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[Application of](https://core.ac.uk/display/35170616?utm_source=pdf&utm_medium=banner&utm_campaign=pdf-decoration-v1) Pyrolysis–Gas Chromatography– Mass Spectrometry for the Identification of **Polymeric Materials**

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The analytical pyrolysis technique hyphenated to gas chromatography–mass spectrometry (GC–MS) has extended the range of possible tools for the characterization of synthetic polymers and copolymers. Pyrolysis Involves thermal fragmentation of the analytical sample at temperatures of 500–1400 °C. In the presence of an inert gas, reproducible decomposition products characteristic for the original polymer or copolymer sample are formed. The pyrolysis products are chromatographically separated using a fused-silica capillary column and are subsequently identified by interpretation of the obtained mass spectra or by using mass spectra Ilibraries. The analytical technique eliminates the need for pretreatment by performing analyses directly on the solid or liquid polymer sample. In this article, application examples of analytical pyrolysis hyphenated to GC-MS for the identification of different polymeric materials in the plastic and automotive industry, dentistry, and occupational safety are demonstrated. For the first time, results of identification of commercial light-curing dental filling material and a car wrapping foil by pyrolysis-GC-MS are presented.

Structural analysis and the study of degradation properties are important to understand and improve performance characteristics of synthetic polymers and copolymers in many industrial applications. Traditional analytical techniques used for characterization of polymers and copolymers such as thermal analysis and Fourier transform infrared (FT–IR) spectroscopy have limitations or are not sufficiently sensitive (1). Pyrolysis techniques hyphenated to gas chromatography–mass spectrometry (GC–MS) have extended the range of possible tools for the characterization of synthetic polymers and copolymers. Under controlled conditions, at elevated temperatures (500–1400 °C) in the presence of an inert gas, reproducible decomposition products characteristic for the original polymer or copolymer sample are formed. The pyrolysis products are chromatographically separated using a fused-silica capillary column and subsequently identified by interpretation of the obtained mass spectra or by using mass spectra libraries (such as the National Institute of Standards and Technology [NIST] or Wiley). Pyrolysis methods eliminate the need for pretreatment by performing analyses directly on the solid polymer or copolymer sample (1). (Please note that this article was presented at the XVII European Conference on Analytical Chemistry, which was held in Warsaw, Poland, on 25–29 August 2013).

Most of the thermal degradation results from free radical reactions initiated by bond breaking and depends on the relative strengths of the bonds that hold the molecules together. A large molecule will break apart and rearrange in a characteristic

way (2–4). If the energy transfer to the sample is controlled by temperature, heating rate, and time, the fragmentation pattern is reproducible and characteristic for the original polymer or copolymer. Another sample of the same composition, heated at the same rate to the same temperature for the same period of time, will produce the same decomposition products. Therefore, the essential requirements of the apparatus in analytical pyrolysis are reproducibility of the final pyrolysis temperature, rapid temperature rise, and accurate temperature control. Depending on the heating mechanism, pyrolysis systems have been classified into two groups: the continuous-mode pyrolyzer (furnace pyrolyzer) and pulse-mode pyrolyzer (flash pyrolyzer, such as the heated filament, Curie-point, and laser pyrolyzer). The pyrolysis unit is directly connected to the injector port of a gas

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- Pyrolysis–GC–MS is a valuable technique for the analysis and identification of synthetic polymers and copolymers.
- The technique described allows the direct analysis of very small sample amounts (5–200 μg) without the need for time-consuming sample preparation.
- Commercial light-curing dental filling material and car wrapping foil were identified using this method.

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Figure 2: Pyrolysis–GC–MS chromatogram of plastic particles from industrial filter fins at 700 °C obtained with apparatus 1. Fused-silica GC capillary column: 60 m \times 0.25 mm, 0.25-um d_f Elite-5ms. GC conditions: programmed column temperature: 60 °C for 1 min, then 60–100 °C at 2.5 °C/min and then 100–280 °C at 10 °C/min (20 min hold at 280 °C); split– splitless injector temperature: 250 °C; split flow: 50 cm³/ min; helium programmed pressure: 70 kPa for 1 min, then 70–110 kPa at 1 kPa/min (hold at 110 kPa to the end of analysis). For peak identification, see Table 1.

chromatograph. A flow of an inert carrier gas, such as helium, flushes the pyrolyzates into the fused-silica capillary column. Figure 1 shows the schematic view of the furnace pyrolyzer used in our investigation. The detection technique of the separated compounds is typically MS, but other GC detectors have also been used depending on the intentions of the analysis (1,4).

