

## Original Article

# Reproductive biology of *Gagora catfish (Arius gagora)* at Meghna river system, Kushiara River, Bangladesh

Md. Arifur Rahman<sup>1</sup>, Mohammad Amzad Hossain<sup>\*2,3</sup>, Md. Rahamat Ullah<sup>1</sup>, Mohammed Mahbub Iqbal<sup>2</sup>

<sup>1</sup>Department of Fisheries Biology and Genetics, Patuakhali Science and Technology University, Patuakhali-8602, Bangladesh.

<sup>2</sup>Department of Fish Biology and Genetics, Sylhet Agricultural University, Sylhet-3100, Bangladesh.

<sup>3</sup>Coastal Marine Ecosystems Research Centre, CQ University, Gladstone Marina, QLD 4680, Australia.

**Abstract:** The gagora catfish, *Arius gagora* is a species of Ariidae family normally exist in estuaries and tidal rivers of Bangladesh and India. In the present study, the reproductive biology of *A. gagora* was assessed in Meghna River system at Kushiara River. Length-weight relationship was found strongly correlated for male ( $r^2 = 0.98$ ), female ( $r^2 = 0.97$ ) and combined sex ( $r^2 = 0.81$ ), respectively. Chi-square tests revealed no significant difference between sex ratios round the year and not deviated from the expected value of 1:1 ( $\chi^2 = 5.57$ ) and the critical chi-square value calculated was 143.25. The fecundity found to be ranged from  $43333.12 \pm 829.83$  to  $53948.81 \pm 1743.28$  in April to August. The relationship of fecundity with ovary weight ( $r^2 = 0.79$ ), ovary length ( $r^2 = 0.88$ ), total length ( $r^2 = 0.85$ ) and total weight ( $r^2 = 0.68$ ) were found positively correlated. Monthly study of gonadosomatic index (GSI) revealed that the breeding season for this fish species may be between July to August with a single peak spawning month in August. Size at sexual maturity was estimated as 30.01 cm total length. Fulton's condition factor (K) value varied in both males and females and indicates that the overall health condition of both the male and female were excellent. Breeding season was found significantly correlated with rainfall, but not with other biological, meteorological, and hydrological parameters. Ovarian development of *A. gagora* was assessed macroscopically and three broad gonad developmental stages were identified. Maturity was specified based on the presence of green egg in the peritoneal cavity. These findings should benefit and improve the sustainable management and conservation of this species both in Bangladesh and its bordering countries.

### Article history:

Received 15 March 2020

Accepted 24 September 2020

Available online 25 December 2020

### Keywords:

Gonadosomatic index

Sexual dimorphism

Dobriyal index

Macroscopic observation

## Introduction

The Ariidae family comprised of 14 genera and 120 species of tropical and sub-tropical marine catfishes (Kailola, 1999). The gagora catfish, *Arius gagora* (Hamilton, 1822) is found in estuaries and tidal rivers of Bangladesh and India (West Bengal and Orissa) (Talwar and Jhingran, 1991) and reported from the Hooghly Estuary (Talwar and Jhingran, 1991) and Meghna River (Rahman, 1989; Rahman, 2005). According to Rahman (2005), gagora catfish is found in estuaries, tidal rivers, and the Bay of Bengal.

In respect to their gonadal conditions, generative potential of a species is one of the basic demands to designate the individuals of that species (Jhingran and Verma, 1972). The reproductive biology of a fish could be defined by its reproductive characters and it

also expresses the combination of the species-specific reproductive mode (Winemiller and Rose, 1992; Murua and Rey, 2003; Morgan, 2008). In addition, for success in fish culture, assessment of the yearly breeding sequence of culturable fishes are very important (Stoumoundi et al., 1993). According to Murua and Rey (2003), information about the reproductive biology and the estimation of fecundity and egg production are fundamental topics in the study of the biology and population dynamics of fish species. The fecundity and egg production of a fish may vary annually and spatially both within and among stocks (Hunter and Leong, 1981) in relation to their condition and environmental factors (Somarakis et al., 2000; Takasuka et al., 2005). Reproducing of a fish may occurs during a certain phase of reproductive

\*Correspondence: Mohammad Amzad Hossain  
E-mail: m.a.hossain@cqu.edu.au

cycle; some breeds once annually while others at regular intermissions throughout the year. Knowledge of gonadal maturation and spawning season of a species permit subsequent studies on spawning frequency, which is important for its management (Chakrabarti and Barun, 2017).

Gonado somatic index (GSI) is used to determine the spawning season, (Verghese, 1975; Jacob and Nair, 1983; Ha and Kinzie, 1996; Shankar and Kulkarni, 2005) and considered as a sensitive and proved noteworthy parameter to monitor gonadal maturation (Belsare, 1962; Lehri 1968; Hong-Yang and Yon, 1992; Shashi and Akela, 1996). The cyclical factors might greatly impact the maturation of ovary results the consecutive changes in the gonads and body weights (Lincoln et al., 1980). Condition factor states the well-being of a certain species (Radkhah and Eagderi, 2015; Zamani-Faradonbeh et al., 2015) and its degree of fatness, the state of sexual maturity, the degree of food sources availability, age and sex of some species and the system of environment which depends on weight of the fish sampled (Pauly, 1983; Fafioye and Oluajo, 2005; Isa et al., 2010; Alam et al., 2014; Keivany et al., 2015; Abbasi et al., 2019).

Reproductive biology parameters such as fecundity, spawning, sex ratio etc. are among the important aspects of the biology of fishes which must be understood to explain the dissimilarities in the level of populations to make efforts to increase the amount of harvest (Azadi and Siddique, 1996). Fish reproduction may be influenced by various factors, including seasonal changes in photoperiod, rainfall and water temperature, along with physio-chemical features of the water body (Bromage et al., 2001; Glasser et al., 2004; Dorostghoal et al., 2009). Therefore, length-weight relationship (LWR), condition factor (K), gonadosomatic index (GSI), hepatosomatic index (HSI), fecundity and macroscopic gonadal maturity were considered as rudimentary parameters for assessing the reproductive condition of *A. gagora*. Despite ecological significance of freshwater fishes, length-weight relationship and condition factor data are often lessened (Mouludi-Saleh and Eagderi, 2019). The

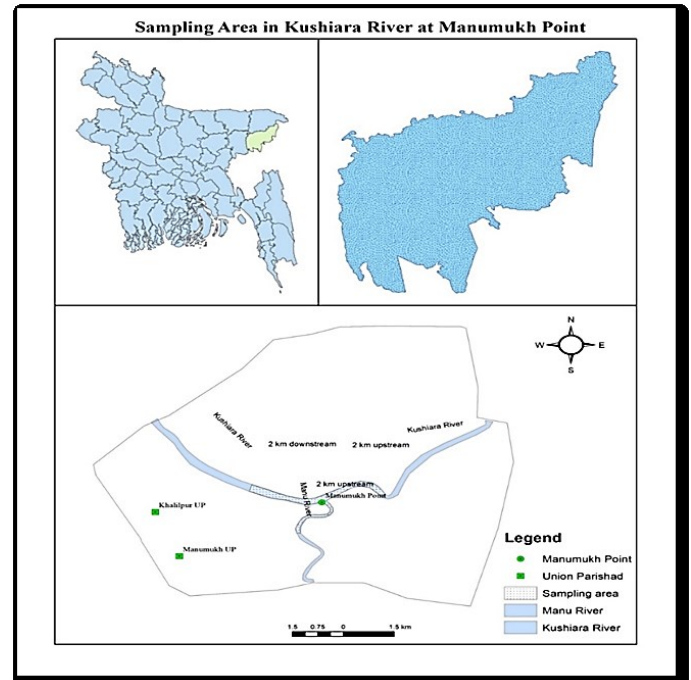


Figure 1. Map showing study area of Meghna river system at Kushiara River, Southeast Bangladesh.

evidence on the general biology of *A. gagora* is also scant in literature from Bangladesh and elsewhere. This is one of the utmost important target species for small-scale fisheries.

