

ANDROLOGY

Anogenital distance as a marker of androgen exposure in humans

Journal:	<i>Andrology</i>
Manuscript ID	ANDR-2015-0369.R1
Manuscript Type:	Review Article
Date Submitted by the Author:	06-Dec-2015
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Key Words:	anogenital distance, endocrine disrupters, prenatal androgen exposure, masculinising programming window, testis development, testicular dysgenesis syndrome, TDS
<p>Note: The following files were submitted by the author for peer review, but cannot be converted to PDF. You must view these files (e.g. movies) online.</p> <p>Fig 1 _Schematic Diagram AGD landmarks.wmf Fig3_AGD hypocrypto_.EMF Fig.4 AGD hypocrypto.EMF</p>	

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1 **Anogenital distance as a marker of androgen exposure in humans**

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2
3 16 **Abstract:**
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5 17 Abnormal fetal testis development has been proposed to underlie common disorders of the male reproductive
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7 18 system such as cryptorchidism, hypospadias, reduced semen quality and testicular germ cell tumour, which
8
9 19 are regarded as components of a 'testicular dysgenesis syndrome'. The increasing trends and geographical
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11 20 variation in their incidence have been suggested to result from *in utero* exposure to environmental chemicals
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13 21 acting as endocrine disruptors. In rodents, the anogenital distance (AGD), measured from the anus to the base
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15 22 of genital tubercle, is a sensitive biomarker of androgen exposure during a critical embryonic window of testis
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17 23 development. In humans, several epidemiological studies have shown alterations in AGD associated with
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19 24 prenatal exposure to several chemicals with potential endocrine disrupting activity. However, the link between
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21 25 AGD and androgen exposure in humans is not well defined. This review focuses on the current evidence for
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23 26 such a relationship. As in rodents, a clear gender difference is detected during fetal development of the AGD
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25 27 in humans which is maintained thereafter. Reduced AGD in association with clinically relevant outcomes of
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27 28 potential environmental exposures, such as cryptorchidism or hypospadias, is in keeping with AGD as a
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29 29 marker of fetal testicular function. Furthermore, AGD may reflect variations in prenatal androgen exposure in
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31 30 healthy children as shorter AGD at birth is associated with reduced masculine play behaviour in preschool
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33 31 boys. Several studies provide evidence linking shorter AGD with lower fertility, semen quality and
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35 32 testosterone levels in selected groups of adults attending andrology clinics. Overall, the observational data in
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37 33 humans are consistent with experimental studies in animals and support the use of AGD as a biomarker of
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39 34 fetal androgen exposure. Future studies evaluating AGD in relation to reproductive hormones in both infants
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41 35 and adults, and to gene polymorphisms, will help to further delineate the effect of prenatal and postnatal
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43 36 androgen exposures on AGD.
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3 37 **Introduction:**
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5 38 Links between disorders of male reproductive system, such as reduced semen quality and testicular germ cell
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7 39 tumour (TGCC), as well as genital abnormalities at birth, such as cryptorchidism and hypospadias, are well
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9 40 established (Skakkebaek et al., 2001). These observations led to the hypothesis of ‘testicular dysgenesis
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11 41 syndrome’ (TDS) which proposes that abnormal testis development during fetal life is an important
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13 42 mechanism underlying common disorders of the male reproductive tract which manifest during infancy or
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15 43 adult life (Skakkebaek et al., 2001). Experimental animal studies, which used anti-androgens to alter fetal
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17 44 testis development, provide a compelling model to support the hypothesis (Dean and Sharpe, 2013). Although
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19 45 TGCC, one component of TDS, has not been replicated in animal models, this is perhaps due to species
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21 46 specificity (Juul et al., 2014). Rare disorders of sex development (DSD) due to a primary defect in testis
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23 47 determination, androgen secretion or androgen action manifest phenotypic features of TDS, supporting the
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25 48 relevance of the TDS model in humans (Hughes et al., 2007). The early origins of male reproductive disorders
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27 49 hypothesis is of relevance to public health as environmental exposure to potential endocrine disruptors *in*
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29 50 *utero* have been proposed to explain the increasing trends in the incidence and their marked geographical
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31 51 variation (Acerini and Hughes, 2006; Hauser et al., 2015). Estimating the burden of prenatal exposure to
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33 52 potential endocrine disruptors is a challenge, as congenital disorders of the male reproductive tract are
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35 53 relatively rare and disturbed reproductive function is likely to manifest a long time after the chemical
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37 54 exposure. In economic terms, an estimated annual cost to the European Union of 15 billion Euro has been
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39 55 calculated for the consequences of male reproductive disorders (e.g., treatments for infertility, orchidopexies,
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41 56 testis cancer) using the ‘Intergovernmental Panel on Climate Change’ weight-of-evidence characterization
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43 57 model for probability of causation (Hauser et al., 2015).
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47
48 59 The anogenital distance (AGD) measured from the anus to the genital tubercle is a sensitive biomarker of
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50 60 prenatal androgen action in animals (McIntyre et al., 2001; Mylchreest et al., 2000; Wolf et al., 2004). AGD
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52 61 signifies perineal growth and androgen-dependent caudal migration of the genital tubercle in rodents
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54 62 (Bowman et al., 2003). It is influenced by exposure to anti-androgens during a critical period of testis
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56 63 development known as the ‘masculinisation programming window’ (MPW) (van den Driesche et al., 2012;
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58 64 Welsh et al., 2008). Consequently, measurement of AGD has been used to study the effects of prenatal
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3 65 exposure to a variety of chemicals with potential endocrine disrupting activity. Several epidemiological
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5 66 studies have shown reduced AGD in association with exposure to phthalates (Adibi et al., 2015; Bornehag et
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7 67 al., 2015; Suzuki et al., 2012; Swan et al., 2005), dioxins (Vafeiadi et al., 2013), bisphenol A (Miao et al.,
8
9 68 2011) and high fat diet (Papadopoulou et al., 2013), the latter indicative of organic pollutant exposure. It is not
10
11 69 possible to conduct experimental toxicological studies in humans and therefore, the evidence for a causal
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13 70 effect of prenatal androgen exposure on AGD in humans is indirect (Welsh et al., 2007). This review focuses
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15 71 on the evidence linking AGD and androgen exposure in humans.
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19 73 **Physiological aspects of androgen exposure in the male**

