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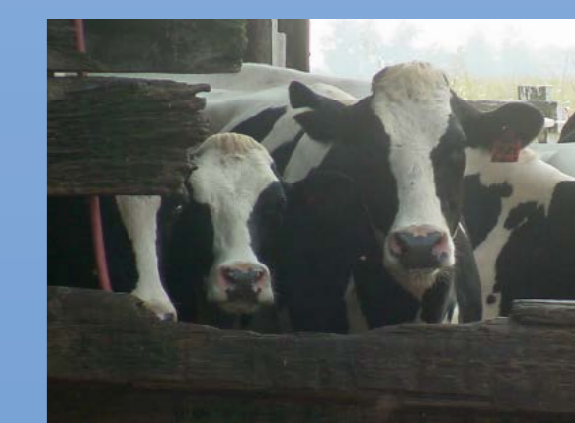
Use Of Py-GC/MS Analysis Techniques In Animal Waste Management: A Preliminary Survey Of Dairy Manures

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Introduction

The increasing practice of industrial-scale agriculture tends to concentrate large masses of animal waste in relatively compact areas, potentially leading to excessive release of polluting nutrients into waterways during major storms. Anaerobic treatment conditions are generally favored to conserve nitrate N as an agricultural commodity. However, overall N contents in waste are often in excess of crop fertilization needs: storing excess N in soluble nitrate form increases pollution potential. Thus the perceived needs of agriculture and society-at-large become at odds. Organic nitrogen forms (e.g., proteins) are more environmentally stable and are less subject to unintentional release. Although U.S. farmers tend to view it with disfavor, non-aqueous (aerobic) treatment such as composting holds potential for storing nitrogen in a more stable, environmentally-benign form.



Photos: Farm Sanctuary (www.factoryfarming.com)

Methods

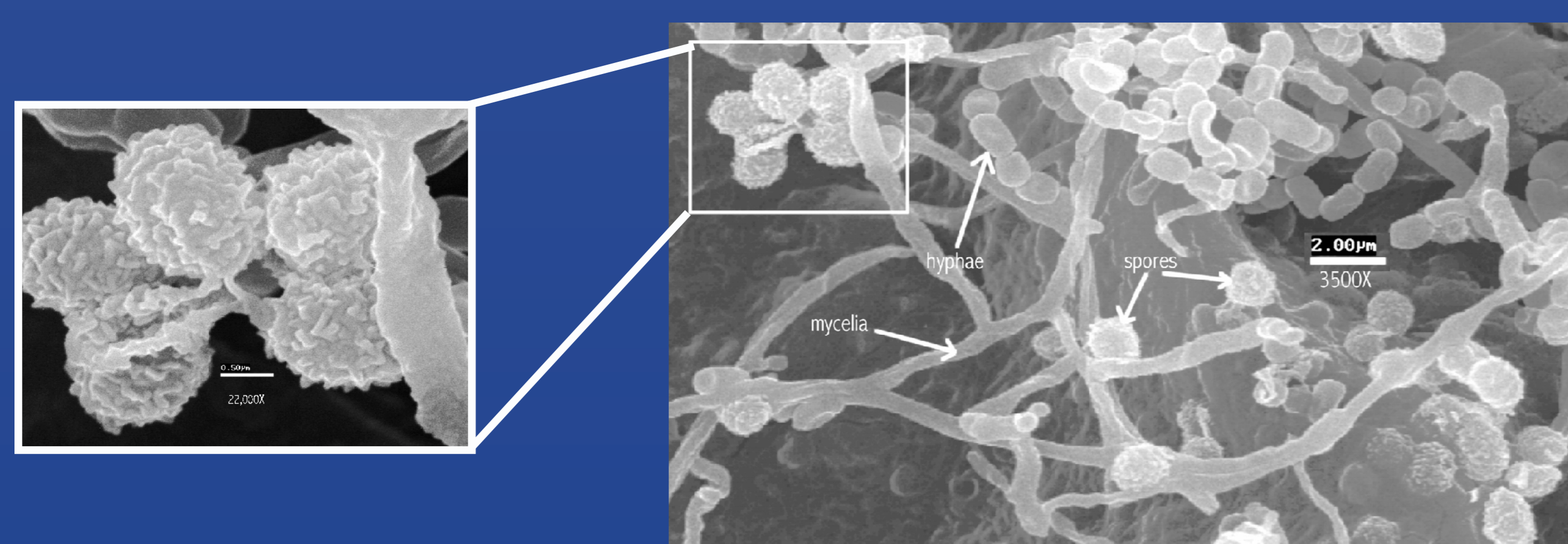
Six dairy farm waste samples were collected. The three samples discussed herein (fresh manure, a waste lagoon deposit, and a dry compost) were chosen as representative of a waste source and two biochemically different waste treatment scenarios: anaerobic versus aerobic. The fresh sample was a grab sample and represents manure in the unaltered state. The anaerobically-degraded lagoon sample, taken from a six-week old pit sludge at a depth of 80cm, was olive green and exhibited a strong odor. The aerobically-degraded compost was taken from older, mixed-age, lot-edge spillover. The location was open to the air and well-drained, but partly sheltered. The compost was browner in color than the lagoon sample and had an earthy odor. Sub-samples were dried, crushed, and cryo-fractured as preparation for pyrolysis.