### Materials and Methods

The Kushiara (Kushiyara) River is originated from India. The original Barak River separated into two rivers named Surma and Kushiara after entering Bangladesh and thereafter both the rivers combined near the Bhairab Bazar, and later joins to massive river Meghna. A yearlong research was conducted in Maghna river system at Kushiara River (Fig. 1) in five successive sampling points and monthly samples were collected.

A total of 117 individual of *A. gagora* procured from Kushiara River through bush trap. The following extents were made for assessing the general biology of *A. gagora*: total length (TL) with an accuracy of 0.1 mm from the tip of mouth to the distal end with digital slide caliper (Mitutoyo, CD-6"CSX), total weight (TW) with accuracy of 0.01 gm (AND EK-6100i); and gonad, liver and intra-peritoneal fat weight (GW) with accuracy of 0.01 gm (AND EK-6100i). Sex ratio (female/male) was calculated monthly. Length-weight

Table 1. Length (cm) and weight (gm) measurement for male, female, and combined sexes of *Arius gagora*.

Sex	Parameter	Min.	Max.	Yearly Mean	Equation	r <sup>2</sup>	P values
Male	TL (cm)	19.2	39.65	29.98±5.45	BW=a×TL <sup>b</sup>	0.98	0.000***
	BW (gm)	235.09	492.69	362.42±62.36			
Female	TL (cm)	21.20	40.40	31.87±5.23	BW=a×TL <sup>b</sup>	0.97	0.000***
	BW (gm)	272.37	688.65	479.3±109.64			
Combined	TL (cm)	19.2	40.4	30.98±5.39	BW=a×TL <sup>b</sup>	0.81	0.000***
	BW (gm)	235.09	688.65	424.36±107.53			

P-value shows level of significance; \*\*\* highly significant.

relationship was estimated through the equation:  $W=aL^b$ , where W is the total body weight (g), and L is the total length (cm). Fulton's condition factor (1904) was estimated by the equation:  $K=100 \times (W/L^3)$ , where W is the total body weight (BW, g), L is the total length (TL, cm) and scaling factor of 100 was used to fetch the K close to the unit factor (Hossain et al., 2012).

To identify gonad maturation stages two indices were considered: (a) gonadosomatic index (GSI) =  $(GW/BW) \times 100$  (Nikolsky, 1963) and (b) Dobriyal index (DI) =  $\sqrt[3]{GW}$  (Dobriyal et al., 1999). The total length size at sexual maturity was assessed by the relationships of total length against GSI and total length against DI. Breeding season of *A. gagora* was estimated based on the monthly differences of GSI value. Hepatosomatic index (HSI) and Intra-peritoneal fat index (IPFI) were calculated to find out energy investment in the breeding event by the equation: Liver weight (gm)/Body weight (gm) x 100 (Cek and Yilmaz, 2009) and Intra-peritoneal fat weight (gm)/Body weight (gm) x 100, respectively.

The ventral incision was made longitudinally from the anus towards the lower jaw by using scissors and eviscerated. The gonads were removed; sex was determined and weighed to the nearest 0.01 gm and preserved in Bouin's fixatives. Mature fishes were subjected to calculate absolute fecundity as stated by Ahyaudin (1999). Absolute fecundity was calculated by gravimetric method described by Babiker and Ibrahim (1979) and Narejo et al. (2002). The numbers of eggs were counted under a binocular microscope (JP Selecta, 3,000-B LED) for each sample. Total numbers of oocytes (N) in both ovaries were calculated using the following formula: Weight of two

ovaries (gm)/Weight of sub-sample × No. of counted oocytes in sub-sample × 100. This research has focused more on ovaries because ovarian development typically defines the breeding season (De Martini and Fountain, 1981). Monthly rainfall, air temperature and photoperiod/total sunshine data were obtained from the MET report logbook of Meteorological Department of Bangladesh (Khatun et al, 2017) and water temperature was determined *in situ*. Then monthly mean meteorological data were correlated with monthly mean GSI of *A. gagora* using Spearman rank-correlation test.

**Statistical Analysis:** SPSS software (version 23.0) was for used data processing and analysis. The Spearman rank-correlation test was applied to analyze the relationship of GSI with HSI, IPFI, meteorological and hydrological data. Monthly mean HSI and environmental parameters, including rainfall, air temperature, water temperature and photoperiod were correlated with the breeding season (mean GSI) of *A. gagora* using Spearman rank-correlation test Chi-square tests ( $\chi^2$ ) were conducted to observe the deviation of sex ratio from the expected value of 1:1. All statistical analyses were considered significant at 5 % level of significance ( $P<0.05$ ).

## Results

A total of 117 *A. gagora* were collected from Meghna river system at Kushiara River for 12 months. This species exhibited overall sex ratio of 1:1.12 (male: female) and the percentage occurrence was 52.99% for females and 47.01 % for male. Chi-square tests revealed no significant different between sex ratios round the year and not deviated from the expected value of 1:1 ( $\chi^2 = 5.57$ ,  $P = 0.65$ ) and the critical chi-

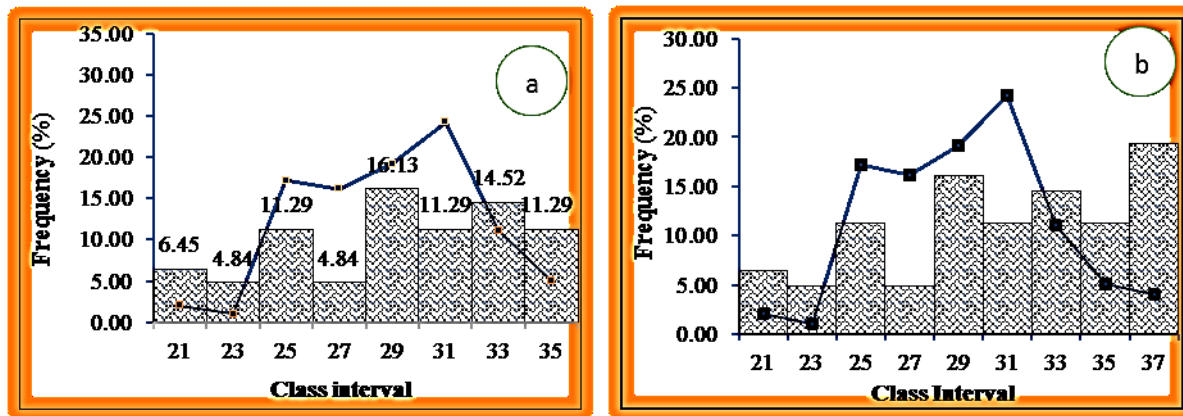
Table 2. K value standards adopted by Barnham et al. (1998).

K value	Comments
1.6	Excellent condition, trophy class fish
1.4	A good, well-proportioned fish
1.2	A fair fish, acceptable to many anglers
1	A poor fish, long and thin
0.8	Extremely poor fish, big head, and narrow, thin body

Table 3. Descriptive statistics showing length (cm) and weight (gm) measurement for male, female, ovipositor diameter (vertical and horizontal) and ovipositor color of female *Arius gagora*.

