

# Prospects & Overviews

## Cross-talk between circadian clocks, sleep-wake cycles, and metabolic networks: Dispelling the darkness

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Integration of knowledge concerning circadian rhythms, metabolic networks, and sleep-wake cycles is imperative for unraveling the mysteries of biological cycles and their underlying mechanisms. During the last decade, enormous progress in circadian biology research has provided a plethora of new insights into the molecular architecture of circadian clocks. However, the recent identification of autonomous redox oscillations in cells has expanded our view of the clockwork beyond conventional transcription/translation feedback loop models, which have been dominant since the first circadian period mutants were identified in fruit fly. Consequently, non-transcriptional timekeeping mechanisms have been proposed, and the antioxidant peroxiredoxin proteins have been identified as conserved markers for 24-hour rhythms. Here, we review recent advances in our understanding of interdependencies amongst circadian rhythms, sleep homeostasis, redox cycles, and other cellular metabolic networks. We speculate that systems-level investigations implementing integrated multi-omics approaches could provide novel mechanistic insights into the connectivity between daily cycles and metabolic systems.

### Keywords:

circadian rhythms; metabolic networks; non-transcriptional oscillator; peroxiredoxin; redox regulation; sleep-wake cycle; systems biology

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### Introduction

Circadian (approx. 24 hour) clocks are believed to exist at almost all levels of life and play a key role in the maintenance of physiological and behavioral processes in accordance with the day/night cycle [1, 2]. Similarly, sleep is thought to be a critical process in higher organisms [3]. However, our view of sleep as being a product of the brain, or even neuronal populations, may obscure underlying principles and function of sleep. For example, in mammals, sleep is invariably measured using electroencephalograms (EEGs), which may not be the best way to characterize or quantify sleep in the molecular era. This is particularly pertinent in organisms such as the fruit fly, in which electrical recordings are correlated with behavioral activity [4], but not yet clearly to sleep-wake cycles. However, it may equally be the case in mammals, in which wide ranging changes in gene expression are seen in the livers of sleep-deprived mice [5], and in the responsiveness of adipose tissue to insulin signaling in humans under sleep restriction [6]. The circadian clockwork and sleep-wake cycles closely interact with each other, which is most obviously seen by the gating of sleep at distinct phases of the 24-hour cycle. Thus, artificially separating these cycles may obscure underlying principles that unite both phenomena.

Adequate sleep is an essential requirement for health. However, a significant proportion of the adult population suffers from trouble sleeping at night, and staying awake during the daytime, most likely due to the aberrations in the switching mechanism that controls transitions between wake and sleep [7]. Understanding the neurobiological

### Abbreviations:

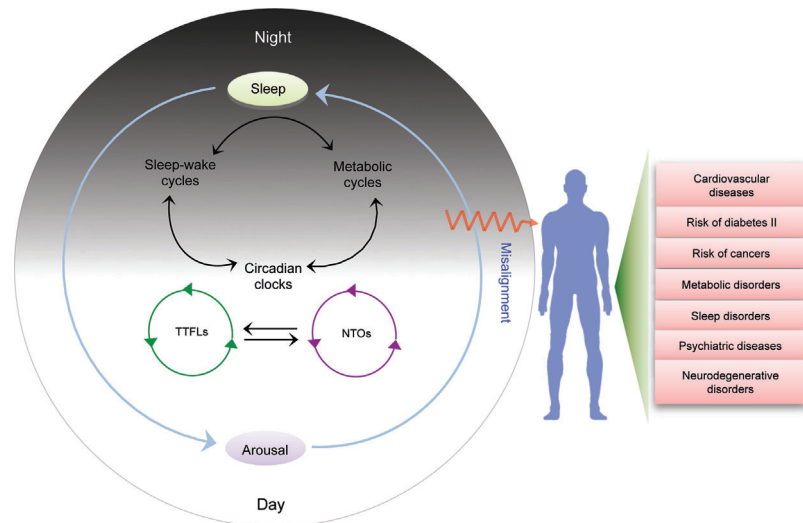
**CLOCK**, circadian locomotor output cycles kaput; **EEG**, electroencephalograms; **NTO**, non-transcriptional oscillator; **PRX**, peroxiredoxin; **RBC**, red blood cell; **SCN**, suprachiasmatic nucleus; **TTFL**, transcription-translation feedback loop.

mechanisms underlying the circadian system and sleep, and their interconnectivity, thus has profound implications for translational healthcare research, since circadian misalignment or aberrations in sleep homeostasis through old age, neurological diseases, and even shift work, are a rising cause of considerable morbidity [8–11]. Moreover, several studies indicate an intimate association of circadian dysfunction and sleep disruption with different human diseases including cancers, heart disease, diabetes, metabolic, vascular, and mental disorders (reviewed in [12–14]).

It is clear that there is a rhythmic pattern in cell function and cycles of energy utilization in accordance with a daily rhythm [15, 16], while sleep plays a crucial role in maintaining metabolic homeostasis [17]. However, the mechanism of bidirectional communication between the sleep centers and the circadian pacemaker, and their regulation of diverse metabolic networks is unclear (Fig. 1). In this article, we will outline the recent advances in our understanding of interdependencies amongst the circadian rhythms, sleep homeostasis, redox cycles, and other cellular energy metabolism networks. Potential applications of systems-level investigations, applying integrated multi-omics approaches to unravel cross-talk between day/night cycles and metabolic systems will also be discussed.

## Beyond transcription/translation feedback loop (TTFL) mechanisms of the clockwork

Since the 1980s, transcriptional/translational feedback loops (TTFLs), wherein rhythmicity in the expression patterns of specific genes are controlled by the periodic expression of “clock” gene products, were considered as the principal drivers of circadian periodicity in multiple biological systems [1, 2, 18, 19]. TTFL models indicate the presence of both positive and negative components in circadian clocks, where the positive loops activate transcription, while the negative elements inhibit the positive components in a cyclic manner. In the mammalian clock, BMAL1 and CLOCK proteins serve as the positive elements by forming a heterodimeric transcription factor complex that promotes expression of members of the *Period* (*Per*) and *Cryptochrome* (*Cry*) families [20]. Subsequently, after entering the nucleus, the PER-CRY heterodimers inhibit their own transcription by repressing the activity of the BMAL1-CLOCK complex [21, 22]. Eventually, a drop in the level of PERs and CRYs de-represses BMAL1-CLOCK activity to initiate a new cycle.



**Figure 1.** Interplay among circadian rhythmicity, metabolic cascades and sleep homeostasis: some opaque zones in circadian biology research. Cross-talk among the circadian oscillators, sleep-wake cycle and metabolic oscillations are important components of daily biological time-keeping mechanisms. However, the precise mechanisms by which sleep-wake centers, circadian clocks, and metabolic pathways communicate with each other have not been clearly demarcated. Interaction between non-transcriptional oscillators (NTOs) and transcription-translation feedback loop (TTFL)-based oscillators is also largely unclear. Circadian or sleep disturbance, and misalignments between their phasing can lead to different types of diseases and disorders, most notably metabolic derangements.

Information obtained through the TTFL-based models is undoubtedly essential for understanding various aspects of circadian rhythms, and in particular tissue functions, since transcriptome and proteome alterations are extensive and cyclical in a range of studies [23–30]. However, in the past decade, identification of molecular rhythmicity in both prokaryotic and eukaryotic cells when transcription and translation are absent cannot be explained by existing models of TTFL mechanisms [31–34]. Moreover, previous observations based on analysis of clock gene mutants in both mammals and fruit flies require re-interpretation in view of recent results demonstrating persistent circadian rhythms in these systems in which negative feedback within the TTFL loop is abolished [15, 35]. Taken together, it is now apparent that TTFL-based models for rhythmicity cannot provide a complete explanation for all features of circadian rhythmicity (reviewed in [36]).