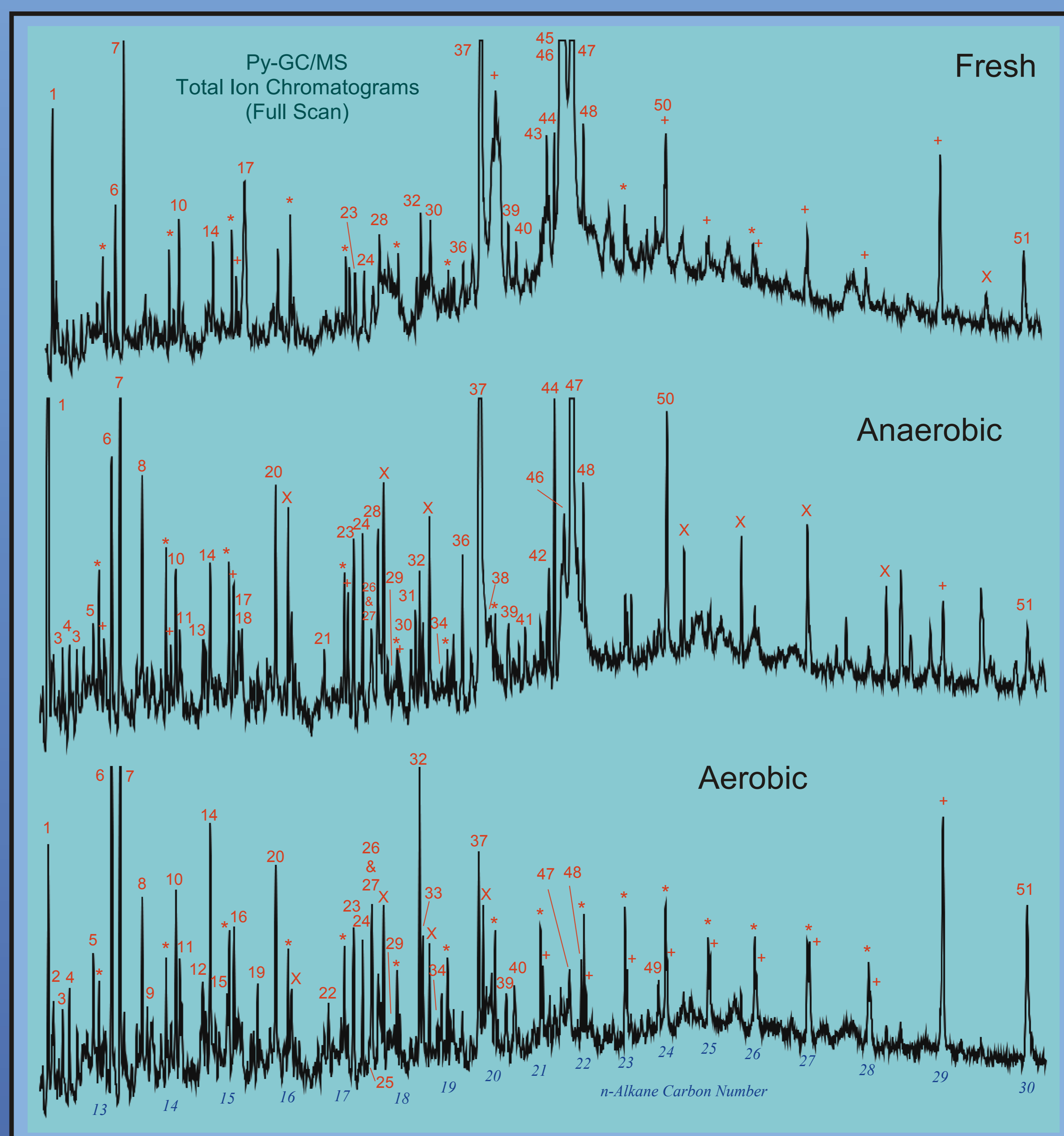
Fresh manure, bedding and spilled feed initially accumulate on the stable floor.



