http://www.ehu.es/histol-histopathol

Cellular and Molecular Biology

Studies on the involvement of endogenous neuropeptides in the control of thymocyte proliferation in the rat

M. Trejter¹, J.B. Warchol², R. de Caro³, R. Brelinska¹, G.G. Nussdorfer³ and L.K. Malendowicz¹

¹Department of Histology and Embryology and ²Department of Radiology and Cell Biology, School of Medicine, Poznan Poland and ³Department of Human Anatomy and Physiology, Section of Anatomy, University of Padua, Padua, Italy

Summary. The possible involvement of endogenous vasoactive intestinal peptide (VIP), cholecystokinin (CCK) and neurotensin (NT) in the control of thymocyte proliferation has been investigated in vivo in the immature rat. For this task, we have studied the effects of the administration of selective antagonists of the receptors of the three neuropeptides on the mitotic index (% of metaphase-arrested cells after vincristin injection) of thymocytes. Both CCK- and TN-receptor antagonists were ineffective. In contrast, two VIP receptor antagonists (VIP-As) enhanced the mitotic index of thymocytes. VIP reversed the effect of VIP-As, but when administered alone it did not alter the mitotic activity of thymocytes. In light of these findings, we conclude that endogenous VIP exerts a maximal tonic inhibitory influence on the basal proliferative activity of rat thymocytes, while endogenous CCK and NT do not play a relevant modulatory role in this process.

Key words: Thymus, Cell proliferation, Vasoactive intestinal peptide, Cholecystokinin, Neurotensin, Rat

Introduction

Several regulatory neuropeptides and their receptors are present in the various components of the immune system, where they are able to modulate some important steps of the immune response. One of these steps is the proliferation of immune competent cells, among which are thymocytes (Savino et al., 1990; Dardenne and Savino, 1994; Head et al., 1998).

The characteristic cytoarchitecture of the thymus is responsible for the maintenance of a unique microenvironment, which seems to be essential for the correct development of T cells (Brelinska and Warchol, 1997). This makes the use of the cell culture techniques unsuitable in the studies on the control of thymocyte proliferation. Moreover, the *in vivo* administration of exogenous neuropeptides do not provide reliable information on the possible physiological role of their endogenous counterparts in the control of thymocyte proliferation.

Bearing these limitations in mind, we designed experiments where the involvement of endogenous vasoactive intestinal peptide (VIP), cholecystokinin (CCK)/gastrin and neurotensin (NT), three neuropeptides contained in the immune system (see Discussion), in the control of thymocyte proliferation has been investigated by administering rats with their selective receptor antagonists.

Materials and methods

Animals and reagents

Immature (20-day-old) female Wistar rats were kept under a 12:12 h light-dark cycle (illumination onset at 8:00 a.m.) at 23 ± 1 °C, and maintained on a standard diet and tap water *ad libitum*.

VIP(1-28) rat, the VIP antagonists neurotensin(6-11)/VIP(7-28) (Moody et al., 1993) (VIP-A1), and [Ac-His¹,D-Phe²,Lys¹⁵,Arg¹⁶]VIP(3-7)GRF(8-27)-NH₂ (VIP-A2) (Gourlet et al., 1997), and pentagastrin (PG) were purchased from Bachem (Bubendorf, Switzerland). The pituitary adenylate cyclase-activating polypeptide (PACAP) antagonist PACAP(6-38) (Dickinson et al., 1997) was obtained from Peninsula (St. Helens, UK). The CCK receptor A and B antagonists PD140,548 (CCKA-A) and PD135,158 (CCKB-A), respectively (Hughes et al., 1990; Higginbottom et al., 1993) were obtained from Research Biochemical International (Natick, Mass., USA). NT and the NT antagonist (NT-A) [D-Trp¹]-NT (Quirion et al., 1980), as well as other laboratory reagents were supplied by Sigma Chemical Co. (St. Louis, MO, USA). Vincristin was obtained from Gedeon-Richter (Budapest, Hungary).

