were rejected from further analysis. These epochs 253 constituted 0.6 ±1.8% (mean ±SD across all conditions and participants) of the total number 254 of epochs. Remaining epochs were then averaged for each condition, resulting in four 255 average ERP waveforms for each participant.

256 The N2-P2 complex was measured at the vertex (Cz), and it was defined as the largest 257 negative-positive deflection occurring after stimulus onset. The amplitude of both the N2 258 and P2 peaks were calculated for each condition and participant, and tested for the effect of 259 capsaicin injection, using the same three-way ANOVA described for the behavioural data 260 (Figure 1, lower panel). As two peaks (N2 and P2) were tested, the statistical threshold, P =261 0.05/2 peaks = 0.025, was determined by Bonferroni correction accounting for the number 262 of peaks. Where effects were significant, the same post-hoc analyses described for the 263 behavioural data (i.e., two-way repeated measures ANOVA) were performed for each group, 264 and the same statistical threshold, Bonferroni corrected (P = 0.05/2 groups = 0.025), was 265 used to determine the significance of the post-hoc results. The latency of the N2 and P2 266 peaks were analysed using the same procedure.

To test the predictive value of ERP amplitude for the presence of central sensitisation, we plotted the receiver operating characteristic (ROC) curves obtained using the interaction term (i.e. (PostRH-PreRH)-(PostLH-PreLH)) calculated for the N2-wave and P2-wave peak

270 amplitudes. The true positive rate (Sensitivity) is plotted against the false positive rate (100-271 Specificity) for different cut-off values of the interaction terms. Each point on the ROC curve 272 represents a sensitivity/specificity pair corresponding to a particular decision threshold for 273 the interaction term. Above each of these thresholds the individual is predicted to be a 274 responder, and vice versa. If interaction terms had perfect classification performance, their 275 ROC curves would pass through the upper left corner (100% sensitivity, 100% specificity). 276 The closer the ROC curve is to the upper left corner, the higher the overall accuracy of the 277 interaction term is in distinguishing responders and non-responders (Zweig and Campbell 278 1993). The Area Under Curve (AUC) is typically used to quantify the classification 279 performance. An AUC value of 0.5 corresponds to a random classification (i.e. to a useless 280 test), whereas an AUC of 1.0 indicates that the test performs perfectly. We calculated the 281 AUC for the interaction terms obtained from the amplitude of the N2 and P2 peaks, to 282 assess their sensitivity and specificity for detecting the presence of a CS state. We tested 283 whether the AUC size of each measure was significantly greater than 0.5 (Hanley and McNeil 284 1982).

285

#### 286 **Results**

287 Six out of twelve participants developed robust secondary hyperalgesia on the capsaicin-

treated hand and were therefore classified as responders (Figure 2, upper panel).

### 289 Capsaicin-induced spontaneous pain

290 The time courses of the capsaicin-induced pain for all subjects are shown in the lower panel

of Figure 2. In the first few seconds after the injection, capsaicin induced a very intense

sensation of burning pain, which decreased exponentially over time (Lee et al. 2008; Magerl et al. 1998). The time course of spontaneous pain ratings for each subject was summarised as area-under-curve (AUC). The AUC for responders and non-responders were compared using a two-sample t test. The result showed no significant difference in capsaicin-induced spontaneous pain between the two groups ( $T_{10} = 0.39$ , P = 0.70). This observation suggests that both groups perceived the conditioning stimulus (i.e. the intra-epidermal injection of capsaicin) similarly.

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# 300 Psychophysics of intra-epidermal stimulation of the area of secondary mechanical 301 punctate hyperalgesia

302 All subjects correctly reported whether the IES was delivered to the right or to the left hand, 303 in all trials. The three-way ANOVA on the subjective ratings of perceived IES intensity 304 showed a two-way interaction between Group and Hand ( $F_{1.10}$  = 9.02, P = 0.01), and more importantly, a clear three-way interaction between Group, Session and Hand ( $F_{1,10}$  = 59.27, 305 306 P = 0.000016) (Figure 3). No other significant effects were detected (Table 1). This finding indicates that right hand stimulation was perceived as more painful than left hand 307 308 stimulation in the responders, but only after capsaicin was injected in the right hand. The 309 results of all post hoc two-way ANOVAs are shown in Table 2. Both responders ( $F_{1.5}$  = 49.79, 310 P = 0.001) and non-responders ( $F_{1,5} = 15.19$ , P = 0.01) showed significant interactions between Session and Hand, but in opposite directions – the responders had clearly 311 312 increased ratings on their treated hand after capsaicin injection, while the non-responders 313 showed mildly decreased ratings on their treated hand after capsaicin injection (Figure 3).

The results demonstrate a clear secondary hyperalgesia from both IES and mechanical punctate stimulation after capsaicin injection.

