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The Wnt Signalling Pathway: A Tailored Target in Cancer

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1 Review

2 The Wnt signalling pathway: a tailored target in

3 cancer

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- 8 Abstract: Cancer is one of the greatest public health challenges. According to the World Health 9 Organization (WHO) 9.6 million cancer deaths have been reported in 2018. The most common 10 cancers include lung, breast, colorectal, prostate, skin cancer (non-melanoma) and stomach. The 11 unbalance of physiological signalling pathways due to the acquisition of mutations in tumour cells 12 is considered the most common cancer driver. The Wingless-related integration site (Wnt)/ β -catenin 13 pathway is crucial for tissue development and homeostasis in all animal species and its 14 dysregulation is one of the most relevant events linked to cancer development and dissemination. 15 The canonical and the non-canonical Wnt/β-catenin pathways are known to control both
- physiological and pathological processes including cancer. Herein the impact of the Wnt/β-catenin cascade in driving cancers from different origin has been examined. Finally, based on the impact of Extracellular Vesicles (EVs) on tumour growth, invasion and chemoresistance, and their role as tumour diagnostic and prognostic tools, an overview of the current knowledge linking EVs to the
- Wnt/ β -catenin pathway is also discussed.
 - **Keywords:** Wnt/ β -catenin dependent pathway; Wnt/ β -catenin independent pathway; colorectal cancer; breast cancer; ovarian cancer; extracellular vesicles

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Introduction

The human wingless-related integration site (Wnt) genes encode 19 evolutionarily conserved glycoproteins with 22-24 Cys residues. In the endoplasmic reticulum (ER), the Wnt ligands are post-translationally acetylated by porcupine, a membrane associated O-acyl transferase. Acetylation leads to palmitoylation which is required for the release and binding of Wnt to the frizzled (*FZD*) receptors. This, on turn, drives the biological response[1].

The Wnt signalling pathway regulates crucial cellular processes including cell fate determination, organogenesis during embryonic development, normal adult homeostasis, motility, polarity and stem cell renewal[2]. Moreover, its contribute in cancer has been extensively investigated[3].

The Wnt pathway has been widely studied and reviewed, and a general understanding of the transduction cascade has been clarified. The Wnt cascade has been subdivided into different branches due to its complexity[4,5]. They include the canonical Wnt/ β -catenin (Wnt/ β -catenin dependent pathway) and the non-canonical Wnt/ β -catenin pathway (β -catenin-independent pathway). The latter was further allocated into two additional branches, the planar cell polarity (PCP) and the Wnt/calcium pathways[2]. Both of them contribute to cancer development and dissemination.

The aim of the present review is to provide an overview of the current knowledge about the Wnt signalling pathway in tumour development and progression. Tumours from different origin are discussed. Although the canonical and the non-canonical Wnt/ β -catenin pathway work together to control physiological and pathological processes[2], data related to each one are independently

debated. Finally, the contribute of extracellular vesicles (EVs) in triggering the Wnt/ β -catenin cascade is also analyzed.

Wnt canonical pathway: β-catenin dependent

The canonical pathway turns around the β -catenin intracellular level (Figure 1). In the absence of Wnt proteins the β -catenin "destruction complex" keeps low β -catenin in the cell. The "destruction complex" mainly consists of two kinases: casein kinase 1α ($CK1\alpha$), glycogen synthase kinase 3β ($GSK-3\beta$) and two scaffolds: axis inhibition (Axin), and adenomatous polyposis coli (APC). Firstly, β -catenin undergoes phosphorylation by $CK1\alpha$ at serine 45 (Ser45), Ser33, Ser37 and threonine 41 (Thr41) by $GSK-3\beta$. Then, the E3 ubiquitin ligase, denoted as β -transducin repeat—containing protein ($\beta TrCP$), marks β -catenin ubiquitination and degradation [1]. This prevents β -catenin nuclear translocation while allows histone deacetylation and chromatin compaction by the Groucho repressor, translating into the inhibition of gene transcription[6] (Figure 1a).

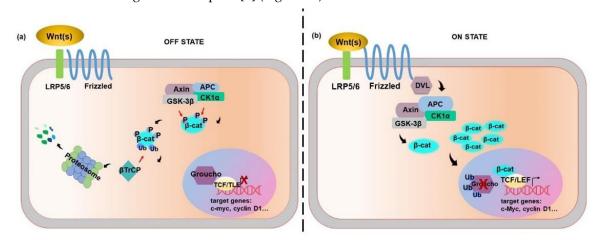


Figure 1. The Canonical Wnt signalling pathway. (a) **OFF STATE**. In the absence of Wnt ligands β-catenin moves to the "destruction complex" consisting of casein kinase 1α (*CK1α*), glycogen synthase kinase 3β (*GSK-3β*) and two scaffolds: axis Inhibition (*Axin*), and adenomatous polyposis coli (*APC*). β-catenin undergoes phosphorylation at Ser45 residue by *CK1α* and at Ser33, Ser37 and Thr41 residues by *GSK-3β*. Then, the E3 ubiquitin ligase β-transducin repeat–containing protein ($\beta TrCP$), marks β-catenin ubiquitination and proteasomal degradation. This prevents β-catenin nuclear accumulation while allows chromatin compaction and Groucho-mediated promoter repression. (b) **ON STATE**. The Wnt ligands bind to frizzled (*FZD*) receptor and the low-density-lipoprotein-related protein 5/6 (*LRP5/LRP6*), this results in dishevelled (*DVL*) phosphorylation and β-catenin release from the "destruction complex", allowing β-catenin accumulation and nuclear translocation. In the nucleus, the Groucho repressor undergoes displacement allowing β-catenin to interact with T-cell factor/lymphoid enhancer factor (*TCF/LEF*), chromatin remodeling and transcription of genes such as *c-myc* and *cyclin D1*.

The activation of the canonical Wnt signal requires both the FZD family receptors and the low-density-lipoprotein-related protein 5/6 (LRP5/LRP6) co-receptors which phosphorylation is essential for receptor activation. Wnt binding to its receptor results in dishevelled (DVL) phosphorylation, leading to Axin de-phosphorylation and decline of its cytoplasmic content [7]. Thereby, β -catenin can be released from the "destruction complex", and its degradation prevented while stabilization allowed. Accumulation of β -catenin turns into its nuclear translocation [7].

Although several nuclear β -catenin binding partners have been involved in the control of gene transcription, the most relevant β -catenin partners are the members of the T-cell factor/lymphoid enhancer factor (TCF/LEF) family of transcription factors [7]. This complex binds to the promoter region of target genes and regulates their transcription.

Once in the nucleus, the engagement of β -catenin transiently converts the *TCF/LEF* into transcriptional activators which displace Groucho and induces chromatin remodelling and transcriptional activity (Figure 1b).

A number of genes are targeted by Wnt- β -catenin. Among them, genes involved in positive- and negative-feedback regulation, cell-cycle progression, and stem cell homeostasis are the most commonly included genes.

Wnt non-canonical pathways: Wnt/planar cell polarity (PCP) and Wnt/Calcium

To date, the canonical Wnt/ β -catenin pathway is much better characterized than the non-canonical one (Figure 2).

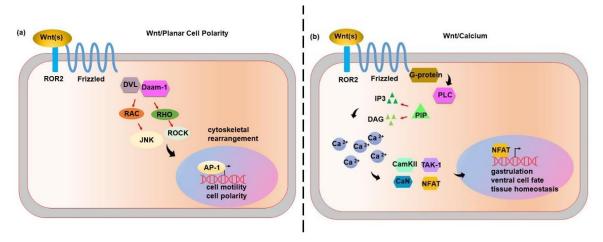


Figure 2. The Wnt non-canonical signalling pathways. (a) Wnt/planar cell polarity (PCP) pathway. Wnt ligands bind to *FZD* receptors and co-receptor RAR-related orphan receptor (*ROR*) and convey the signal to *DVL*. *DVL* forms the Disheveled associated activator of morphogenesis 1 (*DVL-Daam-1*) complex, which triggers *RhoA*, *RHO* and *ROCK* to control cytoskeletal rearrangement. On the other hand, *DVL* triggers *RAC*, *JNK* and *AP-1* involved in cell motility and polarity. (b) Wnt/Calcium pathway. Wnt ligands bind to *FZD* and activate the phospholipase C (*PLC*), which hydrolyses the phosphatidylinositol (4,5)-biphosphates (*PIP2*) to inositol (1,4,5)-triphosphates (*IP3*) and diacylglycerol (*DAG*). This translates into intracellular calcium release and the activation of *CaN* and *CamKII*. The calmodulin activation stimulates *TAK-1* and *NLK* activity. *CaN* activates the *NFAT*, which moves to the nucleus and modulates the expression of genes involved in the control of gastrulation, ventral cell fate and tissue homeostasis.

In the non-canonical PCP pathway, Wnt ligands bind to *FZD* receptors and co-receptor protein tyrosine kinase 7 (*PTK7*), RAR-related orphan receptor (*ROR*) or the receptor like tyrosine kinase (*RYK*) and convey the signal to DVL. On the one side, *DVL* forms the disheveled associated activator of morphogenesis 1 (*DVL-Daam-1*) complex, which triggers a small guanosine-5'-triphosphate (GTP) GTPase, such as ras homolog gene family member A (*RhoA*), RHO and RHO-associated kinase (*ROCK*). *DVL* also triggers ras-related C3 botulinum toxin substrate (*RAC*), JUN-N-terminal kinase (*JNK*) and the activator protein-1 (*AP-1*).[7] The PCP pathway is involved in the cytoskeletal rearrangement, cell motility and co-ordinates cell polarity. In vertebrates, the PCP pathway is also required for morphology and migration of dorsal mesodermal cells undergoing gastrulation, hair follicle organization, and orientation of stereocilia in the sensory epithelium of the inner ear [8] (Figure 2a).

In the calcium-dependent pathway Wnt ligands bind to FZD and activate the phospholipase C (PLC) which hydrolyses the phosphatidylinositol (4,5)-biphosphates (PIP2) to inositol (1,4,5)-triphosphates (IP3) and diacylglycerol (DAG). This translates into the release of the intracellular calcium and the activation of both calcineurin (CaN) and calcium/calmodulin-dependent kinase II (CamKII). Moreover, the activation of calmodulin promotes the activation of the TGF- β -Activated kinase 1 (TAK-1) and nemo-like kinase (NLK), thereby antagonizing and neutralizing the canonical Wnt/ β -catenin cascade. CaN activates the nuclear factor of activated T-cells (NFAT), which moves to

the nucleus and regulates the expression of target genes [7] (Figure 2b). The calcium-dependent pathway plays a crucial role in several processes, including early pattern formation during gastrulation [2], ventral cell fate [9], dorsal axis formation [10], and tissue homeostasis [11].

COLORECTAL CANCER

Colorectal cancer (CRC) is one of most common cancer worldwide and represents a deep cause of cancer mortality [12] with a rapid increase in incidence and death rate [13]. Dienstmann et al. [14] established a new classification of CRCs into four consensus molecular subtypes (*CMSs*). Among them *CMS2*, *CMS3*, and *CMS4* have a higher rate of *APC* mutations (over 50%) compared to *CMS1*. Each *CMS* has unique features: *CMS1* (MSI Immune, 14%): hyper- mutated, microsatellite instability, strong immune activation; *CMS2* (Canonical, 37%): epithelial, chromosomally unstable, marked Wnt and myc signalling activation; *CMS3* (Metabolic, 13%): epithelial, metabolic dysregulation; and *CMS4* (Mesenchymal, 23%): a prominent transforming growth factor β (*TGF\beta*) activation, stromal invasion, and angiogenesis. Samples with combined features (13%) represent transition phenotypes or are supposed to reflect the intra-tumour heterogeneity [14].

The heterogeneous genetic ground underlying CRC initiation and progression mainly involves gene fusion, deletion or amplification, somatic gene mutations and epigenetic alterations. Wnt/ β -catenin signalling has emerged as one of the most significant biological pathways in both physiological setting and CRC development. Almost all CRC are characterized by a hyper-active Wnt/ β -catenin pathway, which, in many cases, is considered the most critical cancer initiating and driving event. Proteins and miRNAs guiding the Wnt/ β -catenin pathway and proposed as potential CRC therapeutic targets are discussed.

Canonical Wnt/β-catenin pathway and CRC

Ring finger protein 6 (*RNF6*) is an oncogene frequently upregulated by gene amplification in primary CRC. Moreover, *APC* mutation and *RNF6* copy number amplification were commonly found in CRC patients. *RNF6* is a RING-domain E3 ubiquitin ligase and exerts its pro-metastatic effects by promoting CRC cell growth, cell-cycle progression, and epithelial to mesenchymal transition (EMT). Furthermore, *RNF6* expression and its gene amplification have been considered independent patients' prognostic factors. *RNF6* mediates the polyubiquitination of the transducin-like enhancer of split 3 (*TLE3*), a transcriptional repressor of the β -catenin/*TCF4* complex, and its proteasome degradation. The lack of *TLE3/TCF4/LEF* interaction enhances the Wnt/ β -catenin transcriptional activity, and the expression of its downstream target genes [15] (Table 1).

Table 1. Proteins/EVs involved in several tumours, their alteration, targets, and impact on tumours.