Stable floor litter is transferred to a lagoon, where it undergoes anaerobic degradation.



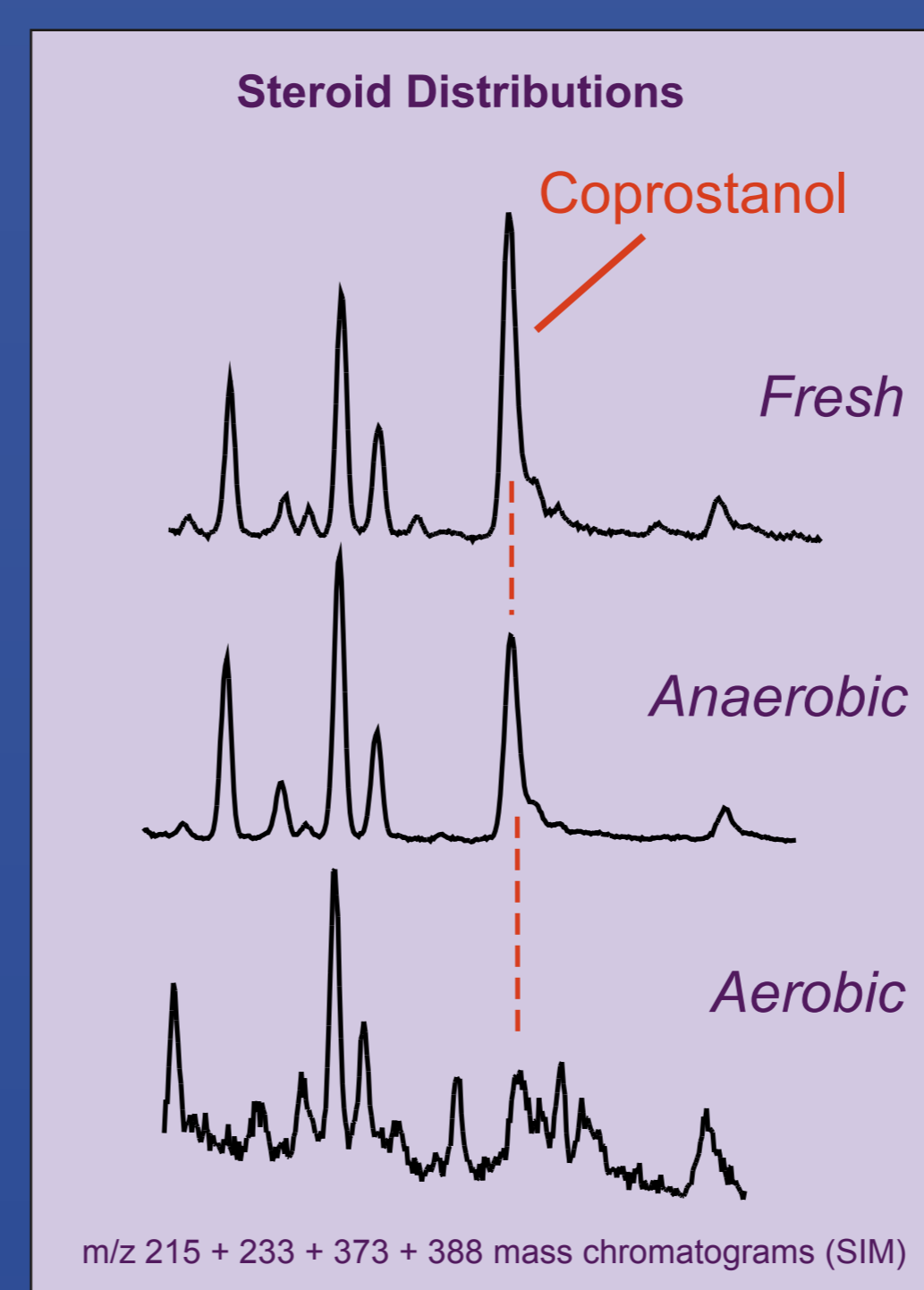
Scanning electron micrographs of composted (aerobically-degraded) manure showing abundant actinomycete hyphae, mycelia and spores.



