

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

A review of the nature, role and control of lithobionts on stone cultural heritage: weighing-up and managing biodeterioration and bioprotection

This is a pre print version of the following article:

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1755615> since 2020-09-17T13:51:03Z

Published version:

DOI:10.1007/s11274-020-02878-3

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

1 **The nature, role and control of lithobionts on stone cultural heritage: weighing-up and**
2 **managing biodeterioration and bioprotection**

3 Sergio Enrico Favero-Longo^{1,*} & Heather A Viles²

4 ¹Università degli Studi di Torino
5 Dipartimento di Scienze della Vita e Biologia dei Sistemi
6 Viale Mattioli 25, 10125 Torino, Italy
7 Tel. +390116705972
8 Fax +390116705962
9 sergio.favero@unito.it
10 orcid.org/0000-0001-7129-5975

11
12 ²University of Oxford,
13 School of Geography and the Environment
14 South Parks Road, Oxford, OX1 3QY, Oxford, UK
15 Tel: +44 1865 285070
16 heather.viles@ouce.ox.ac.uk
17 orcid.org/0000-0002-2444-1295

18
19 *corresponding author: sergio.favero@unito.it
20

21

22 **Acknowledgements**

23 The authors thank Prof. em. Ian Maddox for the kind invitation to contribute this review article.

24 SEFL and HV are also grateful to institutions and authorities which allowed their research work in
25 the following heritage sites represented in Figures 1 and 3: Residences of the Royal House of Savoy
26 (Polo Museale del Piemonte, Italy); Archaeological Park of Seradina and Bedolina (Comune di
27 Capo di Ponte, Valle Camonica, Italy), Soprintendenza Archeologia della Lombardia (Milano,
28 Italy); Gardens of Boboli (Gallerie degli Uffizi, Italy); the graveyard of Gressoney-La-Trinité
29 (Comune di Gressoney-La-Trinité, Italy); North Grotto Temple (Dunhuang Academy, China).

30 SEFL also thanks prof. Rosanna Piervittori, dr. Enrica Matteucci (Università di Torino), dr. Ada
31 Roccardi (Istituto Superiore per la Conservazione e il Restauro, Roma), and all the colleagues of the
32 Working Group for Cultural Heritage of the Italian Lichen Society for many stimulating discussions
33 on lichens and other lithobionts in the last two decades.

34

35 **Abstract**

36 Lithobionts (rock-dwelling organisms) have been recognized as agents of aesthetic and physico-
37 chemical deterioration of stonework, and removal of microorganisms from cultural heritage stone
38 surfaces (CHSS) is widely considered a necessary step in conservation interventions. On the other
39 hand, lithobiontic communities can help integrate CHSS with their environmental setting and
40 enhance biodiversity. Moreover, in some cases bioprotective effects have been reported and even
41 interpreted as potential biotechnological solutions for conservation.

42 This paper reviews the plethora of traditional and innovative methodologies to characterize
43 lithobionts on CHSS in terms of biodiversity, interaction with the stone substrate and impacts on
44 durability. In order to develop the best management and conservation strategies for CHSS, such
45 diagnosis should be acquired on a case-by-case basis, as generalized approaches are unlikely to be
46 suitable for all lithobionts, lithologies, environmental and cultural contexts or types of stonework.

47 Strategies to control biodeteriogenic lithobionts on CHSS should similarly be based on
48 experimental evaluation of their efficacy, including long term monitoring of their effects on
49 bioreceptivity. This review examines what is known about the efficacy of control methods based on
50 traditional-commercial biocides, as well as those based on innovative application of substances of
51 plant and microbial origin, and physical techniques. A framework for providing a balanced
52 scientific assessment of the role of lithobionts on CHSS and integrating this knowledge into
53 management and conservation decision-making is presented.