PROTEIN	RELATED	EXPRESSION	PATHWAY	IMPACT ON TUMOUR	REF.
	CANCERs	LEVEL	INTERACTION		
RNF6	CRC	Upregulated	β-catenin	cell growth	[15]
				cell cycle progression	
				EMT	
				metastasis	
LGR5	CRC, BC	Upregulated	β-catenin	proliferation	[16, 65 68]
		. 0	,	migration	
TNIK	Gastric	<mark>Upregulated</mark>	<mark>β-catenin</mark>	cell growth	[19, 2 0]
KYA1797K	CRC	Upregulated	β-catenin	tumour growth	[19<mark>22</mark>]
			,	stem cell features	
BCL6	CRC	Upregulated	β-catenin	cellular proliferation	[22-24] [25-
			-	tumour development	

				tumour progression	
ZEB2 and	CRC	Upregulated	β-catenin	tumour progression	[27,28] <mark>[30,3</mark>
ZEB1				invasion	1] [64,65]
					[67,68]
XIAP	CRC	Upregulated	β-catenin	proliferation	[27,28]
			·	chemoresistance	[30,31]
RHBDD1	CRC	Upregulated	β-catenin	metastasis	[31][<mark>34</mark>]
			'	stemness	2 31
				EMT	
				migration	
				invasiveness	
SLC35C1	CRC	Downregulated	β-catenin	cell proliferation	[32][<mark>35</mark>]
				cell progression	
NPTX2	CRC	Upregulated	β-catenin	tumour stages	[33][<mark>36</mark>]
				lymphatic invasion	
				metastasis	
KLHL22	CRC	Downregulated	β-catenin	invasion	[37][<mark>40</mark>]
				migration	
CCL2	CRC	Upregulated	Non-canonical	progression	[45][<mark>48</mark>]
LGR4	ВС	Upregulated	β-catenin	tumorigenesis	[63-66]
			·	metastasis	[<mark>65-68</mark>]
				CSC maintenance	,
ST7L	ВС	Downregulated	GSK-3β	proliferation	[67][<mark>69</mark>]
				invasion	
TMED	ВС	Upregulated	β-catenin	cell cycle progression	[68][<mark>70</mark>]
				colony formation	
				migration	
Wnt5a	ВС	Downregulated	β-catenin	migration	[73,74]
				lactate production	[<mark>75,76]</mark>
				invasion	
Wnt5a	ВС	Downregulated	β-catenin	cell proliferation	[76-78]
			-cyclin D1	aggressiveness	[<mark>79-80</mark>]
			-TGF-β		
Wnt5a	ВС	Upregulated	ALCAM	vessel invasion	[79][<mark>81</mark>]
				tumour size	
				migration	
Nek2B	TNBC	Upregulated	β-catenin	chemoresistance	[89][<mark>91</mark>]
VANGL2	TNBC	Upregulated	p62/SQSTM1 (PCP)	migration	[90][<mark>92</mark>]
				anchorage-dependent	
				and independent cell	
				proliferation	
HePTP	TNBC	Upregulated	-GSK-3β	metastasis	[91][<mark>93</mark>]

			β-catenin		
DLC-3	TNBC	Downregulated	β-catenin	proliferation colony formation migration invasion	[92][<mark>94</mark>]
DKK1	OC	Downregulated	β-catenin	stemness	[100][<mark>102</mark>]
SFRP1	OC	Downregulated	β-catenin	cell growth stem-like phenotype	[101][<mark>103</mark>]
AXIN2	OC	Downregulated	β-catenin	stem-like phenotype	[101][<mark>103</mark>]
LGR6	OC	Upregulated	β-catenin	stemness chemoresistance	[105,106] [<mark>107,108</mark>]
RAB14	OC	Upregulated	β-catenin	proliferation chemoresistance invasion	[97,107] [<mark>109,110</mark>]
FZD7	OC	Upregulated	Non-canonical	EMT cell cycle progression migration	[109][<mark>112</mark>]
ITGBL1	OC	Upregulated	Non-canonical	migration adhesion	[110][<mark>113</mark>]
ALPL	OC	Upregulated	Non-canonical	EMT migration invasion	[115][<mark>118</mark>]
VDR	Melanoma	Upregulated	β-catenin	tumour growth immune response	[119][<mark>122</mark>]
EVs	CRC	Upregulated	β catenin	— migration, — metastasis — tumour growth	[123]
14 3 3ζ in EVs	HEK293T, COS 7, SW480, HeLa, Huh7, HEK293 EBNA- PurR and L Wnt3a cells	Upregulated	— β catenin — GSK 3β — DVL2	— survival — migration	[124 126]
EVs	CRC	Upregulated	β catenin	dedifferentiationdrug resistancecolony formation	[127,128]
EVs	HCC	Silenced	β catenin	— proliferation — tumour growth	[132,133]

DKK 1 in EVs	MM	Upregulated	β catenin	osteoclast activity	[134]
			-	osteoblast	
				differentiation	
EVs	OSCC	Upregulated	β catenin	metastasis	[136]
			·	stemness	
				— cell reprogramming	
				chemoresistance	
Wnt5b in EVs	PANC 1	Upregulated	β catenin	proliferation	[138]
	Caco-2 cell		dependent and	- migration	
	lines		independent		
EVs	BC	Upregulated	Wnt PCP	- cell growth and	[139]
				motility	_
149 The	e leucine-rich r	epeat-containing G-	protein coupled recept	or 5 ($LGR5$) is a Wnt/ β -catenin t	arget

The leucine-rich repeat-containing G-protein coupled receptor 5 (*LGR5*) is a Wnt/ β -catenin target gene implicated in cancer cell proliferation and migration. It has been reported that LGR5 is highly expressed in CRC tissues compared to the healthy ones. A decline in β -catenin and *c-myc* mRNA expression were detected by knocking-down LGR5 expression, suggesting that it may regulate the Wnt/ β -catenin activity by modulating the expression of β -catenin. Furthermore, since targeting *LGR5* improves the response to chemotherapy, LGR5 has been proposed as a novel therapeutic target in CRC [16] (Table 1).

The β-catenin and RAS signalling pathways are frequently associated to the development and progression of several different cancers. They mainly act on cancer stem cell (CSC) expansion. High levels of β-catenin and RAS proteins are considered the major drivers of CSC expansion and cancer dissemination and are associated with poor patient's outcome [17].

Targeting the CSC pool without affecting the somatic stem cell (SSC) niche is one of the major goals of the last decades. As reported by Lenz et al. [18], the β-catenin antagonist molecule, ICG-001, effectively prevented the interplay between β-catenin and its coactivator cAMP response element binding protein (CREB)-binding protein (CBP). Moreover, ICG-001 effectively and without side effects abrogated drug-resistant cells. On the same line, PRI-724, a second generation of CBP/βcatenin antagonist, was found safe in pre-clinical studies and displayed an acceptable toxicity profile.

Yu et al. [19] investigated the traf2- and nck-interacting kinase (TNIK) amplification and its role in tumor progression by applying siRNA technology, while Masuda et al. [20] have generated a small molecule denoted as NCB-0846 acting as TNIK inhibitor. TNIK selectively binds both to TCF4 and β catenin in order to promote cancer cell growth via Wnt/β-catenin cascade and drives colorectal CSC expansion. The NCB-0846 inhibitor was effective in interfering with *TNIK* activity tumour growth.

KYA1797K, a small molecule identified by Cha et al. [21], was found effective in suppressing CRC growth due to the activation of $GSK-3\beta$ via Axin binding and β -catenin/RAS destabilization. In line with this observation, treatment with KYA1797K abrogated CRC stem cell features both in vitro and in vivo. Mechanistically, KYA1797K pushes β-catenin and RAS towards the Axin binding [22] (Table 1).

In the last decade miRNAs have gained particular attention in cancer [23]. miRNA profiling has been linked to cancer types, stage, and invasion [24]. Moreover, oncogenic or tumour suppressive actions have been linked to miRNA expression. For these reasons, miRNAs are considered valuable tools for cancer diagnosis and prognosis and therefore useful therapeutic targets (Table 2).

Table 2. miRNAs involved in the tumours, their alteration and tumour impact.

miRNA	RELATED	EXPRESSION	IMPACT ON	REF.
	CANCER	LEVEL	TUMOUR	
miR-144-3p	CRC	Downregulated	cell proliferation	[22][<mark>25-27</mark>]

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miR-377-3p	CRC	Upregulated	cell expansion	[25][<mark>28</mark>]
			EMT	
			repression of	
			apoptosis	
miR-377-3p	CRC	Downregulated	proliferation	[26][<mark>29</mark>]
			migration	
			chemoresistance	
miR-520e	CRC	Downregulated	cell proliferation	[29,30][<mark>32,33</mark>]
			colony formation	
			invasion	
miR106a	ВС	Upregulated	cell growth	[58][<mark>61</mark>]
			cisplatin sensitivity	
miR-5188	ВС	Upregulated	tumour cell	[60][<mark>63</mark>]
			proliferation	
			metastasis	
			formation	
			EMT	
			chemoresistance	
miR-148a	ВС	Downregulated	cell migration	[61][<mark>64</mark>]
			invasion	
miR-6838-	ВС	Downregulated	cell invasion	[92][<mark>95</mark>]
5p			migration	
			EMT	
miR-27a-3p	ВС	Upregulated	proliferation	[93][<mark>96</mark>]
			migration.	
miR-1207	OC	Upregulated	tumorigenicity	[100][<mark>103</mark>]
			stem cell-like traits	
			stemness	
miR-590-3p	OC	Upregulated	cell growth	[101,102][<mark>104,105</mark>]
			migration,	
			invasion	
miR-1180	OC	Upregulated	cell proliferation	[103][<mark>106</mark>]
			glycolysis	
miR-939	PCa	Downregulated	tumour stage	[116][<mark>120</mark>]
			metastasis	
miR-92a-3p	CRC EVs	Upregulated	cancer progression	[129]
			stemness	
			EMT	
			drug resistance	
miR-1273f	HCC EVs	Upregulation	cell proliferation	[134]
			migration	
			invasiveness	
			EMT	

miR-1260b	LAC EVs	Upregulation	cell invasion	[137][<mark>136</mark>]
			metastasis	
miR-214-3p	TEC EVs	Upregulation	neovessel formation	[139][<mark>139</mark>]
miR-24-3p	TEC EVs	Downregulation	neovessel formation	[139][<mark>139</mark>]

Sun and co-workers [25] identified miR-144-3p as a new biomarker for CRC diagnosis and response to treatment. miR-144-3p was found downregulated and associated with CRC pathological stages in CRC patients. Interestingly, miR-144-3p overexpression reduced CRC cell proliferation by delaying G1/S phase transition in tumour cells. On the contrary, the B-cell lymphoma 6 protein (*BCL6*), a nuclear protein belonging to the BTB/POZ/zinc finger (*ZF*) family of transcription factors, was found upregulated and surprisingly post-transcriptionally regulated by miR-144-3p. Previous studies revealed that *BCL6* is involved in the control of cell cycle progression and differentiation [26,27]. Indeed, miR-144-3p/*BCL6* co-operate to inhibit cellular proliferation, development, and progression of CRC by interfering with *c-myc* and *cyclin D1* expression [25] (Table 1).

miR-377-3p displays an ambiguous role in CRC. Liu and colleagues [28] uncovered that upregulation of miR-377-3p promotes G1-S phase transition, cell expansion and EMT, while represses apoptosis in CRC patients. Moreover, GSK-3 β , a direct miR-377-3p target, was found upregulated upon miR-377-3p overexpression. These data suggest that a complex regulatory network boosting tumour progression is associated with the expression of miR-377-3p in CRC.

Conversely, in a recent study, Huang et al. [29] have shown that miR-377-3p, significantly reduced in CRC patients, is involved in the control of proliferation, migration and chemo resistance, particularly at advanced tumour stage. The authors investigated miR-377 functions and mechanism of action in CRC cells. The zinc finger E-box binding homeobox 2 (ZEB2) and the X-linked inhibitor of apoptosis protein (XIAP) are two positive regulators of the Wnt/ β -catenin cascade [30,31]. In CRC, ZEB2 enables tumour progression and invasion, whereas XIAP promotes cell proliferation and chemoresistance. De facto, miR-377-3p overexpression was found to suppress the malignant CRC phenotype, as well as cell proliferation, invasion and drug resistance by directly targeting the 3' UTR sequence of both ZEB2 and XIAP mRNAs. Since miR-377-3p/ZEB2-XIAP inhibited CRC progression by reducing Wnt/ β -catenin-associated gene expression (e.i. *cyclin D1*, Axin2, TCF1, SOX2, c-myc, matrix metalloproteinase-2 (MMP-2), MMP-9, CD44, vascular endothelial growth factor (VEGF), and Twist) approaches increasing its expression have been proposed for novel therapeutic options (Table 1).

Functional experiments showed that miR-520e plays a pivotal role in regulating CRC cell proliferation, colony formation and invasion [32]. Moreover, it has been reported that low miR-520e expression is associated with the increased CRC growth and migration. The astrocyte elevated gene-1 (AEG-1), which acts as an oncogene [33], is a direct miR-520e target in CRC. Cells overexpressing miR-520e displayed lower $GSK-3\beta$ phosphorylation and β -catenin expression. Mechanistically, it was found that miR-520e regulates cancer cell behaviour by targeting AEG-1 which on turn inactivate the Wnt/ β -catenin signalling and the transcription of its downstream genes. Hence, miR-520e overexpression could represent a promising therapeutic target in CRC by AEG-1 suppression.

Approximately 40–50% of CRC patients develop metastasis, mostly to the liver and lung. In cancer patients, metastases are associated with 90% of all cancer-related death, thereby the mechanisms accounting for the metastatic spread have been deeply investigated. Zhang et al. [34] demonstrated that the rhomboid domain containing 1 (RHBDD1) plays a crucial role in driving metastasis formation in CRC patients, via the Wnt/β-catenin pathway. It has been shown that RHBDD1 is able to influence the Wnt/β-catenin cascade by increasing the phosphorylation of β-catenin at the Ser552 and Ser675 residue without affecting its nuclear translocation. Moreover, it promotes EMT, stemness, migration and invasiveness. RHBDD1 also improves the expression of the β-catenin target gene, ZEB1. Furthermore, the protein level of RHBDD1 positively correlated with ZEB1. Thereby, RHBDD1 has been proposed as a novel therapeutic target and/or a clinically useful biomarker for metastatic CRC (Table 1).

SLC35C1, or GDP-fucose transporter 1, is a member of the solute carrier (SLC) superfamily of solute carriers. The Deng's group [35] explored the mechanism throughout SLC35C1 regulates the canonical Wnt/β-catenin pathway in CRC. They demonstrated a reduction of SLC35C1 and an increase of β-catenin at all tumour stages. Indeed, silencing SLC35C1 resulted in the increased release of Wnt3a and c-myc, Axin2 and cyclin-D1 expression. This suggests that SLC35C1 is involved in the control of the canonical Wnt/β-catenin pathway, and thereby in tumour cell proliferation and tumour progression (Table 1).

Neuronal pentraxin 2 (*NPTX2*) is a member of the neuronal pentraxin family and is essential for the formation of synapsis. *NPTX2* was found overexpressed at both mRNA and protein level in CRC, particularly in metastatic lesions [36]. *NPTX2*, which was found to positively correlate with tumour stages, lymphatic invasion, distant metastasis, and poor patients' outcome, promotes β -catenin nuclear translocation and the expression of *c-myc*, *cyclin D1*, *Snail*, and *N-cadherin*. No *NPTX2* receptors have been identified in CRC, however, its cellular internalization was found mediated by the Wnt/ β -catenin receptor, *FZD6*. Additionally, it has been reported that *NPTX2/FZD6* interaction translates in cancer cell proliferation and metastasis formation by triggering the Wnt/ β -catenin pathway [36] (Table 1).

Aberrant gene expression and DNA methylation profiles are considered hallmarks of CRC initiation and progression [37]. Due to the APC inactivating mutations, the Wnt/ β -catenin pathway plays a key role in CRC metastatic spread [35][38]. Bruschi el al. [39] investigated the early transcriptional and epigenetic changes resulting from APC inactivation in intestinal crypts in crypt base columnar (CBC) cells. The authors have found that APC disruption rapidly induces changes in DNA methylation, indicating that focal remodelling of the DNA methylation profile occurs early and concomitantly with the first oncogenic event. Moreover, it has been demonstrated that the hyperactivation of the Wnt/ β -catenin pathway associated with the APC loss-of-function turns out in a rapid increase of intestinal stem cell commitment towards differentiation. Again, it was correlated with the remodelling of the DNA methylation profile. This study unveils that early changes in DNA methylation are crucial for the impaired fate decision program associated with APC loss-of-function.