Offprint requests to: Prof. G.G. Nussdorfer, Department of Human Anatomy and Physiology, Section of Anatomy, Via Gabelli 65, I-35121 Padova, Italy. Fax: (+39) 049-827-2319. e-mail: ggnanat@ ipdunidx.unipd.it

Neuropeptides and the control of thymocyte proliferation

Experimental procedures

Groups of immature rats (n=6) were given three subcutaneous injections (28, 16 and 4 h before sacrifice) of the following chemicals dissolved in 0.2 ml 0.9% NaCl: (i) VIP and/or VIP-A1, VIP-A2 and PACAP(6-38) (20 nmol/kg); (ii) CCKA-A or CCKB-A alone or with PG (20 nmol/kg); and (iii) NT and/or NT-A (40 nmol/kg). Control rats were injected with the saline vehicle. All groups of animals received an intraperitoneal injection of vincristin (0.1 mg/100 g) 180 min before the autopsy. Rats were decapitated at 11:00 a.m.

Cell-proliferation assay

Thymuses were promptly removed, and capsuleadjacent fragments were fixed in 2.5% glutaraldehyde, postfixed in 1% osmium tetroxide, and embedded in Araldite. Then, 0.5 μ m-thick sections were cut, and stained with toluidine blue. The mitotic index (% of metaphase-arrested cells) was calculated at x400, by counting 5,000 cells in the subcapsular zone (4-5 layers of cells) of each thymus.







Fig. 2. Lack of effect of endogenous NT on the proliferative activity of immature rat thymocytes. Bars are means \pm SEM (n=6).

Statistical analysis

Individual results were averaged per experimental group, and SEM was calculated. The statistical comparison of the data was done by ANOVA, followed by the Multiple Range test of Duncan.

Results

Neither PG nor CCKA-A and CCKB-A, administered alone or together, affected the proliferative activity of rat thymocytes (Fig. 1). Likewise, NT and/or NT-A did not change thymocyte mitotic index (Fig. 2). VIP-A1 and VIP-A2 evoked a marked increase in the mitotic index of thymocytes, and the effect was prevented by the simultaneous injection of VIP. VIP and PACAP(6-38) were ineffective (Fig. 3).

Discussion

The present results indicate that of the three neuropeptides investigated only VIP plays a physiological role in the control of thymocyte proliferation.

CCK is a 33-amino acid residue peptide, that is widely distributed in the central nervous system and peripheral organs and tissues, where it acts through two main subtypes of receptors, named CCKA and CCKB



Fig. 3. Effect of endogenous VIP on the proliferative activity of immature rat thymocytes. Bars are means \pm SEM (n=6). *: p<0.01 from the respective control group.

receptors belonging to the G protein-coupled receptor superfamily. PG is a selective and potent agonist of CCKB receptors (Wank et al., 1992; Pisegna et al., 1993; Crawley and Corvin, 1994). The following evidence indicates that CCK is involved in immunomodulation. CCK and its receptors are present in the lymphoid tissue, including thymus (Felten et al., 1985; Weinberg et al., 1997), and CCK restores thymus-dependent immune response in thymectomized mice and stimulates IgMplaque forming cells (Belokrylov et al., 1990; Molchanova et al., 1992). Soder and Hellstrom (1987) reported that neither CCK nor PG affect the proliferative activity of human thymocytes or guinea pig T and B lymphocytes cultured in vitro. In contrast, De la Fuente et al. (1998) demonstrated an inhibitory effect of CCK on mitogen-induced proliferation of murine lymphocytes in vitro. Our previous in vivo studies (Malendowicz et al., 1999) showed a potent CCKA receptor-mediated stimulatory effect of exogenous CCK on the mitotic activity of rat thymocytes. However, our present findings strongly suggest that this effect of CCK has to be considered pharmacological in nature. In fact, neither CCKA-A and CCKB-A nor PG administration significantly affect thymocyte proliferation, thereby ruling out the possibility that endogenous CCK and gastrin play a relevant physiological modulatory role in this process.