Month	BL (cm)	BW (gm)	OPD (V)	OPD (H)	OPC
July	37.74±1.14	623.28±38.28	0.56±0.09	0.35±0.1	P-LP
August	38.4±1.35	648.87±23.42	0.58±0.07	0.36±0.04	R
September	22.34±0.76	304.65±15.55	0.29±0.03	0.24±0.02	LP
October	23.97±2.38	325.23±44.64	0.29±0.02	0.24±0.01	LP
November	25.96±0.38	362.58±8.43	0.4±0.1	0.23±0.03	LP
December	27.7±1.14	398.53±20.65	0.35±0.04	0.25±0.03	LP-P
January	29.74±0.6	429.97±14.26	0.41±0.05	0.26±0.04	LP-P-R
February	31.1±0.9	449.49±14.34	0.41±19.57	0.26±0.04	LP-RP
March	32.95±0.92	486.42±14.76	0.39±0.01	0.26±0.05	LP-P-DP
April	34.37±0.94	508.04±15.31	0.47±0.05	0.28±0.025	P-DP-R
May	34.35±1.09	510.2±16.04	0.48±0.025	0.3±0.0	LP-LR-R
June	36.78±2.14	552.42±42.27	0.56±0.08	0.35±0.1	LP-LR-R

OPD (V) = Ovipositor diameter (vertical), OPD (H) = Ovipositor diameter (horizontal), OPC = Ovipositor color, LP = Light Pink, P = Pink, DP = Dark Pink, LR=Light Red, R=Red

Figure 2. Total length frequency distributions of male (a) and female (b) *Arius gagora* in Meghna river system at Kushiara River round the year.

square value calculated was 143.25.

The length and weight measurement of *A. gagora* reveal that the females were significantly larger than males both in terms of length and weight (Table 1) and showed predominance over male. In addition, LFDs showed that, here 37.00-40.99 cm total length size groups were numerically dominant for female constituted together 57.70% of the total population and 29.0-30.99 size group for male (Fig. 2). The relationship between body weight and total length

provided a positive correlation (Table 1) for male ( $r^2 = 0.98$ ), female ( $r^2 = 0.97$ ) and combined sex ( $r^2 = 0.81$ ).

The monthly K-values varied for both sexes calculated as ranging from lowest average value  $1.12 \pm 0.49$  (June) and maximum  $2.74 \pm 0.16$  (September) for female and lowest  $0.81 \pm 0.02$  (August) and uppermost  $2.94 \pm 0.36$  (September) for male, respectively. The yearly average condition factor for male *A. gagora* was found  $1.50 \pm 0.61$  and

Table 4. Relationship between dependent variables (genital papilla and ovipositor diameter) and independent variables of both sexes of *Arius gagora*.

Sex	Dependant variable	Independent variable	Pearson squares (r <sup>2</sup> Values)
Male	Genital papilla length vertical	Total length	r <sup>2</sup> = 0.66
	Genital papilla length horizontal	Total length	r <sup>2</sup> = 0.26
	Genital papilla length vertical	Body weight	r <sup>2</sup> = 0.67
	Genital papilla length horizontal	Body weight	r <sup>2</sup> = 0.26
Female	Ovipositor diameter length vertical	Total length	r <sup>2</sup> = 0.73
	Ovipositor diameter length horizontal	Total length	r <sup>2</sup> = 0.46
	Ovipositor diameter length vertical	Body weight	r <sup>2</sup> = 0.71
	Ovipositor diameter length horizontal	Body weight	r <sup>2</sup> = 0.46

Table 5. Monthly mean ± SD fecundity of female *Arius gagora* collected from the Meghna river system at Kushiara River, southeast Bangladesh.

Month	Body Length (cm)	Body Weight (gm)	Absolute Fecundity
Jul'13	37.95±0.98	625.25±39.19	50761.44±1586.21
Aug'13	38.33±9.99	648.87±163.07	53948.81±1743.28
Sep'13	21.34±11.36	352.39±189.48	Uncountable
Jan'14	29.74±9.09	489.97±152.68	Uncountable
Apr'14	34.37±9.05	568.04±150.97	43333.12±829.83
May'14	34.35±9.14	570.2±152.47	45195.56±1754.85
June'14	36.78±9.32	612.42±155.67	49860.67±2111.97

The mean values were calculated as mean ±standard deviation (SD).

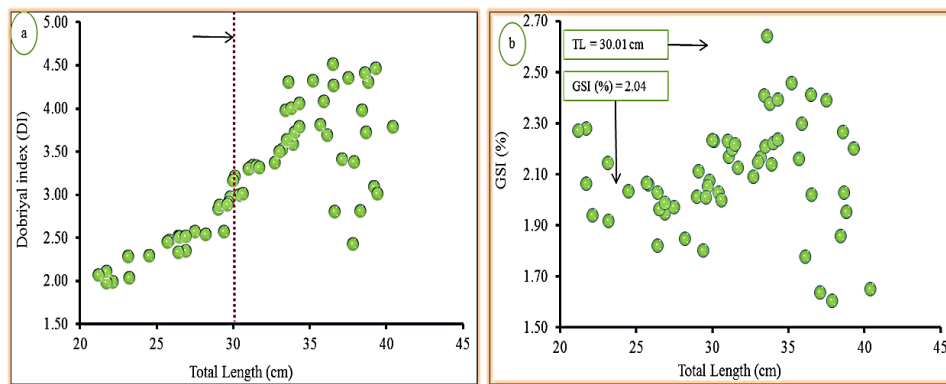


Figure 3. Relationships between (a) Dobriyal index (DI) and total length, and (b) gonadosomatic index (GSI) with total length of female *Arius gagora* in the Meghna river system at Kushiara river, southeast Bangladesh.

1.57±0.50 for female which indicates that the overall health condition of both the male and female were excellent (Table 2).

The interactions between total length against gonadosomatic index and total length against Dobriyal index (DI) of female *A. gagora* are shown in Figure 3. During this study period, GSI ranged from 1.13 to 3.95 with a yearly average 1.97±0.45 and DI ranged from 1.98 to 4.51 with yearly average value of 3.21±0.72. The GSI (>2.04%) and DI (>3.21) rose sharply at ~30.01 cm SL for most of the female population.

The results of Spearman rank-correlation test indicated significant results between rainfall and breeding season ( $r_s = 0.606, P < 0.05$ ), but no relation was found between HSI and GSI ( $r_s = -0.231, P = 0.471$ ), air temperature and GSI ( $r_s = 0.517, P = 0.085$ ), water temperature and GSI ( $r_s = 0.460, P = 0.133$ ) and photoperiod and GSI ( $r_s = -0.336, P = 0.286$ ) (Fig. 4).

Male brood fishes were identified based on pointed muscular genital papilla and oozing of milt when a slight pressure is applied on the abdomen (Fig. 5). The