The kelch-like family member 22 (KLHL22) is a tumour suppressor protein involved in the development/progression of several cancers [40]. Low expression of KLHL22 was found in CRC tissues. KLHL22 overexpression was associated with decreased migration, invasion and reduced expression of the EMT markers, vimentin, N-cadherin, Twist1 and Snail1. Intriguingly, KLHL22 knockdown led to β -catenin and LEF increased expression, while KLHL22 overexpression translates into $GSK-3\beta$ upregulation and β -catenin downregulation [40] (Table 1).

Non-canonical Wnt pathway and CRC

The canonical and non-canonical Wnt family members play discrete roles in CRC. The activation of the Wnt/calcium pathway turns into stimulation of sensitive proteins such as *CamKII* and *PKC* [38][41]. A Ror family of receptor tyrosine kinases, the *ROR*2 has been shown to act as a Wnt5a receptor or co-receptor [42]. Wnt5a has different roles in CRC. It can act as antagonist or agonist of the canonical Wnt/ β -catenin pathway, depending on the cellular context. Lee et al. [43] noticed that the antagonism between the canonical and the non-canonical Wnt/ β -catenin signalling pathways is linked to Wnt5a. Mechanistically, Wnt5a suppressed the canonical Wnt/ β -catenin cascade by acting as ligand on the *ROR* α [42]. After *PKC* α -mediated phosphorylation, ROR α modifies its affinity and interacts with the armadillo repeat domains of β -catenin, thus supressing its transcriptional activity.

Three relevant goals have been recently achieved by Voloshanenko et al. [44] supporting the role of Wnt5a/b in cell growth, via the non-canonical β -catenin pathway. First, they identified the procollagen-lysine,2-oxoglutarate 5-dioxygenase 2 (*PLOD2*), the hydroxyacyl-CoA dehydrogenase (*HADH*), ligand-dependent corepressor (*LCOR*) and the receptor expression-enhancing protein 1 (*REEP1*) as candidate genes regulated by the non-canonical Wnt/ β -catenin pathway. Second, these genes were found regulated by Wnt5a/b, as well as by *ROR2*, the *DVL2*, the activating transcription factor 2 (*ATF2*) and *ATF4* in a non-canonical Wnt/ β -catenin independent manner. Lastly, Wnt5a/b silencing was found to impair cancer cell proliferation.

Among several soluble Wnt proteins, Wnt11 was found upregulated in CRC patients [45]. Recently, Gorroño-Etxebarria and colleagues [46] have shown that increased Wnt11 and its FZD6, RYK, PTK7 receptors, positively correlate with poor prognosis. Additionally, Wnt11 downregulated β -catenin transcriptional activity and increased ATF2 via the non-canonical Wnt signalling pathway. Thereby, Wnt11 has been proposed as a prognostic biomarker and therapeutic target in CRC patients.

Tumour micro environment (TME) has a pivotal role in cancer development [47]. Liu et al.[48] reported that, unlike CRC cells, tumour associate macrophages (TAMs), and in particular M2-like cells express Wnt5a. Furthermore, it has been shown that Wnt5a positive TAMs regulate macrophages infiltration, tumour cell proliferation and migration. Wnt5a pro-tumour activity was found associated with the overexpression of the C-C motif chemokine ligand 2 (*CCL2*) in Wnt5a-treated macrophages. Consistently, Wnt5a knockdown reduced *CCL2* expression in TAMs and their cancer-promoting activity. In Wnt5a-treated macrophages both *CaMKII* and ERK1/2 undergo phosphorylation and lead to *CCL2* secretion. This study provided evidence for a new role of Wnt5a in CRC and describes a potential novel therapeutic target (Table 1).

BREAST CANCER

Breast cancer (BC) is the most diagnosed cancer in women [46][49], the first cause of cancer death in women worldwide [47][50], and one of the most expensive health care cost [46][49]. Both the canonical and non-canonical Wnt/β-catenin pathways are essential for mammary gland development [51] and for BC growth and dissemination [52]. Hyper-active Wnt/β-catenin was reported in breast tumours [50][53]. In human BC, elevated intracellular β-catenin level has been associated with high tumour grade [54] and poor prognosis. Moreover, up to 90% of metaplastic carcinomas and non-metastasizing fibromatosis have been associated with the highest β-catenin expression level [55]. Moreover, proteins such as Wnt3a [56] and xenopus frizzled 7 (*Xfz7*) [57] have been involved in the activation of both the canonical and the non-canonical Wnt signalling pathways.

Canonical Wnt pathway and BC

Dysregulation of the Wnt/β-catenin cascade has been associated with cancer initiation and metastasis formation [56][58]. Moreover, high β-catenin expression has been reported in basal-like BC subtype [50][53]. Additionally, it has been demonstrated that loss of secreted frizzled-related protein 1 (sFRP1) is an early event in BC patients and is associated with poor prognosis [59]. Furthermore, the activation of the Wnt/β-catenin cascade has been associated with radio resistance of progenitor cells. Thereby, the Wnt/β-catenin pathway has been proposed as a target to harm the self-renewal potential of stem/progenitors [60].

A recent study demonstrated that high β -catenin level is associated with miR106a overexpression and involved in BC cell growth. Additionally, high level of miR106a was reported to reduce cisplatin sensitivity. Major results were obtained exploiting the Wnt inhibitor, FH535. In fact, FH535 treatment reduced the expression of β -catenin, *cyclin D1*, *c-myc* and *Ki67*, impaired tumour growth and induced apoptosis [61].

In a different study [62], the impact of the Wnt/ β -catenin canonical pathway in cisplatin resistance was investigated by silencing β -catenin via small interfering RNA (siRNA). The authors demonstrated that upon β -catenin silencing, the cells become more sensitive to cisplatin treatment. These effects were associated with the increased expression of the apoptotic proteins caspase 3/9.

A recent study demonstrated that miR-5188, aberrantly expressed in breast cancer patients, positively correlates with poor prognosis. The molecular analyses revealed that miR-5188 directly targets the forkhead box protein O1 (FOXO1). In physiological setting, FOXO1 binds β -catenin and induces its degradation. This implies that miR-5188 overexpression leads to β -catenin nuclear accumulation and transcription of its downstream target genes, mainly involved in EMT, tumour cell proliferation, metastasis formation and chemo resistance. Moreover, the authors elegantly showed that miR-5188 expression is under the control of c-Jun, which directly binds to its promoter region. This on turn generates a positive loop accelerating tumour progression. Clinically, miR-5188 has been proposed as a diagnostic or prognostic factor and/or a direct target for anti-cancer therapy [63].

The upregulation of the lncRNA hoxa transcript at the distal tip (*HOTTIP*) has been also linked to poor prognosis in BC patients. Overexpression of *HOTTIP* correlates with the expansion of breast CSCs (BCSCs) and the expression of the stem cell markers, *OCT4* and *SOX2*. Han et al. [64] demonstrated a reduced expression of differentiation markers, such as *CK18* and *CK14* and that miR-148a inhibits BC cell migration and invasion by directly targeting Wnt1. Moreover, it has been reported that *HOTTIP* controls miR-148a-3p by acting as a competing endogenous RNA (ceRNA). Thereby, *HOTTIP* promotes expansion of CSCs *in vitro* and tumorigenesis *in vivo* by regulating the miR-148a-3p/Wnt1/β-catenin axis [64]. These data are summarized in Table 2.

The *LGR4* was identified as a prognostic marker in breast tumours displaying poor prognosis [65]. A tight molecular interplay between *LGR4* and Wnt/β-catenin signalling has been reported to control stemness. Indeed, *LGR4* binding to the soluble R-spondin proteins eases the Wnt/β-catenin cascade [64][66]. Previous studies have proven that upregulation of *ZEB1* by *SLUG* (the protein product of *SNAI2*), increased EMT [67]. As a matter of fact, *LGR4* knockdown leads to *SLUG* and *ZEB1* downregulation, thereby impairs invasion and metastasis [68]. A correlation with poor outcome and the expression of the *LGR4* homolog *LGR5* was also reported. *LGR5* maintains the pool of BCSCs and promotes tumour progression and invasiveness by activating the Wnt/β-catenin canonical pathway [68] (Table 1).

Wang et al. [69] first demonstrated that the expression of the suppression of tumorigenicity 7 like (ST7L) is downregulated in BC cells, and more importantly, that ST7L acts as an antitumor supervisor by reducing $GSK-3\beta$ phosphorylation and inducing β -catenin degradation. However, the mechanisms through which ST7L controls $GSK-3\beta$ phosphorylation is still missing (Table 1).

A recent study [70] reported the overexpression of the transmembrane emp24 domain (TMED) in BC and its correlation to poor prognosis. Aberrant level of TMED boosts cell cycle progression, colony formation, migration and invasion and the expression of CDK2, CDK4, CDK6, cyclin E, β -catenin, cyclin D1, c-myc, MMP-7 and TCF4. Conversely, silencing TMED3 drastically reduced migration and invasion. Moreover, the observation that β -catenin knockdown translates in the reduction of its regulated genes supports the notion that the oncogenic effect of TMED goes through the Wnt/ β -catenin pathway (Table 1).

Cryptotanshinone (CTS) is an herbal medicine derived from roots of salvia miltiorrhiza which displays anti-tumour properties. It has been shown that *in vitro* CTS reduces tumour cell growth, migration and invasion by downregulating the pyruvate kinase muscle isozyme M2 (PKM2), a protein involved in glycolysis, and more importantly in β -catenin activation [71].

Wnt non-canonical pathway and BC

Among the Wnt ligands, the most extensively studied ligand, activating the β -catenin independent pathway, is Wnt5a. However, its different biological actions are enlightened by the observation that it can also initiate the canonical β -catenin signalling cascade [70][72].

Wnt5a is an evolutionarily conserved Wnt ligand, which plays an important role in developmental processes. Wnt5a^{-/-} knockout mice showed perinatal lethality, due to developmental defects [73].

In tumorigenesis, Wnt5a signalling is central and displays multiple intriguing and opposite roles mainly acting as a β -catenin antagonist. These data are discussed.

The Wnt5a suppressive properties detected in tumours connoted by β -catenin hyper-activation has been linked to the shift towards the stimulation of the β -catenin independent signalling pathway.

Foxy5 is a Wnt5a mimicking hexapeptide able to decrease BC cell migration and invasion [74]. More recently Prasad et al. [75] confirmed these data and added new information on the role of Wnt5a in the regulation of the expression of the phosphofructokinase platelet-type (PFKP). They have shown that low PFKP level correlates to cancer cell migration and poor patients' survival. The growth and expansion of tumour cells also rely on glucose consumption resulting in the accumulation of lactate. Cancer cell metabolism was also associated with β -catenin activation [76]. At this regard, it has been shown that Wnt5a affects the aerobic glycolysis by inhibiting the activation of β -catenin. Therefore, an onco-suppressive role was proposed for PFKP.

According to the study of Borcherding et al. [77], Serra Roarty et al. [76][78] demonstrated that the paracrine activity of Wnt5a suppresses the expression of both β -catenin and *cyclin D1*. The authors have shown that Wnt5a supports TGF- β -mediated tumour suppressive functions by antagonising Wnt/ β -catenin signalling and limiting tumour cell proliferation.

Moreover, Leris and colleagues [79] proved that Wnt5a mRNA level was significantly lower in tumour than in normal tissues, particularly in those displaying a more aggressive behaviour. Again, this observation has suggested a suppressive role of Wnt5a in cancers. It has been also reported that loss of Wnt5a associates with a higher histological tumour grade, increased risk of recurrence, and a shorter recurrence-free survival in invasive BC [80] (Table 1).

On the contrary, Kobayashi et al. [81] reported that Wnt5a is expressed in ER-positive BC cells and positively associates to vessel invasion, tumour size and migration. Mechanistically, Wnt5a induces the expression of the activated leukocyte cell adhesion molecule (*ALCAM*), a protein involved in migration and invasion. Knockdown of either Wnt5a or *ALCAM* inhibited tumour cell migration, confirming the role of the Wnt5a/*ALCAM* axis in the migratory phenotype of ER-positive BC (Table 1).

A relevant role of Wnt5a in reprogramming the TME was also described [82]. It has been shown that under pro-inflammatory conditions the non-canonical Wnt protein induces the expansion of the CD163(+) immunosuppressive macrophages translating in the release of IL-10 and the inhibition of the classical *TLR4*-NF-kB signalling pathway [82].

Moreover, higher level of Wnt5a was found in human monocyte-derived myeloid dendritic cells (Mo-mDCs) than in normal monocytes and macrophages. Wnt5a was found to inhibit the generation of Mo-mDCs by stimulating BC cells to produce IL-6. In addition, the presence of IL-6 in the conditioned media of Wnt5a stimulated BC cells was found involved in the inhibition of Mo-mDC differentiation [83]. Consistently, overexpression of Wnt5a mRNA was detected in metastases derived from primary BC cells and in BC cell lines [84].

Wnt5a signalling is also able to modify the CD44-AKT signalling pathway, leading to a reduced BC cell migration and invasion. In epithelial BC cells, silencing of Wnt5a drives EMT-like changes without altering the expression of common EMT markers. On the contrary, it interferes with CD44 expression and induces pAKT downregulation, thereby acting via a EMT-independent mechanism [85].

The dual activity of Wnt5a has been also ascribed to the Wnt5a isoforms. Bauer et al. [86] have shown that the Wnt5a gene encodes for two distinct isoforms: the Wnt5a-long (*Wnt5a-L*) and Wnt5a-short (*Wnt5a-S*) isoform. When analysed in several cell lines *Wnt5a-L* reduced tumour progression, while *Wnt5a-S* promoted tumour growth.

Overall, Wnt5a may play multiple roles. Whether it acts as a tumour suppressor or a tumour promoter remains elusive and depends on the availability of essential receptors, the TME, and the activation of discrete signalling pathways.

TRIPLE-NEGATIVE BREAST CANCER

Triple-Negative Breast Cancer (TNBC) is an invasive type of breast carcinoma that lacks the expression of estrogen and progesteron receptor as well of the human epidermal growth factor receptor 2 (HER2) [87] and accounts from 10 to 15% of all BC [88].

TNBC patients have poor outcome due to the high grade of proliferation, early tumour dissemination, and the lack of targeting approaches [89,90]. The malignancy is associated with earlier age of onset, aggressive clinical course, and dismal prognosis [88]. TNBC gained attention due to the aggressiveness and the lack of effective treatment options. Therefore, the most relevant data on this breast cancer subtype are independently discussed.

Gene expression omnibus (GEO) databases were applied by Shen et al. [91] to gather gene expression data in TNBC patients who underwent chemotherapy. They reported that co-expression of NIMA-related kinase 2 (Nek2) and β -catenin correlated with patients' poor prognosis. β -catenin binds to and is phosphorylates by the Nek2B isomer. Thereby, in TNBC, Nek2B functions as a β -catenin regulator by activating the Wnt signalling pathway and its downstream target genes. In

addition, it has been suggested that Nek2B and β -catenin may synergize to promote resistance to chemotherapy. However, further studies are required to better elucidate the relationship between β -catenin and Nek2 and its possible implications in cancer development (Table 1).