CDS Pyroprobe, 600oC, 20 sec., HP 5980 GC; 50m/0.2mm/0.3um DB5-ms col.; 40°C (5 min.), 40-300°/min., 300° (20 min.); HP 5780 MSD, 50-440 Da, 0.9 scans/sec.

Observations

Significant differences between the pyrolyzates were noted: The lagoon sample is strongly enriched in fatty acids relative to post. As anaerobic systems tend to be lower in pH, this is expected. A more detailed examination of the pyrolyzates might be that, while both have many compounds in common, the lagoon sample reveals methoxyphenols derived from the plant matter in the feed, relative proportions may vary considerably. The compost pyrolyzate appears to be relatively enriched in indoles and dipeptides, as well as organonitrogen compounds (indoles and dipeptides), as well as organonitrogen compounds carbons. This conservation of organic long-chain aliphatic hydrocarbons that aerobic manure management, N lends support to the idea that a more desirable agricultural practice, such as by composting, could be a more desirable agricultural practice.



The fecal steroid coprostanol is relatively much less abundant in the aerobically-degraded compost.

Py-GC/MS peak identification. *: n-alkenes, +: n-alkanes, x: silane & phthalate contaminants. Numbered peaks identified below.

- | | |
|--|---|
| 1) dihydrobenzofuran | 25) dipeptide (Pro-Ala) |
| 2) 2-methoxy-4-methylphenol | 26) bisphenol |
| 3) methoxyphenol | 27) acetosyringol |
| 4) benzenepropanenitrile | 28) tetradecanoic acid |
| 5) 2-methoxy-4-ethylphenol | 29) Pro-Gly, Pro-Lys |
| 6) indole | 30) pentadecanoic acid |
| 7) 2-methoxy-4-vinylphenol | 31) unknown acid |
| 8) 2,6-dimethoxyphenol | 32) phtyadiene |
| 9) 2-methoxy-4-(prop-1-enyl)phenol | 33) phytene |
| 10) methylindole | 34) Pro-Val, Pro Arg |
| 11) 2-methoxy-4-formylphenol | 35) tetradecanitrile + unspec. acid |
| 12) 1,2-hydroxyphenyl-ethanone | 36) hexadecanoic acid methyl ester |
| 13) C ₈ H ₈ O ₄ (?) | 37) hexadecanoic acid |
| 14) isoeugenol | 38) hexadecanitrile |
| 15) indoleidone | 39) methylpyridindole |
| 16) 2-methoxy-4-(propane-2-one)-phenol | 40) pyridindole |
| 17) levoglucosan | 41) heptadecanoic acid |
| 18) tetrahydronaphthalenamine | 42) octadecanitrile |
| 19) 1-(4-hydroxy-methoxyphenyl)-ethanone | 43) nonadecan-2-one |
| 20) 2,6-dimethoxy-4-vinyl-phenol | 44) octadecanoic acid, methyl ester |
| 21) unknown | 45) octadecadienoic acid |
| 22) 1-phenyl-1H-pyrrole-2,5-dione | 46) octadecenoic acid |
| 23) 2,6-dimethoxy-4-(pro-2-enyl)-phenol (trans) | 47) octadecanoic acid |
| 24) prist-1-ene | 48) hexadecanamide |
| | 49) biphenoldiol |
| | 50) octadecanamide |
| | 51) C ₃₀ alkene or alcohol ? |

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