54

55 **Keywords:** biocide, biodiversity, biofilm, bioreceptivity, conservation of cultural heritage stone
56 surfaces, stone cleaning, stone durability

57 **The issue of lithobionts on the stone cultural heritage: an introduction**

58 The preservation and transmission to future generations of stone cultural heritage (objects, buildings
59 and sites carved in, or built from, stone), including rock art, archaeological and historical
60 monuments and artistic stonework, are globally shared and signed duties, which need to be
61 combined with the protection and conservation of natural heritage (Convention Concerning the
62 Protection of the World Cultural and Natural Heritage 1972). As with other global issues in today's
63 changing world, scientific advances can inform effective and environmentally conscious
64 management, based on correct identification of threats, and development of sustainable solutions
65 (Ferretti and Comino 2015; Charter and Tischner 2017). The growth of lithobionts (i.e. rock-
66 dwelling organisms) on cultural heritage stone surfaces (CHSS; Fig. 1) has long been associated
67 with biodeterioration, which can be defined as any undesirable change in the properties of a
68 material caused by the vital activities of organisms (Hueck 1965), and recognized as a threat to
69 conservation (Caneva et al. 2008). This is not surprising given widespread evidence of
70 biogeophysical processes (such as mechanical fracturing and disruption of rocks and minerals by
71 living structures) and biogeochemical processes (chemical destabilization and compositional change
72 of rocks and minerals by metabolic processes and products), which are vital to pedogenesis
73 (Silverman 1979; Totsche et al. 2010). Plenty of field and experimental data demonstrate rock
74 weathering by macro- and micro-organisms (bioweathering), such as vascular plants (Pawlik et al.
75 2016) and bryophytes (Ricci and Altieri 2008), lichenized- (Seaward 2015) and non-lichenized
76 fungi (Gadd 2017), photo- (Albertano 2012) and chemo- lithotrophic bacteria (Mapelli et al. 2012).
77 Coupled with the desire to remove lithobionts from heritage stone surfaces for aesthetic reasons
78 (because they can produce unsightly discoloration and obscure important carved details), such
79 biodeteriorative roles strengthen the case for conservation interventions (Pinna 2017).

80 On the other hand, lithobiontic communities have also been seen as enhancing biodiversity, and in
81 some cases bringing positive aesthetic characteristics to CHSS (see section *Factors affecting the*
82 *opportunity to remove (or preserve) lithobionts*). Furthermore, the last two decades have seen an

83 increasing interest in whether lithobiontic communities can in some circumstances also act as a
84 bioprotective layer, covering stone surfaces and limiting weathering processes driven by abiotic
85 factors, such as meteorological forces and air pollutants (Carter and Viles 2005; McIlroy de la Rosa
86 et al. 2012). Such findings, mixed with advances in investigating biomediated approaches
87 (microbial biocementation) which can promote the consolidation of stone (Fernandes 2006; Wang
88 et al. 2016; Shraddha and Darshan 2019), have contributed to the recent proposal that bioprotection
89 of buildings and cultural heritage by lithobionts may be a sustainable strategy (Gadd and Dyer
90 2017). It is important not to see contrasting research findings on biodeterioration and bioprotection
91 as conflicting positions, as they likely represent different aspects of complex interactions within and
92 between natural or cultural heritage ecosystems (see section *Biodeteriorative and/or bioprotective*
93 *effects*). Misunderstandings on this point may indeed critically impact decisions on the management
94 of lithobionts on CHSS, which should not be based on generalized views of biodeterioration vs
95 bioprotection as ideologically opposed ‘schools of thought’, but on diagnostic analyses targeting
96 each case. Advances in molecular, microscopy and spectroscopy methods since the 1990s have
97 strongly improved the likelihood of characterizing the diversity of lithobionts on CHSS, and their
98 related biogeophysical and biogeochemical processes (Piñar and Sterflinger 2018; Sanmartín et al.
99 2018; Schröder et al. 2019). In the following section, colonization patterns and physico-chemical
100 processes driven by the major groups of lithobionts are exemplified, together with a critical
101 evaluation of what diagnostic approach is required to reliably unveil their biodeterioration and/or
102 bioprotection potential on CHSS. In the second part, approaches to manage lithobionts on CHSS are
103 outlined, and a potential decision framework based on experimental assessment and monitoring of
104 both the roles of lithobionts and the efficacy of control strategies is introduced.