## Transcription-translation feedback is not required for circadian oscillations in cyanobacteria

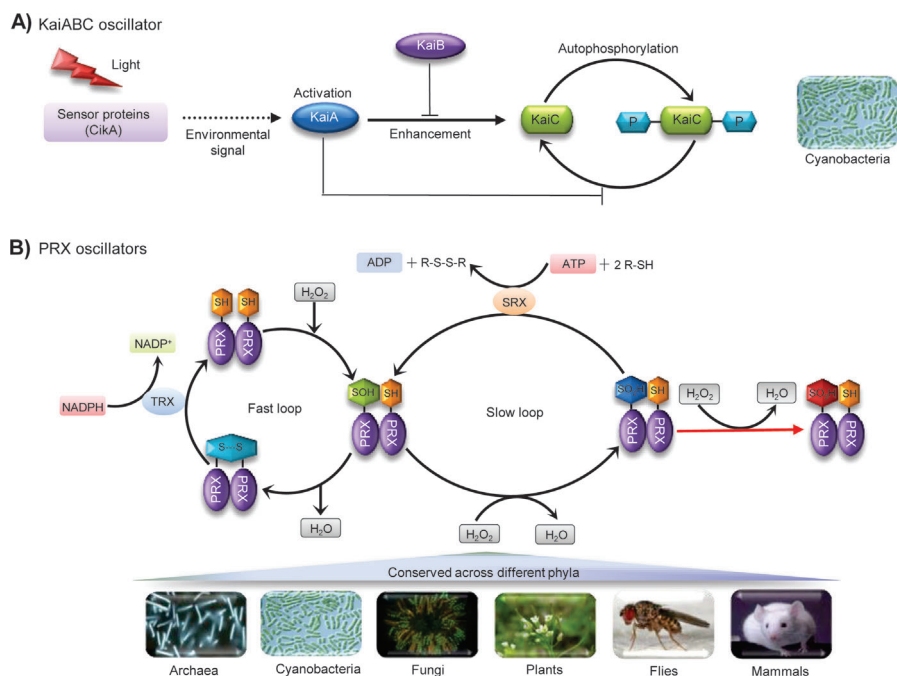
Nearly 10 years ago, the major shortcomings of the TTFL-based mechanism became obvious when circadian rhythms were found to be persisting in cyanobacteria even in the absence of transcription-translation feedback [31]. Identification of temperature-compensated, self-sustainable and robust oscillation in cyanobacterial KaiC phosphorylation evidently indicated transcription-translation feedback could be important, but not indispensable for circadian rhythmicity [32]. Intriguingly, subsequent studies provided valuable

information regarding the dynamics of the circadian KaiABC oscillator and its modulation by redox-active co-factor, which helped to decipher the precise mechanism of this entirely non-transcriptional phosphorylation-based rhythm in cyanobacteria [37, 38]. Identification of the cyanobacterial KaiABC oscillator indicated the existence of non-transcriptional oscillators (NTOs); nevertheless, the analysis of such an NTO was only restricted to cyanobacteria, as Kai proteins are not conserved across distinct phyla (Fig. 2A).

### Peroxiredoxins serve as the conserved biomarkers of circadian clocks

Another peculiarity about circadian systems previously was the apparent lack of molecular phylogeny of mechanisms controlling circadian rhythms. Virtually all known “clock” genes and proteins are not conserved across the various domains of life, although there are some homologous components in fruit flies and mammals. This previously suggested that the “logic” of the clockwork always involved a TTFL, but the players were different in each model system that has been studied [39]. Of late, the oxidation-reduction status of peroxiredoxin (PRX) proteins has been found to be regulated in a rhythmic fashion in anucleate human red blood cells (RBCs) without involving any transcriptional-translational mechanisms, representing the presence of autonomous oscillations in the redox status of the cell [33, 34]. Subsequently, oscillations of peroxiredoxin proteins (PRX) have been established as evolutionarily conserved markers of the clockwork, pointing to redox cycles as a likely unifying principle among disparate organisms [40], which was not observed in case of KaiABC oscillators (Fig. 2B).

PRXs are thiol-dependent peroxidases, which serve as an antioxidant in the maintenance of intracellular levels of hydrogen peroxide ( $H_2O_2$ ), peroxynitrite, and hydroperoxides to protect organisms against diverse oxidative stresses. Interestingly, these abundant cellular antioxidant proteins, which are probably evolved from a thioredoxin-like ancestor [41], are present in almost all living organisms [42]. On the basis of the number and bonding pattern of the catalytic cysteine residues, PRXs are classified in two distinct groups. The 1-Cys family contains only the  $NH_2$ -terminal Cys that can become oxidised, while the 2-Cys type contains both the  $NH_2$ - and  $COOH$ -terminal Cys residues and can therefore form either *intermolecular* (typical 2-Cys Prx) or *intramolecular*

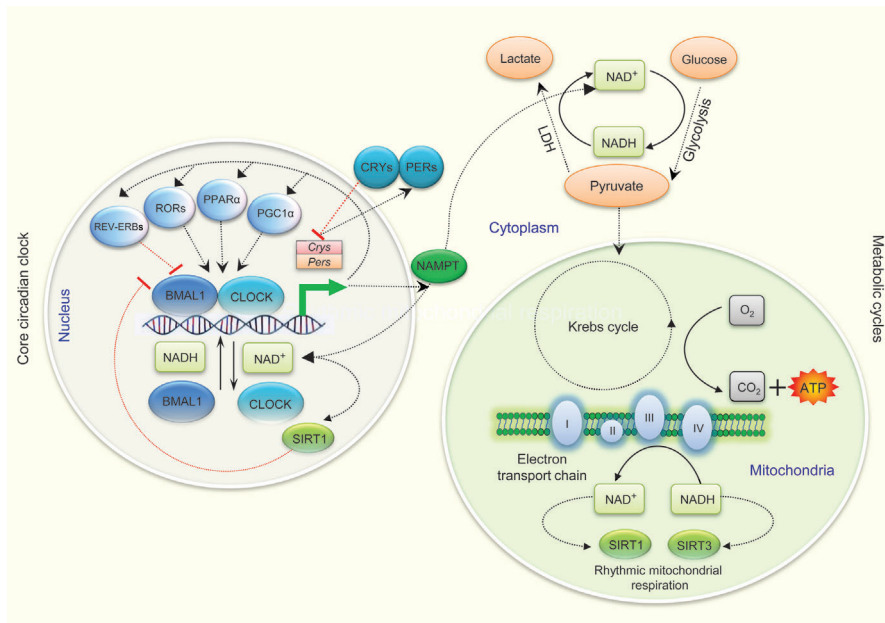


**Figure 2.** Molecular architecture of non-transcriptional oscillators (NTOs) in prokaryotic and eukaryotic organisms. **A:** Autonomous oscillation in cyanobacterial KaiC phosphorylation. Environmental sensors such as CikA transfer signal cascades to initiate the interaction between KaiA and dephosphorylated KaiC hexamers, which subsequently stimulates autokinase activity (phosphorylation happens at multiple residues of KaiC). KaiC autokinase activity is inhibited by KaiB binding, which aids in maintaining the equilibrium state between non-phosphorylated and phosphorylated forms of KaiC. **B:** Oxidation-reduction cycles of the peroxiredoxin (PRX) proteins as a conserved biomarker of circadian clocks across distinct phyla. There are two interconnected cycles (fast and slow loop) in the catalytic mechanism of typical 2-Cys PRXs. In the first cycle (fast loop) peroxidation of catalytic cysteine of PRXs leads to the formation of sulfenic acid (Cys-SOH), followed by disulfide bond (S–S) formation. The recycling step is catalyzed by Thioredoxin (TRX). Further oxidation of sulfenic moiety of PRXs (Cys-SOH) to sulfinic acid form (Cys-SO<sub>2</sub>H) happens in the second cycle (slow loop). Overoxidized Cys-SO<sub>2</sub>H residue can be slowly recycled through a reduction reaction carried out by sulfiredoxin (SRX) in an ATP-dependent manner. The sulfinic form (Cys-SO<sub>2</sub>H) can also be hyperoxidized into a sulfonic acid (Cys-SO<sub>3</sub>H), but this transformation is thought to be irreversible (modified from [15]).

(atypical 2-Cys Prx) disulfide bonds during oxidation. Apart from the detoxification of the various peroxide substrates, eukaryotic 2-Cys peroxiredoxins (2-Cys PRXs) play a significant role in hydrogen peroxide-mediated signal transduction pathways [43]. In addition to cyclic oxidation of peroxiredoxin proteins, it is highly likely that redox oscillations impact directly on many other susceptible proteins in cells. Specifically, so-called hyper-reactive cysteine residues represent particularly attractive targets [44].

### Do the non-transcriptional and TTFL oscillators and redox state regulate each other?

Redox state may impact on transcriptional activity of clock components [45] and neuronal activity within the master pacemaker in mammals, the suprachiasmatic nuclei (SCN) [46]. Interestingly, in turn circadian clocks also control the cellular redox status, since expression levels of many reactive oxygen species (ROS) responsive genes, or antioxidant enzymes,



**Figure 3.** Cross-talk between the circadian and metabolic clocks. Coupling mechanisms between the circadian and metabolic oscillators are miscellaneous. There are a series of transcription/translation feedback loops in the core clock mechanism. CRY proteins (along with the PER proteins) function as the negative regulators for maintenance of circadian rhythms. PPAR $\alpha$  and PGC-1 $\alpha$  stimulates expressions of clock genes, while RORs regulate Bmal1 transcription through formation of a feedback loop involving ROR $\alpha$  and REV-ERB $\alpha$ . Core clock proteins such as BMAL1 and CLOCK (NPAS2 substitutes for CLOCK in some brain regions (not shown)) regulate the rate limiting steps of NAD<sup>+</sup> biosynthesis [58, 62], while the DNA binding affinity of BMAL1 and CLOCK is controlled by the intracellular NAD<sup>+</sup>/NADH ratio [45]. LDH plays a crucial role in increasing the cellular concentration of NAD<sup>+</sup>. NAD<sup>+</sup>-dependent deacetylases, for example SIRT1 or SIRT3 regulate circadian clock gene expression [59, 64]. NAMPT acts as a rate-limiting enzyme in mammalian NAD<sup>+</sup> biosynthesis and its expression is also regulated by the core clock genes [62]. (Details for these possible connecting components between the circadian oscillators and various metabolic processes have been summarised in Table 1). Abbreviations: BMAL1, brain and muscle ARNT-Like 1; CLOCK, circadian locomotor output cycles kaput; Cry, cryptochrome; LDH, lactate dehydrogenase; NAD, nicotinamide adenine dinucleotide; NAMPT, nicotinamide phosphoribosyl-transferase; NPAS2, neuronal PAS domain protein 2; Per, period; PPAR, peroxisome proliferators-activated receptor; PGC-1 $\alpha$ , PPAR gamma coactivator-1 alpha; ROR, retinoic acid orphan receptors; SIRT 1, sirtuin 1; SIRT 3, sirtuin 3.

are frequently regulated by the clock genes [47, 48]. Consequently, mechanistic interactions between redox and circadian components suggest that the redox state of a cell and clocks are influenced and regulated by each other.