NT, a 13-amino acid residue peptide that acts through G protein-coupled receptors (Vincent et al., 1999), is contained in the immune system. NTimmunoreactivity has been detected in the chicken (Atoji et al., 1996) and human thymus (Vanneste et al., 1997), and NT receptor mRNA in the medulla and epithelial cells of the chicken thymus (Atoji et al., 1996). NT was found to stimulate [³H]-thymidine incorporation in human thymocytes and to inhibit it in guinea pig lymph node cells (Soder and Hellstrom, 1987). Our present findings cast doubts on the involvement of endogenous NT in the regulation of thymocyte mitogenic activity in the rat, inasmuch as neither NT nor NT-A affect it. This observation appears to accord well with the reported lack of NT-immunoreactivity in the rat thymus (Muller and Weihe, 1991; Atoji et al., 1996).

VIP is a 28-amino acid residue peptide, that displays a remarkable amino acid-sequence homology with PACAP (Nussdorfer and Malendowicz, 1998). VIP and PACAP act through G protein-coupled receptors, named VIP-PACAP receptors. Three subtypes of VIP-PACAP receptors have been identified so far: the PAC1, VPAC1 and VPAC₂ receptors (Harmar et al., 1998). Their binding potency is as follows: PAC1, PACAP>>>VIP; VPAC1, VIP>PACAP; and VPAC2, VIP=PACAP (Nussdorfer and Malendowicz, 1998). VIP is abundant in the immune system (Martinez et al., 1999) and VIPergic fibers are present in the subcapsular cortex, interlobular septa and accompanying vasculature of the thymus (Gomariz et al., 1990, 1993; Kendall and Al-Shawaf, 1991; Müller and Weihe, 1991; Bellinger et al., 1997). Lymphocytes and thymocytes are provided with

VPAC₁ and VPAC₂ receptors (Delgado et al., 1996). VIP has been reported to inhibit $[{}^{3}\text{H}]$ -thymidine uptake by non-activated and activated human thymocytes, as well as basal DNA synthesis in rat spleen cells (Soder and Hellstrom, 1987; Yiangou et al., 1990; Boudard and Bastide, 1991; De la Fuente et al., 1996; Ganea, 1996; Pankhaniya et al., 1998).

The present demonstration that VIP-As enhance the mitotic activity of rat thymocytes, while VIP per se is ineffective strongly suggests that endogenous VIP exerts a maximal tonic inhibition of the thymocyte proliferative activity. This tonic inhibition is conceivably mediated by the VPAC1 receptor subtype. In fact, the selective antagonist of VPAC1 receptors VIP-A2 (Gourlet et al., 1997) raises thymocyte proliferation, while the selective antagonist of PAC1 receptors PACAP(8-36), that also interacts with VPAC2 receptors (Dickinson et al., 1997), is ineffective. The physiological and physiopathological relevance of this VIP effect on the rat thymus growth remains to be investigated.