105

106

107

108 **Patterns and impact of lithobiontic colonization**

109 *Stone bioreceptivity and biogeophysical and biogeochemical impacts of lithobionts*

110 Natural rock surfaces and CHSS provide interfaces between geological materials, air and water
111 which are invariably colonised by lithobionts (Gorbushina and Broughton 2009). Intrinsic physico-
112 chemical properties of stone materials (e.g. surface roughness, porosity, mineral, composition) are
113 the primary factors determining their bioreceptivity, i.e. their aptitude to be colonized by living
114 organisms (Guillitte 1995; Miller et al. 2010). The rate of supply of biological particles, usually
115 resistant structures (spores, pollens) dispersed as bioaerosol, is the main complementary factor
116 needed to start and support colonization dynamics (Mandrioli et al. 2003). Other extrinsic factors,
117 such as climatic conditions and the availability of organic nutrients, influence the composition and
118 structure of lithobiontic communities on CHSS, with their network of nutritional interactions
119 (autotrophs/producers; heterotrophs/destroyers and consumers) (Caneva et al. 2008).

120 An index to quantify bioreceptivity has been proposed to evaluate the performance of any
121 construction material and support management decisions in the ornamental stone industry which
122 can be applied to CHSS (Vázquez-Nion et al. 2018). In the suggested protocol, the growth of a
123 standard photoautotrophic community (bryophyta, green algae, cyanobacteria) on coupons of
124 different lithologies is compared by fluorimetry (to quantify the amount of chlorophyll *a*) and
125 spectrophotometry (to quantify colour change. Such an approach reflects the fact that aesthetic
126 appearance (colour, visibility of artistic details) is important for perceptions of the value of CHSS
127 (Brimblecombe and Grossi 2005). On the other hand, such an index does not take account of the
128 three-dimensional phenomenon of lithobiontic colonization of CHSS, in which the hidden
129 dimension, below the surface, represents the major interface between biotic and mineral
130 components.

131 Across a range of spatial scales, higher plants, bryophytes, lichens and microorganisms anchor or
132 adhere to mineral substrates, including CHSS, in order to provide stability, exploit water and/or
133 nutrients, and as a consequence modify the physico-chemical properties of the substrates. The root

134 systems of woody plants growing directly on pavements or masonries, and, in the case of tree, in
135 proximity to building foundations and hypogean structures, can exert mechanical forces and cause
136 superficial and structural damage, especially in archaeological sites (Caneva et al. 2009; Bartoli et
137 al. 2017). The penetration of slender rhizoids and protonemata of mosses (even though not enforced
138 by lignin) have been related to mechanical damage of mosaics and wall paintings (Saiz-Jimenez et
139 al. 1991; Ricci and Altieri 2008). In the case of lichens (Fig. 2 a-d), the penetration of mycobiont
140 hyphae beneath epilithic crustose thalli, and of the anchoring points of rhizinae and haptera of
141 foliose and fruticose lichens, respectively, has been microscopically characterized within many
142 different stone materials (de los Ríos and Ascaso 2005; Salvadori and Casanova-Municchia 2016).
143 Different patterns of penetration are found depending on the mineralogical and microstructural
144 features of different lithologies, and the species involved, with hyphal organization, spread, and
145 depths ranging from a few microns to several millimeters (Favero-Longo et al. 2005; Scarciglia et
146 al. 2012; Sohrabi et al. 2017). Moreover, some lichen species display an endolithic habit, with the
147 thallus (including the photobiont layer) growing entirely within the rock substrate, exploiting
148 internal cracks (chasmo-endolithic) or intrinsic porosities (crypto-endolithic), or actively dissolving
149 minerals (eu-endolithic) (Pinna et al. 1998; Casanova-Municchia et al. 2014; Favero-Longo et al.
150 2015).