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21 74 Leydig cells differentiate at approximately 8 weeks of gestation and secrete testosterone to mediate
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23 75 differentiation of the internal and external genitalia (Grinspon et al., 2014). Testosterone production is initially
24
25 76 regulated during the first trimester by human chorionic gonadotrophin (hCG), which reaches a peak
26
27 77 concentration at 12-17 weeks and subsequently declines (Cole, 2010). Development of the male genitalia is
28
29 78 completed by 14-16 weeks (Grinspon et al., 2014; Welsh et al., 2008). Fetal luteinising hormone (LH)
30
31 79 regulates testosterone secretion from the second trimester which induces further growth of the phallus and
32
33 80 scrotum, and together with insulin-like factor 3 (INSL-3), promotes testis descent (Asa et al., 1991; Bay et al.,
34
35 81 2007). After a surge of LH and testosterone secretion following delivery, testosterone levels decline in the
36
37 82 first week of life (Corbier et al., 1990). Further activation of the hypothalamic-pituitary-gonadal axis known
38
39 83 as the mini-puberty, starts at the end of the first week, peaking at 1-3 months of age before declining to low or
40
41 84 undetectable levels by 6 months of age (Bergada et al., 2006; Forest et al., 1974; Kuiri-Hanninen et al., 2014).
42
43 85 Subsequently, the axis remains quiescent until the onset of puberty. The observation of a MPW in the rat
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45 86 (equivalent to 8-14 weeks of gestation in the human) suggests that alterations in androgen action during a
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47 87 critical window of fetal development results in TDS and permanent changes in AGD (Welsh et al., 2008).
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51 89 **Definition and measurement methods**