TNBC aggressiveness also relies on the activation of the non-canonical Wnt/PCP pathway. Indeed, the aberrant activation of downstream genes activated by the non-canonical Wnt/PCP pathway has been implicated in tumour growth and poor prognosis. Results from Puvirajesinghe and colleagues [92] revealed that van gogh-like 2 (*VANGL2*), a core Wnt/PCP component, plays a crucial role in cancer cell migration, anchorage-dependent and independent cell proliferation, as well as in tumour growth. Since, the scaffold p62/SQSTM1 protein, a *VANGL2*-binding partner, has a key role in *VANGL2*-p62/SQSTM1–*JNK* pathway, the possibility to exploit p62/SQSTM1 as a potential therapeutic target has been proposed. This would be of particular relevance since the *JNK* targeting approaches are associated with major side effects in clinical setting (Table 1).

Yu and colleagues [93] demonstrated that the hematopoietic protein tyrosine phosphatase (HePTP) stabilizes β -catenin in the cytoplasm and allows its nuclear translocation by regulating the phosphorylation of GSK-3 β . This results in the transcriptional activation of target genes leading to cell migration and invasion. Since knockdown of HePTP significantly suppresses metastases formed by TNBC cells, HePTP has been also proposed for therapeutic approaches in TNBC (Table 1).

Recently, Kong et al. [94] have shown that a Rho-GTPase-activating protein, the deleted in liver cancer gene 3 (DLC-3), is downregulated in TNBC and its expression is linked to lymphatic metastases. DLC-3 overexpression leads to β -catenin and c-myc downregulation as well as in reduced in vitro cell proliferation, colony formation, migration, and invasion. Hence, a tumour-suppressor role related to the inhibition of the Wnt/ β -catenin signalling pathway has been postulated (Table 1).

Liu and colleagues [95] have reported a low expression of miR-6838-5p in TNBC compared to normal cells. miR-6838-5p overexpression reduced cell invasion, migration, EMT, β -catenin, *c-myc* and *cyclin D1* expression by post-transcriptionally controlling Wnt3a expression.

Recently, miR-27a-3p was found overexpressed in tumour cells and linked to poor prognosis in TNBC patients. miR-27a-3p leads to the activation of Wnt/ β -catenin cascade and enhances cell proliferation and migration by directly targeting the 3'-UTR region of *GSK-3* β [96] (Table 2).

OVARIAN CANCER

Ovarian Cancer (OC) is a global issue representing the fourth most common cancer in the female population, particularly in developed countries [97]. The poor survival rate is mainly due to the lack of screening methods at the early stages along with the absence of effective treatment options for advanced stages [96][98]. Among different OC subtypes, the epithelial subtype (EOC) holds about 90% of the overall ovarian malignancies [97][99].

Canonical Wnt pathway and OC

Wnt/ β -catenin signalling pathway play a crucial role in carcinogenesis of all OC subtypes [98][100]. In particular, several transcription factors, proteins and miRNAs acting on this pathway have been explored [99][101].

Chen and co-workers [102] investigated the role of the Wnt/ β -catenin pathway antagonist dickkopf-related protein 1 (*DKK1*). They showed that *DKK1* is involved in the control of OC stemness. Mechanistically, it has been shown that STAT3 directly activates the transcription of miRNA-92a, translating in *DKK1* downregulation [102]. Moreover, overexpression of miR-1207 was found to correlate with high nuclear β -catenin level [103]. Wu et al. [103] investigated the effects of miR-1207 on the expression of the *SFRP1-AXIN2* and the inhibitor of β -catenin and T cell factor 4 (*ICAT*). They found that miR-1207 overexpression was associated with a reduced *SFRP1-AXIN2* and *ICAT* expression and the appearance of a stem-like phenotype (Table 1).

Salem et al. [104] proved that miR-590-3p promotes OC growth and metastasis, by targeting *FOXA*2. Moreover, it has been shown that miR-590-3p upregulation significantly increase cell growth, migration, and invasion in EOC cells, both *in vitro* and *in vivo* [105]. Similarly, *FOXA*2, which exhibits suppressive activity on EOC cells, has been identified as a miR-590-3p target [105]. The cyclin

G2 gene (CCNG2) has been also reported to display several repressive actions on EOC-derived tumour cell lines. It inhibits cell proliferation, migration, invasion and EMT. Thereby, since miR-590-3p post-transcriptionally regulates FOXA2, FOXO3, CCNG2 and DDK1 expression, miR-590-3p has been proposed as a potential target in EOC patients [105]. A crucial role of SFRP1 in OC growth has been also proposed. Since miR-1180 is highly expressed in neoplastic tissues, Hu et al. [106] explored the relationship between miR-1180 and the SFRP1/Wnt/ β -catenin signalling pathway in this context, demonstrating that miR-1180 triggers the activation of the Wnt/ β -catenin cascade by targeting SFRP1.

The members of the R-spondin ligand family have been reported as positive effectors of the Wnt/ β -catenin signalling [107]. *LGR4-6* plays crucial roles in the activation of the Wnt/ β -catenin cascade [107,108]. Moreover, Ruan et al. [107] have reported that LGR6 induces stemness and chemo resistance via the Wnt/ β -catenin pathway in OC cells. Restrain of the stem phenotype and increased sensitivity to chemotherapy have been proved by *LGR6* silencing (Table 1).

A recent study established that the overexpression of the Rab GTPase family member, Rab14, regulates $GSK-3\beta$ phosphorylation and β -catenin nuclear accumulation [109,110]. Moreover, high level of Rab14 was found associated with higher expression of Wnt/ β -catenin target genes including MMP-7 and c-myc [110](Table 1).

Jiang et al. [97] have demonstrated that tetrandrine (TET) enhances the anti-tumour effect of paclitaxel (PTX) by decreasing c-myc and cyclin D1 and increasing p21 expression, resulting in cell cycle arrest. The pro-apoptotic effects of PTX+TET have been also investigated. TET was found to inhibit β -catenin downstream target genes by enhancing PTX activity and conferring sensitivity to PTX in resistant cells [97].

Barghout and co-workers [111] demonstrated a more active Wnt/ β -catenin signalling in carboplatin-resistant cells than in sensitive ones. Unlike the Wnt ligands, the negative Wnt regulators *DKK1*, *SFRP1*, and the *FRZB* have been found downregulated in cisplatin-resistant cells. These findings suggest that Wnt/ β -catenin blockade may be effective on resistant EOC.

Non-canonical Wnt pathway and OC

The *FZD7* is highly expressed in OC [112] and its overexpression in mesenchymal (Mes) and Stem-A OC subtypes, has been associated with the induction of EMT. The PCP pathway, which activates the *Rho-ROCK* axis, was found involved in the activation of actomyosin contractility, cadherin-based cell-cell adhesion and migration, while the Wnt/calcium pathway in the metastatic spread and cytoskeleton changes in this clinical setting [112]. Therefore, it has been proposed that the *FZD7* controls both cell cycle progression and cell migration via the non-canonical Wnt/PCP pathway (Table 1).

The integrin beta like 1 subunit (*ITGBL1*) was found highly overexpressed in OC [113]. It has been shown that *ITGBL1* promotes cell migration and adhesion via Wnt/PCP, *RhoA*, the focal adhesion kinase, and the steroid receptor coactivator (*FAK/src*) pathway (Table 1).

The *PTK7* which interacts with Wnt5A, *LRP6* and *FZD7* [114,115] may act as tumour suppressor or oncogene [116,117]. In EOC, *PTK7* downregulation is indeed associated with a poor prognosis [116].

Luo and colleagues [118] have investigated the role of the alkaline phosphatase (*ALPL*) in OC. They demonstrated that *ALPL* overexpression inhibits EMT, migration and invasion of high grade serous OCs (HGSOC) and *FZD2* correlates with a poor survival rate [118]. Mechanistically they have shown that ALPL overexpression represses Wnt5a/*FZD2*-mediated EMT activation possibly by interfering with STAT3 activation [118] (Table 1).

WNT PATHWAY AND OTHER CANCERS

Glioma is an aggressive tumour of the nervous system displaying rapid progression and poor prognosis. Zhao et al. [119] have found that overexpression of β -catenin and *cyclin D1* is associated with high level of the long noncoding RNA, *FGD5* antisense RNA 1 (lncRNA FGD5-AS1). A close relationship between them was straitened by the observations that inhibition of FGD5-AS1 reduced

β-catenin and *cyclin D1* expression while β-catenin downregulation decrease lncRNA FGD5-AS1 expression. This results in the impaired tumour cell migration and invasion.

Prostate cancer (PCa) is among the most common tumour in male. A recent study by Situ et al. [120] provided evidence for the involvement of the microRNA-939 (miR-939) in PCa. Downregulation of miR-939 was found in tumour tissues at advanced tumour stage, in distant lesions as well as associated with poor prognosis. Molecularly, it was demonstrated that miR-939 upregulation interferes with the Wnt/ β -catenin cascade by directly targeting the hepatoma-derived growth factor (HDGF).

Osteosarcoma (OS) is a common bone paediatric tumour displaying high rate of lung metastasis. The inhibition of β -catenin activation, metastasis formation and chemo-resistance were found modulated by tegavivint (a Wnt/ β -catenin inhibitor) which has been proposed as an alternative therapeutic option in OS [121].

Melanoma is among the most immunogenic tumours displaying increased lymphocytic infiltration. Low 1α ,25-dihydroxyvitamin D3 and vitamin D receptor (*VDR*) level correlates to increased cancer incidence and melanoma progression, respectively. Recently, it has been shown that high *VDR* expression correlated with the inhibition of tumour growth, low Wnt/ β -catenin activation and the induction of the immune response [122] (Table 1).

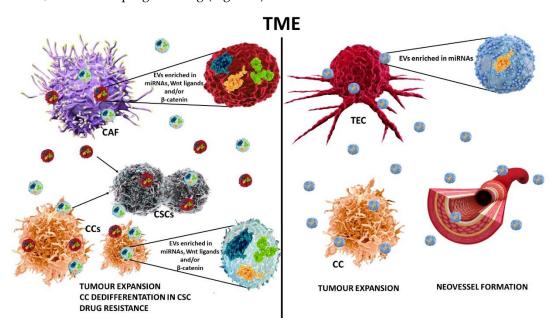
The long non-coding RNA00261 (Linc00261) has been shown to display onco-suppressor properties in Pancreatic Cancer (PC). Linc00261 overexpression inhibits PC cell proliferation, invasion, EMT and metastasis. Bioinformatics analysis revealed that Linc0026 inhibits the activation of the β -catenin/TCF4 cascade and the metastatic spread by regulating the miR-552 5p/FOXO3 axis [123].

EXTRACELLULAR VESICLES AND THE WNT PATHWAY

EVs are heterogeneous small membrane-bound carriers with complex cargoes released under both physiological and pathological conditions. Almost any cell can release EVs, which act as intercellular mediators modifying target cell fate at closed or distant sites [121][124].

Based on the biogenesis, size, content, mechanisms of release and function, three discrete EV subtypes are recognized: microvesicles (MVs), exosomes, and apoptotic bodies [121][124].

EVs-mediated transfer of specific molecules are known to dictate the phenotype of the recipient cell. They can act on proliferation, motility, EMT, migration, invasion, immune evasion, chemoresistance, and TME reprogramming (Figure 3).



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Figure 3. Schematic representation of cell-to-cell communication in the TME by EVs. EVs are released by almost all cell types in the TME. EVs serve as inter-cellular mediators transferring specific molecules (proteins including Wnt ligands and β-catenin, and miRNAs) to recipient cell thus promoting tumour expansion, cancer cell dedifferentiation in CSCs, chemo-resistance, and neovessel formation. CCs: cancer cells; CSCs: cancer stem cells; TEC: tumour-derived endothelial cell; CAF: cancer associated fibroblasts.

Moreover, EVs derived from serum or other biological fluids have been proposed as tumour biomarkers. More importantly, EVs have gained attention as anti-cancer tools. Indeed, EVs can be used as drug delivery systems or potential cancer vaccines. Moreover, the transfer of Wnt ligands or β -catenin via EVs has been proposed as a Wnt signalling activation mechanism.

Kalra et al. [125] have shown that EVs released by CRC cells and containing the mutant β -catenin and high Wnt/ β -catenin activity boost the expression of target genes as *c-myc* and *cyclin D1* when transferred to recipient cell (Table $\frac{13}{2}$).

Table 3. EVs involved in several tumours, their alteration, targets, and impact on tumours

EV CARGO	EV SOURCE	TARGET CELLs	RELATED CANCERS	EXPRESSION LEVEL	PATHWAY INTERACTION	IMPACT ON TUMOUR CELLs	REF.
Mutant β- catenin in EVs	LIM1215	RKO	CRC	Upregulated	<mark>β-catenin</mark>	migration, metastasis tumour growth	[125]
14-3-3ζ in EVs	НЕК293Т	COS-7, SW480	CRC	Upregulated	β-catenin GSK-3β DVL2	survival migration	[126]
Wnt ligands in EVs	CAFs	CRC	CRC	Upregulated	β-catenin	dedifferentiation drug resistance colony formation	[127, 128]
β-catenin in EVs	milk	HCC	HCC	Silenced	β-catenin	proliferation tumour growth	[131, 132]

DKK-1 in EVs	MM	MM	MM	Upregulated	<mark>β-catenin</mark>	osteoclast activity osteoblast differentiation	[133]
EVs	OSCC	OSCC	OSCC	Upregulated	β-catenin	metastasis stemness chemoresistance	[135]
Wnt5b in EVs	Caco-2 and PANC-1	A549	Lung cancer	Upregulated	β-catenin dependent and independent pathways	proliferation migration	[137]
EVs	CAFs	BC	BC	Upregulated	Wnt-PCP	cell growth and motility	[138]

The 14-3-3 are conserved molecules displaying regulatory functions and promoting cancer progression [126]. The 14-3- 3ζ isoform which binds both β -catenin and GSK- 3β , leads to the nuclear translocation and accumulation of β -catenin and enhance cell motility. Moreover, EVs enriched in 14-3- 3ζ and β -catenin, after internalization, promote cell survival and migration by activating the Wnt/ β -catenin cascade [126] (Table $\frac{1}{3}$).

Hu et al. [127] have investigated the mechanism of drug resistance in CRC and have proven that EVs released by fibroblasts drive dedifferentiation of CRC cells towards CSCs (Figure 3a). Additionally, they found that EVs derived from fibroblasts contain the Wnt ligands which activate the Wnt/ β -catenin pathway in target cells, induce transdifferentiation of CRC cells into CSCs and increase drug resistance. Furthermore, it has been reported that collagen accumulation and the *APC* mutation in CRC cells stimulate the release of EVs and, under hypoxia conditions, fibroblast derived EVs boost CRC colony formation [128] (Table $\frac{13}{2}$).