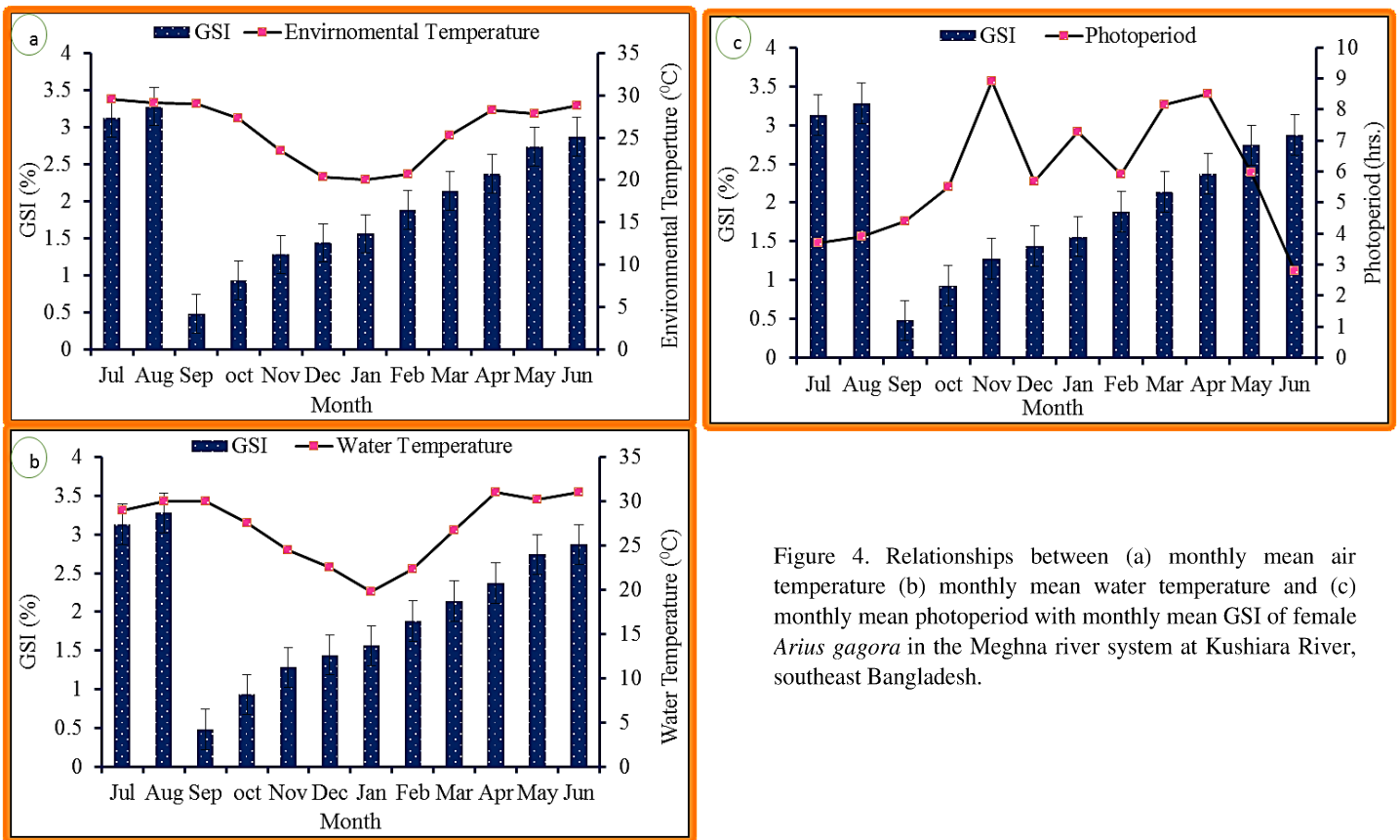


Figure 4. Relationships between (a) monthly mean air temperature (b) monthly mean water temperature and (c) monthly mean photoperiod with monthly mean GSI of female *Arius gagora* in the Meghna river system at Kushiara River, southeast Bangladesh.

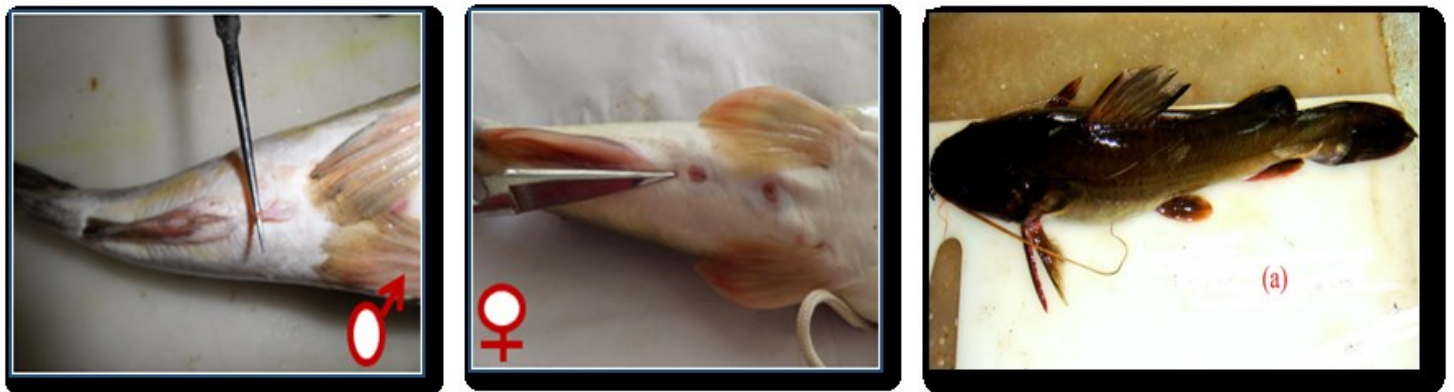


Figure 5. A general view of phenotypic sexual characteristics in male (left, ♂) and female (right, ♀) *Arius gagora* (a).

female broods of *A. gagora* was recognized through a bulging abdomen; soft inflated belly and red genital vent (Fig. 5). The genital opening varied from Red (matured) dark pink (maturing) to light pink (immature) in color for females (Table 3). Vertical ovipositor length of female and genital papillae of male were significant (Table 4) both in terms of total body weight and length (female, BW = 0.71, TL = 0.73; male BW = 0.67, TL = 0.66).

The testes of *A. gagora* were paired, elongated, leaf-like and exposed enormous number of dentations. The ovaries of *A. gagora* were paired tubular organs

and the color of ovaries was found to oscillate from brownish to greenish depending on maturation stages as showed in Figure 6. In mature ovary, noticeable blood vessels were encountered. Ovaries linked only at the posterior where they opened into the genital aperture. The maximum GSI value of female found to be maximum  $3.28 \pm 0.11$  in August and minimum  $0.48 \pm 0.003$  in September. On the other hand, the highest HSI value for female was observed  $0.67 \pm 0.006$  in April and lowest  $0.38 \pm 0.09$  in August (Fig. 7). The monthly mean GSI and HSI values of female *A. gagora* were plotted with rainfall (Fig. 7).

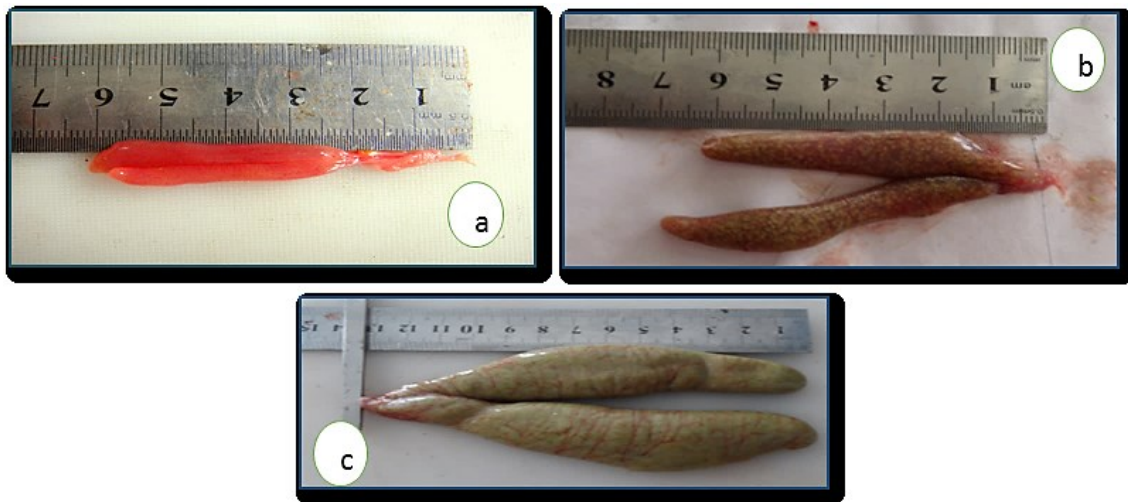


Figure 6. (a) Immature: Sex organs very small and semi-transparent, grayish red in color and possess half or slightly more than the half of abdominal length of enclosure (September-January), (b) Maturing: ovaries become grey brownish in color, partial red blood capillary and occupies more than half of the yard of abdomen, eggs are visible in naked eye (April -June), and (c) Matured: sex organs courtyard filled abdominal and eggs completely round. The color of the ovaries becomes dark green and the red blood capillary was conspicuous (July-August).