The existence of NTOs such as the cyanobacterial KaiABC oscillator, and the more pervasive oxidation-reduction cycles of PRX proteins, thus challenges the paradigm of TTFL-based mechanisms. Now, the imperative question is whether the NTO and TTFL-based oscillators are interlinked, and if so, how they co-exist in cells and collectively interplay to maintain unequivocal cellular time keeping. Although direct connections between the two systems is not yet fully delineated, there is experimental evidence for their interactions, and mathematical modeling indicates that coupling between NTO and TTFLs effectively boosts overall clock performance (reviewed in [36, 49]). First of all, the Kai and PRX system still exhibit oscillation even in the absence of TTFL [32, 33]. Secondly, circadian rhythms are not obliterated in systems with either constitutive expression or deletion of clock genes (in TTFL

knockout mutants) [35, 50, 51]. It is therefore reasonable to speculate that timekeeping is controlled by a biochemical oscillator. To this end, understanding of the reciprocal communication of circadian oscillators with various metabolic and redox cycles could provide valuable insights, as elaborated in the next section.

### Cross-talk between the circadian and metabolic clocks: Reciprocal regulation of the circadian cycles and energetic pathways

Connection between circadian and metabolic systems is one of the most important and enlightening areas of current circadian biology research [52–55]. Adverse effects of circadian disruption or sleep deprivation on metabolic functions clearly indicate the impact of these processes on energy homeostasis [56]. Over a decade ago, McKnight and co-workers demonstrated regulation of two clock proteins (BMAL1 and CLOCK) by the metabolic cofactor NAD(P), and therefore, a connection between the cellular metabolism and clockwork circuitry [57].

With the passage of time, subsequent studies have illustrated several modes of bidirectional regulation of cellular metabolites and clock proteins. For example, there is modulation of CLOCK-mediated chromatin remodeling and regulation of circadian clock gene expression by the NAD<sup>+</sup>-dependent deacetylase SIRT1 [58, 59] and circadian regulation of the NAD<sup>+</sup> salvage pathway by CLOCK-SIRT1 [60]. Similarly, circadian regulation of the enzymatic activity of acetyl-CoA Synthetase 1 (AceCS1) leads to varying intracellular levels of the central metabolite acetyl-CoA [61]. Furthermore, nicotinamide phosphoribosyltransferase (NAMPT), the rate-limiting enzyme in mammalian NAD<sup>+</sup> biosynthesis, exhibits rhythmicity in its expression level [62]. These observations highlight how the clock's downstream transcriptional network extends to metabolic genes. Likewise, other metabolic links have been elaborated, in particular with respect to poly(ADP-ribose) polymerase 1 (PARP-1) [63] and the role of the NAD(+)-dependent deacetylase sirtuin 3 (SIRT3) in rhythmic mitochondrial function [64] (Fig. 3). Connecting hubs between the metabolic and clock networks are diverse, and certainly not yet fully explored. However, in the light of our current understanding of circadian biology, it is likely that NAD-dependent enzymes, nutrient-sensing

transcriptional regulatory proteins, redox transcription factors, and protein kinases serve as the crucial candidates for mediating cross-talk between energetic pathways and circadian cycles [65]. Table 1 summarizes the circadian control of different metabolic processes in eukaryotes.

Oscillation of metabolic pathways is not a new concept. In the 1960s, rapid rhythmicity (over minutes rather than hours) in the glycolytic intermediates glucose-6-phosphate/fructose-6-phosphate and fructose-1,6-diphosphate (FDP) levels was first demonstrated in yeast extracts by Ghosh and Chance [66], and more recent studies have built on this initial work [67, 68]. On a longer timescale, hepatic gluconeogenesis, which plays an important role in maintaining glucose homeostasis in mammals during starvation, is also under circadian regulation, most probably through the control of Cryptochromes (*Cry1* and *Cry2*) by CLOCK and BMAL1 [69]. Similarly, components of the Krebs cycle such as NADP-dependent isocitrate dehydrogenase also exhibit circadian periodicity in their abundance or activity [70].

Intriguingly, several components of central metabolic pathways, such as a number of rate-limiting enzymes involved in glycolysis and the tricarboxylic acid (TCA) cycle, are redox-sensitive, indicating the potential of these metabolic pathways to be regulated by the circadian redox oscillations [71]. A key metabolite at the interface of cytosolic and mitochondrial metabolism is acetyl-CoA, which not only plays a role in metabolism itself, but also regulates protein function by participating in acetylation reactions. Such post-translational modification of proteins, which include histones, can thus regulate gene expression, in addition to modulating enzyme function. In this vein, recent work highlights that clock-driven acetylation modulates a considerable number of mitochondrial proteins involved in multiple metabolic networks [72]. Moving beyond the analysis of transcriptional processes and gene expression patterns is essential to address connectivity between the clock and metabolism, but has been challenging because of the technical challenges posed by performing such analyses, which rely heavily on mass spectrometry based metabolomics.

Recently, comprehensive metabolomics analyses of different biological fluids (saliva and blood) have provided insights regarding the circadian regulation of various human metabolic pathways [73], and consequences of sleep deprivation on the human metabolome [74]. Of note, it has been demonstrated that nearly 15% of all metabolites identified in human plasma and saliva are controlled by circadian clocks [73]. Similarly, a good number of metabolites in exhaled human breath also exhibit circadian rhythmicity [75]. Importantly, sleep deprivation in humans adversely affects the oscillatory behavior of many blood metabolites including tryptophan, serotonin, taurine, acylcarnitines, glycerophospholipids, and sphingolipids [74], indicating a connection between sleep restriction and circadian clock disruption and metabolic dysfunction. Further work is, however, needed to delineate the exact mechanism for these observations. These studies have thus been important in creating an avenue towards the recognition of new physiological/metabolic pathways which are controlled by circadian clocks or sleep-wake cycles. Moreover, such metabolomic profiling has the potential to identify

novel noninvasive biomarkers of circadian disruption, sleep deprivation and associated metabolic and neurological disorders.

## How might the clock and sleep be connected?

In mammalian circadian system, the brain's suprachiasmatic nucleus (SCN), which is considered as the master circadian "clock," orchestrates synchronization of oscillators in peripheral tissues [76, 77]. The neural circuits involved in the regulation of sleep-wake states and circadian rhythms are becoming established, as are the vital roles of circadian and homeostatic processes in regulation of the sleep and arousal-promoting circuitry [7, 78]. Intriguingly, there is physiological evidence that indicate sleep centers can also regulate the circadian pacemaker [79]. In addition, sleep plays a role in the clearance of potentially neurotoxic waste products from the central nervous system, and thereby maintains metabolic homeostasis [17]. Circadian clocks regulate different aspects of sleep, suggesting that redox and metabolism may affect sleep homeostasis through their impact on the state of the circadian system. However, the exact mechanism by which sleep-wake centers communicate with the SCN and metabolic cycles has not been untangled.

Components of the central circadian pacemaker are known to regulate sleep onset and control the transition to wakefulness, while the quantity of sleep appears to be controlled by homeostatic centers [80]. At the molecular level, the interactions between clocks and sleep-wake cycles have been investigated using mutant mice and fruit fly models lacking core circadian clock genes to try to unravel the possible functions of clock genes in sleep homeostasis (reviewed in [81, 82]). For example, targeted disruption or deletion of the core clock components such as *Bmal1* (*Cycle* in *Drosophila*), *Clock/Npas2*, *Per1/Per2*, and *Cry1/Cry2* result in various phenotypes including increased sleep fragmentation and rebound following deprivation of sleep, in addition to an enhanced tendency to switch between non-rapid eye movement (NREM) and rapid eye movement (REM) sleep [83]. Conversely, sleep deprivation can impinge on the expression levels of core circadian transcriptional regulators and their DNA-binding capabilities [84], demonstrating the existence of a reciprocal regulation between the central clock machinery and sleep-wake cycle. Interestingly, as discussed in the previous section, binding of certain clock proteins may be regulated by intracellular redox potential, indicating the potential for cross-talk between the circadian clock machinery, energy metabolism, and sleep regulation.