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#### 317 ERP waveforms

ERPs elicited by IES stimuli showed a clear N2-P2 complex maximal at electrode Cz, in all four conditions of each group. Grand-average waveforms and scalp maps at N2 and P2 peak latencies are shown in Figure 4. The ERP amplitude increased after capsaicin injection in the right hand of the responders, compared with all other conditions. Statistical comparisons of peak amplitude and latency of the N2 and P2 waves across different conditions and groups are reported below, and summarized in Tables 1 and 2.

324 N2 peak amplitude. The three-way ANOVA of N2 peak amplitudes showed a three-way 325 interaction between Group, Session and Hand ( $F_{1,10} = 7.84$ , P = 0.019). No other significant 326 effects were detected (Table 1). Hence, N2 peak amplitudes at Cz were greater following 327 right-hand IES compared to left-hand IES in the responders, but only when IES were 328 delivered to the hand where capsaicin had been injected (i.e. the right hand). Post hoc two-329 way ANOVAs (Table 2) revealed that only responders showed an interaction between 330 Session and Hand ( $F_{1,5}$  = 15.15, P = 0.011) indicating increased N2 amplitudes on their treated hand after capsaicin injection. Figure 5 shows the single-subjects N2 peak 331 332 amplitudes, as well as the statistical results.

*P2 peak amplitude.* The three-way ANOVA of P2 peak amplitudes showed that there was a two-way interaction between Group and Session ( $F_{1,10} = 11.13$ , P = 0.008). This effect was caused by an overall increased P2 amplitude in the post-capsaicin session of responders, but a decreased P2 amplitude in the post-capsaicin session of non-responders. No other significant effects were detected (Table 1). Post hoc two-way ANOVAs (Table 2) showed that there was a trend for an interaction between Session and Hand which, however, did not survive correction for multiple comparisons in responders ( $F_{1,5} = 9.77$ , P = 0.026): in this group, P2 amplitudes in the post-capsaicin session were, compared to the pre-capsaicin session, increased following right hand stimulation and slightly decreased following left hand stimulation.

N2 peak latency. The three-way ANOVA of N2 peak latencies showed a main effect of Hand ( $F_{1,10} = 7.41$ , P = 0.022). No other significant effects were detected (Table 1). Post hoc twoway ANOVAs (Table 2) fail to detect any effects in either responders or non-responders that survived correction for multiple comparisons.

*P2 peak latency.* The three-way ANOVA on the P2 peak latencies did not detect any
 significant effect. Therefore, post hoc analyses were not performed.

*ROC curves.* The ROC curves obtained from N2 and P2 peak amplitudes are plotted in Figure 6. The AUC (±standard error) for N2 and P2 were 0.92 ±0.09 and 0.72 ±0.16, respectively. Only the AUC for N2 was significantly greater than 0.5 (N2: P = 0.016; P2: P = 0.200). This suggests that the N2 peak amplitude has adequate sensitivity and specificity for detecting the presence of CS induced by intra-epidermal injection of capsaicin.

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355

# 356 **Discussion**

357 Developing a biomarker for secondary hyperalgesia, a cardinal symptom of central 358 sensitization (CS), would be useful for both drug discovery and clinical therapy. Such a 359 biomarker would help analgesic drug discovery in early phase trials, facilitate diagnosis of 360 neuropathic pain, and allow objective monitoring of drug treatments in patients.

361 IES is a technically simple and inexpensive method to selectively stimulate type II A $\delta$  skin 362 nociceptors (Inui and Kakigi 2012; Inui et al. 2002; Mouraux et al. 2010). Importantly, IES 363 elicits clear time-locked EEG responses, thus allowing quantification of CS. However, 364 mechanical punctate hyperalgesia is known to be mediated by I-AMH units, rather than II-365 AMH units (Magerl et al. 2001). Given that IES selectively activates II-AMH units (Mouraux et 366 al. 2010), we tested (1) whether secondary hyperalgesia from IES co-exists with secondary 367 mechanical punctate hyperalgesia, and (2) whether such hyperalgesia is reflected in a 368 corresponding increase in EEG responses.

369 We obtained several interesting results. First, the intensity of the sensation elicited by IES 370 was significantly increased after intra-epidermal injection of capsaicin in those participants 371 who developed robust mechanical punctate hyperalgesia – clearly showing that 372 hyperalgesia from IES occurs and coexists with mechanical hyperalgesia. Second, the peak 373 amplitude of the N2 wave elicited by IES was significantly increased in responders, similarly 374 to the intensity of the sensation elicited by IES. This increased respons only occurred when 375 IES were delivered to the hand where capsaicin was injected. Third, ROC analysis showed 376 that the N2 peak amplitude offers the ability to predict the presence of CS with high 377 sensitivity and specificity. These findings suggest that the EEG responses elicited by IES 378 reflect secondary hyperalgesia and thus are a reliable neural correlate of CS.