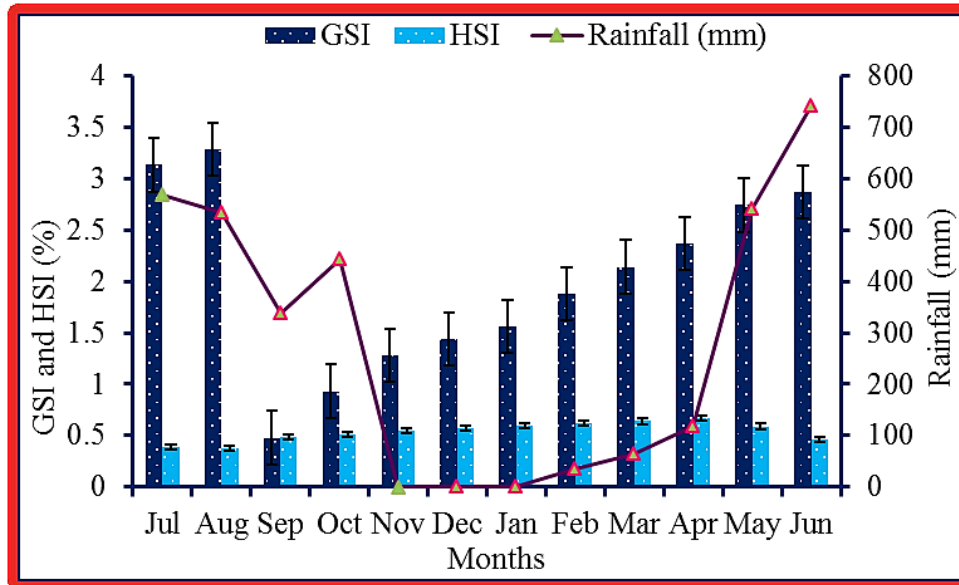


Figure 7. Monthly variation of GSI and HSI of female *Arius gagora* with mean monthly rainfall in the Meghna river system at Kushiara River, southeast Bangladesh.

The ovary of *A. gagora* is tabular shaped and comprising two lobes in which right sided one is found to greater in length than left one found in the peritoneal cavity (Fig. 6). According to gross morphology of the gonad, the complete reproductive cycle of female fish was categorized into three stages.

In the present study, for the assessment of absolute fecundity 28 randomly scooped up matured female fish were used (Table 5). The absolute fecundity was found to differ from  $43333.12 \pm 829.83$  (in April) to  $53948.81 \pm 1743.28$  (in August). The relationship

between fecundity and total length, fecundity, and body weight (Table 6) provided a positive correlation ( $r^2 = 0.85$  and  $r^2 = 0.68$ , respectively). Moreover, fecundity and ovary length, fecundity, and ovary weight (Table 5) revealed a positive correlation ( $r^2 = 0.88$  and  $r^2 = 0.79$  respectively).

The food components found in the guts were only small fishes (Fig. 8). The monthly mean GSI and IPFI values of female *A. gagora* were plotted in the Figure 9. The highest IPFI value was observed  $0.84 \pm 0.03$  in April and lowest  $0.08 \pm 0.03$  in August for female (Fig.

Table 6. Relationship between fecundity and independent variables of female *Arius gagora* collected from the Meghna river system at Kushiara River, southeast Bangladesh.

Dependant variable	Independent variable	Pearson squares ( $r^2$ Values)
Fecundity	Total length	$r^2 = 0.85$
Fecundity	Body weight	$r^2 = 0.68$
Fecundity	Ovary length	$r^2 = 0.88$
Fecundity	Ovary weight	$r^2 = 0.79$



Figure 8. Clear image showing food materials in gut (a) and intra-peritoneal fat (b) of female *Arius gagora* collected from the Meghna river system at Kushiara River, southeast Bangladesh

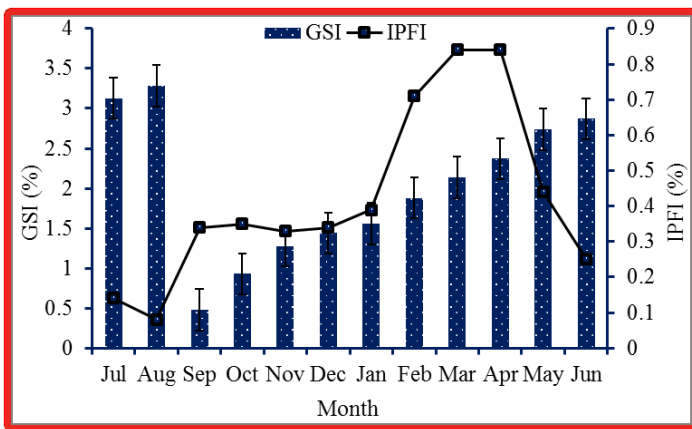


Figure 9. Monthly correlation between GSI and IPFI of female *Arius gagora* in the Meghna river system at Kushiara River, southeast Bangladesh.

reported by Shinkafi et al. (2002, 2011) in *S. eupterus* and *S. clarias* (Shinkafi et al., 2002) and *S. schall* and *S. nigrita* (Laleye et al., 2006).

The relationship between body weight and total length provided a positive correlation. Noor (2013) assessed TL-BW relationships in *R. rita* and found to be highly significant with correlation coefficient values of 0.97 and 0.95 for male and female, respectively which correlates the present findings. The investigators could not sample individuals of smaller size group because of biased selection of fishing technique (bush trap), or fishermen did not set trap where the smaller fish were (Hossain et al., 2012, 2016; Azad et al., 2018; Khatun et al., 2018), or perhaps the small fish’s absence on the sampling areas (Hossain, 2010a, b, 2012, 2013, 2017).

The monthly K-values was lowest in June and maximum in September for female and lowest in August and uppermost in September for male. The condition factor of a fish states physical condition, fatness, or well-being (Mir et al., 2012) of that fish, based on the hypothesis that weightier fish of a given length are in better condition. Konan et al. (2014) found lowest K values in female *C. buettikoferi* during

9).

**Discussions**

According to Nikolsky (1963), the sex ratio may vary from year to year in the same population, but in most fish species, it is close to one. However, the sex ratio of *A. gagora* showed a predominance of female, a similar trend also reported from *M. gulis* (Islam et al., 2008) and *O. pabo* (Gupta et al., 2014). In *A. gagora*, the females were larger than males, similar results also



April  $0.75 \pm 0.09$  which support the present findings. Brito and Bazzoli (2003) studied *P. coruscans* and found lowest K value  $1.15 \pm 0.19$  in spent and highest  $1.20 \pm 0.23$  in resting female which also corroborates the present findings.

The mean GSI varied from 0.48 (September) to 3.28 (August). Monthly study of gonadosomatic index (GSI) revealed that, the breeding season for this fish species may be between July to August with a single peak spawning month in August. The size at sexual maturity was considered to be at 30.01 cm of total length, and individuals with a GSI  $\geq 2.04\%$  and DI  $\geq 3.21$  could be roughly defined as mature *A. gogora* females in the Meghna river system at Kushiara River, southeast Bangladesh.

The sharp fall in IPFI was encountered onset of breeding. Spearman rank-correlation test shows no significant correlation between GSI and IPFI ( $r_s = -0.253$ ,  $P = 0.428$ ). The present finding is found to be congruent with Brito and Bazzoli (2003) in catfish *P. coruscans*.