52
53 90 In rodents, AGD is measured from the anus to the posterior base of the genital tubercle (Gallavan et al., 1999).
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55 91 In contrast to rodents, the external genitalia are well developed at birth in humans with the genital tubercle
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57 92 transformed into the penis in males and clitoris in females. Investigators have used different landmarks to
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3 93 measure AGD in humans to replicate the measurement in rodents. In males, AGD has been measured from the
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5 94 anus to the perineoscrotal junction (anoscrotal distance) (Salazar-Martinez et al., 2004), to the posterior base
6
7 95 or to the anterior base of the penis (Hsieh et al., 2008) (Fig.1). Measurements in females use the distance from
8
9 96 the anus to the anterior fourchette (anofourchettal distance) (Salazar-Martinez et al., 2004) or to the base of
10
11 97 the clitoris (anoclitoral distance) (Liu et al., 2014). The method described by Salazar-Martinez et al. is
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13 98 commonly used; it is more reliable and has a lower inter-observer variability (Dean and Sharpe, 2013;
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15 99 Papadopoulou et al., 2013; Salazar-Martinez et al., 2004). In this review, the term AGD describes ‘anoscrotal
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17 100 distance’ in males and ‘anofourchettal distance’ in females unless otherwise stated (Salazar-Martinez et al.,
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19 101 2004). Although AGD has been widely used as a marker of potential endocrine disruption *in utero*, its
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21 102 limitations include a lack of standardisation of methodology and information on reproducibility (Table-1) and
22
23 103 insufficient data on normative references, including ethnic differences (Dean and Sharpe, 2013). AGD is
24
25 104 associated with birth weight to a varying degree depending on the population studied (regression coefficient
26
27 105 adjusted for gestation ranges from 1.5 to 3.0 mm/kg) (Papadopoulou et al., 2013; Romano-Riquer et al., 2007;
28
29 106 Salazar-Martinez et al., 2004) and there is no consensus for adjusting AGD for the variations in body size. In
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31 107 addition, low birth weight is itself a risk factor for TDS as it is associated with hypospadias, cryptorchidism,
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33 108 male infertility and TGCC (Francois et al., 1997; Juul et al., 2014; Michos et al., 2007; Toppari et al., 2010).
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37 110 Fig. 1

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39 111 Table -1
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43 113 **Associations between AGD, gender and age in healthy individuals**

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45 114 The AGD in rodents is approximately twice as long in males compared to females, and is routinely used to
46
47 115 determine sex (Dean and Sharpe, 2013). We and others have reported that AGD is also sexually dimorphic in
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49 116 humans and displays a similar relative magnitude of difference (male: female ratio ranges from 1.4:1 to 2.2:1)
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51 117 (Table-1) (Huang et al., 2008; Papadopoulou et al., 2013; Salazar-Martinez et al., 2004; Sathyanarayana et al.,
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53 118 2010; Sathyanarayana et al., 2015; Thankamony et al., 2009). The sexual dimorphism is already present by
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55 119 gestation 11-13 weeks based on fetal imaging and, by weeks 17-20, it is of the same magnitude as that
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3 120 observed at birth, with a male: female ratio of 2:1 (Fowler et al., 2011). The latter time point corresponds to
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5 121 the completion of differentiation of the external genitalia in humans (Welsh et al., 2008).
6
7 122 A large cohort (n=925) from the Cambridge Birth Growth Study (CBGS) had AGD measurements performed
8
9 123 at birth and at ages 3, 12, 18 and 24 months (Thankamony et al., 2009). AGD increased rapidly during the first
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11 124 3 months and plateaued after 1 year of age (Fig. 2). Using longitudinal and cross-sectional data from two birth
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13 125 cohorts in Greece and Spain, Papadopoulou et al. confirmed this pattern of growth. The sex dimorphism was
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15 126 maintained throughout to a similar degree to that observed at birth (male: female ratio; at birth, 2.2:1; age 24
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17 127 months; 2.3:1) (Thankamony et al., 2009). The similar proportional increase in AGD in boys and girls during
18
19 128 the first two years suggests the growth of perineum in proportion to overall body size. However, the period of
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21 129 rapid increase in AGD and penile length during the first three months of life corresponds to the mini-puberty
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23 130 (Kurtoglu and Bastug, 2014). We also found a modest association between increments in AGD and penile
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25 131 length during this period independent of the changes in body size, suggesting the postnatal surge in
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27 132 testosterone production may also contribute to changes in AGD (Thankamony et al., 2009). A positive
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29 133 association has been found between penile growth and serum testosterone levels during the first three months
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31 134 of life (Boas et al., 2006). This observation, coupled with animal data showing changes in AGD when
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33 135 postnatal androgen exposure is altered (Mitchell et al., 2015), supports the hypothesis that the mini-puberty
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35 136 plays a part in postnatal AGD development. Further studies are needed to delineate mini-puberty and its AGD
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37 137 component using detailed anthropometry, as well as more frequent hormone measurements which may require
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39 138 novel methods such as dried blood spot analytical technology (McDade et al., 2007).
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48 142 Longitudinal data for AGD measurements from infancy to adulthood are not available. However, cross-
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50 143 sectional data in young adults show that large increases occur in later life and sexual dimorphism is
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52 144 maintained to a lesser degree (ranges; males, 48.3 - 51.3 mm; females, 34.8 - 37.7 mm; male: female ratio
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54 145 calculated from the means from different studies, 1.4:1) (Lee et al., 2015; Mendiola et al., 2012; Mendiola et
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56 146 al., 2011; Parra et al., 2015). When these changes occur is not known. We speculate that large increases in
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58 147 AGD occur during puberty in association with the development of the external genitalia. AGD remain
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3 148 unchanged in adult males, however, there are data to suggest that AGD decreases in females following the
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5 149 menopause (Eisenberg et al., 2013; Lee et al., 2015).
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8 9 151 **Cryptorchidism and hypospadias**