379

#### 380 Peripheral afferents mediating secondary hyperalgesia from IES

381 Whilst our observations clearly indicate that secondary hyperalgesia elicited by IES appears 382 to co-exist with secondary hyperalgesia elicited by mechanical punctate stimuli, it remains 383 unclear whether the two phenomena are mediated by similar populations of  $A\delta$ 384 nociceptors. There is strong physiological evidence that secondary mechanical punctate 385 hyperalgesia is mediated by I-AMH nociceptors. For example, Magerl and colleagues (2001) 386 demonstrated that secondary mechanical punctate hyperalgesia still occurs in skin that was 387 rendered devoid of II-AMH epidermal terminals by application of high concentrations of 388 topical capsaicin (Magerl et al. 2001). In contrast, Mouraux and colleagues (2010) showed 389 that both sensations and EEG responses elicited by IES were abolished in skin that was 390 similarly treated with high-concentration capsaicin, suggesting that IES activates mostly II-391 AMH nociceptors (Mouraux et al. 2010). It follows that the secondary hyperalgesia from IES 392 observed in this study is likely to be mediated mainly by II-AMH, rather than I-AMH 393 nociceptors. However, further experiments are required to confirm whether hyperalgesia 394 from IES and mechanical punctate stimulation are truly mediated by different populations of 395 Aδ afferents. Nonetheless, it is plausible that, after capsaicin injection, inputs from both I-396 AMH and II-AMH nociceptors are heterosynaptically facilitated via a common central 397 mechanism, and account for the co-existence of secondary hyperalgesia from IES and 398 mechanical punctate stimulation (Ziegler et al. 1999).

399 Variability in capsaicin-induced secondary hyperalgesia

We observed considerable variability in the degree of punctate hyperalgesia that developed
 after intra-epidermal capsaicin injection. Only half of the subjects developed robust

402 punctate hyperalgesia (i.e. a two-fold increase of pain ratings when stimulating the injected
403 hand with respect to the control hand; Figure 2).

404 It is unlikely that this difference between responders and non-responders was related to the 405 strength of conditioning stimulus, i.e., the activation of C-nociceptors by intra-epidermal 406 injection of capsaicin. Indeed, both groups reported similar intensities and durations of 407 burning pain following intra-epidermal injection of capsaicin, which suggests that the 408 conditioning stimulus was similar for both groups. We note that the development of 409 secondary hyperalgesia can be highly variable even with a highly standardized electrical 410 conditioning stimulus, which suggests considerable differences in the development of CS 411 responses between individuals (Pfau et al. 2011). Furthermore, there is clear evidence that 412 genetic variability contributes to variability in hyperalgesic response following intra-413 epidermal capsaicin injection (Tegeder et al. 2008).

#### 414 Brain potentials evoked by IES and central sensitization: advantages and limitations

Previous studies have suggested that brain potentials elicited by punctate mechanical stimulation may be recorded and employed as a potential objective correlates of the CS states (Davies et al. 2010; Iannetti et al. 2013; Kohlloffel et al. 1991). However, as detailed in the Introduction, evoked potentials elicited by punctate mechanical stimuli have significant technical and physiological constrains that may hamper clinical translation.

In contrast, IES have several advantages over mechanical punctate stimulation. When delivered at low currents, they are fully selective for A $\delta$  nociceptors, and allow for accurate timing and standardization of stimuli. The stimulating electrode is affordable and can be affixed to any part of the body without difficulty.

424 The current results show that the amplitude of the ERP elicited by IES of the skin with secondary hyperalgesia clearly reflects that the somatosensory system is centrally 425 426 sensitised. The amplitude of the N2 wave was significantly larger when IES were delivered to 427 the hand in which capsaic in injection resulted in a clear secondary hyperalgesia (Figures 4 428 and 5, Tables 1 and 2). Moreover, the areas under the ROC curves indicate that the change 429 in N2 peak amplitude was significantly predictive of the presence of secondary hyperalgesia 430 (Figure 6). This result suggests that the changes in N2 amplitude may be developed as a 431 potentially useful biomarker of CS.

432 Several limitations to IES remain. First, we were unable to isolate the early, contralateral N1 433 wave typically observed in the brain potentials evoked by nociceptive laser stimuli (Treede 434 et al. 1988; Valentini et al. 2012), most likely because of its lower signal-to-noise ratio. 435 Compared to the subsequent N2-P2 complex, the N1 wave has been shown to better reflect the afferent nociceptive drive (Lee et al. 2009) and appears less susceptible to top-down 436 437 modulation, for example placebo manipulation (Martini et al. 2015). These characteristics 438 make the N1 wave a potentially more robust marker for central sensitisation. Second, the 439 selective activation of A $\delta$  nociceptors by IES relies on the use of strictly low-intensity 440 currents. This limitation prevents recording stimulus response functions, as higher-intensity 441 currents necessarily entail a coactivation of tactile AB afferents, and therefore a loss of 442 specificity for A $\delta$  fibre stimulation (Mouraux et al. 2010). Stimulus response functions are 443 particularly useful when assessing the analgesic potential of novel drugs as they can divulge 444 interactions between stimulus or pain intensity and dose effects. Recording of stimulus 445 response function using the brain response elicited by mechanical punctate stimuli is similarly problematic because, as detailed earlier, when high forces are exerted the 446