Accumulating evidence shows that EVs enriched in miRNAs are key determinants of human cancer cell growth, invasion and metastasis [129][129]. CAF-derived EVs enclose miR-92a-3p which contribute to cancer progression, stemness, EMT, and drug resistance. Moreover, miR-92a-3p enriched EVs correlated with the activation of the Wnt/ β -catenin pathway [129] (Figure 3a).

Long non-coding RNA-*APC1* (lncRNA-APC1) is a negative regulator of CRC. Low level of lncRNA-APC1 correlates with metastasis, advanced clinical stage and poor prognosis in CRC patients. *APC*, via lncRNA-APC1 promotes cell-cycle arrest and suppresses angiogenesis by lowering the release of CRC cell-derived EVs. Finally, it has been shown that EV-derived from CRC are enriched in Wnt1 and enhance CRC cell proliferation and migration via the non-canonical Wnt/PCP signalling [130].

Hepatocellular carcinoma (HCC) is one of the most common cause of cancer-related deaths worldwide. Constitutive activation of the Wnt/ β -catenin pathway turns into the expression of the epithelial cell adhesion molecule (*EpCAM*) [131]. Ishiguro et al. [132] provided evidence that loss in β -catenin and reduced proliferation and invasion can be obtained by *EpCAM* positive liver cancer stem cells (LCSC) targeted by EVs engineered with a β -catenin specific siRNA (Table $\frac{13}{3}$).

Multiple myeloma (MM) is a hematopoietic malignancy associated with an altered homeostasis of bone formation/resorption. MM-derived EVs enriched in DKK-1 were found to boost the Wnt/ β -catenin signalling and contribute to the abnormal osteogenesis. The inhibition of EV shedding

combined to chemotherapy were found to impair tumour load, angiogenesis and osteolysis [133] (Table 43).

Furthermore, a recent study noticed that the release of EVs from HCC cells is increased in hypoxic conditions and linked to cancer cell proliferation, migration, invasiveness and EMT. Mechanistically they have shown that miR-1273f enriched in EVs activates the Wnt/ β -catenin signalling cascade by targeting the Wnt/ β -catenin inhibitor LHX6 [134].

Chen et al. [135] proved that EVs released from oral squamous cell carcinoma (OSCC) cells correlate with the increased level of β -catenin, the expression of several oncogenic markers, the reprogramming of normal gingival fibroblasts into CAFs, the increased metastasis, stemness reprogramming, chemoresistance, and patients' poor survival (Table $\frac{13}{2}$).

Xia et al. [136] have demonstrated the uptake of EVs and the delivery of functional miRNAs in different cell lines. The exosomal-miR-1260b was found crucial for the activation of the Wnt/ β -catenin signalling and the invasivness of lung adenocarcinoma cells.

Harada et al. [137] purified and characterized Wnt5b-associated EVs. In pancreatic PANC-1 and colorectal Caco-2 cell lines Wnt5a carried by EVs displays the ability to enhance cancer progression (Table 43).

Luga et al. [138] demonstrated that EV shedding by fibroblasts boosts BC cell growth and motility via the Wnt/PCP signalling. CAF-derived EVs were found crucial drivers of cell migration during metastasis formation. Moreover, they found that EVs secreted from fibroblast L cells promote the autocrine Wnt11-PCP cascade in tumour cells increasing their motility and metastatic properties (Table 13).

Lombardo et al. [139] provided evidence that EVs released by tumour-derived endothelial cells (TECs-EVs) boost *in vivo* TEC-derived neovessels. Mechanistically they showed that EV released by naive TECs-EVs regulate the expression of *APC*, *GSK-3\beta* and drive β -catenin nuclear accumulation via miR-214-3p and miR-24-3p (Figure 3b). Overall, this study revealed a key role of the Wnt/ β -catenin cascade in TEC-derived neovessel formation. Moreover, they recently showed that naïve TEC-EVs were also able to boost TNBC metastatic spread and lung metastasis formation when injected intravenously [140] (Table 2).

Overall these data indicate a crucial contribute of EVs released by different cell sources in driving tumor development and dissemination. Several data suggest that these effects mainly rely on the transfer of their specific cargo into target cells. Therefore, approaches able to modify their cargo, particularly miRs and proteins involved in their tumor promoting action, have been proposed as useful therapeutic options. EV engineering by using siRNA for mutated protein has been tested and their effectiveness demonstrated in pancreatic cancer [141]. This suggests that using siRNA for mutant β catenin should be considered as an alternative option for CRC. Likewise, siRNA for different Wnt proteins or rearrangement of dysregulated EV miRs can be used to targeting the Wnt/ β catenin cascade. Alternatively, EVs loaded with Wnt/ β catenin inhibitors can be used as naturally delivery tools.

CONCLUSIONS

Cell-to cell communication is part of the evolutional processes. Wnt ligands are essential for the homeostasis and in the last 30 years genetic, biochemical, and molecular investigations have uncovered several Wnt signalling components [2,3]. Driving interest on this topic mainly relies on dysregulation of the Wnt/ β -catenin signalling and cancer development/progression [3]. Moreover, Wnt/ β -catenin cascade seems to contribute to the TME shape, which play a crucial role in the control of tumour progression and immune regulation. Many different Wnt proteins have been described, and among them Wnt5a, plays a critical role taking part in both the canonical and the non-canonical Wnt/ β -catenin pathway [77,78].

The identification of specific tools able to interfere with the Wnt/ β -catenin cascade has been a hotspot for many years. This is particularly true for CRC, in which almost 70% of CRC patients display *APC* mutations [15]. Apart from CRC, the Wnt/ β -catenin pathway is gaining attention in several malignancies, such as breast, ovarian, melanoma, prostate and paediatric osteosarcoma [117–

119]. At this regard, BC and in particular TNBC are featured by the abnormal activation of both the canonical and non-canonical Wnt/ β -catenin pathway [89,90]. Likewise, a hyper-active Wnt/ β -catenin cascade has been shown to play a crucial role in the progression, stemness, and drug resistance in OC [101,105]. Several miRNAs have been identified to modulate this cascade and thereby widely studied as screening markers or targets in different tumour settings [142].

In the TME, intercellular communication has been recently reported as mediated by the transfer of EV molecular cargo and revised in [143]. Their cargo also includes a number of Wnt components. Of note, wild-type and mutant β -catenin able to promote survival and proliferation of recipient cell and, in several instance dedifferentiation towards a CSC phenotype, have been detected in EVs (Figure 3a). Moreover, their role in mediating drug resistance has been reported. Furthermore, since EVs are released within the TME their contribute in cancer growth and progression has been extensively investigated [144]. EV shedding, blockade, or engineering have been proposed as innovative anti-tumour instrument to fine-tuning the Wnt/ β catenin pathway [142,145].

In the last decades several efforts have been directed to the development of Wnt/ β catenin targeting approaches in order to interfere with tumour progression. However, these efforts have been limited by the crucial role of the Wnt/ β catenin pathway in preserving tissue homeostasis. Therefore, future energies should be directed to clearly dissect the mechanisms driving the unbalanced Wnt/ β catenin pathway in cancer, and EV mechanism of action should be considered amid them. Should identified, targeting approaches would become a suitable anti-cancer option.

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683 REFERENCES

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- Jackstadt, R.; Hodder, M.C.; Sansom, O.J. WNT and β-Catenin in Cancer: Genes and Therapy. *Annu. Rev. Cancer Biol.* 2020, 4, 177–196, doi:10.1146/annurev-cancerbio-030419-033628.
- Komiya, Y.; Habas, R. Wnt signal transduction pathways. *Organogenesis* **2008**, 4, 68–75, doi:10.4161/org.4.2.5851.
- Duchartre, Y.; Kim, Y.M.; Kahn, M. The Wnt signaling pathway in cancer. *Crit. Rev. Oncol. Hematol.* **2016**, 99, 141–149, doi:10.1016/j.critrevonc.2015.12.005.
- Thrasivoulou, C.; Millar, M.; Ahmed, A. Activation of intracellular calcium by multiple Wnt ligands and translocation of β -catenin into the nucleus: A convergent model of Wnt/Ca2+and Wnt/ β -catenin pathways. *J. Biol. Chem.* **2013**, 288, 35651–35659, doi:10.1074/jbc.M112.437913.
- 5. Florian, M.C.; Nattamai, K.J.; Dörr, K.; Marka, G.; Überle, B.; Vas, V.; Eckl, C.; Andrä, I.; Schiemann, M.;
 Oostendorp, R.A.J.; et al. A canonical to non-canonical Wnt signalling switch in haematopoietic stemcell ageing. *Nature* **2013**, *503*, 392–396, doi:10.1038/nature12631.
- 696 6. MacDonald, B.T.; Tamai, K.; He, X. Wnt/β-Catenin Signaling: Components, Mechanisms, and Diseases.
 697 Dev. Cell 2009, 17, 9–26.
- 698 7. Gajos-Michniewicz, A.; Czyz, M. WNT Signaling in Melanoma; 2020; Vol. 21; ISBN 4842272570.
- Wang, Y.; Nathans, J. Tissue/planar cell polarity in vertebrates: New insights and new questions.

 Development 2007, 134, 647–658.
- 9. Saneyoshi, T.; Kume, S.; Amsaki, Y.; Mikoshiba, K. The wnt/calcium pathway activates nf-at and promotes ventral cell fate in xenopus embryos. *Nature* **2002**, *417*, 295–299, doi:10.1038/417295a.
- Tao, Q.; Yokota, C.; Puck, H.; Kofron, M.; Birsoy, B.; Yan, D.; Asashima, M.; Wylie, C.C.; Lin, X.; Heasman, J. Maternal Wnt11 activates the canonical Wnt signaling pathway required for axis formation

- 705 in Xenopus embryos. *Cell* **2005**, *120*, 857–871, doi:10.1016/j.cell.2005.01.013.
- 706 11. Steinhart, Z.; Angers, S. Wnt signaling in development and tissue homeostasis. *Development* **2018**, 145, 1–8, doi:10.1242/dev.146589.
- Rawla, P.; Sunkara, T.; Barsouk, A. Epidemiology of colorectal cancer: Incidence, mortality, survival, and risk factors. *Prz. Gastroenterol.* **2019**, *14*, 89–103, doi:10.5114/pg.2018.81072.
- 710 13. Bhandari, A.; Woodhouse, M.; Gupta, S. Colorectal cancer is a leading cause of cancer incidence and mortality among adults younger than 50 years in the USA: A SEER-based analysis with comparison to other young-onset cancers. *J. Investig. Med.* **2017**, *65*, 311–315, doi:10.1136/jim-2016-000229.
- 713 14. Dienstmann, R.; Wang, X.; de Reyni, lien; Schlicker, A.; Soneson, C.; Marisa, L.; Roepman, P.; Nyamundanda, G.; Angelino, P.; Bot, B.M.; et al. The consensus molecular subtypes of colorectal cancer. 2015, doi:10.1038/nm.3967.
- Liu, L.; Zhang, Y.; Wong, C.C.; Zhang, J.; Dong, Y.; Li, X.; Kang, W.; Chan, F.K.L.; Sung, J.J.Y.; Yu, J.
 RNF6 promotes colorectal cancer by activating the Wnt/b-catenin pathway via ubiquitination of TLE3.
 Cancer Res. 2018, 78, 1958–1971, doi:10.1158/0008-5472.CAN-17-2683.
- 519 16. Study of the role of leucine-rich repeat-containing g-protein coupled receptor 5 (LGR5) and WNT pathway in colon cancer | Egyptian Journal of Biochemistry and Molecular Biology Available online: https://www.ajol.info/index.php/ejbmb/article/view/191481 (accessed on Sep 11, 2020).
- 722 17. Moon, B.S.; Jeong, W.J.; Park, J.; Kim, T. II; Min, D.S.; Choi, K.Y. Role of oncogenic K-Ras in cancer stem cell activation by aberrant wnt/β-catenin signaling. *J. Natl. Cancer Inst.* **2014**, 106, 1–10, doi:10.1093/jnci/djt373.
- T25 18. Lenz, H.J.; Kahn, M. Safely targeting cancer stem cells via selective catenin coactivator antagonism.

 Cancer Sci. 2014, 105, 1087–1092, doi:10.1111/cas.12471.
- 727 19. Yu, D.H.; Zhang, X.; Wang, H.; Zhang, L.; Chen, H.; Hu, M.; Dong, Z.; Zhu, G.; Qian, Z.; Fan, J.; et al. 728 The essential role of TNIK gene amplification in gastric cancer growth. *Oncogenesis* **2014**, 3, doi:10.1038/oncsis.2014.2.
- 730 20. Masuda, M.; Uno, Y.; Ohbayashi, N.; Ohata, H.; Mimata, A.; Kukimoto-Niino, M.; Moriyama, H.; 731 Kashimoto, S.; Inoue, T.; Goto, N.; et al. TNIK inhibition abrogates colorectal cancer stemness. *Nat. Commun.* 2016, 7, 1–7, doi:10.1038/ncomms12586.
- Cha, P.H.; Cho, Y.H.; Lee, S.K.; Lee, J.; Jeong, W.J.; Moon, B.S.; Yun, J.H.; Yang, J.S.; Choi, S.; Yoon, J.; et
 al. Small-molecule binding of the axin RGS domain promotes β-catenin and Ras degradation. *Nat. Chem.*Biol. 2016, 12, 593–600, doi:10.1038/nchembio.2103.
- Cho, Y.H.; Ro, E.J.; Yoon, J.S.; Kwak, D.K.; Cho, J.; Kang, D.W.; Lee, H.Y.; Choi, K.Y. Small molecule-induced simultaneous destabilization of β-catenin and RAS is an effective molecular strategy to suppress stemness of colorectal cancer cells. *Cell Commun. Signal.* 2020, *18*, 1–11, doi:10.1186/s12964-020-0519-z.
- 739 23. Tan, W.; Liu, B.; Qu, S.; Liang, G.; Luo, W.; Gong, C. MicroRNAs and cancer: Key paradigms in molecular therapy (Review). *Oncol. Lett.* **2018**, *15*, 2735–2742, doi:10.3892/ol.2017.7638.
- 741 24. Lee, Y.S.; Dutta, A. MicroRNAs in Cancer Contents: *Annu Rev Pathol* 2009, 4, 199–227,
 742 doi:10.1146/annurev.pathol.4.110807.092222.MicroRNAs.
- Sun, N.; Zhang, L.; Zhang, C.; Yuan, Y. MiR-144-3p inhibits cell proliferation of colorectal cancer cells by
 targeting BCL6 via inhibition of Wnt/β-catenin signaling. *Cell. Mol. Biol. Lett.* 2020, 25,
 doi:10.1186/s11658-020-00210-3.
- Polo, J.M.; Dell'Oso, T.; Ranuncolo, S.M.; Cerchietti, L.; Beck, D.; Da Silva, G.F.; Prive, G.G.; Licht, J.D.;
 Melnick, A. Specific peptide interference reveals BCL6 transcriptional and oncogenic mechanisms in B-