Male brood fishes were identified based on pointed muscular genital papilla. Presence of genital papilla in males of catfish species has been perceived earlier before and reported from *Mystus gulio* (Mookherjee et al., 1941; David, 1963), *Gagata* spp. and *Batasio* spp. (Hora and Law, 1941) and *G. reticulam* (Mukherji, 1936). The testes of sexually mature catfish are paired elongated structures located dorsally in the body cavity and the left testis is usually longer than the right one (Vandyk and Pieterse, 2008) which supports the present findings. Spawning period of *A. gogora* was determined by the monthly mean Gonadosomatic Index (GSI) values. The studies on seasonal variation suggested that, the HSI values gradually increase before the onset of the breeding season and its values exhibited a sharp decline with the onset of the breeding activities.

Fish reproduction may be influenced by various factors, including seasonal changes in photoperiod, rainfall and water temperature, along with physico-chemical features of the water body and relating to holding conditions (Bromage et al., 2001; Glasser et al., 2004; Dorostghoal et al., 2009). Isa et al. (2012)

noted that male *A. argyropleuron* reached higher GSI value in November, whereas females reached maturity in April followed by June. It does not support the present findings. Kabir et al. (2012) noted that GSI of *P. hypophthalmus*, increased significantly with each morphological stage until the 7<sup>th</sup> month of culture period coinciding with the final stages of oocyte maturity, during of eight months study in captive condition. But the present findings agreed with the findings *M. tengara* (Banu et al., 1992). On the other hand, if HSI values did not decrease during ovary development, indicates that the stored energy in liver was not utilized during this period and this is likely due to adequate provision of food to the fish (Cek and Yilmaz, 2009; Reidel et al., 2010). But in the present study, HSI values decreased during ovary development, indicating that the stored energy in liver was utilized during this period and this is likely due to inadequate provision of food to the fish. Kabir et al. (2012) noted peak of GSI and HSI of *P. hypophthalmus* in November which is not congruent with the present findings.

Based on macroscopic evaluation, the gonadal development stages of *A. gogora* were divided into 3 broad stages. Five gonad developmental stages were identified including spent phase in *Rasbora tawarensis* (Muchlisin et al., 2010) and *Clarias gariepinus* (Tyor and Pahwa, 2017). Fish with gonad stage three (matured) were considered sexually mature in case of *A. gogora*.

Musa and Bhuiyan (2007) stated fecundity range of 4,652-57,932 in *M. bleekeri*; Bhatt et al. (1977) 20,064-46,443 and 3,314-63,135 for *M. seenghala* and *M. cavasius*, respectively. The current research findings were found to be correlated with the above-mentioned findings. Fecundity correlates with total body length, body weight and ovary weight (Sultan and Khan, 1982). The relation concerning fecundity and ovary length was found to be conspicuous among all the relationships. Present findings found to correlate with the findings of Gupta and Banerjee (2013) in *M. tengara* with a bit exception in case of total body weight.

Generally, catfishes are strong carnivorous species,

the food item taken by them generally being reported as remain intact in shape inside the stomach. Samples collected from bush trap fishing were dissected in the laboratory. Earlier studies on *Arius dussumieri* (Devanesan and Chidambaram, 1953; Suseelan and Nair, 1969; Menon, 1979) and *O. militaris* (Devanesan and Chidambaram, 1953; Venkataraman, 1960; Srikrishna, 1981) revealed these species to be carnivorous which is congruent with the present findings. Fishes mainly catfishes start to store fats as energy reserve in the coelomic cavity which is known as intra-peritoneal fat or coelomic fat during the pre-spawning period. Thereafter, this reserved energy gradually started to be decreasing prior to pre-spawning-onset of spawning.

### Conclusion

Information on reproductive biology for planning, conservation and management is important for a threatened fish species. The findings of the current study would be effective tools towards scheming management and conservation plans for sustainable conservation and captive maturation. The information gathered through the current study may be used for better management of Gagla catfish fishery in this country towards conservation and for induction of a potential species into aquaculture. However, manipulation of environmental parameters resembled to the month of July-August can be effective for artificial breeding and beginning of gonadal maturation. Further study should be carried out for more reliable information on the breeding biology of *A. gagora*. Studies are needed to culture the species in captive condition with other species

It is a prodigious inclination for the authors to express earnest appreciation to Bangladesh Fisheries Research Institute (BFRI), Mymensingh, Bangladesh for providing all the laboratory facilities.

### References

Abbasi K., Mouludi-Saleh A., Eagderi S., Sarpanah A. (2019). Length-weight relationship and condition factor of eight species of the genera *Capoeta*, *Garra*, *Chondrostoma*, *Schizothorax* and *Paraschistura* from

- Iranian inland waters. Iranian Journal of Ichthyology, 6(4): 264-270.
- Ahyaudin B.A. (1999). Aspects of the reproductive biology of female snakehead (*Channa striatus*) obtained from irrigated rice agro-ecosystem, Malaysia. Hydrobiologia, 411: 71-77.
- Alam M.M., Rahman M.T., Parween S. (2014). Morphometric characters and condition factors of five freshwater fishes from Pagla river of Bangladesh. International Journal of Aquatic Biology, 2: 14-19.
- Azad M.A.K., Hossain M.Y., Khatun D., Parvin M.F., Nawer F., Rahman O., Hossen M.A. (2018). Morphometric relationships of the tank goby *Glossogobius giuris* (Hamilton, 1822) in the Gorai River using multi-linear dimensions. Jordan Journal of Biological Sciences, 11: 81-85.
- Azadi M.A., Siddique M.S. (1996). Fecundity of catfish (in Bangladesh) *Heteropneustes fossilis* (Bloch). Bangladesh Journal of Zoology, 14: 33-39.
- Babiker M.M., Ibrahim H. (1979). Studies on the biology of reproduction in the cichlid *Tilapia nilotica*: gonadal development and fecundity. Journal of Fish Biology, 14: 437-448.
- Banu N., Ali S., Alam M.N. (1992). Studies on the fecundity of *Mystus tengara* (Hamilton) of Agargaon region, Dhaka University. Journal of Biological Sciences, 1: 49-51.
- Barnham P.S.M., Baxter A. (1998). Condition factor 'Kn' for Salmonid fish. Fisheries Notes, 5:1-3.
- Belsare D.K. (1962). Seasonal changes in the ovary of *Ophiocephalus punctatus*. Indian Journal of Fisheries, 9: 140-156.
- Bhatt V.S., Dalal S.G., Abidi S.A.H. (1977). Fecundity of the freshwater catfishes *Mystus seenghala* (Sykes), *Mystus cavasius* (Ham.), *Wallago attu* (Bloch) and *Heteropneustes fossilis* (Bloch) from the plains of northern India. Hydrobiologia, 54 (3): 219-224.
- Brito M.F.G., Bazzoli N. (2003). Reproduction of the surubim catfish (Pisces, Pimelodidae) in the Sao Francisco River, Pirapora Region, Minas Gerais, Brazil. Arquivo Brasileiro de Medicina Veterinária e Zootecnia, 55: 624-633.
- Bromage N.R., Porter M.J.R., Randall C.F. (2001). The environmental regulation of maturation in farmed finfish with special reference to the role of photoperiod and melatonin. Aquaculture, 197: 63-98.
- Cek S., Yilmaz E. (2009). The effect of varying dietary energy on gonad development at first sexual maturity of the Sharp tooth catfish (*Clarias gariepinus* Burchell, 1822). Aquaculture International, 17: 553-563.