The applications of analytical pyrolysis–GC–MS range from research and development of new materials, quality control, characterization and competitor product evaluation, medicine, biology and biotechnology, geology, airspace, and environmental analysis to forensic purposes or conservation and restoration of cultural heritage. These applications cover analysis and identification of polymers, copolymers, and additives in components of automobiles, tyres, packaging materials, textile fibres, coatings, half-finished products for electronics, paints or varnishes, lacquers, leather, paper or wood products, food, pharmaceuticals, surfactants, and fragrances.

Our earlier publications (1,5–12) presented the analysis and identification of degradation products of commercially available synthetic polymers and copolymers by using analytical pyrolysis hyphenated to gas chromatography with flame ionization detection (GC–FID) and GC–MS. In this work, new examples of applications of this analytical technique for the identification of different polymeric materials are demonstrated.

Experimental

Samples: Plastic particles from industrial filter fins, a car wrapping foil, unknown fibres, and commercial light-curing dental filling material were used in the investigation.

Instrumentation and Analytical Conditions: Approximately 100–200 µg of solid sample was cut out with a scalpel and inserted without any further preparation into the bore of the pyrolysis solids-injector and then placed with the plunger on the quartz wool of the quartz tube of the furnace pyrolyzer Pyrojector II (SGE Analytical Science). Three spots on each sample were analyzed in duplicate. The pyrolyzer was operated at a constant temperature of 550, 600, 700, or 900 °C. The pressure of helium carrier gas at the inlet to the furnace was 95 kPa.

Pyrolysis–GC–MS measurements were made using two apparatus. In the first apparatus (1), the pyrolyzer was connected to a Trace 2000 gas chromatograph (ThermoQuest, CE Instruments) with a quadrupole mass spectrometer Voyager (ThermoQuest, Finnigan, MassLab Group) operated in electron ionization (EI) mode. A 60 m \times 0.25 mm, 0.25-um Elite-5ms fused-silica GC capillary column (PerkinElmer Instruments) was used. The GC conditions were as follows: programmed

material at 600 °C obtained with apparatus 1. Fused-silica GC capillary column 60 m \times 0.25 mm, 0.25-µm $d_{\rm f}$ Elite-5ms. GC conditions: programmed column temperature: 60 °C for 1 min, then 60–100 °C at 2.5 °C/min and then 100–280 °C at 10 °C/ min (20 min hold at 280 °C); split–splitless injector temperature: 250 °C; split flow: 50 cm³/min; helium programmed pressure: 70 kPa for 1 min, then 70–110 kPa at 1 kPa/min (hold at 110 kPa to the end of analysis). For peak identification, see Table 2.

Figure 4: Pyrolysis–GC–MS chromatogram of polyaramid fibers at 900 °C obtained with apparatus 2. Fused-silica GC capillary column: 59 m \times 0.25 mm, 0.25-µm $d_{\rm f}$ DB-5ms. GC conditions: programmed column temperature: 75 °C for 1 min, then 75–280 °C at 7 °C/min (hold to the end of analysis); programmed pressure of helium carrier gas: 122.2 kPa for 1 min, then 122.2–212.9 kPa at 7 kPa/min (hold at 212.9 kPa to the end of analysis). For peak identification, see Table 3.

column temperature: 60 °C for 1 min, then 60–100 °C at 2.5 °C/ min, 100–280 °C at 10 °C/min (20-min hold at 280 °C). The temperature of the split–splitless injector was 250 °C and the split flow was 50 cm³/min. Helium, grade 5.0 (Westfalen AG), was used as a carrier gas. The helium programmed pressure was 70 kPa for 1 min, then 70–110 kPa at 1 kPa/min (hold at 110 kPa to the end of analysis) was used. The transfer line temperature was 280 °C. The MS EI ion source temperature was kept at 250 °C. The ionization occurred with a kinetic energy of the impacting electrons of 70 eV. The current emission of the rhenium filament was 150 µA. The MS detector voltage was 350 V. Mass spectra and reconstructed chromatograms (total ion current [TIC]) were obtained by automatic scanning in the mass range m/z 35-450 u. Pyrolysis–GC–MS data were processed with the Xcalibur software (ThermoQuest) and the NIST 05 mass spectra library.