447 mechanical punctate stimulus becomes less selective for A\delta fibre activation (Treede et al. 448 2002; van den Broeke et al. 2015). More recent data reveal that stimulus response functions can be constructed using IES, by varying the number of pulses delivered in quick succession 449 450 (5 ms intervals) to normal skin – increasing the number of pulses increases the intensity of 451 sensation and EEG amplitudes without changing reaction times or response latencies 452 (Mouraux et al. 2014). Further experiments are required to ascertain if this remains the case 453 after capsaicin-induced hyperalgesia. Moreover, although our present results suggest the 454 potential usefulness of EEG responses to IES as an objective measure of CS, the small sample 455 size used in the present study limits statistical power for detection of smaller effects. Future 456 studies with large samples are needed to confirm the predictive value of IES brain potentials 457 for the state of CS.

458 Conclusion

Our study demonstrates that secondary hyperalgesia to IES occurs in a well-recognized experimental model of CS, and that the subjective report was corroborated by increased evoked EEG responses. These findings suggest that EEG responses elicited by low-intensity IES, particularly the change in the peak amplitude of the N2 wave, can be used as an objective, physiological correlate of secondary hyperalgesia. Hence, IES evoked potentials hold promise as a low-cost non-invasive biomarker for CS that can be translated for clinical use with relative ease compared to existing techniques.

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549

# 550 Figure legends

551 Figure 1. Upper panel. Experimental design. The state of central sensitization was induced 552 by intra-epidermal injection of capsaicin (red arrow on the timeline). Capsaicin-induced 553 spontaneous pain lasted between 8 and 35 minutes, during which pain ratings were collected every 10 s during the first 3 minutes and then every 30 s until the pain intensity 554 555 ratings were less than 5 out of 100 (red box). Psychophysical and EEG responses to IES were 556 collected before capsaicin injection (i.e., pre-capsaicin session, green box) and after 557 capsaicin induced spontaneous pain had disappeared (i.e., post-capsaicin session, blue box). 558 The development of secondary hyperalgesia to punctate mechanical stimuli was assessed by 559 the ratio of the subjective intensity ratings of the sensation evoked by stimulation of the 560 right and the left hand (Right/Left) (purple arrow on the timeline). Participants were 561 considered responders if the ratio was  $\geq 2$ , and non-responders otherwise. Lower panel. 562 Schematic of the statistical analysis. A three-way ANOVA, with the factors of Group 563 (responders, non-responders), Session (pre-capsaicin, post-capsaicin) and Hand (left, right), 564 was used to analyse both psychophysical and ERP responses. The three-way interaction 565 (Group x Session x Hand) indicated the effect of central sensitization on these responses. 566 Further *post hoc* two-way ANOVAs with the factors of Session and Hand were performed to 567 define the effect within each group.

568

**Figure 2**. *Upper panel*. Participants were divided into two groups according to the ratio of probe ratings to punctate stimulation of the right and left hands: participants who rated the intensity of right hand stimulation as at least twice that of the left hand stimulation were

classified as responders. Participants were sorted by the ratio of reported intensity ratings, in descending order. *Lower panel*. Time course of capsaicin-induced pain ratings. Single participants are colour coded. Solid lines indicate responders. All participants rated the pain intensity between 90 and 100 at the moment of the injection. Pain ratings decreased fairly quickly over time. *Upper-right inset*. The comparison of the mean area under curve (AUC) between responders and non-responders revealed no significant difference ( $T_{10} = 0.39$ , P =0.70). Coloured symbols indicate single-subject AUC data.

579

580 Figure 3. Subjective intensity ratings of the sensation elicited by the IES (Intra-epidermal 581 Electrical Stimulation) of responders (left column) and non-responders (right column). 582 Upper panel: to highlight the interaction between the factors Session and Hand, the 583 subtracted ratings (Post minus Pre capsaicin injection) are shown for each hand. Colored 584 dots indicate single subjects, and the black dots indicate the group average of each 585 condition. Two-way ANOVA revealed that responders had a highly significant interaction between the factors Session and Hand. This reveals a capsaicin-induced increase of IES 586 587 ratings (Post-Pre) on the right hand. In contrast, in non-responders the two-way ANOVA 588 revealed a decrease of IES ratings on the right hand compared to those on the left hand. 589 These differences in the capsaicin effect on IES ratings between responders and nonresponders were confirmed by the three-way ANOVA, which revealed a highly significant 590 591 triple interaction (Group x Session x Hand; the comparison between the left and the right 592 columns). LH: left hand; RH: right hand. Lower panel: individual values (colored dots) and 593 mean value (the black dots) of each condition. PreLH: Pre-capsaicin, left hand; PostLH: Post-594 capsaicin, left hand; PreRH: Pre-capsaicin, right hand; PostRH: Post-capsaicin, right hand.