10 152 Cryptorchidism and hypospadias are the most common genital anomalies at birth with incidences of 2-9% and
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12 153 0.2-1%, respectively, and provide an important outcome for studies of prenatal exposure to endocrine
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14 154 disrupting chemicals (Toppari et al., 2010). Establishing the relationship between AGD and congenital
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16 155 anomalies of the male reproductive tract at birth is key to determining whether AGD has a role as a biomarker
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18 156 of such prenatal exposure (Thankamony et al., 2014). A cross-sectional population study reported that boys
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20 157 with cryptorchidism have a shorter age-adjusted 'anogenital index' (a derivative of AGD adjusted for weight)
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22 158 (Swan et al., 2005). In boys undergoing surgery for hypospadias (n=26), AGD was measured under
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24 159 anaesthesia and found to be shorter compared with age-matched controls who had other urological conditions
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26 160 (Hsieh et al., 2012). In a larger study, we compared boys aged up to two years with cryptorchidism (n=71) or
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28 161 hypospadias (n=81) referred for surgical treatment with healthy controls from a birth cohort (n=482) by
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30 162 deriving age-specific standard deviation scores (SDS) of AGD and penile length (Thankamony et al., 2014).
31
32 163 AGD measurements in boys with cryptorchidism (-0.48 SDS) or hypospadias (-0.90 SDS) were significantly
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34 164 lower compared with healthy controls (+0.03 SDS) (Fig. 3 & 4). They also had a shorter penile length
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36 165 (cryptorchidism, -0.35 SDS; hypospadias, -1.34 SDS) compared with healthy boys (-0.02 SDS). Boys with
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38 166 hypospadias also had smaller overall body size than controls, consistent with the well-documented prevalence
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40 167 of low birth weight in idiopathic hypospadias (Jensen et al., 2012). However, the reduction in AGD and penile
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42 168 length persisted following adjustment for body size in a multiple regression model. The observations of
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44 169 reduced AGD have also been reported in a large population-based study (boys with cryptorchidism, n=51;
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46 170 controls, n=534) from India involving consecutively born term male neonates evaluated at birth (Jain and
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48 171 Singal, 2013).
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54 173 Fig. 3