In the second apparatus (2), the pyrolyzer was connected to a 7890A gas chromatograph with a series 5975C quadrupole mass spectrometer (Agilent Technologies Inc.) operated in EI mode. A 59 m \times 0.25 mm, 0.25-µm $d_{\rm f}$ DB-5ms fused-silica GC capillary column (J&W Scientific) was used. Helium, grade 5.0 (Westfalen AG), was used as a carrier gas. The GC conditions were as

follows: programmed column temperature: 60 °C for 1 min, then 60–280 °C at 7 °C/min (hold at 280 °C to the end of analysis); programmed helium pressure: 122.2 kPa for 1 min, then 122.2– 212.9 kPa at 7 kPa/min (hold at 212.9 kPa to the end of analysis). Second set of GC conditions: programmed column temperature: 75 °C for 1 min, then 75–280 °C at 7 °C/min (hold at 280 °C to the end of analysis); programmed helium pressure: 122.2 kPa for 1 min, then 122.2–212.9 kPa at 7 kPa/min (hold at 212.9 kPa to the end of analysis).

The temperature of the split–splitless injector was 250 °C and the split ratio was 50:1. The transfer line temperature was 280 °C. The MS EI ion source temperature was kept at 230 °C. The ionization occurred with a kinetic energy of the impacting electrons of 70 eV. The quadrupole temperature was 150 °C. Mass spectra and reconstructed chromatograms (total ion current) were obtained by automatic scanning in the mass range m/z 35–750 u. Pyrolysis–GC–MS data were processed with the ChemStation software (Agilent Technologies) and the NIST 05 mass spectra library.

Results and Discussion

Pyrolysis-GC-MS of Plastic Particles from Industrial Filter Fins: A sample of plastic particles from industrial filter fins was pyrolyzed at 700 °C to identify its composition. Figure 2 shows the obtained pyrolysis–GC–MS chromatogram of the sample. Based on the decomposition products summarized in Table 1, the plastic particles were identified as a mixture of poly(hexamethylene adipamide) (nylon 6-6) and polypropylene glycol with a small amount of styrene–butadiene rubber (SBR). The peaks of propylene and acetone indicate the presence of polypropylene glycol. The main decomposition product of nylon 6-6 is cyclopentanone (retention time $[t_R] = 10.51$ min). Other peaks in Figure 2, like butene/1,3-butadiene ($t_R = 5.58$ min), benzene ($t_R = 7.43$ min), toluene ($t_R = 9.72$ min), and styrene $(t_R = 14.39$ min), are typical pyrolysis products of SBR (1,2,5,6,18). The small peak of 4-isopropylphenol ($t_R = 28.15$ min) may be a clue to the presence of polycarbonate or bisphenol A epoxy resin (5,6). All of the pyrolysis products and the materials identified from pyrolysis products in filter fins are summarized in Table 1.

Pyrolysis-GC-MS of a Car Wrapping Foil: The next object of identification was a car wrapping foil pyrolyzed at 600 °C. Figure 3 shows the obtained pyrolysis–GC–MS chromatogram of the car wrapping foil. Based on the decomposition products summarized in Table 2, the plastic material was identified as a mixture of flexible poly(vinyl chloride) (PVC) with bis(2-ethylhexyl) phthalate (BEHP) plasticizer and poly(hexamethylene adipamide) (nylon 6-6). The chromatogram in Figure 3 shows the typical pyrolysis products of PVC, like hydrogen chloride ($t_B = 5.49$ min), benzene ($t_p = 7.48$ min), and naphthalene ($t_p = 25.93$ min). This is the result of the formation of double bonds by the elimination of hydrogen chloride from the poly(vinyl chloride) macromolecules, followed by the breaking of the carbon chain with or without cyclization reaction (2). The detected cyclopentanone $(t_R = 10.46$ min) is generally known as a characteristic pyrolysis product of nylon 6-6 (2,3,6). Methyl methacrylate ($t_{\text{R}} = 8.20$ min) identified in pyrolyzate is formed from poly(methyl methacrylate) (6) and most likely comes from the adhesive film. Thus, the identified 3,3-diphenylacrylonitrile ($t_{\text{B}} = 36.69$ min) may be from the adhesive layer of the foil.

The thermal decomposition of the plasticizer bis(2-ethylhexyl) phthalate identified in car wrapping foil leads to the formation at 600 °C of 2-ethyl-1-hexene ($t_B = 10.29$ min), 2-ethylhexanal

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 $(t_R = 17.33$ min), 2-ethyl-1-hexanol ($t_R = 20.57$ min), and phthalic anhydride ($t_P = 28.63$ min) (1.7). In the car wrapping material, the rest of the tetrahydrofuran solvent ($t_{\text{R}} = 6.99$ min) was also detected. Table 2 shows the identified ingredients of the pyrolyzed car wrapping foil.