- 748 cell lymphoma cells. *Nat. Med.* **2004**, *10*, 1329–1335, doi:10.1038/nm1134.
- 749 27. Shaffer, A.L.; Yu, X.; He, Y.; Boldrick, J.; Chan, E.P.; Staudt, L.M. BCL-6 represses genes that function in
- 750 lymphocyte differentiation, inflammation, and cell cycle control. *Immunity* **2000**, *13*, 199–212, doi:10.1016/S1074-7613(00)00020-0.
- 752 28. Liu, W.Y.; Yang, Z.; Sun, Q.; Yang, X.; Hu, Y.; Xie, H.; Gao, H.J.; Guo, L.M.; Yi, J.Y.; Liu, M.; et al. miR-
- 377-3p drives malignancy characteristics via upregulating GSK-3β expression and activating NF-κB pathway in hCRC cells. *J. Cell. Biochem.* **2018**, *119*, 2124–2134, doi:10.1002/jcb.26374.
- Huang, L.; Liu, Z.; Hu, J.; Luo, Z.; Zhang, C.; Wang, L.; Wang, Z. MiR-377-3p suppresses colorectal cancer
- through negative regulation on Wnt/β-catenin signaling by targeting XIAP and ZEB2. *Pharmacol. Res.* 2020, *156*, 104774, doi:10.1016/j.phrs.2020.104774.
- 758 Qi, S.; Song, Y.; Peng, Y.; Wang, H.; Long, H.; Yu, X.; Li, Z.; Fang, L.; Wu, A.; Luo, W.; et al. ZEB2 mediates
- multiple pathways regulating cell proliferation, migration, invasion, and apoptosis in glioma. *PLoS One* **2012**, *7*, 1–12, doi:10.1371/journal.pone.0038842.
- Ng, V.H.; Hang, B.I.; Sawyer, L.M.; Neitzel, L.R.; Crispi, E.E.; Rose, K.L.; Popay, T.M.; Zhong, A.; Lee,
- L.A.; Tansey, W.P.; et al. Phosphorylation of XIAP at threonine 180 controls its activity in Wnt signaling.

 2018, doi:10.1242/jcs.210575.
- 764 32. Lv, S.; Zhang, J.; He, Y.; Liu, Q.; Wang, Z.; Liu, B.; Shi, L.; Wu, Y. MicroRNA-520e targets AEG-1 to
- suppress the proliferation and invasion of colorectal cancer cells through Wnt/GSK-3β/β-catenin signalling. *Clin. Exp. Pharmacol. Physiol.* **2020**, *47*, 158–167, doi:10.1111/1440-1681.13185.
- Hu, G.; Wei, Y.; Kang, Y. The multifaceted role of MTDH/AEG-1 in cancer progression. *Clin. Cancer Res.* 2009, 15, 5615–5620, doi:10.1158/1078-0432.CCR-09-0049.
- 769 34. Zhang, M.; Miao, F.; Huang, R.; Liu, W.; Zhao, Y.; Jiao, T.; Lu, Y.; Wu, F.; Wang, X.; Wang, H.; et al.
- RHBDD1 promotes colorectal cancer metastasis through the Wnt signaling pathway and its downstream target ZEB1. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 1–14, doi:10.1186/s13046-018-0687-5.
- Deng, M.; Chen, Z.; Tan, J.; Liu, H. Down-regulation of SLC35C1 induces colon cancer through overactivating Wnt pathway. *J. Cell. Mol. Med.* **2020**, 24, 3079–3090, doi:10.1111/jcmm.14969.
- 774 36. Xu, C.; Tian, G.; Jiang, C.; Xue, H.; Kuerbanjiang, M.; Sun, L.; Gu, L.; Zhou, H.; Liu, Y.; Zhang, Z.; et al.
- NPTX2 promotes colorectal cancer growth and liver metastasis by the activation of the canonical Wnt/βcatenin pathway via FZD6. *Cell Death Dis.* **2019**, *10*, doi:10.1038/s41419-019-1467-7.
- 777 37. Vymetalkova, V.; Vodicka, P.; Vodenkova, S.; Alonso, S.; Schneider-Stock, R. DNA methylation and
- 778 chromatin modifiers in colorectal cancer. *Mol. Aspects Med.* **2019**, *69*, 73–92, doi:10.1016/j.mam.2019.04.002.
- Tanaka, N.; Mashima, T.; Mizutani, A.; Sato, A.; Aoyama, A.; Gong, B.; Yoshida, H.; Muramatsu, Y.
- Nakata, K.; Matsuura, M.; et al. APC mutations as a potential biomarker for sensitivity to tankyrase
- 782 inhibitors in colorectal cancer. *Mol. Cancer Ther.* **2017**, *16*, 752–762, doi:10.1158/1535-7163.MCT-16-0578.
- 783 39. Bruschi, M.; Garnier, L.; Cleroux, E.; Giordano, A.; Dumas, M.; Bardet, A.F.; Kergrohen, T.; Quesada, S.;
- Cesses, P.; Weber, M.; et al. Loss of Apc Rapidly Impairs DNA Methylation Programs and Cell Fate Decisions in Lgr5+ Intestinal Stem Cells; 2020; Vol. 80; ISBN 3343435929.
- 786 40. Song, Y.; Yuan, H.; Wang, J.; Wu, Y.; Xiao, Y.; Mao, S. KLHL22 regulates the EMT and proliferation in
- 787 colorectal cancer cells in part via the Wnt/β-catenin signaling pathway. *Cancer Manag. Res.* **2020**, *12*, 3981–788 3993, doi:10.2147/CMAR.S252232.
- 789 41. Proto, M.C.; Fiore, D.; Piscopo, C.; Franceschelli, S.; Bizzarro, V.; Laezza, C.; Lauro, G.; Feoli, A.; Tosco,
- A.; Bifulco, G.; et al. Inhibition of Wnt/β-Catenin pathway and Histone acetyltransferase activity by

- 791 Rimonabant: A therapeutic target for colon cancer. *Sci. Rep.* **2017**, *7*, 1–14, doi:10.1038/s41598-017-11688-792 x.
- Liu, Y.; Rubin, B.; Bodine, P.V.N.; Billiard, J. Wnt5a induces homodimerization and activation of Ror2 receptor tyrosine kinase. *J. Cell. Biochem.* **2008**, *105*, 497–502, doi:10.1002/jcb.21848.
- Lee, J.M.; Kim, I.S.; Kim, H.; Lee, J.S.; Kim, K.; Yim, H.Y.; Jeong, J.; Kim, J.H.; Kim, J.Y.; Lee, H.; et al.
 RORα Attenuates Wnt/β-Catenin Signaling by PKCα-Dependent Phosphorylation in Colon Cancer. *Mol.* Cell 2010, 37, 183–195, doi:10.1016/j.molcel.2009.12.022.
- Voloshanenko, O.; Schwartz, U.; Kranz, D.; Rauscher, B.; Linnebacher, M.; Augustin, I.; Boutros, M. β catenin-independent regulation of Wnt target genes by RoR2 and ATF2/ATF4 in colon cancer cells. *Sci. Rep.* 2018, 8, 1–14, doi:10.1038/s41598-018-20641-5.
- Nishioka, M.; Ueno, K.; Hazama, S.; Okada, T.; Sakai, K.; Suehiro, Y.; Okayama, N.; Hirata, H.; Oka, M.; Imai, K.; et al. Possible involvement of Wnt11 in colorectal cancer progression. *Mol. Carcinog.* **2013**, *52*, 207–217, doi:10.1002/mc.21845.
- 46. Gorroño-Etxebarria, I.; Aguirre, U.; Sanchez, S.; González, N.; Escobar, A.; Zabalza, I.; Quintana, J.M.; dM Vivanco, M.; Waxman, J.; Kypta, R.M. Wnt-11 as a potential prognostic biomarker and therapeutic target in colorectal cancer. *Cancers* (*Basel*). **2019**, *11*, 1–19, doi:10.3390/cancers11070908.
- Vitale, I.; Manic, G.; Coussens, L.M.; Kroemer, G.; Galluzzi, L. Macrophages and Metabolism in the Tumor Microenvironment. *Cell Metab.* 2019, *30*, 36–50.
- 48. Liu, Q.; Song, J.; Pan, Y.; Shi, D.; Yang, C.; Wang, S.; Xiong, B. Wnt5a/caMKII/ERK/CCL2 axis is required for tumor-associated macrophages to promote colorectal cancer progression. *Int. J. Biol. Sci.* **2020**, *16*, 811 1023–1034, doi:10.7150/ijbs.40535.
- Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer* 1. Clin. 2018, 68, 394–424, doi:10.3322/caac.21492.
- Du, X.L.; Fox, E.E.; Lai, D. Competing Causes of Death for Women With Breast Cancer and Change Over Time From 1975 to 2003 NIH Public Access. *Am J Clin Oncol* 2008, 31, 105–116, doi:10.1097/COC.0b013e318142c865.
- 51. Yu, Q.C.; Verheyen, E.M.; Zeng, Y.A. Mammary development and breast cancer: A Wnt perspective.

 819 *Cancers (Basel).* **2016**, *8*, 1–26, doi:10.3390/cancers8070065.
- Katoh, M. Canonical and non-canonical WNT signaling in cancer stem cells and their niches: Cellular heterogeneity, omics reprogramming, targeted therapy and tumor plasticity (Review). *Int. J. Oncol.* 2017, 51, 1357–1369.
- 823 53. Khramtsov, A.I.; Khramtsova, G.F.; Tretiakova, M.; Huo, D.; Olopade, O.I.; Goss, K.H. Wnt/β-catenin 824 pathway activation is enriched in basal-like breast cancers and predicts poor outcome. *Am. J. Pathol.* 825 2010, 176, 2911–2920, doi:10.2353/ajpath.2010.091125.
- 826 54. Sormunen, R.T.; Leong, A.S.Y.; Vääräniemi, J.P.; Fernando, S.S.E.; Eskelinen, S.M. Immunolocalization of the fodrin, E-cadherin, and β-catenin adhesion complex in infiltrating ductal carcinoma of the breast Comparison with an in vitro model. *J. Pathol.* **1999**, *187*, 416–423, doi:10.1002/(sici)1096-829 9896(199903)187:4<416::aid-path255>3.0.co;2-d.
- 830 55. Lacroix-Triki, M.; Geyer, F.C.; Lambros, M.B.; Savage, K.; Ellis, I.O.; Lee, A.H.S.; Reis-Filho, J.S. β-831 Catenin/Wnt signalling pathway in fibromatosis, metaplastic carcinomas and phyllodes tumours of the breast. *Mod. Pathol.* **2010**, 23, 1438–1448, doi:10.1038/modpathol.2010.141.
- Samarzija, I.; Sini, P.; Schlange, T.; MacDonald, G.; Hynes, N.E. Wnt3a regulates proliferation and

- migration of HUVEC via canonical and non-canonical Wnt signaling pathways. *Biochem. Biophys. Res. Commun.* **2009**, *386*, 449–454, doi:10.1016/j.bbrc.2009.06.033.
- Medina, A.; Reintsch, W.; Steinbeisser, H. Xenopus frizzled 7 can act in canonical and non-canonical Wnt
- signaling pathways: implications on early patterning and morphogenesis. *Mech. Dev.* **2000**, *92*, 227–237, doi:10.1016/S0925-4773(00)00240-9.
- 839 58. Michaelson, J.S.; Leder, P. β-catenin is a downstream effector of Wnt-mediated tumorigenesis in the mammary gland. *Oncogene* **2001**, *20*, 5093–5099, doi:10.1038/sj.onc.1204586.
- Klopocki, E.; Kristiansen, G.; Wild, P.J.; Klaman, I.; Castanos-Velez, E.; Singer, G.; Stöhr, R.; Simon, R.;
- Sauter, G.; Leibiger, H.; et al. Loss of SFRP1 is associated with breast cancer progression and poor prognosis in early stage tumors. *Int. J. Oncol.* **2004**, *25*, 641–649, doi:10.3892/ijo.25.3.641.
- 844 60. Woodward, W.A.; Chen, M.S.; Behbod, F.; Alfaro, M.P.; Buchholz, T.A.; Rosen, J.M. WNT/β-catenin
- mediates radiation resistance of mouse mammary progenitor cells. *Proc. Natl. Acad. Sci. U. S. A.* **2007**, 846 104, 618–623, doi:10.1073/pnas.0606599104.
- 447 You, F.; Li, J.; Zhang, P.; Zhang, H.; Cao, X. MiR106a promotes the growth of transplanted breast cancer and decreases the sensitivity of transplanted tumors to cisplatin. *Cancer Manag. Res.* **2020**, *12*, 233–246, doi:10.2147/CMAR.S231375.
- 850 62. Zhu, X.; Feng, J.; Fu, W.; Shu, X.; Wan, X.; Liu, J. Effects of cisplatin on the proliferation, invasion and apoptosis of breast cancer cells following β-catenin silencing. *Int. J. Mol. Med.* **2020**, *45*, 1838–1850, doi:10.3892/ijmm.2020.4543.
- 853 Cou, Y.; Lin, X.; Bu, J.; Lin, Z.; Chen, Y.; Qiu, Y.; Mo, H.; Tang, Y.; Fang, W.; Wu, Z. Timeless-Stimulated miR-5188-FOXO1/β-Catenin-c-Jun Feedback Loop Promotes Stemness via Ubiquitination of β-Catenin in Breast Cancer. *Mol. Ther.* **2019**, *28*, 313–327, doi:10.1016/j.ymthe.2019.08.015.
- Han, L.; Yan, Y.; Zhao, L.; Liu, Y.; Lv, X.; Zhang, L.; Zhao, Y.; Zhao, H.; He, M.; Wei, M. LncRNA HOTTIP facilitates the stemness of breast cancer via regulation of miR-148a-3p/WNT1 pathway. *J. Cell. Mol. Med.*2020, 24, 6242–6252, doi:10.1111/jcmm.15261.
- 859 65. Yue, Z.; Yuan, Z.; Zeng, L.; Wang, Y.; Lai, L.; Li, J.; Sun, P.; Xue, X.; Qi, J.; Yang, Z.; et al. LGR4 modulates breast cancer initiation, metastasis, and cancer stem cells. FASEB J. 2018, 32, 2422–2437, doi:10.1096/fj.201700897R.
- 862 Ruffner, H.; Lle Sprunger, J.; Charlat, O.; Leighton-Davies, J.; Grosshans, B.; Salathe, A.; Zietzling, S.; Rie Beck, V.; Therier, M.; Isken, A.; et al. R-Spondin Potentiates Wnt/b-Catenin Signaling through Orphan Receptors LGR4 and LGR5. 2012, doi:10.1371/journal.pone.0040976.
- Wels, C.; Joshi, S.; Koefinger, P.; Bergler, H. Transcriptional Activation of ZEB1 by Slug Leads to Cooperative Regulation of the EMT like Phenotype in Melanoma. *J Invest Dermatol* **2011**, 131, 1877–1885, doi:10.1038/jid.2011.142.Transcriptional.
- 868 Yang, L.; Xie, X.; Tang, H.; Kong, Y.; Xie, X.; Chen, J.; Song, C.; Liu, X.; Ye, F.; Li, N.; et al. LGR5 promotes breast cancer progression and maintains stem-like cells through activation of wnt/β-catenin signaling.