There is a longstanding belief that sleep happens only at the level of the whole organism. However, recent work highlights that even within the brain, there is local and use-dependent sleep of subsets of neural circuitry, which forces a reappraisal of what sleep is and what it might be for [85, 86]. For example, electrophysiological analysis of the sleep-wake cycle indicates the concurrent existence of different sleep intensities within distinct regions of human brain [87]. Likewise, slow wave activity (SWA) in local cortical EEG

Table 1. Possible connecting components between the circadian oscillators and metabolic processes

Candidates	Plausible roles/ involvement in cross-talks	References
CLOCK/NPAS2	Circadian transcription factor CLOCK/NPAS2 controls NAD <sup>+</sup> biosynthesis through regulation of NAMPT expression	[62]
PER proteins	NAD(P) <sup>+</sup> /NAD(P)H ratio regulates the binding of CLOCK/NPAS2-BMAL1 heterodimers to their E box cognate sequence PER proteins regulate expression of the core clock gene Bmal1	[59, 115]
CRY proteins	Binding of SIRT1 to CLOCK-BMAL1 complexes promotes PER2 deacetylation and degradation PER proteins regulate lipid and glycogen metabolism through their interactions with diverse nuclear receptors CRY proteins (along with the PER proteins) function as the negative regulators for maintenance of circadian rhythms They regulate circadian rhythmicity of cAMP signaling and hepatic gluconeogenesis AMPK directly phosphorylates CRY proteins and reduces their half-life	[69, 116]
NAD <sup>+</sup>	NAD(P) <sup>+</sup> and NAD(P)H reflect the metabolic and redox status of the cell NAD <sup>+</sup> serves as a metabolic oscillator and controls the core clock machinery primarily through SIRT1	[58, 62]
SIRT1	NAD <sup>+</sup> -dependent SIRT1 controls expression the circadian clock genes (Bmal1, Per2, and Cry1) through PER2 deacetylation	[58–60]
	CLOCK-SIRT1 regulates circadian control of the NAD <sup>+</sup> salvage pathway it regulates circadian transcription also by the deacetylation of histone H3 tails	
SIRT3	SIRT3 maintains rhythms in the acetylation and activity of oxidative enzymes and respiration	[64]
NAMPT	Core clock components regulate its activity through control of concentrations of NAD <sup>+</sup> NAMPT is the rate-limiting enzyme in mammalian NAD <sup>+</sup> biosynthesis Its expression is regulated by the core clock genes	[62]
PARP1	Its inhibition leads to the oscillation of Per2 by releasing CLOCK; BMAL1 PARP1 modifies clock components in response to feeding-fasting cycles	[63, 117]
PRXs	It regulates the binding of CLOCK-BMAL1 to DNA and interaction of CLOCK-BMAL1 with PER and CRY repressor proteins SP1, a nuclear target protein of PARP-1, regulate its expression PRX proteins exhibit self-sustained oscillation in their oxidation–reduction cycles	[34, 40]
AceCS1	PRX cycle provides feedback to regulate the core clock transcriptional network probably through the oscillation of ROS Perturbation of their functions causes a long-period phenotype or leads to a depression in the amplitude of circadian oscillations Circadian control of intracellular levels of acetyl-CoA and thereby fatty acid elongation is regulated through the enzymatic activity of AceCS1	[118]
AMPK	AceCS1 activity is controlled by acetylation, and its rhythmic acetylation is regulated by SIRT1 AMPK serves as the major sensor of the AMP/ATP ratio, activates stress-promoted transcription and regulates clock gene expression	[119]
PPAR $\alpha$ and PGC-1 $\alpha$	It regulates stability (promotes degradation) of core clock proteins (CRY and PER) Transcriptional coactivator PGC-1 $\alpha$ stimulates expressions of clock genes (Bmal1 and Rev-erb $\alpha$ )	[120, 121]
ALAS1	It has association with the SirT1 histone deacetylase complex, can serve as a sensor for the metabolic state of the cell, and also induces the expression of gluconeogenic genes PPAR $\alpha$ regulates fatty acid oxidation and apolipoprotein synthesis ALAS1, the rate limiting enzyme in haem biosynthesis, is a target gene for the NPAS2/BMAL1 heterodimer	[122]
HSF1	Circadian rhythmicity in the cellular haem levels is maintained through the regulation of the expression of ALAS1 by the core clock genes Reciprocally, haem regulates activity of the BMAL1-NPAS2 transcription complex HSF1 plays an important role in transporting nutrient signals to the circadian circuitry	[123]
CREB	Phosphorylation by diverse protein kinases regulates its activity It also functions as a key regulator of temperature-dependent expression of heat shock protein/ chaperone genes associated with circadian oscillators cAMP signaling via CREB and other transcriptional oscillator is imperative for the molecular circadian oscillators CREB-dependent transcription supports steady cycling of the core clock transcriptional loop	[124, 125]

(Continued)

**Table 1.** (Continued)

Candidates	Plausible roles/ involvement in cross-talks	References
FOXO	Nutrient and stress sensor FOXO regulates sensitivity of the circadian clock to stress conditions; its effects on circadian rhythms are non-cell-autonomous	[126, 127]
RORs	SIRT1 regulates FOXO transcription factors in a stress-dependent manner Expression of several gluconeogenic genes is directly regulated by FOXO1 RORs are components of the master oscillator in mammalian circadian system that regulate Bmal1 transcription through formation of a feedback loop involving ROR $\alpha$ and REV-ERB $\alpha$ RORs can alter PER2 activity by direct physical interactions	[128, 129]

AceCS1, Acetyl-CoA Synthetase 1; ALAS1, aminolevulinatase synthase 1; AMPK, AMP-dependent protein kinase; CREB, cAMP response element-binding protein; Cry, cryptochrome; FOXO, Forkhead homeobox type O; HSF1, heat shock transcription factor 1; NAD, nicotinamide adenine dinucleotide; NAMPT, nicotinamide phosphoribosyl-transferase; NPAS2, neuronal PAS domain protein 2; PARP1, poly (ADP-ribose) polymerase 1; Per, period; PPAR, peroxisome proliferators-activated receptor; PGC-1 $\alpha$ , PPAR gamma coactivator-1 alpha; PRX, peroxiredoxin; ROR, retinoic acid orphan receptors; ROS, reactive oxygen species; SIRT, sirtuin.

recordings from brain regions of awake animals demonstrates that neuronal subsets may enter “off” states during a long stretch of wakefulness, probably due to the falling in levels of arousal-promoting neuromodulators [88]. Similarly, Krueger and colleagues have demonstrated sleep as a property of local neuronal assemblies, and hypothesized that the local-network sleeps, which are perhaps controlled by the oscillation in the levels of sleep-regulatory molecules, are connected by central mechanisms and serve as the fundamental basis of whole-organism sleep [89]. Therefore, local populations of neurons might sleep at the cellular level in a use-dependent manner, which could be a means of saving energy, particularly during prolonged wakefulness. To this end, it would be interesting to decipher the possible functional consequences of such local characteristics of sleep.

Akin to circadian clocks, it is now imperative to study the self-sustained mechanism of sleep at the cellular level, which is often difficult to execute *in vivo* in the mammalian brain due to the presence of the entangled web of neuronal networks that are controlled by signals from both local and global sources. Of note, a recent study on primary mouse cortical cultures demonstrated sleep as a prominent characteristics of simple neuronal networks grown *in vitro* [90]. Interestingly, the authors observed electrophysiological, metabolic and transcriptomic similarities between the *in vitro* neuronal networks and those determined from sleep-deprived mice. The presence of important features of the sleep-wake cycle in an *in vitro* setting opens up new avenues for molecular level investigations of the local nature of sleep and its multifaceted interplay with diverse metabolic and circadian oscillators, which has not been possible to achieve previously.

### The promise of systems level multi-omics approaches to unraveling the interconnectivity of circadian clockworks, metabolic oscillators, and the sleep-wake switch

Although identification of NTOs and metabolic oscillations has enhanced our understanding of circadian rhythmicity, there are still several opaque areas that entail further investigation (Fig. 1). The most contemporary questions are: (1) In what manner do non-transcriptional and transcriptional clock mechanisms interact with each other? (2) What are the mechanisms underlying the reciprocal regulation of the non-transcriptional circadian clocks and energetic pathways? (3) Do circadian redox oscillations impact on the sleep-wake switch? (4) How are dysfunctions in circadian clocks/sleep-wake cycles linked to diverse diseases, including metabolic disorders such as diabetes mellitus?