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56 174 Fig. 4

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3 176 Hypospadias and cryptorchidism are relevant clinical disorders to evaluate the possible role of endocrine
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5 177 disruption, for which there is ample evidence from animal studies (van den Driesche et al., 2012; Welsh et al.,
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7 178 2008). The evidence to support a link between these disorders and altered testis function *in utero* in humans
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9 179 relies on epidemiological studies (Toppari et al., 2010). These common disorders are sometimes the
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11 180 manifestations of rare, but, well-defined disorders of androgen production or action such as androgen receptor
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13 181 mutations. However, in the majority of cases the cause is unknown (Toppari et al., 2010). The link between
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15 182 reduced semen quality and TGCC (Skakkebaek et al., 2001), and the association between cryptorchidism and
16
17 183 lower INSL3 or higher gonadotrophin levels (Bay et al., 2007; Suomi et al., 2006), attest to underlying testis
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19 184 dysfunction. Cryptorchidism is considered to be the result of a milder defect in androgen function as most of
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21 185 the cases are due to impaired inguinoscrotal descent which normally occurs between 26 and 40 weeks of
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23 186 gestation, compared with hypospadias which occurs earlier during the MPW (between 8 and 20 weeks)
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25 187 (Thorup et al., 2010). Reported trends towards a shorter AGD and penile length with increasing severity of
26
27 188 hypospadias (Thankamony et al., 2014), and higher testis position in cryptorchidism (Jain and Singal, 2013),
28
29 189 further suggest that AGD is a marker of the severity of impairments in androgen production. It is now
30
31 190 recognised that approximately half of all cases of cryptorchidism are “acquired” (i.e. following normal
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33 191 positioning of the testes at birth), hitherto referred to as ‘the ascending testis’ (Hack et al., 2012). It is possible
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35 192 that this form of cryptorchidism may be associated with suboptimal testosterone production during mini-
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37 193 puberty (Acerini et al., 2009; Wohlfahrt-Veje et al., 2009). Larger studies of boys with cryptorchidism may
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39 194 indicate whether AGD measurements sub-divided according to congenital versus acquired forms reflect
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41 195 altered androgen action occurring *in utero* or during the early postnatal period.
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197 **Gender-typical behaviour**

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47 198 A relationship between prenatal androgen exposure and gender-related behaviour is well established in
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49 199 animals and humans (Hines et al., 2015). The evidence to support this in humans is mainly derived from
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51 200 studies of girls with congenital adrenal hyperplasia (CAH) who have been exposed to higher androgen levels
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53 201 during intrauterine life. Females with CAH show greater preference for boys’ toys and activities during
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55 202 childhood, have lower heterosexual orientation and are less feminine in their gender identity (Berenbaum and
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57 203 Hines, 1992; Hines et al., 2004; Pasterski et al., 2011; Pasterski et al., 2007; Pasterski et al., 2005). In
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3 204 addition, androgen production during early infancy, as measured by urinary testosterone levels, is related to
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5 205 increased masculine play behaviour in healthy boys at 14 months of age (Lamminmaki et al., 2012). It is not
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7 206 clear, however, whether the observation is solely related to postnatal androgen exposure. We studied gender-
8
9 207 typed play behavior in healthy boys at age 3-4 years (n=81) in relation to longitudinal measurements of AGD
10
11 208 and penile length during their first two years of life (Pasterski et al., 2015). Gender- related play behaviour
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13 209 was evaluated using the 'Preschool Activities Inventory' which consists of a validated 24-item parent
14
15 210 questionnaire that assesses gender-typed toy and activity preferences (Golombok and Rust, 1993). Both AGD
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17 211 at birth, and penile growth during the first 3 months of life, independently predicted masculine behavior in a
18
19 212 regression model controlling for overall changes in body size. As expected, AGD was not related to gender
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21 213 type play behaviour in girls. These findings provide evidence that AGD in healthy boys at birth reflects
22
23 214 prenatal androgen exposure, whereas the rate of penile growth in early infancy reflects more the effect of early
24
25 215 postnatal androgen exposure during mini-puberty. Taken together, both parameters have potential use as
26
27 216 biomarkers of endocrine disruption during pre- and early post-natal development.
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218 **Disorders of sex development (DSD)**