Identification of Unknown Plastic Fibres: A sample of unknown plastic fibres was pyrolyzed at 700 °C and 900 °C, respectively, to identify its composition. Figure 4 shows the pyrolysis–GC–MS chromatogram of the sample pyrolyzed at 900 °C. Based on the decomposition products summarized in Table 3, the fibres were identified as polyaramid $[poly(p -phenylene terephthalamide)]$ (Figure 5). The main identified degradation products of polyaramid at 900 °C are benzene (t_{R} = 7.56), aniline (t_{R} = 11.61 min), and benzonitrile (t_{R} = 11.81 min). Currently, polyaramid fibres have only been characterized in a few publications using thermal analysis

Figure 5: Chemical structure of poly(p-phenylene terephthalamide) (polyaramid).

(thermogravimetry, derivative thermogravimetry, and differential thermal analysis), infrared spectroscopy techniques (13–15), and pyrolysis–GC–MS (2,16–19).

Polyaramid fibres are a class of heat-resistant, strong synthetic fibres. They are used in aerospace and military applications for ballistic-rated body armour, fabric, ballistic composites, and fire fighters protective clothing as well as in bicycle tyres and as an asbestos substitute.

Identification of Commercial Light-Curing Dental Filling Material: A number of dental filling materials are presently available for tooth restorations. The four main groups of these materials, which dentists have used for about 35 years, are the conventional glass-ionomer cements, resin-based composites, resin-modified glass-ionomer cements, and polyacid-modified resinous composites (20). Light-curing glass-ionomer cements contain polyacrylic acid, chemically or photo-curing monomers (multifunctional methacrylates, like triethylene glycol dimethacrylate or 2-hydroxyethyl methacrylate), an ion-leaching glass, and additives (initiators, inhibitors, stabilizers, and others) (20). Resin-modified glass-ionomer cements are now widely used in dentistry as direct filling materials, liners, bases, luting cements, and fissure sealants (21). These materials mainly consist of polymer matrix and glass-ionomer parts. The polymer matrix is based on a monomer system and different multifunctional methacrylates with additives (21). Methacrylic monomers, such as bisphenol A glycidyl methacrylate (Bis-GMA), urethane

Figure 6: Pyrolysis–GC–MS chromatogram of commercial light-curing dental filling material at 550 °C obtained with apparatus 2. Fused-silica GC capillary column: 59 m × 0.25 mm, 0.25-um d DB-5ms. GC conditions: programmed column temperature: 60 °C for 1 min, then 60–280 °C at 7 °C/min (hold at 280 °C to the end of analysis); programmed helium pressure: 122.2 kPa for 1 min, then 122.2–212.9 kPa at 7 kPa/min (hold at 212.9 kPa to the end of analysis). For peak identification, see Table 4.

Table 4: Pyrolysis products of commercial light-curing dental filling material.

dimethacrylate (UDMA), triethylene glycol dimethacrylate (TEGDMA), and 2-hydroxyethyl methacrylate (HEMA), are the main components of resin-based dental filling materials. The presence of additives such as initiators, activators, inhibitors, and plasticizers in uncured dental material mixture is necessary (21).

Figure 6 shows the total ion current pyrolysis–GC–MS chromatogram of commercial light-curing dental filling material pyrolyzed at 550 °C. The pyrolysis products identified by using mass spectra library NIST 05 are summarized in Table 4.

The carbon dioxide ($t_R = 6.85$ min) identified in pyrolyzate is formed from polyacrylic acid (2,18). The identified substances 2-hydroxyethyl methacrylate (HEMA) ($t_R = 13.65$ min), ethylene glycol dimethacrylate (EGDMA) ($t_R = 19.48$ min), and triethylene glycol dimethacrylate (TEDMA) ($t_R = 28.72$ min) are known as standard composites of dental filling materials (1). Other compounds in Table 4, such as bisphenol A (t_R = 33.10 min) or bisphenol A diglycidyl ether ($t_R = 42.42$ min), are probably formed by thermal degradation of bisphenol A diglycidyl monoor dimethacrylates. The presence of the additives, such as the antioxidant 2,6-bis(1,1-dimethylethyl)-4-methylphenol (BHT) (t_P) = 23.17 min) or the UV-absorber drometrizol (t_R = 31.95 min) was also confirmed. The triphenylantimony ($t_R = 34.55$ min) identified in pyrolyzate is used as catalyst in the UV-induced polymerization (1).

Conclusion

Analytical pyrolysis–GC–MS has been proven as a valuable technique for the analysis and identification of organic polymeric materials in the plastic and rubber industry. For the first time pyrolysis–GC–MS was used for the identification of commercial light-curing dental filling material and for the identification of a car wrapping foil. This technique allows the direct analysis of very small sample amounts (5–200 µg) without the need for time-consuming sample preparation.

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