### Proteome level analyses can serve as an excellent complementary platform for mRNA level observations

While transcriptomic studies have previously provided valuable insights into which gene networks and tissue-

specific programmes are controlled by the circadian clock [23] and sleep [5, 91], there has been a dearth of proteome level investigation in circadian biology [92]. As with metabolomics analyses, technological limitations have previously hampered progress in global protein profiling. Initial proteomics studies implemented gel-based approaches, and consequently, were only able to measure expression patterns of a limited number of rhythmic proteins due to the poor coverage of the entire proteomes [28, 93, 94]. Application of next-generation quantitative proteomics approaches involving ultra-sensitive mass spectrometers, which are presently at the pinnacle of promising proteomics technologies [95], could be extremely useful in decoding the mechanisms and extent of protein oscillations and the links to the sleep-wake switch (Fig. 4A). With the broadening availability of mass spectrometry, a number of quantitative proteomics datasets have emerged investigating the mechanism of synchronization of circadian rhythms by the SCN [96, 97], post-transcriptional mechanisms of circadian regulation [30], and diurnal oscillations in the mammalian hepatic proteome [29]. To date, such approaches have not been applied to characterize sleep-wake cycles, or to determine proteome level changes upon sleep deprivation, although gel-based techniques have been employed to characterize the latter in the past [98, 99].

In recent years, ribosome profiling, which provides genome-wide information on protein synthesis through deep sequencing of ribosome protected mRNA fragments, is emerging as an efficient technique to track *in vivo* translation. Circadian clocks play some vital roles in coordinating transcription and translation steps which are essential for ribosome biogenesis. Intriguingly, some very recent studies have demonstrated the promising applications of ribosome profiling for studying translational control in circadian gene expression and for mapping rhythmic transcriptome [100, 101]. Proteomic measurements and ribosome profiling collectively can provide comprehensive information regarding rhythmic proteins and can complement the technical limitations of each others.

### **Redox proteomics and characterization of post-translational modifications provide novel mechanistic insights into circadian biology**

Cross-talk between the cellular redox state and the circadian clocks has been studied extensively in different organisms [45, 102, 103]. Following the identification of oxidation-reduction cycles of peroxiredoxin proteins as the conserved markers of circadian rhythms [40], there has been an emerging interest in profiling redox oscillations at a global level to investigate the consequences of metabolic cycles on circadian rhythmicity and sleep-wake switching. Recent studies indicate that the circadian rhythm of redox state controls excitability in SCN neurons [46, 104]. Consequently, it can be speculated that redox homeostasis and neuronal activity are coupled non-transcriptional circadian oscillators intertwined in neuronal physiology. Peroxiredoxin proteins may not be unique in their ability to undergo redox oscillations since many other proteins are susceptible to oxidation of their cysteine residues by peroxide [44]. Thus,

redox proteomics analyses will undoubtedly provide a novel mechanistic insight into the effects of brain disruption of redox processes/signaling on sleep and circadian rhythms and vice versa [71].

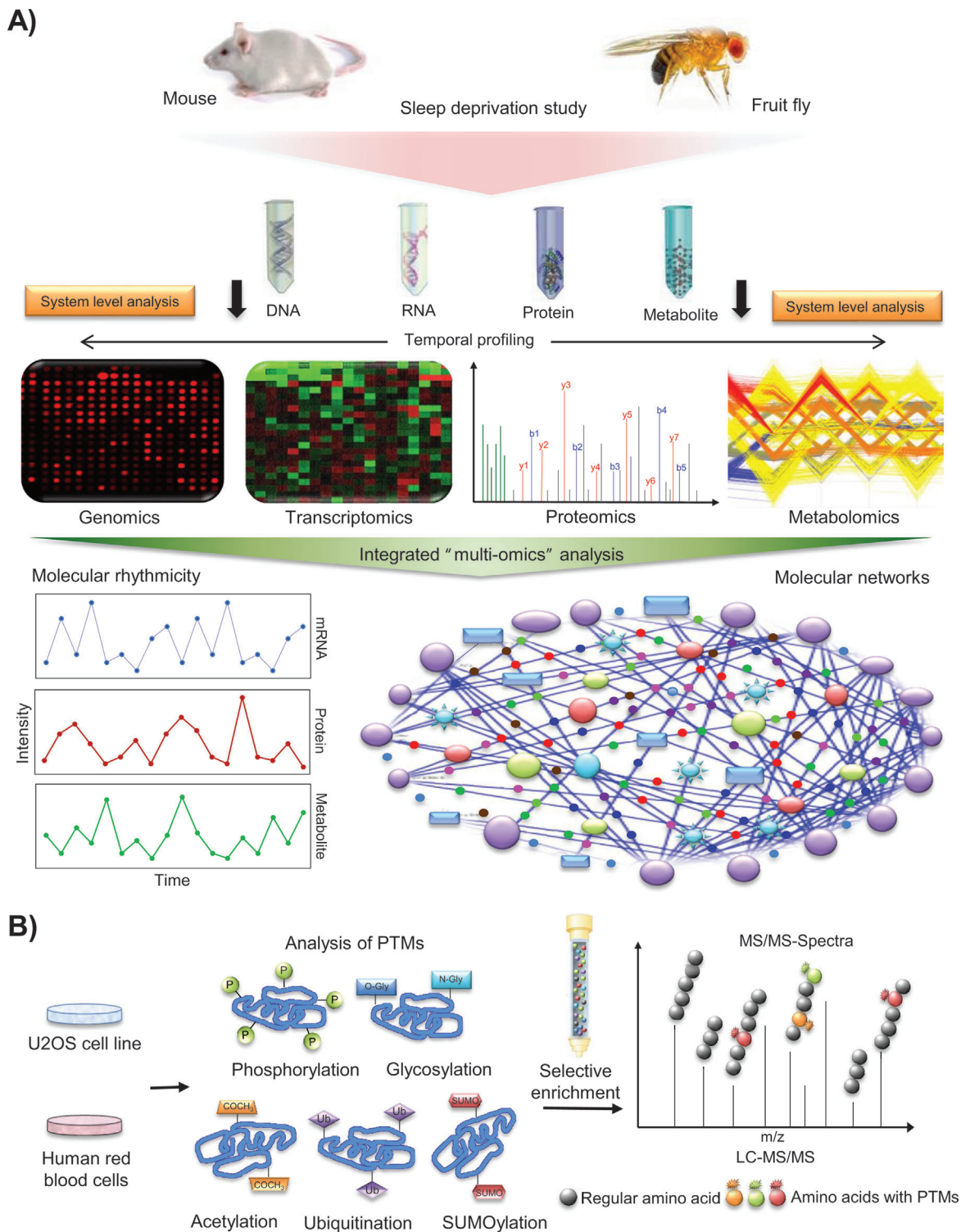
A complementary set of analyses could also be used to probe specific post-translational modifications (PTMs) of proteins across the day, and also under sleep restriction paradigms. Such modifications regulate the recruitment, recognition, assembly/disassembly, translocation, and consequently the eventual fates of the majority of eukaryotic proteins [105]. In the context of the clockwork, there is substantial evidence that the control of functional activity and stability of well characterized clock proteins is driven by diverse types of PTMs, including phosphorylation [106], acetylation [58, 59], ubiquitylation [107], and SUMOylation [108]. We thus anticipate that comparative analysis of PTMs in non-transcriptional models (e.g. anucleate red blood cells) and nucleated cell culture lines (e.g. human osteosarcoma U2OS cell line) have the potential to identify dynamic modifications in proteins that might be known to be associated with the clock, but also others that are completely novel. Moreover, examining non-transcriptional and transcriptional models will establish nodes of interconnectivity between these mechanisms (Fig. 4B).

### **Integration of different “omics” datasets is promising for studying functional interactions between circadian and metabolic cycles**

A recent study by Sassone-Corsi and colleagues demonstrated synergistic links between oscillations in the circadian transcriptome and metabolic pathways [109]. Furthermore, the same research group has developed the CircadiOmics platform (<http://circadiomics.igb.uci.edu/>), which is a computational framework that could serve as a repository for metabolomics and other “omics” level high-throughput data associated with circadian clocks [110]. Nevertheless, existing tools such as this do not yet integrate proteome level information, which is critical for building a comprehensive view of biological rhythmicity. Thus far, similar integrative endeavors have not been attempted for sleep datasets, and therefore tying together common molecular pathways between sleep and the circadian clock at a systems level remains an unrealized goal.

In order to unravel the transient snapshots of dynamic circadian rhythmicity and sleep homeostasis, there is an urgent need to amalgamate the findings obtained from mRNA, protein and metabolite level investigations to get a true understanding of these multiple dynamic facets. In the future, we anticipate that an integrated multi-omics analysis (specifically proteomics and metabolomics) of sleep-deprived models (using different complementary model organisms such as the mouse and fruit fly) will collectively provide a complete representation of bidirectional molecular communication between the sleep centers and the circadian pacemaker. Eventually, this may also lead to the identification of novel conserved markers of circadian rhythms and sleep deprivation (Fig. 4A). Consequently, integrated quantitative multi-omics analyses may also highlight molecular pathways affected by disruption of circadian clocks and





**Figure 4.** Systems level multi-omics approaches to uncover the bi-directional communications between the sleep centers and the circadian pacemaker. **A:** Schematic representation of an integrated multi-omics (genomics, transcriptomics, proteomics, and metabolomics) analysis for unraveling the interconnectivities among circadian clocks, metabolic cycles and the sleep-wake switch. Systems level analyses of sleep-deprived models (fruit flies and mice) could collectively provide an inclusive representation regarding the temporal physiological states of organisms, and alterations in molecular oscillations and networks as a consequence of sleep deficiency. **B:** Schematic illustration of an analysis of post-translational modifications (PTMs) in a non-transcriptional (anucleate red blood cell) model and a nucleated cell line (human osteosarcoma U2OS cell line) in vitro for studying the connectivity between the non-transcriptional and TTFL mechanisms of circadian rhythmicity. Selective enrichment of the post-translationally modified peptides and subsequent mass spectrometry-based profiling of diverse PTM patterns might provide additional mechanistic insights into the dynamic modifications of core clock proteins and their interactors.

sleep-wake cycles and their association with different adverse health consequences.