219 Disorders of sex development are defined as congenital conditions in which development of chromosomal,
220 gonadal, or anatomical sex is atypical and include several disorders with well-defined alterations in production
221 or action of sex steroids (Hughes et al., 2006). Although such presentations provide opportunities to study
222 AGD as part of a phenotype, there are difficulties in identifying the landmarks for measurement of AGD,
223 particularly in the in severe forms where there are genital ambiguities. An increased AGD has been reported
224 in a small study of girls with CAH (Callegari et al., 1987). The complete form of androgen insensitivity
225 syndrome (CAIS) is an example of XY sex reversal due to mutations in the androgen receptor gene resulting
226 in resistance to androgen action (Hughes et al., 2012). As expected, an androgen receptor knock-out mouse
227 model (the ARKO mouse) showed a female phenotype in ARKO males, including AGD similar to female
228 mice (Yeh et al., 2002). No data are available for AGD in women with CAIS using the commonly used
229 method of measurement discussed in this review. However, in a heterogenous group of 19 women with a
230 clinical diagnosis of CAIS examined during a routine clinic visit, clitoral length was reduced but the clitoral to
231 urethral distance (measured from the base of the clitoris to the anterior aspect of the external urethral meatus)

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3 232 was similar to a control group of women undergoing a gynaecological examination under anaesthesia (Crouch
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5 233 et al., 2011). It is not clear whether the segment of perineum represented by this measurement truly reflects
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7 234 the AGD parameter which is accepted as being androgen dependent in both animal models and humans.
8
9 235 Studies are required in the partial form of androgen insensitivity syndrome (PAIS) or hypogonadotropic
10
11 236 hypogonadism (where impaired androgen production occurs in late gestation) to determine if these, and other
12
13 237 examples of DSD, provide information on assessing AGD as a quantitative measure of prenatal androgen
14
15 238 exposure. Female rhesus monkeys exposed to androgens early in gestation exhibited hyperandrogenism,
16
17 239 oligomenorrhea, large polyfollicular ovaries and other features consistent with the polycystic ovarian disease
18
19 240 (PCOS) phenotype seen in humans (Abbott et al., 2005). The effects of such exposure *in utero* to higher
20
21 241 androgens on AGD in female offspring of mothers with conditions such as CAH or PCOS merit study. A
22
23 242 cross-sectional study of healthy young women showed greater follicular numbers were associated with longer
24
25 243 AGD which, in turn, was also positively associated with higher serum testosterone levels (Mendiola et al.,
26
27 244 2012; Mira-Escolano et al., 2014).

245

246 **Testis function in adults and AGD**

247 Evidence has been accumulating in several countries since it was first reported that sperm quality in men is in
248 decline (Carlsen et al., 1992). Impaired semen quality, an important marker of testis function and component
249 of TDS, is now reported in up to 20% of otherwise healthy young men (Andersson et al., 2008; Jorgensen et
250 al., 2006). Shorter AGD is associated with lower sperm concentration, total sperm count, sperm motility and
251 also testosterone levels in men attending andrology clinics (Eisenberg et al., 2011; Eisenberg and Lipshultz,
252 2015; Eisenberg et al., 2012; Mendiola et al., 2015). While a longer AGD is associated with a higher sperm
253 count and better semen quality, the strength of association is insufficient to predict male fertility in an
254 individual (Eisenberg and Lipshultz, 2015). A short AGD does distinguish men with non-obstructive causes of
255 azoospermia (which is associated with TDS) from those with obstructive azoospermia (Eisenberg et al., 2012).
256 These findings lend support for AGD as a biomarker for TDS and testis function in selected populations, but,
257 few studies have explored the link between AGD and the larger range in testis size and semen quality in
258 healthy males. A shorter AGD in association with lower semen quality has been reported in a US study of
259 healthy young men (Mendiola et al., 2011); however, the results were not replicated in a similar study