596 Figure 4. Group-average ERP waveforms and scalp maps elicited by IES in responders (left 597 panel) and non-responders (right panel). Waveforms at the channel Cz in different 598 conditions are shown in different color. The ERP elicited by IES stimuli clearly increased after 599 capsaicin injection only on the right hand in responders. Scalp maps at the N2 peak latencies 600 show a central distribution, slightly lateralized to the hemisphere contralateral to the 601 stimulated hand, maximal at the vertex (upper part of each panel). Scalp maps at the P2 602 peak latencies show a central distribution, maximal at the vertex (lower part of each panel). 603 The color bar shows the ERP amplitude in scalp maps.

604

605 Figure 5. ERP amplitudes (N2) of IES of responders (left column) and non-responders (right 606 column). Upper panel: to highlight the interaction between Session and Hand in each group, 607 the subtracted ERP amplitudes (Post *minus* Pre capsaicin injection) are shown for each hand. 608 Colored dots indicate single subjects, and the black dots indicate the group average of each 609 condition. Two-way ANOVA revealed that responders had a significant interaction between 610 the factors Session and Hand. This reveals a capsaicin-induced increase of IES ERP 611 amplitudes (Post-Pre) on the right hand. In contrast, in non-responders the two-way ANOVA 612 did not show any significant effect. These differences in the capsaicin effect on ERP 613 amplitudes between responders and non-responders were confirmed by the three-way ANOVA, which revealed a significant triple interaction (Group x Session x Hand; the 614 615 comparison between the left and the right columns). LH: left hand; RH: right hand. Lower 616 panel: individual values (colored dots) and mean value (the black dot) of each condition.

PreLH: Pre-capsaicin, left hand; PostLH: Post-capsaicin, left hand; PreRH: Pre-capsaicin, right
hand; PostRH: Post-capsaicin, right hand.

620	Figure 6. ROC curves and their corresponding area under curve (AUC) obtained using the
621	interaction term for N2 peak amplitude (left panel) and P2 peak amplitude (right panel) as
622	the predictive factor. Although both measures show predictive ability, only the AUC of N2
623	ROC was significantly greater than 0.5, indicating that it is therefore a predictor for the state
624	of central sensitisation.

# **Table 1.** Results of the three-way ANOVA of the psychophysical and EEG responses elicited

626 by intraepidermal electrical stimulation	(IES).
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3-way ANOVA	Pain intensity	ERP peak	amplitude	ERP peak latency	
	ratings	N2	P2	N2	P2
Main effect of	$F_{1,10} = 0.20$	$F_{1,10} = 0.08$	<i>F</i> <sub>1,10</sub> = 0.15	$F_{1,10} = 0.008$	<i>F</i> <sub>1,10</sub> = 1.51
Group	<i>P</i> = 0.665	<i>P</i> = 0.778	<i>P</i> = 0.705	<i>P</i> = 0.930	<i>P</i> = 0.247
Main effect of	$F_{1,10} = 0.05$	$F_{1,10} = 0.26$	<i>F</i> <sub>1,10</sub> = 2.65	$F_{1,10} = 0.50$	$F_{1,10} = 0.38$
Session	<i>P</i> = 0.833	<i>P</i> = 0.618	<i>P</i> = 0.134	<i>P</i> = 0.498	<i>P</i> = 0.553
Main effect of	$F_{1,10} = 4.52$	$F_{1,10} = 1.60$	$F_{1,10} = 0.52$	$F_{1,10} = 7.41$	$F_{1,10} = 0.11$
Hand	<i>P</i> = 0.059	<i>P</i> = 0.234	<i>P</i> = 0.487	<i>P</i> = 0.022	<i>P</i> = 0.742
2-way interaction	$F_{1,10} = 3.04$	$F_{1,10} = 1.40$	<i>F</i> <sub>1,10</sub> = 11.13	$F_{1,10} = 0.05$	<i>F</i> <sub>1,10</sub> = 0.53
Group x Session	<i>P</i> = 0.112	<i>P</i> = 0.265	<i>P</i> = 0.008	<i>P</i> = 0.827	<i>P</i> = 0.484
2-way interaction	$F_{1,10} = 9.02$	$F_{1,10} = 0.45$	<i>F</i> <sub>1,10</sub> = 1.42	$F_{1,10} = 0.11$	$F_{1,10} = 0.0003$
Group x Hand	<i>P</i> = 0.013	<i>P</i> = 0.517	<i>P</i> = 0.261	<i>P</i> = 0.751	<i>P</i> = 0.987
2-way interaction	$F_{1,10} = 4.31$	$F_{1,10} = 0.19$	<i>F</i> <sub>1,10</sub> = 1.27	$F_{1,10} = 4.33$	<i>F</i> <sub>1,10</sub> = 1.19
Session x Hand	<i>P</i> = 0.065	<i>P</i> = 0.674	<i>P</i> = 0.286	<i>P</i> = 0.064	<i>P</i> = 0.301
3-way interaction	<i>F</i> <sub>1,10</sub> = 59.27	<i>F</i> <sub>1,10</sub> = 7.84	<i>F</i> <sub>1,10</sub> = 2.04	$F_{1,10} = 0.37$	$F_{1,10} = 0.06$
Group x Session x	<i>P</i> = 0.000016	<i>P</i> = 0.019	<i>P</i> = 0.184	<i>P</i> = 0.559	<i>P</i> = 0.813
Hand					

627 Significant effects are highlighted in bold.

- 628 Table 2. Psychophysical and EEG responses elicited by IES for each condition, and results of
- the post hoc two-way ANOVAs for each group.