- Chakrabarti P., Barun S. (2017). Histological organization and surface ultrastructure of ovaries of *Gudusia chapra* during different phases of reproduction (Teleostei: Clupeidae). Iranian Journal of Ichthyology, 4: 41-53.
- David A. (1963). Sexual dimorphism, fecundity and food of the estuarine bagrid, *Mystus gulio* (Ham.). Proceedings of the National Academy of Sciences, India, 33: 385-410.
- De Martini E.E., Fountain R. (1981). Ovarian cycling frequency and batch fecundity in the queen fish, *Seriphus politus*: Attributes representative of serial spawning fishes. Fisheries Bulletin, 79: 547-559.
- Devanesan D.W., Chidambaram K. (1953). The common food fishes of the Madras Presidency. Madras Govt. Publication. 207 p.
- Dobriyal A.K., Rautela K.K., Rautela A.S. (1999). Invention of a new index for the determination of sexual maturity in fishes. Uttar Pradesh Journal of Zoology, 19: 207-209.
- Dorostghoal M., Peyghan R., Papan F., Khalili. (2009). Macroscopic and microscopic studies of annual ovarian maturation cycle of Shirbot (*Barbus grypus*) in Karoon river of Iran. Iranian Journal of Veterinary Research, 10: 172-179.
- Fafioye O.O., Oluajo O.A. (2005). Length-weight relationships of five fish species in Epelagoon, Nigeria. African Journal Biotechnology, 4(7): 749-751.
- Fulton F. (1904). The rate of growth of fishes. 22<sup>nd</sup> Annual Report of the Fishery Board of Scotland 3: 141-241
- Glasser F., Mikolajczyk T., Jalabert B., Baroillerand J.F., Breton B. (2004). Temperature effects along with the reproductive axis during spawning induction of grass carp (*Ctenopharyngodon idella*). General and comparative Endocrinology, 136: 171-179.
- Gupta B.K., Sarker U.K., Bhardwaj S.K. (2014). Pabda in river Gomti. Journal of Environmental Biology, 35(2): 345-51.
- Gupta S., Banerjee S. (2013). Studies on reproductive biology of *Mystus tengara* (Ham. -Buch., 1822), a freshwater catfish of West Bengal, India. International Journal of Aquatic Biology, 1: 175-184.
- Ha P.Y., Kinzie R.A. (1996). Reproductive biology of *Awaous guamensis*, an amphibian Hawaiian Goby. Environmental Biology of Fishes, 45: 383-396.
- Hamilton F. (1822). An account of the fishes found in the river Ganges and its branches. Constable and Co, Edinburgh, UK. 405 p.
- Hong-Yang P., Yon J.M. (1992). Histological changes of ovary, testis and pituitary gland in reproductive period of rainbow trout. *Oncorhynchus mykiss*, Aquaculture, 92: 21-25.
- Hora S.L., Law N.C. (1941). The freshwater fish of Travancore. Records of the Indian Museum, 43: 254-255.
- Hossain M.Y. (2010a). Morphometric relationships of length-weight and length-length of four cyprinid small indigenous fish species from the Padma River (NW Bangladesh). Turkish Journal of Fisheries and Aquatic Sciences, 10: 213-216.
- Hossain M.Y. (2010b). Length-weight, length-length relationships and condition factor of three schibid catfishes from the Padma River, northwestern Bangladesh. Asian Fish Science, 23: 329-339.
- Hossain M.Y., Hossen M.A., Ali M.M., Pramanik M.N.U., Nower F., Rahman M.M., Sharmin S., Khatun D., Bahkali A.H., Elgorban A.M., Yahya K. (2017). Life-history traits of the endangered carp *Botia dario* (Cyprinidae) from the Ganges River in northwestern Bangladesh. Pakistan Journal of Zoology, 49: 801-809.
- Hossain M.Y., Jewel M.A.S., Nahar L., Rahman M.M., Naif A., Ohtomi J. (2012) Gonadosomatic index-based size at first sexual maturity of the catfish *Eutropiichthys vacha* (Hamilton, 1822) in the Ganges River (NW Bangladesh). Journal of Applied Ichthyology, 28: 601-605.
- Hossain M.Y., Khatun M.M., Jasmine S., Rahman M.M., Jewel M.A.S., Ohtomi J. (2013). Life history traits of the threatened freshwater fish *Cirrhinus reba* (Hamilton, 1822) (Cypriniformes: Cyprinidae) in the Ganges River, northwestern Bangladesh. Sains Malaysiana, 42: 1219-1229.
- Hossain M.Y., Naser S.M.A., Bahkali A.H., Yahya K., Hossen M.A., Elgorban A.M., Islam M.M., Rahman M.M. (2016). Life history traits of the flying barb *Esomus danricus* (Cyprinidae) in the Ganges River, northwestern Bangladesh. Pakistan Journal of Zoology, 48: 399-408.
- Hunter J.R., Leong R. (1981). The spawning energetics of female northern anchovy, *Engraulis mordax*. Fishery Bulletin, 79: 215-229.
- Isa M.M., Noor N.S.M., Yahya K., Nor S.A.M. (2012). Reproductive biology of estuarine catfish, *Arius argyropleuron* (Siluriformes: Ariidae) in the northern part of Peninsular Malaysia. Journal of Biology, Agriculture and Healthcare, 2: 14-28.
- Isa M.M., Rawi M.C.S., Rosla R., Shah M.S.A., Shah M.A.S.R. (2010). Length-weight Relationships of Freshwater Fish Species in Kerian River Basin and Pedu Lake. Research Journal of Fisheries and Hydrobiology, 5(1): 1-8.

- Islam M.A., Begum M., Pal H.K., Alam M.J. (2008). Studies on the Gonadosomatic Index and Fecundity of *Mystus gulio* (Ham.). *Progressive Agriculture*, 19(2): 161-166.
- Jacob S.S., Nair N.B. (1983). Reproductive biology of the Larvivorous fish *Macropodus Capanus* (CuvandVal). *Proceedings of the Indian Academy of Sciences*, 92: 159-170.
- Jhingran A.G., Verma D.N. (1972). Sexual maturity and spawning of *Gudusia chapra* (Ham.) in Ganga river system. *Proceedings of the Indian National Science Academy*, 42: 207-224.
- Kabir M.A., Ghaedi A. Hashim R. (2012). Ovarian development and sexual maturation of female striped catfish, *Pangasianodon hypophthalmus* (Sauvage, 1878) reared in captivity. *Asian Fisheries Science*, 25: 232-244.
- Kailola P.J. (1999). Family Ariidae. Species identification guide for fisheries purposes. The living marine resources of the western central Pacific. Batoid fishes, chimeras and bony fishes part 1 (Elopidae to Linophrynidae), 1827-1879.
- Keivany Y., Nezamoleslami A., Dorafshan S., Eagderi S. (2015). Length-weight and length-length relationships in populations of *Garra rufa* from different rivers and basins of Iran. *International Journal of Aquatic Biology*, 3(6): 409-413.
- Khatun D., Hossain M.Y., Parvin M.F., Ohtomi J. (2018). Temporal variation of sex ratio, growth pattern and physiological status of *Eutropiichthys vacha* (Schilbeidae) in the Ganges River, NW Bangladesh. *Zoology and Ecology*, 28: 343-354.
- Khatun M.A., Rashid M.B., Hygen H.O. (2017). Climate of Bangladesh. MET report published by Bangladesh meteorological department, Dhaka and Norwegian meteorological institute, Norway, (7-8): 9-66.
- Konan Y.A., Kone T., Bamba M., Kone M. (2014). Reproductive Strategies of the Catfish *Clarias buettikoferi* (Pisces, Clariidae) in the Tanoe-Ehy Swamp Forest (South-Eastern Côte d'Ivoire). *World Journal of Fish and Marine Sciences*, 6 (1): 16-23.
- Laleye P., Chikou A., Gnohossou P., Vandewalle P., Philippart J.C., Teugels G. (2006). Studies on the biology of two species of catfish *Synodontis schall* and *Synodontis nigrita* (Ostariophysi: Mochokidae) from the Oueme River, Benin. *Belgian Journal of Zoology*, 136: 193-201.
- Lehri G.K. (1968). Cyclical Changes in the ovary of catfish *Clarius batrachus*. *Acta Anatomica*, 69: 105-124.
- Lincoln G.A., Racey P.A., Share P.J., Kland H. (1980). Endocrine changes associated with spring And Autumn sexually of rook. *Corvus Frugilegus Journal*, 190: 137-153.
- Menon N.G. (1979). Studies on the Biology and Fishery of Giant Marine Catfish *Tachysurus thalassinus* (Rupel!). Unpubl. Ph.D. Thesis, University of Cochin, India.
- Mir J.I., Sarkar U.K., Dwivedi A.K., Gusain O.P., Pal A., Jena J.K. (2012). Pattern of Intrabasin variation in condition factor, relative condition factor of an Indian major carp, *Labeo rohita* (Hamilton-Buchanan, 1822) in the Ganges Basin India. *Journal of Biological Sciences*, 4: 126-135.
- Mookherjee H.K., Mazumdar S.R., Dasgupta B. (1941). Sexual dimorphism in *Macrones gulio* (Ham.). *Indian Journal of Veterinary Science*, 10(3): 295.
- Morgan J. (2008). Integrating Reproductive Biology into Scientific Advice for Fisheries Management. *Journal of Northwest Atlantic Fishery Science*, 41: 37-51.
- Mouludi-Saleh A., Eagderi S. (2019). Length-weight relationship and condition factor of ten fish species (Cyprinidae, Sisoridae, Mugilidae, Cichlidae, Gobiidae and Channidae) from Iranian inland waters. *Journal of Wildlife and Biodiversity*, 3(4): 12-15.
- Muchlisin Z.A., Musman M., Azizah M.N.S. (2010). Spawning seasons of *Rasbora tawarensis* (Pisces: Cyprinidae) in Lake Laut Tawar, Aceh Province, Indonesia. *Reproductive Biology and Endocrinology*, 8: 49.
- Mukherji D.D. (1936). Report on fishes, Part II, Sisoridae and Cyprinidae. *Memoirs of the Connecticut Academy of Arts and Sciences*, 10: 328-329.
- Murua H., Rey F.S. (2003). Female Reproductive Strategies of Marine Fish Species of the North Atlantic. *Journal of Northwest Atlantic Fishery Science*, 33: 23-31.
- Musa A.S.M., Bhuiyan A.S. (2007). Fecundity of *Mystus bleekeri* (Day, 1877) from the River Padma near Rajshahi city. *Turkish Journal of Fisheries and Aquatic Sciences*, 7: 161-162.
- Narejo N.T., Rahmatullah S.M., Rashid M.M. (2002). Studied the reproductive biology of freshwater spiny eel, *Mastacembelus armatus* (Lacepede) reared in the concerned cistern. *Journal of Aquaculture in the Tropics*, 17(4): 251-259.
- Nikolsky G.V. (1963). The ecology of fishes. Academic Press. London and New York. 352 p.
- Noor M.A.S. (2013). Length-length and length-weight relationships of critically endangered striped catfish *Rita rita* (Hamilton) from the Padda River near Rajshahi of Northwestern Bangladesh. *Journal of Pharmacy and*