151 Similar ranges of growth, from fully epilithic to fully endolithic, also characterize autotrophic and
152 heterotrophic microorganisms usually organized as biofilms, which are adapted to every kind and
153 level of environmental stress in terms of temperature, water and nutrient availability, and solar
154 irradiation (Gorbushina and Broughton 2009). Archaea, bacteria and eukaryotic microbes, as non-
155 lichenized fungi, widely live embedded in an extracellular matrix of biopolymers, usually known as
156 as EPS (Extracellular Polymeric Substances), which is dominated by polysaccharides, but also
157 contains (glyco-)proteins, glycolipids and DNA (Flemming et al. 2007, 2017). High hydration of
158 EPS makes biofilms functionally active and resistant environments, favouring the nutrition,
159 communication and defence of microbes, and also drives their physico-chemical interactions with

160 the substrate. Cyclic hydration and dehydration of biofilms cause volume changes and, as a
161 consequence, pressures at their points of adhesion, causing disaggregation and detachments (Negi
162 and Sarethy 2019). A similar mechanical action is exerted by mosses and lichens, whose water
163 status varies passively with environmental conditions (poikilohydric organisms; Seaward 2015).

164 The covering of rock surfaces and CHSS by coloured dark patinas and thalli also influences the
165 thermal behaviour of the surface and consequent physical stress (Garty 1990; Carter and Viles
166 2004). In parallel, water availability at the lithobiont-rock interface favours the mobilization of
167 metabolites and mineral ions, supporting chemical modification and sometimes dissolution of the
168 original mineral constituents and the precipitation of new (bio-)minerals (Banfield et al. 1999).

169 Accordingly, protons excreted by plant root tips induce cationic exchange and contribute to ion
170 mobilization from the contacted minerals, and their chemical modification (Caneva et al. 2008).

171 Bryophytes, like algae and cyanobacteria, can favour the precipitation of carbonates, as a
172 consequence of photosynthetic removal of CO₂ and their ability to solubilize and bind Ca, a
173 phenomenon reported for natural springs, but also for fountains and monuments in humid regions
174 (Bolívar and Sánchez-Castillo 1997; Ortega-Morales et al. 2000; Crispim and Gaylarde 2005; Ricci
175 and Altieri 2008). Several lichenized and non-lichenized fungi secrete a large variety of primary
176 and secondary metabolites with acidic and chelating functions, supporting acidolysis and
177 complexolysis (Piervittori et al. 2009; Gadd 2017). The biomineralization at the lichen-substrate
178 interface of different oxalates (e.g. Ca-, Mg-, Fe-, Mn-oxalate), depending on the rock metal
179 contents, has frequently been cited as evidence of the direct influence of the mycobiont secretion of
180 oxalic acid on colonized surfaces (Adamo and Violante 2000; Chen et al. 2000; Gadd et al. 2014).

181 In particular, calcium oxalates (whewellite and weddellite) have been reported on a wide range of
182 lithologies, even those poor in Ca, because of their very low solubility in comparison with other
183 oxalate species (Fig. 2 e-f; Favero-Longo et al. 2005), and they are often found on CHSS colonized
184 by certain lichens (Edwards et al. 2003; Pena-Pozo et al. 2018; Tonon et al. 2019). The insolubility
185 of Ca-oxalates also accounts for their long persistence on CHSS, where their biological origin has

186 been suggested as an alternative to a chemical origin (e.g. due to degradation of restoration
187 products) even in the absence of viable lithobiontic communities (Caneva et al. 1993). Mineral
188 leaching by other fungal primary metabolites (e.g. citric and malic acids) has also been documented
189 (Wei et al. 2012; Sazanova et al. 2016), but they have generally received less attention in the field
190 of CHSS in comparison to oxalates because of the absence of clear or persistent traces. Mineral
191 leaching activity by complexation has also been reported for some depsides, depsidones and
192 pulvinic acid derivatives (secondary metabolites exclusive to lichens) although their low solubility
193 may limit their impact (Ascaso et al. 1976; Haas and Purvis 2006; Favero-Longo et al. 2013). All
194 these metabolites, as well as the involvement of EPS, seem to contribute to the weathering of
195 silicate minerals into clays, as TEM investigations have showed for the lichen-driven
196 vermiculization of biotite and other cases (Cuadros 2017), and in several cases to the dissolution of
197 carbonates (Pinheiro et al. 2019).