## Conclusions and outlook

We now have a vast array of knowledge about the molecular underpinnings of the circadian clock, particularly at the level of transcriptional networks. The influence of the clock is pervasive, with ~40% of transcripts in the mouse genome displaying daily cycles [111]. Similarly, transcriptional changes in the brain under sleep deprivation are extensive. However, how sleep and the clock are linked at the molecular level remains a fascinating question. Novel insights into metabolic cycles and their connectivity with the circadian clockwork promise to offer potential routes to link the two processes, since sleep is also influenced by metabolism [112, 113] and it regulates metabolic processes within the body [114]. A considerable amount of “dark matter” still, however, exists in our understanding of the clockwork and sleep, which stems from a lack of tools to assess other “omes” at a global, quantitative level. In particular, we vitally need proteomic datasets that not only quantify protein abundance, but also a diversity of post-translational modifications that can modulate their function. Moreover, we must concurrently assay metabolites to get a complete picture of the end products of metabolic pathways that are known to be regulated by the circadian and sleep systems. Multi-omics approaches that are now becoming widely accessible will thus change the way that high-throughput temporal profiling can be performed, and thus offers a conduit to insights that have previously been beyond our reach.

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## References

- Dunlap JC. 1999. Molecular bases for circadian clocks. *Cell* **96**: 271–90.
- Bell-Pedersen D, Cassone VM, Earnest DJ, Golden SS, et al. 2005. Circadian rhythms from multiple oscillators: lessons from diverse organisms. *Nat Rev Genet* **6**: 544–56.
- Siegel JM. 2005. Clues to the functions of mammalian sleep. *Nature* **437**: 1264–71.
- Nitz DA, van SB, Tononi G, Greenspan RJ. 2002. Electrophysiological correlates of rest and activity in *Drosophila melanogaster*. *Curr Biol* **12**: 1934–40.
- Maret S, Dorsaz S, Gurcel L, Pradervand S, et al. 2007. Homer1a is a core brain molecular correlate of sleep loss. *Proc Natl Acad Sci USA* **104**: 20090–5.
- Broussard JL, Ehrmann DA, Van CE, Tasali E, et al. 2012. Impaired insulin signaling in human adipocytes after experimental sleep restriction: a randomized, crossover study. *Ann Intern Med* **157**: 549–57.
- Saper CB, Scammell TE, Lu J. 2005. Hypothalamic regulation of sleep and circadian rhythms. *Nature* **437**: 1257–63.
- Schwartz JR, Roth T. 2006. Shift work sleep disorder: burden of illness and approaches to management. *Drugs* **66**: 2357–70.
- Reddy AB, O'Neill JS. 2010. Healthy clocks, healthy body, healthy mind. *Trends Cell Biol* **20**: 36–44.
- Durgan DJ, Young ME. 2010. The cardiomyocyte circadian clock: emerging roles in health and disease. *Circ Res* **106**: 647–58.
- Vetter C, Fischer D, Matera JL, Roenneberg T. 2015. Aligning work and circadian time in shift workers improves sleep and reduces circadian disruption. *Curr Biol* **25**: 907–11.
- Hastings MH, Reddy AB, Maywood ES. 2003. A clockwork web: circadian timing in brain and periphery, in health and disease. *Nat Rev Neurosci* **4**: 649–61.
- Wulff K, Gatti S, Wettstein JG, Foster RG. 2010. Sleep and circadian rhythm disruption in psychiatric and neurodegenerative disease. *Nat Rev Neurosci* **11**: 589–99.
- Sahar S, Sassone-Corsi P. 2009. Metabolism and cancer: the circadian clock connection. *Nat Rev Cancer* **9**: 886–96.
- Reddy AB, Rey G. 2014. Metabolic and non-transcriptional circadian clocks: eukaryotes. *Annu Rev Biochem* **83**: 165–89.
- Rey G, Reddy AB. 2013. Physiology. Rhythmic respiration. *Science* **342**: 570–1.
- Xie L, Kang H, Xu Q, Chen MJ, et al. 2013. Sleep drives metabolite clearance from the adult brain. *Science* **342**: 373–7.
- Takahashi JS, Hong HK, Ko CH, McDearmon EL. 2008. The genetics of mammalian circadian order and disorder: implications for physiology and disease. *Nat Rev Genet* **9**: 764–75.
- Harmer SL, Panda S, Kay SA. 2001. Molecular bases of circadian rhythms. *Annu Rev Cell Dev Biol* **17**: 215–53.
- Chen R, Schirmer A, Lee Y, Lee H, et al. 2009. Rhythmic PER abundance defines a critical nodal point for negative feedback within the circadian clock mechanism. *Mol Cell* **36**: 417–30.
- Sangoram AM, Saez L, Antoch MP, Gekakis N, et al. 1998. Mammalian circadian autoregulatory loop: a timeless ortholog and mPer1 interact and negatively regulate CLOCK-BMAL1-induced transcription. *Neuron* **21**: 1101–13.
- Kume K, Zylka MJ, Sriram S, Shearman LP, et al. 1999. MCRY1 and mCRY2 are essential components of the negative limb of the circadian clock feedback loop. *Cell* **98**: 193–205.
- Akhtar RA, Reddy AB, Maywood ES, Clayton JD, et al. 2002. Circadian cycling of the mouse liver transcriptome, as revealed by cDNA microarray, is driven by the suprachiasmatic nucleus. *Curr Biol* **12**: 540–50.
- Panda S, Antoch MP, Miller BH, Su AI, et al. 2002. Coordinated transcription of key pathways in the mouse by the circadian clock. *Cell* **109**: 307–20.
- Storch KF, Lipan O, Leykin I, Viswanathan N, et al. 2002. Extensive and divergent circadian gene expression in liver and heart. *Nature* **417**: 78–83.
- Ueda HR, Chen W, Adachi A, Wakamatsu H, et al. 2002. A transcription factor response element for gene expression during circadian night. *Nature* **418**: 534–9.
- Ueda HR, Hayashi S, Chen W, Sano M, et al. 2005. System-level identification of transcriptional circuits underlying mammalian circadian clocks. *Nat Genet* **37**: 187–92.
- Reddy AB, Karp NA, Maywood ES, Sage EA, et al. 2006. Circadian orchestration of the hepatic proteome. *Curr Biol* **16**: 1107–15.
- Mauvoisin D, Wang J, Jouffe C, Martin E, et al. 2014. Circadian clock-dependent and -independent rhythmic proteomes implement distinct diurnal functions in mouse liver. *Proc Natl Acad Sci USA* **111**: 167–72.
- Robles MS, Cox J, Mann M. 2014. In-vivo quantitative proteomics reveals a key contribution of post-transcriptional mechanisms to the circadian regulation of liver metabolism. *PLoS Genet* **10**: e1004047.
- Nakajima M, Imai K, Ito H, Nishiwaki T, et al. 2005. Reconstitution of circadian oscillation of cyanobacterial KaiC phosphorylation in vitro. *Science* **308**: 414–5.
- Tomita J, Nakajima M, Kondo T, Iwasaki H. 2005. No transcription-translation feedback in circadian rhythm of KaiC phosphorylation. *Science* **307**: 251–4.
- O'Neill JS, van OG, Dixon LE, Troein C, et al. 2011. Circadian rhythms persist without transcription in a eukaryote. *Nature* **469**: 554–8.
- O'Neill JS, Reddy AB. 2011. Circadian clocks in human red blood cells. *Nature* **469**: 498–503.
- Lakin-Thomas PL. 2006. Transcriptional feedback oscillators: maybe, maybe not. *J Biol Rhythms* **21**: 83–92.
- van OG, Millar AJ. 2012. Non-transcriptional oscillators in circadian timekeeping. *Trends Biochem* **37**: 484–92.