	Group	PreRH	PostRH	PreLH	PostLH	Main Effect of	Main effect of	Interaction		
$\pm$ SD) $\pm$ SD) $\pm$ SD) $\pm$ SD) $(Pre vs. Post)$ $(LH vs. RH)$ Pain intensity ratingsResponders39.073.341.529.0 $F_{1.5} = 0.87$ $F_{1.5} = 11.59$ $F_{1.5} = 49.79$ $\pm$ 21.3 $\pm$ 15.9 $\pm$ 27.1 $\pm$ 16.3 $P = 0.394$ $P = 0.019$ $P = 0.009$ Non-61.534.151.651.1 $F_{1.5} = 2.90$ $F_{1.5} = 0.44$ $F_{1.5} = 15.19$ Responders $\pm$ 15.7 $\pm$ 20.0 $\pm$ 23.6 $\pm$ 25.3 $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplitudesResponders-8.6-14.0-9.9-8.9 $F_{1.5} = 1.66$ $F_{1.5} = 2.55$ $F_{1.5} = 1.515$ Non12.9-9.7-10.0-11.5 $F_{1.5} = 0.204$ $P = 0.171$ $P = 0.012$ Non12.9-9.7-10.0-11.5 $F_{1.5} = 0.20$ $F_{1.5} = 0.14$ $F_{1.5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders10.68.37.6 $F_{1.5} = 4.32$ $F_{1.5} = 1.87$ $F_{1.5} = 9.77$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1.5} = 0.48$ $P = 0.723$ $P = 0.875$ $P = 0.875$ Non-10.77.410.87.9 $F_{1.5} = 0.42$ $P = 0.755$ $P = 0.875$ Non-10.7 </td <td>Group</td> <td>FIERI</td> <td>FOSLINT</td> <td>FIELII</td> <td>FUSILIT</td> <td></td> <td></td> <td>meraction</td>	Group	FIERI	FOSLINT	FIELII	FUSILIT			meraction		
Pain intensity ratingsPain intensity ratingsResponders39.073.341.529.0 $F_{1,5} = 0.87$ $F_{1,5} = 11.59$ $F_{1,5} = 49.79$ $\pm 21.3$ $\pm 15.9$ $\pm 27.1$ $\pm 16.3$ $P = 0.394$ $P = 0.019$ $P = 0.0009$ Non- $61.5$ $34.1$ $51.6$ $51.1$ $F_{1,5} = 2.90$ $F_{1,5} = 0.44$ $F_{1,5} = 15.19$ Responders $\pm 15.7$ $\pm 20.0$ $\pm 23.6$ $\pm 25.3$ $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplitudes $23.6$ $\pm 25.3$ $P = 0.150$ $P = 0.535$ $P = 0.011$ Non- $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non- $-12.9$ $-9.7$ $-10.0$ $-11.5$ $F_{1.5} = 0.20$ $F_{1.5} = 0.14$ $F_{1.5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders $7.9$ $10.6$ $8.3$ $7.6$ $F_{1.5} = 4.32$ $F_{1.5} = 1.87$ $F_{1.5} = 9.77$ P2 peak amplitudesResponders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.735$ $P = 0.026$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{1.5} = 7.42$ $F_{1.5} = 0.11$ $F_{1.5} = 0.03$ Non- $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latency $=165$ $136$ <		(Mean	(Mean	(Mean	(Mean	Session	Hand	Session x Hand		
Responders39.073.341.529.0 $F_{1,5} = 0.87$ $F_{1,5} = 11.59$ $F_{1,5} = 49.79$ Non- $\pm 15.9$ $\pm 7.1$ $\pm 16.3$ $P = 0.394$ $P = 0.019$ $P = 0.009$ Non- $51.5$ $34.1$ $51.6$ $51.1$ $F_{1,5} = 2.90$ $F_{1,5} = 0.44$ $F_{1,5} = 15.19$ Responders $\pm 15.7$ $\pm 20.0$ $\pm 23.6$ $\pm 25.3$ $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplitudeResponders $-8.6$ $-14.0$ $-9.9$ $-8.9$ $F_{1,5} = 1.66$ $F_{1,5} = 2.55$ $F_{1,5} = 15.15$ Non- $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non- $-12.9$ $-9.7$ $-10.0$ $-11.5$ $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.254$ $P = 0.723$ $P = 0.250$ P2 peak amplitudeResponders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudeResponders $1.9$ $10.6$ $8.3$ $7.6$ $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ Responders $1.9$ $10.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Non- $10.7$ $7.4$ $10.8$		±SD)	±SD)	±SD)	±SD)	(Pre vs. Post)	(LH vs. RH)			