 870 Stem Cells 2015, 33, 2913–2924, doi:10.1002/stem.2083.
- 871 69. Wang, H.; Sun, L.; Jiang, J.; Yu, S.; Zhou, Q. Suppression of the proliferation and invasion of breast cancer cells by ST7L occurs through inhibition of activation of Wnt/GSK-3β/β-catenin signalling. *Clin. Exp. Pharmacol. Physiol.* **2020**, *47*, 119–126, doi:10.1111/1440-1681.13166.
- Zhang, X.; Luo, Y.; Li, Q. TMED3 promotes proliferation and migration in breast cancer cells by
 activating Wnt/β-catenin signaling. Onco. Targets. Ther. 2020, 13, 5819–5830, doi:10.2147/OTT.S250766.
- Zhou, J.; Su, C.M.; Chen, H.A.; Du, S.; Li, C.W.; Wu, H.; Tsai, S.H.; Yeh, Y.T. Cryptanshinone inhibits the

- 877 glycolysis and inhibits cell migration through PKM2/β-catenin axis in breast cancer. *Onco. Targets. Ther.*878 **2020**, *13*, 8629–8639, doi:10.2147/OTT.S239134.
- 879 72. Mikels, A.J.; Nusse, R. Purified Wnt5a protein activates or inhibits β-catenin-TCF signaling depending on receptor context. *PLoS Biol.* **2006**, *4*, 570–582, doi:10.1371/journal.pbio.0040115.
- Yamaguchi, T.P.; Bradley, A.; McMahon, A.P.; Jones, S. A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo. *Development* **1999**, *126*, 1211–1223.
- Säfholm, A.; Tuomela, J.; Rosenkvist, J.; Dejmek, J.; Härkönen, P.; Andersson, T. The wnt-5a-derived hexapeptide Foxy-5 inhibits breast cancer metastasis in vivo by targeting cell motility. *Clin. Cancer Res.* **2008**, *14*, 6556–6563, doi:10.1158/1078-0432.CCR-08-0711.
- Prasad, C.P.; Södergren, K.; Andersson, T. Reduced production and uptake of lactate are essential for the ability of WNT5A signaling to inhibit breast cancer cell migration and invasion. *Oncotarget* **2017**, *8*, 71471–71488, doi:10.18632/oncotarget.17277.
- Lee, S.Y.; Jeon, H.M.; Ju, M.K.; Kim, C.H.; Yoon, G.; Han, S.I.; Park, H.G.; Kang, H.S. Wnt/snail signaling regulates cytochrome c oxidase and glucose metabolism. *Cancer Res.* **2012**, 72, 3607–3617, doi:10.1158/0008-5472.CAN-12-0006.
- 892 77. Borcherding, N.; Kusner, D.; Kolb, R.; Xie, Q.; Li, W.; Velez, G.; Askeland, R.; Weigel, R.J.; Zhang, W.; 893 City, I.; et al. HHS Public Access. **2016**, *75*, 1972–1982, doi:10.1158/0008-5472.CAN-14-2761.Paracrine.
- 894 78. Roarty, K.; Baxley, S.E.; Crowley, M.R.; Frost, A.R.; Serra, R. Loss of TGF-β or Wnt5a results in an increase in Wnt/β-catenin activity and redirects mammary tumour phenotype. *Breast Cancer Res.* **2009**, 11, 1–11, doi:10.1186/bcr2244.
- Leris, A.C.A.; Roberts, T.R.; Jiang, W.G.; Newbold, R.F.; Mokbel, K. WNT5A expression in human breast cancer. *Anticancer Res.* **2005**, *25*, 731–734.
- 899 80. Jönsson, M.; Dejmek, J.; Bendahl, P.O.; Andersson, T. Loss of Wnt-5a protein is associated with early relapse in invasive ductal breast carcinomas. *Cancer Res.* **2002**, *62*, 409–416.
- 81. Kobayashi, Y.; Kadoya, T.; Amioka, A.; Hanaki, H.; Sasada, S.; Masumoto, N.; Yamamoto, H.; Arihiro, K.; Kikuchi, A.; Okada, M. Wnt5a-induced cell migration is associated with the aggressiveness of estrogen receptor-positive breast cancer. *Oncotarget* **2018**, *9*, 20979–20992, doi:10.18632/oncotarget.24761.
- 904 82. Bergenfelz, C.; Medrek, C.; Ekström, E.; Jirström, K.; Janols, H.; Wullt, M.; Bredberg, A.; Leandersson, K. 905 Wnt5a Induces a Tolerogenic Phenotype of Macrophages in Sepsis and Breast Cancer Patients. *J. Immunol.* 2012, 188, 5448–5458, doi:10.4049/jimmunol.1103378.
- 907 83. Bergenfelz, C.; Janols, H.; Wullt, M.; Jirström, K.; Bredberg, A.; Leandersson, K. Wnt5a Inhibits Human 908 Monocyte-Derived Myeloid Dendritic Cell Generation. *Scand. J. Immunol.* **2013**, *78*, 194–204, 909 doi:10.1111/sji.12075.
- 910 84. Fernandez-Cobo, M.; Zammarchi, F.; Mandeli, J.; Holland, J.F.; Pogo, B.G.T. Expression of Wnt5A and Wnt10B in non-immortalized breast cancer cells. *Oncol. Rep.* **2007**, *17*, 903–907, doi:10.3892/or.17.4.903.
- 912 85. Prasad, C.P.; Chaurasiya, S.K.; Guilmain, W.; Andersson, T. WNT5A signaling impairs breast cancer cell migration and invasion via mechanisms independent of the epithelial-mesenchymal transition. *J. Exp.* 914 *Clin. Cancer Res.* **2016**, *35*, 1–15, doi:10.1186/s13046-016-0421-0.
- 915 86. Bauer, M.; Bénard, J.; Gaasterland, T.; Willert, K.; Cappellen, D. WNT5A encodes two isoforms with distinct functions in cancers. *PLoS One* **2013**, *8*, 1–14, doi:10.1371/journal.pone.0080526.
- 917 87. Abramson, V.G.; Lehmann, B.D.; Ballinger, T.J.; Pietenpol, J.A. Subtyping of triple-negative breast cancer: Implications for therapy. *Cancer* 2015, 121, 8–16, doi:10.1002/cncr.28914.
- 919 88. Marra, A.; Viale, G.; Curigliano, G. Recent advances in triple negative breast cancer: The immunotherapy

- 920 era. *BMC Med.* **2019**, *17*, 1–9, doi:10.1186/s12916-019-1326-5.
- 921 89. Dent, R.; Trudeau, M.; Pritchard, K.I.; Hanna, W.M.; Kahn, H.K.; Sawka, C.A.; Lickley, L.A.; Rawlinson,
- E.; Sun, P.; Narod, S.A. Triple-negative breast cancer: Clinical features and patterns of recurrence. *Clin. Cancer Res.* 2007, *13*, 4429–4434, doi:10.1158/1078-0432.CCR-06-3045.
- 924 90. Geyer, F.C.; Pareja, F.; Weigelt, B.; Rakha, E.; Ellis, I.O.; Schnitt, S.J.; Reis-Filho, J.S. The Spectrum of Triple-Negative Breast Disease: High- and Low-Grade Lesions. *Am. J. Pathol.* 2017, *187*, 2139–2151.
- 926 91. Shen, H.; Yan, W.; Yuan, J.; Wang, Z.; Wang, C. Nek2B activates the wnt pathway and promotes triple-927 negative breast cancer chemothezrapy-resistance by stabilizing β-catenin. *J. Exp. Clin. Cancer Res.* **2019**, 928 38, 1–17, doi:10.1186/s13046-019-1231-y.
- 92. Puvirajesinghe, T.M.; Bertucci, F.; Jain, A.; Scerbo, P.; Belotti, E.; Audebert, S.; Sebbagh, M.; Lopez, M.; 930 Brech, A.; Finetti, P.; et al. Identification of p62/SQSTM1 as a component of non-canonical Wnt VANGL2-931 JNK signalling in breast cancer. *Nat. Commun.* **2016**, 7, doi:10.1038/ncomms10318.
- 93. Yu, L.; Wang, C.; Pan, F.; Liu, Y.; Ren, X.; Zeng, H.; Shi, Y. HePTP promotes migration and invasion in
 933 triple-negative breast cancer cells via activation of Wnt/β-catenin signaling. *Biomed. Pharmacother.* 2019,
 934 118, doi:10.1016/j.biopha.2019.109361.
- 935 94. Kong, B.; Lv, Z.-D.; Xia, J.; Jin, L.-Y.; Yang, Z.-C. DLC-3 suppresses cellular proliferation, migration, and invasion in triple-negative breast cancer by the Wnt/β-catenin pathway. *Int. J. Clin. Exp. Pathol.* **2019**, 12, 937 1224–1232.
- 938 95. Liu, G.; Wang, P.; Zhang, H. MiR-6838-5p suppresses cell metastasis and the EMT process in triple-939 negative breast cancer by targeting WNT3A to inhibit the Wnt pathway. *J. Gene Med.* **2019**, 21, 1–8, 940 doi:10.1002/jgm.3129.
- 94. Wu, R.; Zhao, B.; Ren, X.; Wu, S.; Liu, M.; Wang, Z.; Liu, W. Mir-27a-3p targeting gsk3β promotes triple-942 negative breast cancer proliferation and migration through wnt/β-catenin pathway. *Cancer Manag. Res.* 943 **2020**, *12*, 6241–6249, doi:10.2147/CMAR.S255419.
- 944 97. Jiang, L.; Hou, R. Tetrandrine Reverses Paclitaxel Resistance in Human Ovarian Cancer via Inducing
 945 Apoptosis, Cell Cycle Arrest Through β-Catenin Pathway. Onco. Targets. Ther. 2020, Volume 13,
 946 3631–3639, doi:10.2147/OTT.S235533.
- 947 98. Nagaraj, A.B.; Joseph, P.; Kovalenko, O.; Singh, S.; Armstrong, A.; Redline, R.; Resnick, K.; Zanotti, K.; 948 Waggoner, S.; Di Feo, A. Critical role of Wnt/\(\beta\)-catenin signaling in driving epithelial ovarian cancer platinum resistance. *Oncotarget* 2015, 6, 23720–23734, doi:10.18632/oncotarget.4690.
- 99. Sauriol, A.; Simeone, K.; Portelance, L.; Meunier, L.; Leclerc-Desaulniers, K.; De Ladurantaye, M.;
 Chergui, M.; Kendall-Dupont, J.; Rahimi, K.; Carmona, E.; et al. Modeling the diversity of epithelial
 ovarian cancer through ten novel well characterized cell lines covering multiple subtypes of the disease.

 Cancers (Basel). 2020, 12, 1–26, doi:10.3390/cancers12082222.
- Jeong, J.W.; Lee, H.S.; Franco, H.L.; Broaddus, R.R.; Taketo, M.M.; Tsai, S.Y.; Lydon, J.P.; DeMayo, F.J.
 B-Catenin Mediates Glandular Formation and Dysregulation of B-Catenin Induces Hyperplasia
 Formation in the Murine Uterus. Oncogene 2009, 28, 31–40, doi:10.1038/onc.2008.363.
- 957 101. Ryland, G.L.; Bearfoot, J.L.; Doyle, M.A.; Boyle, S.E.; Choong, D.Y.H.; Rowley, S.M.; Tothill, R.W.; 958 Gorringe, K.L.; Campbell, I.G. MicroRNA Genes and Their Target 3'-Untranslated Regions Are 1059 Infrequently Somatically Mutated in Ovarian Cancers. PLoS One 2012, 7, e35805, doi:10.1371/journal.pone.0035805.
- 961 102. Chen, M.W.; Yang, S.T.; Chien, M.H.; Hua, K.T.; Wu, C.J.; Hsiao, S.M.; Lin, H.; Hsiao, M.; Su, J.L.; Wei, L.H. The STAT3-miRNA-92-Wnt signaling pathway regulates spheroid formation and malignant