37. Wood TL, Bridwell-Rabb J, Kim YI, Gao T, et al. 2010. The KaiA protein of the cyanobacterial circadian oscillator is modulated by a redox-active cofactor. *Proc Natl Acad Sci USA* **107**: 5804–9.
38. Qin X, Byrne M, Mori T, Zou P, et al. 2010. Intermolecular associations determine the dynamics of the circadian KaiABC oscillator. *Proc Natl Acad Sci USA* **107**: 14805–10.
39. Rosbash M. 2009. The implications of multiple circadian clock origins. *PLoS Biol* **7**: e62.
40. Edgar RS, Green EW, Zhao Y, van OG, et al. 2012. Peroxiredoxins are conserved markers of circadian rhythms. *Nature* **485**: 459–64.
41. Copley SD, Novak WR, Babbitt PC. 2004. Divergence of function in the thioredoxin fold superfamily: evidence for evolution of peroxiredoxins from a thioredoxin-like ancestor. *Biochemistry* **43**: 13981–95.
42. Hall A, Karplus PA, Poole LB. 2009. Typical 2-Cys peroxiredoxins-structures, mechanisms and functions. *FEBS J* **276**: 2469–77.
43. Wood ZA, Poole LB, Karplus PA. 2003. Peroxiredoxin evolution and the regulation of hydrogen peroxide signaling. *Science* **300**: 650–3.
44. Weerapana E, Wang C, Simon GM, Richter F, et al. 2010. Quantitative reactivity profiling predicts functional cysteines in proteomes. *Nature* **468**: 790–5.
45. Rutter J, Reick M, Wu LC, McKnight SL. 2001. Regulation of clock and NPAS2 DNA binding by the redox state of NAD cofactors. *Science* **293**: 510–4.
46. Wang TA, Yu YV, Govindaiah G, Ye X, et al. 2012. Circadian rhythm of redox state regulates excitability in suprachiasmatic nucleus neurons. *Science* **337**: 839–42.
47. Krishnan N, Davis AJ, Giebultowicz JM. 2008. Circadian regulation of response to oxidative stress in *Drosophila melanogaster*. *Biochem Biophys Res Commun* **374**: 299–303.
48. Lai AG, Doherty CJ, Mueller-Roeber B, Kay SA, et al. 2012. CIRCADIAN CLOCK-ASSOCIATED 1 regulates ROS homeostasis and oxidative stress responses. *Proc Natl Acad Sci USA* **109**: 17129–34.
49. Wu L, Reddy AB. 2014. Rethinking the clockwork: redox cycles and non-transcriptional control of circadian rhythms. *Biochem Soc Trans* **42**: 1–0.
50. McDearmon EL, Patel KN, Ko CH, Walisser JA, et al. 2006. Dissecting the functions of the mammalian clock protein BMAL1 by tissue-specific rescue in mice. *Science* **314**: 1304–8.
51. Mohawk JA, Baer ML, Menaker M. 2009. The methamphetamine-sensitive circadian oscillator does not employ canonical clock genes. *Proc Natl Acad Sci USA* **106**: 3519–24.
52. Wijnen H, Young MW. 2006. Interplay of circadian clocks and metabolic rhythms. *Annu Rev Genet* **40**: 409–48.
53. Eckel-Mahan K, Sassone-Corsi P. 2009. Metabolism control by the circadian clock and vice versa. *Nat Struct Mol Biol* **16**: 462–7.
54. Bass J. 2012. Circadian topology of metabolism. *Nature* **491**: 348–56.
55. Masri S, Sassone-Corsi P. 2013. The circadian clock: a framework linking metabolism, epigenetics and neuronal function. *Nat Rev Neurosci* **14**: 69–75.
56. Bass J, Takahashi JS. 2010. Circadian integration of metabolism and energetics. *Science* **330**: 1349–54.
57. Rutter J, Reick M, McKnight SL. 2002. Metabolism and the control of circadian rhythms. *Annu Rev Biochem* **71**: 307–31.
58. Nakahata Y, Kaluzova M, Grimaldi B, Sahar S, et al. 2008. The NAD<sup>+</sup>-dependent deacetylase SIRT1 modulates CLOCK-mediated chromatin remodeling and circadian control. *Cell* **134**: 329–40.
59. Asher G, Gatfield D, Stratmann M, Reinke H, et al. 2008. SIRT1 regulates circadian clock gene expression through PER2 deacetylation. *Cell* **134**: 317–28.
60. Nakahata Y, Sahar S, Astarita G, Kaluzova M, et al. 2009. Circadian control of the NAD<sup>+</sup> salvage pathway by CLOCK-SIRT1. *Science* **324**: 654–7.
61. Sahar S, Masubuchi S, Eckel-Mahan K, Vollmer S, et al. 2014. Circadian control of fatty acid elongation by SIRT1 protein-mediated deacetylation of acetyl-coenzyme A synthetase 1. *J Biol Chem* **289**: 6091–7.
62. Ramsey KM, Yoshino J, Brace CS, Abrassart D, et al. 2009. Circadian clock feedback cycle through NAMPT-mediated NAD<sup>+</sup> biosynthesis. *Science* **324**: 651–4.
63. Asher G, Reinke H, Altmeyer M, Gutierrez-Arcelus M, et al. 2010. Poly(ADP-ribose) polymerase 1 participates in the phase entrainment of circadian clocks to feeding. *Cell* **142**: 943–53.
64. Peek CB, Affinati AH, Ramsey KM, Kuo HY, et al. 2013. Circadian clock NAD<sup>+</sup> cycle drives mitochondrial oxidative metabolism in mice. *Science* **342**: 1243417.
65. Asher G, Schibler U. 2011. Crosstalk between components of circadian and metabolic cycles in mammals. *Cell Metab* **13**: 125–37.
66. Ghosh A, Chance B. 1964. Oscillations of glycolytic intermediates in yeast cells. *Biochem Biophys Res Commun* **16**: 174–81.
67. Bier M, Bakker BM, Westerhoff HV. 2000. How yeast cells synchronize their glycolytic oscillations: a perturbation analytic treatment. *Biophys J* **78**: 1087–93.
68. Kloster A, Olsen LF. 2012. Oscillations in glycolysis in *Saccharomyces cerevisiae*: the role of autocatalysis and intracellular ATPase activity. *Biophys Chem* **165–166**: 39–47.
69. Zhang EE, Liu Y, Dentin R, Pongsawakul PY, et al. 2010. Cryptochrome mediates circadian regulation of cAMP signaling and hepatic gluconeogenesis. *Nat Med* **16**: 1152–6.
70. Akimoto H, Kinumi T, Ohmiya Y. 2005. Circadian rhythm of a TCA cycle enzyme is apparently regulated at the translational level in the dinoflagellate *Lingulodinium polyedrum*. *J Biol Rhythms* **20**: 479–89.
71. Guo J, Nguyen AY, Dai Z, Su D, et al. 2014. Proteome-wide light/dark modulation of thiol oxidation in cyanobacteria revealed by quantitative site-specific redox proteomics. *Mol Cell Proteomics* **13**: 3270–85.
72. Masri S, Patel VR, Eckel-Mahan KL, Peleg S, et al. 2013. Circadian acetylation reveals regulation of mitochondrial metabolic pathways. *Proc Natl Acad Sci USA* **110**: 3339–44.
73. Dallmann R, Viola AU, Tarokh L, Cajochen C, et al. 2012. The human circadian metabolome. *Proc Natl Acad Sci USA* **109**: 2625–9.
74. Davies SK, Ang JE, Revell VL, Holmes B, et al. 2014. Effect of sleep deprivation on the human metabolome. *Proc Natl Acad Sci USA* **111**: 10761–6.
75. Martinez-Lozano SP, Tarokh L, Li X, Kohler M, et al. 2014. Circadian variation of the human metabolome captured by real-time breath analysis. *PLoS ONE* **9**: e114422.
76. Yamazaki S, Numano R, Abe M, Hida A, et al. 2000. Resetting central and peripheral circadian oscillators in transgenic rats. *Science* **288**: 682–5.
77. Reppert SM, Weaver DR. 2001. Molecular analysis of mammalian circadian rhythms. *Annu Rev Physiol* **63**: 647–76.
78. Fuller PM, Gooley JJ, Saper CB. 2006. Neurobiology of the sleep-wake cycle: sleep architecture, circadian regulation, and regulatory feedback. *J Biol Rhythms* **21**: 482–93.
79. Deboer T, Vansteensel MJ, Detari L, Meijer JH. 2003. Sleep states alter activity of suprachiasmatic nucleus neurons. *Nat Neurosci* **6**: 1086–90.
80. Borbély AA. 1982. A two process model of sleep regulation. *Hum Neurobiol* **1**: 195–204.
81. Franken P, Dijk DJ. 2009. Circadian clock genes and sleep homeostasis. *Eur J Neurosci* **29**: 1820–9.
82. Franken P. 2013. A role for clock genes in sleep homeostasis. *Curr Opin Neurobiol* **23**: 864–72.
83. Shaw PJ, Tononi G, Greenspan RJ, Robinson DF. 2002. Stress response genes protect against lethal effects of sleep deprivation in *Drosophila*. *Nature* **417**: 287–91.
84. Wisor JP, Pasumarthi RK, Gerashchenko D, Thompson CL, et al. 2008. Sleep deprivation effects on circadian clock gene expression in the cerebral cortex parallel electroencephalographic differences among mouse strains. *J Neurosci* **28**: 7193–201.
85. Krueger JM, Obal F, Jr. 2003. Sleep function. *Front Biosci* **8**: d511–9.
86. Huber R, Ghilardi MF, Massimini M, Tononi G. 2004. Local sleep and learning. *Nature* **430**: 78–81.
87. Ferrara M, De GL. 2011. Going local: insights from EEG and stereo-EEG studies of the human sleep-wake cycle. *Curr Top Med Chem* **11**: 2423–37.
88. Vyazovskiy VV, Olcese U, Hanlon EC, Nir Y, et al. 2011. Local sleep in awake rats. *Nature* **472**: 443–7.
89. Krueger JM, Rector DM, Roy S, Van Dongen HP, et al. 2008. Sleep as a fundamental property of neuronal assemblies. *Nat Rev Neurosci* **9**: 910–9.
90. Hinard V, Mikhail C, Pradervand S, Curie T, et al. 2012. Key electrophysiological, molecular, and metabolic signatures of sleep and wakefulness revealed in primary cortical cultures. *J Neurosci* **32**: 12506–17.
91. Cirelli C, Gutierrez CM, Tononi G. 2004. Extensive and divergent effects of sleep and wakefulness on brain gene expression. *Neuron* **41**: 35–43.
92. Robles MS, Mann M. 2013. Proteomic approaches in circadian biology. *Handb Exp Pharmacol* **2013**: 389–407.
93. Moller M, Sparre T, Bache N, Roepstorff P, et al. 2007. Proteomic analysis of day-night variations in protein levels in the rat pineal gland. *Proteomics* **7**: 2009–18.
94. Deery MJ, Maywood ES, Chesham JE, Sladek M, et al. 2009. Proteomic analysis reveals the role of synaptic vesicle cycling in sustaining the suprachiasmatic circadian clock. *Curr Biol* **19**: 2031–6.