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3 260 conducted in Spanish population (Parra et al., 2015). Report of a lower AGD measured from the anus to the
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5 261 anterior penile base in patients with prostate cancer compared to men with other causes of lower urinary tract
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7 262 symptoms (Castaño-Vinyals et al., 2012) suggest a possible link between alterations in the androgen-
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9 263 dependent fetal development of prostate (Wilson, 2011) and the cancer risk. However, anoscrotal distances
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11 264 were not different, and the findings of this small study need to be confirmed in future studies.
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15 266 Although testis function is a key component of TDS, its relationship with AGD in adults is somewhat
16
17 267 conflicting. Whereas studies in selected groups of men attending andrology clinics showed consistent
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19 268 associations between shorter AGD and lower semen quality or testosterone levels (Eisenberg et al., 2011;
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21 269 Eisenberg and Lipshultz, 2015; Eisenberg et al., 2012; Mendiola et al., 2015), these relations were inconsistent
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23 270 in healthy men with only one of the two studies showing an association with semen quality and none showed a
24
25 271 relationship with testosterone levels (Mendiola et al., 2011; Parra et al., 2015). We speculate that the link
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27 272 between AGD and testis function is more robust in adults at the lower end of the distribution of reproductive
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29 273 health (possibly reflecting a high prevalence of testicular dysgenesis) compared to healthy men.
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31 274

33 275 **Genetic variations and AGD**

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35 276 As previously discussed, establishing a direct link between prenatal androgen exposure and AGD is difficult
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37 277 in humans, and the current evidence has relied mainly on data from observational epidemiological studies.
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39 278 Novel study designs using genes as instruments for causal inferences (i.e., Mendelian Randomisation) as has
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41 279 been applied to the study of cardiovascular disease (Thanassoulis, 2013) could provide additional strength to
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43 280 the evidence linking AGD and androgen exposure. Gene polymorphisms associated with TGCC, testosterone
44
45 281 levels and male fertility and based on large genome-wide association studies (GWAS) are now available and
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47 282 may provide suitable instruments to test this hypothesis (Kosova et al., 2012; Litchfield et al., 2015; Ruth et
48
49 283 al., 2015). A similar approach on the basis of candidate gene analysis has found a variant in the estrogen
50
51 284 receptor alpha gene (*ESR1*) associated with shorter AGD (Sathyanarayana et al., 2012). Polymorphisms in
52
53 285 *ESR1* are associated with hypospadias, male infertility and alterations in semen parameters (Ban et al., 2008;
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55 286 Safarinejad et al., 2010). Measurement of the number of CAG repeats in the androgen receptor gene (*AR*) in a
56
57 287 cohort of adult males attending a urology clinic showed that increased CAG lengths were associated with
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3 288 shorter AGD when the data were analysed in a stratified model (Eisenberg et al., 2013), but this finding was
4
5 289 not replicated in another study (Sathyanarayana et al., 2012). There is an inverse association between the
6
7 290 length of the CAG repeats and transcriptional activity of the androgen receptor as measured *in vitro*
8
9 291 (Chamberlain et al., 1994), and when studied in relation to sperm quality, longer CAG repeats were associated
10
11 292 with reduced sperm quality in some studies (Milatiner et al., 2004; Wallerand et al., 2001) but not in others
12
13 293 (Dadze et al., 2000; Singh et al., 2006).

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17 295 **Conclusion**

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19 296 There is considerable observational evidence in humans that supports a link between AGD and exposure to
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21 297 androgens during fetal life. The findings are consistent with animal data that show a critical MPW during fetal
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23 298 male sex development and the programming of AGD. A plethora of studies are now reporting population data
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25 299 for AGD and applying the methods to assess testicular function and androgen action across a wide range of
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27 300 clinical disorders. AGD also appears to be a valid biomarker to assess the effects of an adverse environment
28
29 301 on human reproductive development from fetal to adult life. Gathering such information will rely on
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31 302 epidemiological studies of birth cohorts followed longitudinally with detailed anthropometric measurements
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33 303 and analysis of targeted chemicals in appropriate biological samples.