References

- Atoji Y., Yamamoto Y. and Suzuki Y. (1996). Neurotensin-containing cells and neurotensin receptor mRNA-expressing epithelial cells in the chicken thymus. Arch. Histol. Cytol. 59, 197-203.
- Bellinger D.L., Lorton D., Horn L., Felten S.Y. and Felten D.L. (1997). Vasoactive intestinal polypeptide (VIP) innervation of rat spleen, thymus, and lymph nodes. Peptides 18, 1139-1149.
- Belokrylov G.A., Molchanova I.V. and Sorochinskaya E.I. (1990). Immunomodulatory properties of certain amino acids influence the immunostimulating properties of specific peptides. Int. J. Immunopharmacol. 12, 841-845.
- Boudard F. and Bastide M. (1991). Inhibition of mouse T-cell proliferation by CGRP and VIP: effects of these neuropeptides on IL-2 production and cAMP synthesis. J. Neurosci. Res. 29, 29-41.
- Brelinska R. and Warchol J.B. (1997). Thymic nurse cells: their functional ultrastructure. Microsc. Res. Tech. 38, 250-266.
- Crawley J.N. and Corwin R.L. (1994). Biological actions of cholecystokinin. Peptides 15, 731-755.
- Dardenne M. and Savino W. (1994). Control of thymus physiology by peptidic hormones and neuropeptides. Immunol. Today 15, 518-523.
- De la Fuente M., Delgado M. and Gomariz R.P. (1996). VIP modulation of immune cell functions. Adv. Neuroimmunol. 6, 75-91.
- De la Fuente M., Carrasco M., Del Rio M. and Hernanz A. (1998). Modulation of lymphocyte functions by sulfated cholecystokinin octapeptide. Neuropeptides 32, 225-233.
- Delgado M., Martinez C., Johnson M.C., Gomariz R.P. and Ganea D. (1996). Differential expression of vasoactive intestinal peptide receptors 1 and 2 (VIP-R1 and VIP-R2) mRNA in murine lymphocytes. J. Neuroimmunol. 68, 27-38.
- Dickinson T., Fleetwood-Walker S.M., Mitchell R. and Lutz E.M. (1997). Evidence for roles of vasoactive intestinal polypeptide (VIP) and pituitary adenylate cyclase-activating polypeptide (PACAP) receptors in modulating the responses of rat dorsal horn neurons to sensory inputs. Neuropeptides 68, 175-185.
- Felten D.L., Felten S.Y., Carlson S.L., Olschowka J.A. and Livnat S. (1985). Noradrenergic and peptidergic innervation of lymphoid tissue. J. Immunol. 135, 755S-765S.

Neuropeptides and the control of thymocyte proliferation

- Ganea D. (1996). Regulatory effects of vasoactive intestinal peptide on cytokine production in central and peripheral lymphoid organs. Adv. Neuroimmunol. 6, 61-74.
- Gomariz R.P., Lorenzo M.J., Cacicedo L., Vicente A. and Zapata A. (1990). Demonstration of immunoreactive vasoactive intestinal peptide (IR-VIP) and somatostatin (IR-SOM) in rat thymus. Brain Behav. Immunol. 4, 151-161.
- Gomariz R.P., Delgado M., Naranjo J.R., Mellstrom B., Tormo A., Mata F. and Leceta J. (1993). VIP gene expression in rat thymus and spleen. Brain Behav. Immun. 7, 271-278.
- Gourlet P., De Neef P., Cnudde J., Waelbroeck M. and Robberecht P. (1997). *In vitro* properties of a high affinity selective antagonist of the VIP₁ receptor. Peptides 18, 1555-1560.
- Harmar A.J., Arimura A., Gozes I., Journot L., Laburthe M., Pisegna J.R., Rawlings S.R., Robberecht P., Said S.I., Sreedharan S.P., Wank S.A. and Waschek J.A. (1998). International union of pharmacology. XVIII. Nomenclature of receptors for vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide. Pharmacol. Rev. 50, 265-270.
- Head G.M., Mentlein R., von Patay B., Downing J.E. and Kendall M.D. (1998). Neuropeptides exert direct effects on rat thymic epithelial cells in culture. Dev. Immunol. 6, 95-104.
- Higginbottom M., Horwell D.C. and Roberts E. (1993). Selective ligands for cholecystokinin receptor subtypes CCK-A and CCK-B within a single structural class. Bioorg. Med. Chem. Lett. 3, 881-884.
- Hughes J., Boden P., Costall B., Domeney A., Kelly E., Horwell D.C., Hunter J.C., Pinnock R.D. and Woodruff G.N. (1990). Development of a class of selective cholecystokinin type B receptor antagonists having potent anxiolytic activity. Proc. Natl. Acad. Sci. USA 87, 6728-6732.
- Kendall M.D. and Al-Shawaf A.A. (1991). Innervation of the rat thymus gland. Brain Behav. Immun. 5, 9-28.
- Malendowicz L.K., Trejter M., de Caro R., Jedrzejczak N., Brelinska R., Markowska A., Nussdorfer G.G. and Nowak M. (1999). Cholecystokinin, acting through the A receptor subtype, stimulates the proliferative activity of adrenocortical cells and thymocytes in the rat. Histol. Histopathol. 14, 439-443.
- Martinez C., Delgado M., Abad C., Gomariz R.P., Ganea D. and Leceta J. (1999). Regulation of VIP production and secretion by murine lymphocytes. J. Neuroimmunol. 93, 126-138.
- Molchanova I.V., Belokrylov G.A., Popova O. and Kalikhevich V.N. (1992). The immunostimulating properties of cholecystokinin octapeptide and its fragments. Bull. Exp. Biol. Med. 114, 631-633.
- Moody T.W., Zia F., Draoui M., Brenneman D.E., Fridkin M., Davidson A. and Gozes I. (1993). A vasoactive intestinal peptide antagonist inhibits non-small cell lung cancer growth. Proc. Natl. Acad. Sci.