$\pm 21.3$ $\pm 15.9$ $\pm 27.1$ $\pm 16.3$ $P = 0.394$ $P = 0.019$ $P = 0.009$ Non- $61.5$ $34.1$ $51.6$ $51.1$ $F_{L,5} = 2.90$ $F_{L,5} = 0.44$ $F_{L,5} = 15.19$ Responders $\pm 15.7$ $\pm 20.0$ $\pm 23.6$ $\pm 25.3$ $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplitudesResponders $-8.6$ $-14.0$ $-9.9$ $-8.9$ $F_{L,5} = 1.66$ $F_{L,5} = 2.55$ $F_{L,5} = 15.15$ $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non- $-12.9$ $-9.7$ $-10.0$ $-11.5$ $F_{L,5} = 0.20$ $F_{L,5} = 0.14$ $F_{L,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders $7.9$ $10.6$ $8.3$ $7.6$ $F_{L,5} = 4.32$ $F_{L,5} = 1.87$ $F_{L,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{L,5} = 7.42$ $F_{L,5} = 0.11$ $F_{L,5} = 0.032$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{L,5} = 0.68$ $F_{L,5} = 8.75$ $P_{L,5} = 5.94$ Non- $165$ $136$ $168$ $178$ $F_{L,5} = 0.68$ $F_{L,5} = 8.75$ $F_{L,5} = 5.94$ Non- $162$ $147$ $169$ $175$ $F_{L,5} = 0.08$ $F_{L,5} = 1.95$ $F_{L,5} = 0.78$ <td>Pain intensity</td> <td colspan="9">Pain intensity ratings</td>	Pain intensity	Pain intensity ratings								
Non-61.534.151.651.1 $F_{1,5} = 2.90$ $F_{1,5} = 0.44$ $F_{1,5} = 15.19$ Responders±15.7±20.0±23.6±25.3 $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplitudesResponders-8.6-14.0-9.9-8.9 $F_{1,5} = 1.66$ $F_{1,5} = 2.55$ $F_{1,5} = 15.15$ Non-±4.4±6.4±5.4±5.1 $P = 0.224$ $P = 0.171$ $P = 0.012$ Non12.9-9.7-10.0-11.5 $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders±5.5±5.1±6.2±2.1 $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders7.910.68.37.6 $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $12.3$ ±2.5±3.5±1.4 $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders±3.6±4.2±3.2±3.8 $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latentResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	39.0	73.3	41.5	29.0	$F_{1,5} = 0.87$	<i>F</i> <sub>1,5</sub> = 11.59	<i>F</i> <sub>1,5</sub> = 49.79		
Responders $\pm 15.7$ $\pm 20.0$ $\pm 23.6$ $\pm 25.3$ $P = 0.150$ $P = 0.535$ $P = 0.011$ N2 peak amplituesResponders $-8.6$ $-14.0$ $-9.9$ $-8.9$ $F_{1,5} = 1.66$ $F_{1,5} = 2.55$ $F_{1,5} = 15.15$ $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non- $-12.9$ $-9.7$ $-10.0$ $-11.5$ $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplituesResponders $7.9$ $10.6$ $8.3$ $7.6$ $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $10.7$ $7.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $P = 0.230$ $P = 0.026$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $P = 0.755$ $P = 0.875$ N2 peak later $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak later $126$ $136$ $168$ $178$ $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $229$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non- $162$ $147$ $169$ $175$ $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$ <td></td> <td>±21.3</td> <td>±15.9</td> <td>±27.1</td> <td>±16.3</td> <td><i>P</i> = 0.394</td> <td><i>P</i> = 0.019</td> <td><i>P</i> = 0.0009</td>		±21.3	±15.9	±27.1	±16.3	<i>P</i> = 0.394	<i>P</i> = 0.019	<i>P</i> = 0.0009		
N2 peak amplitudesInterpretationInterpretationInterpretationInterpretationResponders-8.6-14.0-9.9-8.9 $F_{1,5} = 1.66$ $F_{1,5} = 2.55$ $F_{1,5} = 15.15$ $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non12.9-9.7-10.0-11.5 $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders7.910.6 $8.3$ 7.6 $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Non-	61.5	34.1	51.6	51.1	$F_{1,5} = 2.90$	$F_{1,5} = 0.44$	<i>F</i> <sub>1,5</sub> = 15.19		
Responders-8.6-14.0-9.9-8.9 $F_{1,5} = 1.66$ $F_{1,5} = 2.55$ $F_{1,5} = 15.15$ $\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non12.9-9.7-10.0-11.5 $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders7.910.6 $8.3$ 7.6 $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	±15.7	±20.0	±23.6	±25.3	<i>P</i> = 0.150	<i>P</i> = 0.535	<i>P</i> = 0.011		