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37 305 **Acknowledgements**

38
39 306 The CBGS studies referred to in this review were supported by a European Union Framework V programme,
40
41 307 the World Cancer Research Fund International, the Medical Research Council (UK), the Newlife Foundation,
42
43 308 the Mothercare Foundation, the Evelyn Trust and the NIHR Cambridge Biomedical Research Centre.

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47 310 **Disclosure Summary:** The authors have no conflicts of interest.

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51 312 **Authorship**

52
53 313 AT wrote the original manuscript, IAH substantially modified and restructured the manuscript. VP, KKO and
54
55 314 CA critically reviewed and substantially edited the manuscript.

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For Peer Review

Table-1 Normative data for anogenital Distance (mm) at birth showing gender differences

Reference*	Country	n	Male	Female	Male: Female Ratio
Salazar-Martinez et al., 2004	Mexico	87	21±3	11±2	1.9
Thankamony et al., 2009	UK	564	19.8±6.1	9.1±2.8	2.2
Huang et al., 2009	Taiwan	65	23 (10-36)	16 (7-23)	1.4
Sathyanarayana et al., 2010	USA	169	23±4	15±3	1.5
Papadopoulou et al., 2013	Greece	165	27.1±4.4	14.4±3.0	1.9
Papadopoulou et al., 2013	Spain	187	23.6±5.1	13.8±2.5	1.7
Sathyanarayana et al., 2015	USA	758	24.7±4.5	16.0±3.2	1.5

Means±SD or medians (range); The measurements correspond to the anoscrotal distance in boys and anofourchettal distance in girls. * studies reporting measurements in both genders at birth are included in the table

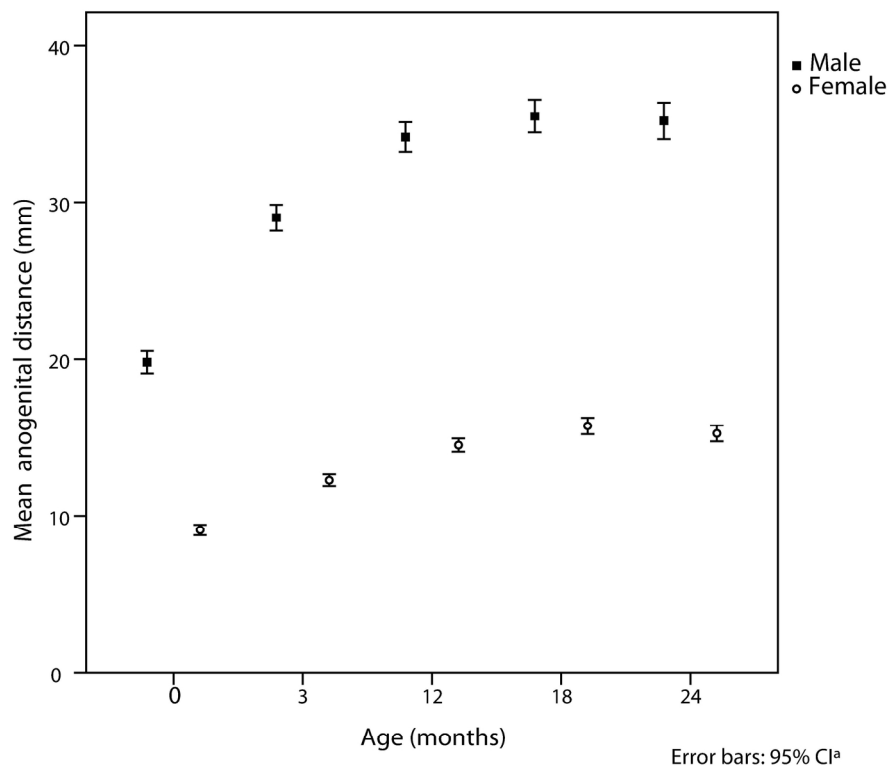


Fig. 2 Longitudinal measurements of AGD in males (n=463) and females (n=426) from birth to 2 years of age. Data presented as means and error bars represent 95% Confidence Intervals. Reproduced with permission from EHP (Thankamony et al., 2009).
172x160mm (300 x 300 DPI)