- Biological Sciences, 4: 32-36.
- Pauly D. (1983). Some simple methods for the assessment of tropical fish stocks. FAO, Rome (234). 52 p.
- Radkhah A., Eagderi S. (2015). Length-weight and length-length relationships and condition factor of six cyprinid fish species from Zarrineh River (Urmia Lake basin, Iran). *Iranian Journal of Ichthyology*, 2(1): 61-64.
- Rahman A.K.A. (1989). *Freshwater Fishes of Bangladesh*, 1<sup>st</sup> edition, Zoological Society of Bangladesh, Department of Zoology, University of Dhaka, Dhaka-1000. 233 p.
- Rahman A.K.A. (2005). *Freshwater Fishes of Bangladesh*, 2<sup>nd</sup> edition, Zoological Society of Bangladesh, Department of Zoology, University of Dhaka, Dhaka-1000. 256 p.
- Reidel A., Boscolo W.R., Feiden A., Romagosa E. (2010). The effect of diets with different levels of protein and energy on the process of final maturation of the gametes of *Rhamdia quelen* stocked in cages. *Aquaculture*, 298: 354-359.
- Shankar D.S., Kulkarni R.S. (2005). Somatic condition of the fish, *Notopterus notopterus* (pallas) during phases of the reproductive cycle. *Journal of Environmental Biology*, 26(1): 49-53.
- Shashi B.S., Akela B.P. (1996). Determination of maturity and spawning period by gonadosomatic index and measurement of mean ova diameter in certain teleosts. *Environment and Ecology*, 14: 399-403.
- Shinkafi B.A., Ipinjolu J.K., Argungu L.A., Abubakar U. (2002). Length-weight relationship and fecundity of *Synodontis clarias* (Linnaeus) in River Rima, Nigeria. *Journal of Agriculture and Environment*, 3(1): 147-154.
- Shinkafi B.A., Ipinjolu J.K., Hassan W.A. (2011). Gonad maturation stages of *Auchenoglanis occidentalis* (Valenciennes 1840) in River Rima, North-Western Nigeria. *Journal of Fisheries and Aquatic Science*, 6: 236-246.
- Somarakis S., Maraveya E., Tsimenides N. (2000). Multispecies ichthyoplankton associations in epipelagic species: is there any intrinsic adaptive function? *Belgian Journal of Zoology*, 130: 125-129.
- Srikrishna. (1981). Studies on the Morphometry and Biology of Catfish *Osteogeneiosus militaris* (Linn.), off Bombay coast. Unpubl. M.Sc. Thesis, University of Mumbai, India. 257 p.
- Stoumboundi M.T., Villwock W., Sela J., Abraham M. (1993). Gonadosomatic index in *Barbus longiceps*, *Capoeta damascina* and their natural hybrid (Pisces, Cyprinidae), versus spermatozoan index in the parental males. *Journal of Fish Biology*, 43: 865-875.
- Sultan S., Khan S.M. (1982). The fecundity of the catfish *Mystus vittatus* (Bloch). *Indian Journal of Animal Research*, 16: 2116-118.
- Suseelan C., Nair K.V.S. (1969). Food and feeding habits of the demersal fishes off Bombay. *Indian Journal of Fisheries*, 16(1): 56-74.
- Takasuka A., Oozeki Y., Kubota H., Tsuruta Y., Funamoto T. (2005). Temperature impacts on reproductive parameters for Japanese anchovy: comparison between inshore and offshore waters. *Fisheries Research*, 76: 475-482.
- Talwar P.K., Jhingran A.G. (1991). *Inland Fishes of India and Adjacent Countries*, Oxford & IBH Publishing Co. Pvt. Ltd. 1158 p.
- Tyor A.K., Pahwa K. (2017). Ovarian Development of African Sharptooth Catfish (*Clarias gariepinus*, Burchell 1822) from Delhi Segment of River Yamuna. *Journal of Fisheries and Aquatic Science*, 12: 117-126.
- Vandyk J.K., Pieterse G.M. (2008). A histo-morphological study of the testies of the sharptooth catfish (*Clarias gariepinus*) as reference for future toxicological assessments. *Journal of Applied Ichthyology*, 24: 415-422.
- Venkataraman G. (1960). Studies on the food and feeding relationship of the inshore fishes off Calicut on the Malabar coast. *Indian Journal of Fisheries*, 7(2): 275-306.
- Vergheese P.U. (1975). Internal rhythm of sexual cycle in a Carp *Cirrhinus reba* (Ham) Under artificial condition of darkness. *Journal of the Inland Fisheries Society of India*, 7: 82-88.
- Winemiller K., Rose K. (1992). Patterns of life-history Diversification in North American fishes: implications for population regulation. *Canadian Journal of Fisheries and Aquatic Sciences*, 49: 2196- 2218.
- Zamani-Faradonbeh M.Z., Eagderi S., Ghojoghi F. (2015). Length-weight relationship and condition factor of seven fish species of Totkabon River (southern Caspian Sea basin), Guilan, Iran. *International Journal of Aquatic Biology*, 3(3): 172-176.