- 963 progression in ovarian cancer. *Cancer Res.* **2017**, 77, 1955–1967, doi:10.1158/0008-5472.CAN-16-1115.
- 964 103. Wu, G.; Liu, A.; Zhu, J.; Lei, F.; Wu, S.; Zhang, X.; Ye, L.; Cao, L.; He, S. MiR-1207 overexpression
- promotes cancer stem cell-like traits in ovarian cancer by activating the Wnt/β-catenin signaling pathway. *Oncotarget* **2015**, *6*, 28882–28894, doi:10.18632/oncotarget.4921.
- 967 104. Salem, M.; Shan, Y.; Bernaudo, S.; Peng, C. miR-590-3p Targets Cyclin G2 and FOXO3 to Promote
- Ovarian Cancer Cell Proliferation, Invasion, and Spheroid Formation. *Int. J. Mol. Sci.* **2019**, *20*, 1810, doi:10.3390/ijms20081810.
- 970 105. Salem, M.; O'brien, J.A.; Bernaudo, S.; Shawer, H.; Ye, G.; Brki, J.; Amleh, A.; Vanderhyden, B.C.; Refky,
- B.; Yang, B.B.; et al. miR-590-3p Promotes Ovarian Cancer Growth and Metastasis via a Novel FOXA2-
- 972 Versican Pathway. **2018**, doi:10.1158/0008-5472.CAN-17-3014.
- 973 106. Hu, J.; Zhao, W.; Huang, Y.; Wang, Z.; Jiang, T.; Wang, L. MiR-1180 from bone marrow MSCs promotes cell proliferation and glycolysis in ovarian cancer cells via SFRP1/Wnt pathway. *Cancer Cell Int.* **2019**, *19*,
- 975 66, doi:10.1186/s12935-019-0751-z.
- 976 107. Ruan, X.; Liu, A.; Zhong, M.; Wei, J.; Zhang, W.; Rong, Y.; Liu, W.; Li, M.; Qing, X.; Chen, G.; et al.
- 977 Silencing LGR6 Attenuates Stemness and Chemoresistance via Inhibiting Wnt/β-Catenin Signaling in
- 978 Ovarian Cancer. *Mol. Ther. Oncolytics* **2019**, *14*, 94–106, doi:10.1016/j.omto.2019.04.002.
- Park, S.; Cui, J.; Yu, W.; Wu, L.; Carmon, K.S.; Liu, Q.J. Differential activities and mechanisms of the four
- 980 r-spondins in potentiating wnt/-catenin signaling. *J. Biol. Chem.* **2018**, 293, 9759–9769, doi:10.1074/jbc.RA118.002743.
- 982 109. Nguyen, V.H.L.; Hough, R.; Bernaudo, S.; Peng, C. Wnt/β-catenin signalling in ovarian cancer: Insights into its hyperactivation and function in tumorigenesis. *J. Ovarian Res.* 2019, *12*, 1–17.
- Hou, R.; Jiang, L.; Yang, Z.; Wang, S.; Liu, Q. Rab14 is overexpressed in ovarian cancers and promotes
- 985 ovarian cancer proliferation through Wnt pathway. *Tumor Biol.* **2016**, *37*, 16005–16013, doi:10.1007/s13277-016-5420-4.
- 987 111. Barghout, S.H.; Zepeda, N.; Xu, Z.; Steed, H.; Lee, C.H.; Fu, Y. Elevated β-catenin activity contributes to
- carboplatin resistance in A2780cp ovarian cancer cells. *Biochem. Biophys. Res. Commun.* **2015**, *468*, 173–989 178, doi:10.1016/j.bbrc.2015.10.138.
- 990 112. Asad, M.; Wong, M.K.; Tan, T.Z.; Choolani, M.; Low, J.; Mori, S.; Virshup, D.; Thiery, J.P.; Huang, R.Y.J.
- 991 FZD7 drives in vitro aggressiveness in stem-A subtype of ovarian cancer via regulation of non-canonical wnt/PCP pathway. *Cell Death Dis.* **2014**, *5*, e1346, doi:10.1038/cddis.2014.302.
- 993 113. Sun, L.; Wang, D.; Li, X.; Zhang, L.; Zhang, H.; Zhang, Y. Extracellular matrix protein ITGBL1 promotes
- ovarian cancer cell migration and adhesion through Wnt/PCP signaling and FAK/SRC pathway. *Biomed*.
- 995 *Pharmacother.* **2016**, *81*, 145–151, doi:10.1016/j.biopha.2016.03.053.
- 996 114. Shin, W.S.; Gim, J.; Won, S.; Lee, S.T. Biphasic regulation of tumorigenesis by PTK7 expression level in
- 997 esophageal squamous cell carcinoma. *Sci. Rep.* **2018**, *8*, 1–9, doi:10.1038/s41598-018-26957-6.
- 998 115. Berger, H.; Breuer, M.; Peradziryi, H.; Podleschny, M.; Jacob, R.; Borchers, A. PTK7 localization and protein stability is affected by canonical Wnt ligands. **2017**, doi:10.1242/jcs.198580.
- 1000 116. Wang, H.; Li, G.; Yin, Y.; Wang, J.; Wang, H.; Wei, W.; Guo, Q.; Ma, H.; Shi, Q.; Zhou, X.; et al. PTK7
- protein is decreased in epithelial ovarian carcinomas with poor prognosis. *Int. J. Clin. Exp. Pathol.* **2014**, 1002 7, 7881–7889.
- 1003 117. Golubkov, V.S.; Chekanov, A. V.; Cieplak, P.; Aleshin, A.E.; Chernov, A. V.; Zhu, W.; Radichev, I.A.;
- Zhang, D.; Dong, P.D.; Strongin, A.Y. The Wnt/planar cell polarity protein-tyrosine kinase-7 (PTK7) is a
- highly efficient proteolytic target of membrane type-1 matrix metalloproteinase: Implications in cancer

- 1006 and embryogenesis. J. Biol. Chem. 2010, 285, 35740-35749, doi:10.1074/jbc.M110.165159.
- 1007 Luo, M.; Zhou, L.; Zhan, S. jie; Cheng, L. juan; Li, R. nan; wang, H.; Liu, B.; Linghu, H. ALPL regulates 118.
- 1008 the aggressive potential of high grade serous ovarian cancer cells via a non-canonical WNT pathway. 1009 Biochem. Biophys. Res. Commun. 2019, 513, 528-533, doi:10.1016/j.bbrc.2019.04.016.
- 1010 119. Zhao, J.B.; Xue, J.F.; Zhang, W.Z.; Ren, Y.L.; Yan, D.M. Long noncoding RNA FGD5-AS1 promotes
- 1011 glioma cell proliferation, migration and invasion by regulating wnt/β-catenin pathway. Cancer Manag. 1012 Res. 2020, 12, 6187-6193, doi:10.2147/CMAR.S250284.
- 1013 Situ, J.; Zhang, H.; Jin, Z.; Li, K.; Mao, Y.; Huang, W. MicroRNA-939 directly targets HDGF to inhibit the
- 1014 aggressiveness of prostate cancer via deactivation of the WNT/β-catenin pathway. Onco. Targets. Ther. 1015 2020, 13, 4257-4270, doi:10.2147/OTT.S250101.
- 1016 121. Nomura, M.; Rainusso, N.; Lee, Y.C.; Dawson, B.; Coarfa, C.; Han, R.; Larson, J.L.; Shuck, R. 1017 Kurenbekova, L.; Yustein, J.T. Tegavivint and the β-Catenin/ALDH Axis in Chemotherapy-Resistant
- 1018 and Metastatic Osteosarcoma. J. Natl. Cancer Inst. 2019, 111, 1216–1227, doi:10.1093/jnci/djz026.
- 1019 122. Muralidhar, S.; Filia, A.; Nsengimana, J.; Pozniak, J.; O'Shea, S.J.; Diaz, J.M.; Harland, M.; Randerson-
- 1020 Moor, J.A.; Reichrath, J.; Laye, J.P.; et al. Vitamin D-VDR signaling inhibits Wnt/b-catenin-mediated 1021 melanoma progression and promotes antitumor immunity. Cancer Res. 2019, 79, 5986-5998,
- 1022 doi:10.1158/0008-5472.CAN-18-3927.
- 1023 Chen, T.; Lei, S.; Zeng, Z.; Zhang, J.; Xue, Y.; Sun, Y.; Lan, J.; Xu, S.; Mao, D.; Guo, B. Linc00261 inhibits
- 1024 metastasis and the WNT signaling pathway of pancreatic cancer by regulating a miR-552-5p/FOXO3 1025 axis. Oncol. Rep. 2020, 43, 930-942, doi:10.3892/or.2020.7480.
- 1026 Joshi, B.S.; De Beer, M.A.; Giepmans, B.N.G.; Zuhorn, I.S. Endocytosis of Extracellular Vesicles and 1027 Release of Their Cargo from Endosomes. ACS Nano 2020, 14, 4444-4455, doi:10.1021/acsnano.9b10033.
- 1028 Kalra, H.; Gangoda, L.; Fonseka, P.; Chitti, S. V.; Liem, M.; Keerthikumar, S.; Samuel, M.; Boukouris, S.;
- 1029 Al Saffar, H.; Collins, C.; et al. Extracellular vesicles containing oncogenic mutant β-catenin activate Wnt 1030 signalling pathway in the recipient cells. J. Extracell. Vesicles 2019, 8, doi:10.1080/20013078.2019.1690217.
- 1031 126. Dovrat, S.; Caspi, M.; Zilberberg, A.; Lahav, L.; Firsow, A.; Gur, H.; Rosin-Arbesfeld, R. 14-3-3 and β-
- 1032 catenin are secreted on extracellular vesicles to activate the oncogenic Wnt pathway. Mol. Oncol. 2014, 8, 1033 894–911, doi:10.1016/j.molonc.2014.03.011.
- 1034 Hu, Y.B.; Yan, C.; Mu, L.; Mi, Y.L.; Zhao, H.; Hu, H.; Li, X.L.; Tao, D.D.; Wu, Y.Q.; Gong, J.P.; et al.
- 1035 Exosomal Wnt-induced dedifferentiation of colorectal cancer cells contributes to chemotherapy 1036 resistance. Oncogene 2019, 38, 1951–1965, doi:10.1038/s41388-018-0557-9.
- 1037 Szvicsek, Z.; Oszvald, Á.; Szabó, L.; Sándor, G.O.; Kelemen, A.; Soós, A.Á.; Pálóczi, K.; Harsányi, L.;
- 1038 Tölgyes, T.; Dede, K.; et al. Extracellular vesicle release from intestinal organoids is modulated by Apc
- 1039 mutation and other colorectal cancer progression factors. Cell. Mol. Life Sci. 2019, 76, 2463-2476, 1040 doi:10.1007/s00018-019-03052-1.
- 1041 129. Hu, J.L.; Wang, W.; Lan, X.L.; Zeng, Z.C.; Liang, Y.S.; Yan, Y.R.; Song, F.Y.; Wang, F.F.; Zhu, X.H.; Liao,
- 1042 W.J.; et al. CAFs secreted exosomes promote metastasis and chemotherapy resistance by enhancing cell
- 1043 stemness and epithelial-mesenchymal transition in colorectal cancer. Mol. Cancer 2019, 18, 1-15,
- 1044 doi:10.1186/s12943-019-1019-x.
- 1045 Wang, F.W.; Cao, C.H.; Han, K.; Zhao, Y.X.; Cai, M.Y.; Xiang, Z.C.; Zhang, J.X.; Chen, J.W.; Zhong, L.P.;
- 1046 Huang, Y.; et al. APC-activated long noncoding RNA inhibits colorectal carcinoma pathogenesis
- 1047 through reduction of exosome production. J. Clin. Invest. 2019, 129, 727-743, doi:10.1172/JCI122478.
- 1048 Yamashita, T.; Budhu, A.; Forgues, M.; Xin, W.W. Activation of hepatic stem cell marker EpCAM by

- 1049 Wnt-β-catenin signaling in hepatocellular carcinoma. *Cancer Res.* **2007**, *67*, 10831–10839, 1050 doi:10.1158/0008-5472.CAN-07-0908.
- 1051 Ishiguro, K.; Yan, I.K.; Lewis-Tuffin, L.; Patel, T. Targeting Liver Cancer Stem Cells Using Engineered
- Biological Nanoparticles for the Treatment of Hepatocellular Cancer. *Hepatol. Commun.* **2020**, *4*, 298–313, doi:10.1002/hep4.1462.
- 1054 133. Faict, S.; Muller, J.; De Veirman, K.; De Bruyne, E.; Maes, K.; Vrancken, L.; Heusschen, R.; De Raeve, H.;
- Schots, R.; Vanderkerken, K.; et al. Exosomes play a role in multiple myeloma bone disease and tumor
- development by targeting osteoclasts and osteoblasts. *Blood Cancer J.* **2018**, *8*, doi:10.1038/s41408-018-1057
- 1058 134. Yu, Y.; Min, Z.; Zhou zhihang; Linhong, M.; Tao, R.; Yan, L.; Song, H. Hypoxia-induced exosomes promote hepatocellular carcinoma proliferation and metastasis via miR-1273f transfer. *Exp. Cell Res.* 2019, 385, doi:10.1016/j.yexcr.2019.111649.
- 1061 135. Chen, J.H.; Wu, A.T.H.; Bamodu, O.A.; Yadav, V.K.; Chao, T.Y.; Tzeng, Y.M.; Mukhopadhyay, D.; Hsiao, 1062 M.; Lee, J.C. Ovatodiolide suppresses oral cancer malignancy by down-regulating exosomal mir-
- 1063 21/STAT3/β-catenin cargo and preventing oncogenic transformation of normal gingival fibroblasts.

 1064 *Cancers (Basel).* **2020**, *12*, doi:10.3390/cancers12010056.
- 1004 *Cancers (Basel).* **2020**, *12*, doi:10.3390/cancers12010056.
- 136. Xia, Y.; Wei, K.; Hu, L.Q.; Zhou, C.R.; Lu, Z. Bin; Zhan, G.S.; Pan, X.L.; Pan, C.F.; Wang, J.; Wen, W.; et al. Exosome-mediated transfer of miR-1260b promotes cell invasion through Wnt/β-catenin signaling pathway in lung adenocarcinoma. *J. Cell. Physiol.* **2020**, 235, 6843–6853, doi:10.1002/jcp.29578.
- 137. Harada, T.; Yamamoto, H.; Kishida, S.; Kishida, M.; Awada, C.; Takao, T.; Kikuchi, A. Wnt5b-associated exosomes promote cancer cell migration and proliferation. *Cancer Sci.* **2017**, *108*, 42–52, doi:10.1111/cas.13109.
- 1071 138. Luga, V.; Zhang, L.; Viloria-Petit, A.M.; Ogunjimi, A.A.; Inanlou, M.R.; Chiu, E.; Buchanan, M.; Hosein, 1072 A.N.; Basik, M.; Wrana, J.L. Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in 1073 breast cancer cell migration. *Cell* 2012, 151, 1542–1556, doi:10.1016/j.cell.2012.11.024.
- 1074 139. Lombardo, G.; Gili, M.; Grange, C.; Cavallari, C.; Dentelli, P.; Togliatto, G.; Taverna, D.; Camussi, G.; 1075 Brizzi, M.F. IL-3R-alpha blockade inhibits tumor endothelial cell-derived extracellular vesicle (EV)-mediated vessel formation by targeting the β-catenin pathway. *Oncogene* 2018, 37, 1175–1191,
- 1077 doi:10.1038/s41388-017-0034-x.
- 140. Lopatina, T.; Grange, C.; Cavallari, C.; Navarro-Tableros, V.; Lombardo, G.; Rosso, A.; Cedrino, M.; 1079

 Pomatto, M.A.C.; Koni, M.; Veneziano, F.; et al. Targeting IL-3Rα on tumor-derived endothelial cells blunts metastatic spread of triple-negative breast cancer via extracellular vesicle reprogramming.

 Oncogenesis 2020, 9, 90, doi:10.1038/s41389-020-00274-y.
- 1082 141. Kamerkar, S.; Lebleu, V.S.; Sugimoto, H.; Yang, S.; Ruivo, C.F.; Melo, S.A.; Lee, J.J.; Kalluri, R. Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. *Nature* **2017**, *546*, 498–503, doi:10.1038/nature22341.
- 1085 142. Matsuda, A.; Ishiguro, K.; Yan, I.K.; Patel, T. Extracellular Vesicle-Based Therapeutic Targeting of β-1086 Catenin to Modulate Anticancer Immune Responses in Hepatocellular Cancer. *Hepatol. Commun.* 2019, 3, 525–541, doi:10.1002/hep4.1311.
- 1088 143. Cavallari, C.; Camussi, G.; Brizzi, M.F. Extracellular Vesicles in the Tumour Microenvironment: Eclectic Supervisors. *Int. J. Mol. Sci.* 2020, 21, 6768, doi:10.3390/ijms21186768.
- 1090 144. Han, L.; Lam, E.W.F.; Sun, Y. Extracellular vesicles in the tumor microenvironment: Old stories, but new tales. *Mol. Cancer* 2019, *18*, 1–14, doi:10.1186/s12943-019-0980-8.

1094

Lucchetti, D.; Tenore, C.R.; Colella, F.; Sgambato, A. Extracellular Vesicles and Cancer: A Focus on. **2020**, 1–19.



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