$\pm 4.4$ $\pm 6.4$ $\pm 5.4$ $\pm 5.1$ $P = 0.254$ $P = 0.171$ $P = 0.012$ Non- $-12.9$ $-9.7$ $-10.0$ $-11.5$ $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders $7.9$ $10.6$ $8.3$ $7.6$ $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non- $10.7$ $7.4$ $10.8$ $7.9$ $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders $165$ $136$ $168$ $178$ $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non- $162$ $147$ $169$ $175$ $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	N2 peak ampl	N2 peak amplitudes								
Non12.9-9.7-10.0-11.5 $F_{1,5} = 0.20$ $F_{1,5} = 0.14$ $F_{1,5} = 1.69$ Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders7.910.6 $8.3$ 7.6 $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ N2 peak latency $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latency165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ Non-162147169175 $F_{1,5} = 0.08$ $F_{3,5} = 1.95$ $F_{3,5} = 0.78$	Responders	-8.6	-14.0	-9.9	-8.9	$F_{1,5} = 1.66$	$F_{1,5} = 2.55$	<i>F</i> <sub>1,5</sub> = 15.15		
Responders $\pm 5.5$ $\pm 5.1$ $\pm 6.2$ $\pm 2.1$ $P = 0.676$ $P = 0.723$ $P = 0.250$ P2 peak amplitudesResponders $7.9$ 10.6 $8.3$ $7.6$ $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.7 $7.4$ 10.8 $7.9$ $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$		±4.4	±6.4	±5.4	±5.1	<i>P</i> = 0.254	<i>P</i> = 0.171	<i>P</i> = 0.012		
P2 peak amplitudesInitial of the second	Non-	-12.9	-9.7	-10.0	-11.5	$F_{1,5} = 0.20$	$F_{1,5} = 0.14$	<i>F</i> <sub>1,5</sub> = 1.69		
Responders7.910.68.37.6 $F_{1,5} = 4.32$ $F_{1,5} = 1.87$ $F_{1,5} = 9.77$ $\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	±5.5	±5.1	±6.2	±2.1	<i>P</i> = 0.676	<i>P</i> = 0.723	<i>P</i> = 0.250		
$\pm 2.3$ $\pm 2.5$ $\pm 3.5$ $\pm 1.4$ $P = 0.092$ $P = 0.230$ $P = 0.026$ Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	P2 peak amplitudes									
Non-10.77.410.87.9 $F_{1,5} = 7.42$ $F_{1,5} = 0.11$ $F_{1,5} = 0.03$ Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	7.9	10.6	8.3	7.6	$F_{1,5} = 4.32$	<i>F</i> <sub>1,5</sub> = 1.87	<i>F</i> <sub>1,5</sub> = 9.77		
Responders $\pm 3.6$ $\pm 4.2$ $\pm 3.2$ $\pm 3.8$ $P = 0.042$ $P = 0.755$ $P = 0.875$ N2 peak latencyResponders165136168178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-162147169175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$		±2.3	±2.5	±3.5	±1.4	<i>P</i> = 0.092	<i>P</i> = 0.230	<i>P</i> = 0.026		
N2 peak latency         Instrume	Non-	10.7	7.4	10.8	7.9	$F_{1,5} = 7.42$	$F_{1,5} = 0.11$	<i>F</i> <sub>1,5</sub> = 0.03		
Responders         165         136         168         178 $F_{1,5} = 0.68$ $F_{1,5} = 8.75$ $F_{1,5} = 5.94$ $\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-         162         147         169         175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	±3.6	±4.2	±3.2	±3.8	<i>P</i> = 0.042	<i>P</i> = 0.755	<i>P</i> = 0.875		
$\pm 29$ $\pm 16$ $\pm 29$ $\pm 47$ $P = 0.447$ $P = 0.032$ $P = 0.059$ Non-       162       147       169       175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	N2 peak latency									
Non-         162         147         169         175 $F_{1,5} = 0.08$ $F_{1,5} = 1.95$ $F_{1,5} = 0.78$	Responders	165	136	168	178	$F_{1,5} = 0.68$	<i>F</i> <sub>1,5</sub> = 8.75	$F_{1,5} = 5.94$		
		±29	±16	±29	±47	<i>P</i> = 0.447	<i>P</i> = 0.032	<i>P</i> = 0.059		
Responders         ±38         ±47         ±44         ±39         P = 0.783         P = 0.221         P = 0.417	Non-	162	147	169	175	$F_{1,5} = 0.08$	$F_{1,5} = 1.95$	$F_{1,5} = 0.78$		
	Responders	±38	±47	±44	±39	<i>P</i> = 0.783	<i>P</i> = 0.221	<i>P</i> = 0.417		

630 Significant effects are highlighted in bold.