95. **Ong SE, Mann M.** 2005. Mass spectrometry-based proteomics turns quantitative. *Nat Chem Biol* 1: 252–62.
96. **Lee JE, Zamdborg L, Southey BR, Atkin N, Jr., et al.** 2013. Quantitative peptidomics for discovery of circadian-related peptides from the rat suprachiasmatic nucleus. *J Proteome Res* 12: 585–93.
97. **Lee JE, Atkins N, Jr., Hatcher NG, Zamdborg L, et al.** 2010. Endogenous peptide discovery of the rat circadian clock: a focused study of the suprachiasmatic nucleus by ultrahigh performance tandem mass spectrometry. *Mol Cell Proteomics* 9: 285–97.
98. **Seo HS, Hirano M, Shibato J, Rakwal R, et al.** 2008. Effects of coffee bean aroma on the rat brain stressed by sleep deprivation: a selected transcript- and 2D gel-based proteome analysis. *J Agric Food Chem* 56: 4665–73.
99. **Pawlyk AC, Ferber M, Shah A, Pack AI, et al.** 2007. Proteomic analysis of the effects and interactions of sleep deprivation and aging in mouse cerebral cortex. *J Neurochem* 103: 2301–13.
100. **Janich P, Arpat AB, Castelo-Szekely V, Lopes M, et al.** 2015. Ribosome profiling reveals the rhythmic liver transcriptome and circadian clock regulation by upstream open reading frames. *Genome Res* 25: 1848–59.
101. **Jang C, Lahens NF, Hogenesch JB, Sehgal A.** 2015. Ribosome profiling reveals an important role for translational control in circadian gene expression. *Genome Res* 25: 1836–47.
102. **Zhou M, Wang W, Karapetyan S, Mwimba M, et al.** 2015. Redox rhythm reinforces the circadian clock to gate immune response. *Nature* 523: 472–6.
103. **Yoshida Y, Iigusa H, Wang N, Hasunuma K.** 2011. Cross-talk between the cellular redox state and the circadian system in *Neurospora*. *PLoS ONE* 6: e28227.
104. **Gillette MU, Wang TA.** 2014. Brain circadian oscillators and redox regulation in mammals. *Antioxid Redox Signal* 20: 2955–65.
105. **Mann M, Jensen ON.** 2003. Proteomic analysis of post-translational modifications. *Nat Biotechnol* 21: 255–61.
106. **Kim EY, Edery I.** 2006. Balance between DBT/CKIepsilon kinase and protein phosphatase activities regulate phosphorylation and stability of *Drosophila* CLOCK protein. *Proc Natl Acad Sci USA* 103: 6178–83.
107. **Hirano A, Yumimoto K, Tsunematsu R, Matsumoto M, et al.** 2013. FBXL21 regulates oscillation of the circadian clock through ubiquitination and stabilization of cryptochromes. *Cell* 152: 1106–18.
108. **Cardone L, Hirayama J, Giordano F, Tamaru T, et al.** 2005. Circadian clock control by SUMOylation of BM AL1. *Science* 309: 1390–4.
109. **Eckel-Mahan KL, Patel VR, Mohnhey RP, Vignola KS, et al.** 2012. Coordination of the transcriptome and metabolome by the circadian clock. *Proc Natl Acad Sci USA* 109: 5541–6.
110. **Patel VR, Eckel-Mahan K, Sassone-Corsi P, Baldi P.** 2012. CircadiOmics: integrating circadian genomics, transcriptomics, proteomics, and metabolomics. *Nat Methods* 9: 772–3.
111. **Zhang R, Lahens NF, Ballance HI, Hughes ME, et al.** 2014. A circadian gene expression atlas in mammals: implications for biology and medicine. *Proc Natl Acad Sci USA* 111: 16219–24.
112. **Petit JM, Burlet-Godinot S, Magistretti PJ, Allaman I.** 2015. Glycogen metabolism and the homeostatic regulation of sleep. *Metab Brain Dis* 30: 263–79.
113. **Benington JH, Heller HC.** 1995. Restoration of brain energy metabolism as the function of sleep. *Prog Neurobiol* 45: 347–60.
114. **Van CE, Spiegel K, Tasali E, Leproult R.** 2008. Metabolic consequences of sleep and sleep loss. *Sleep Med* 9: S23–8.
115. **Grimaldi B, Bellet MM, Katada S, Astarita G, et al.** 2010. PER2 controls lipid metabolism by direct regulation of PPARgamma. *Cell Metab* 12: 509–20.
116. **Van der Horst GT, Muijtjens M, Kobayashi K, Takano R, et al.** 1999. Mammalian Cry1 and Cry2 are essential for maintenance of circadian rhythms. *Nature* 398: 627–30.
117. **Zaniolo K, Desnoyers S, Leclerc S, Guerin SL.** 2007. Regulation of poly(ADP-ribose) polymerase-1 (PARP-1) gene expression through the post-translational modification of Sp1: a nuclear target protein of PARP-1. *BMC Mol Biol* 8: 96.
118. **Hallows WC, Lee S, Denu JM.** 2006. Sirtuins deacetylate and activate mammalian acetyl-CoA synthetases. *Proc Natl Acad Sci USA* 103: 10230–5.
119. **Lamia KA, Sachdeva UM, DiTacchio L, Williams EC, et al.** 2009. AMPK regulates the circadian clock by cryptochrome phosphorylation and degradation. *Science* 326: 437–40.
120. **Liu C, Li S, Liu T, Borjigin J, et al.** 2007. Transcriptional coactivator PGC-1alpha integrates the mammalian clock and energy metabolism. *Nature* 447: 477–81.
121. **Rodgers JT, Lerin C, Haas W, Gygi SP, et al.** 2005. Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature* 434: 113–8.
122. **Kaasik K, Lee CC.** 2004. Reciprocal regulation of haem biosynthesis and the circadian clock in mammals. *Nature* 430: 467–71.
123. **Reinke H, Saini C, Fleury-Olela F, Dibner C, et al.** 2008. Differential display of DNA-binding proteins reveals heat-shock factor 1 as a circadian transcription factor. *Genes Dev* 22: 331–45.
124. **O'Neill JS, Maywood ES, Chesham JE, Takahashi JS, et al.** 2008. CAMP-dependent signaling as a core component of the mammalian circadian pacemaker. *Science* 320: 949–53.
125. **Lee B, Li A, Hansen KF, Cao R, et al.** 2010. CREB influences timing and entrainment of the SCN circadian clock. *J Biol Rhythms* 25: 410–20.
126. **Brunet A, Sweeney LB, Sturgill JF, Chua KF, et al.** 2004. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. *Science* 303: 2011–5.
127. **Frescas D, Valenti L, Accili D.** 2005. Nuclear trapping of the forkhead transcription factor FoxO1 via Sirt-dependent deacetylation promotes expression of glucogenetic genes. *J Biol Chem* 280: 20589–95.
128. **Sato TK, Panda S, Miraglia LJ, Reyes TM, et al.** 2004. A functional genomics strategy reveals Rora as a component of the mammalian circadian clock. *Neuron* 43: 527–37.
129. **Schmutz I, Ripperger JA, Baeriswyl-Aebischer S, Albrecht U.** 2010. The mammalian clock component PERIOD2 coordinates circadian output by interaction with nuclear receptors. *Genes Dev* 24: 345–57.