USA 90, 4345-4349.

- Muller S. and Weihe E. (1991). Interrelation of peptidergic innervation with mast cells and ED1-positive cells in rat thymus. Brain Behav. Immun. 5, 55-72.
- Nussdorfer G.G. and Malendowicz L.K. (1998). Role of VIP, PACAP, and related peptides in the regulation of the hypothalamo-pituitaryadrenal axis. Peptides 19, 1443-1467.
- Pankhaniya R., Jabrane-Ferrat N., Gaufo G.O., Sreedharan S.P., Dazin P., Kaye J. and Goetzl E.J. (1998). Vasoactive intestinal peptide enhancement of antigen-induced differentiation of a cultured line of mouse thymocytes. FASEB J. 12, 119-127.
- Pisegna J.R., de Weerth A., Huppi K. and Wank S.A. (1993). Molecular cloning of the human brain and gastric cholecystokinin receptor: structure, functional expression and chromosomal localization. Biochem. Biophys. Res. Commun. 189, 296-303.
- Quirion R., Rioux F., Regoli D. and St-Pierre S. (1980). Selective blockade of neurotensin-induced coronary vessels constriction in perfused rat hearts by a neurotensin analogue. Eur. J. Pharmacol. 61, 309-312.
- Savino W., Ban E., Villa Verde D.M. and Dardenne M. (1990). Modulation of thymic endocrine function, cytokeratin expression and cell proliferation, by hormones and neuropeptides. Int. J. Neurosci. 51, 201-204.
- Soder O. and Hellstrom P.M. (1987). Neuropeptide regulation of human thymocyte, guinea pig T lymphocyte and rat B lymphocyte mitogenesis. Int. Arch. Allergy Appl. Immunol. 84, 205-211.
- Vanneste Y., Thome A.N., Vandersmissen E., Charlet C., Franchimont D., Martens H., Lhiaubet A.M., Schimpff R.M., Rostene W. and Geenen V. (1997). Identification of neurotensin-related peptides in human thymic epithelial cell membranes and relationship with major histocompatibility complex class I molecules. J. Neuroimmunol. 76, 161-166.
- Vincent J.P., Mazella J. and Kitabgi P. (1999). Neurotensin and neurotensin receptors. Trends Pharmacol. Sci. 20, 302-309.
- Wank S.A., Pisegna J.R. and de Weerth A. (1992). Brain and gastrointestinal cholecystokinin receptor family: structure and functional expression. Proc. Natl. Acad. Sci. USA 89, 8691-8695.
- Weinberg D.S., Ruggeri B., Barber M.T., Biswas S., Miknyocki S. and Waldman S.A. (1997). Cholecystokinin A and B receptors are differentially expressed in normal pancreas and pancreatic adenocarcinoma. J. Clin. Invest. 100, 597-603.
- Yiangou Y., Serrano R., Bloom S.R., Pena J. and Festenstein H. (1990). Effects of prepro-vasoactive intestinal peptide-derived peptides on the murine immune response. J. Neuroimmunol. 29, 65-72.

Accepted October 4, 2000

158