198 Different processes need to be invoked to explain the euendolithic behaviour of certain lichen
199 species, as they do not secrete either oxalic acid or the above mentioned lichen secondary
200 metabolites (Pinna et al. 1998). In some cases respiration-induced acidification of the substrate has
201 been shown to be sufficient to provoke pitting activity (formation of sub-millimetric cavities on the
202 surface) (Weber et al. 2011), but the involvement of siderophore-like compounds has also been
203 hypothesized, as these complexing compounds, involved in iron nutrition, also scavenge calcium if
204 iron availability is low (Favero-Longo et al. 2011). Similarly to endolithic lichens, microcolonial
205 fungi (MCF), which are a group of fungi tolerant of the extreme stress of exposed substrates and
206 thus colonizing bare rock surfaces, including CHSS (Sterflinger 2010), are responsible for
207 dissolution and pitting phenomena on carbonate and silicate rocks, but the processes responsible
208 still need to be clarified (De Leo et al. 2019). The rigidity of their cell wall due to melanin
209 deposition has been related to their penetration ability (Sterflinger and Kumbein 1997), but ion
210 mobilization by chelating molecules (Favero-Longo et al. 2011) and/or corrosive EPS containing
211 pullulan and galactofuromannan as the main constituents (Breitenbach et al. 2018) may be involved.

212 The strong negative charge of cyanobacterial EPS, due to uronic acids and sulphate groups, also
213 contributes to the dissolution of cations from colonized substrata and their microbial adsorption
214 (Bellezza et al. 2006; Albertano 2012). Sulfuric, nitrous and nitric acids are released by
215 chemolithotrophic (sulphur-oxidizing, nitrifying) bacteria and (ammonia-oxidizing) archaea,
216 through enzyme catalyzed oxidation of inorganic compounds, such as ammonia, elemental sulphur
217 and hydrogen sulphide, and are likely to have impacts on CHSS (Warscheid and Braams 2000;
218 Zhang et al. 2019).

219

220 *Biodeteriorative and/or bioprotective effects*

221 All the above-mentioned (and many other) patterns of lithobiont-substrate interactions, in terms of
222 biogeophysical and biogeochemical processes, have been increasingly detailed in the last two-three
223 decades and supported by a huge number of study cases, including CHSS. This has certainly helped
224 clarify the worldwide distribution and impacts of certain phenomena, but, in some cases,
225 experimental evidence obtained by advanced, 21st century technologies simply confirms early
226 insights dating back to the first part of the 20th century and before. For example, early descriptions
227 and accurate microscopic observations of lichen growth within rocks (Gümbel, 1856; Fry 1924
228 1927), as well as the recognition of lichen-driven chemical processes due to oxalic acid secretion
229 and/or a respiration-induced acidification (Slater, 1856; Uloth 1861; Sollas, 1880; Smith 1921),
230 provided convincing evidence of lichen biogeophysical and biogeochemical processes and are still
231 widely quoted today. However, the advent in recent years of a wide range of portable, non-
232 destructive methods now permits sophisticated measurements (such as spectroscopic analysis)
233 which can characterize the presence of metabolites and biominerals on/within CHSS in situ
234 (Maguregui et al. 2012; Costantini et al. 2018). Advances in molecular biology, and in particular the
235 diagnostic power of next-generation sequencing, have highlighted the huge variety of
236 microorganisms which can occur together on and beneath rock surfaces and CHSS, only partially

237 detectable by microscopy and culturing approaches, and not always directly related to more visible
238 surface colonizers (Bjelland et al. 2011; Piñar and Sterflinger 2018; Trovão et al. 2019).

239 On the other hand, whilst the observation and detection of (micro-)organisms, their structures and
240 metabolites, or their molecular signs, certainly implies present or past interactions with rock
241 substrates and CHSS, it does not necessarily mean that they play a biodeteriorative role as is often
242 routinely thought. In this context, DNA-metabarcoding analyses particularly need careful evaluation
243 to decipher which microorganisms are growing on and actively interacting with the substrates, and
244 which simply occur as past traces, dormant structures or episodic (dust) contaminants (Marvasi et
245 al. 2019). Implementation of molecular analysis pipelines, including the storage and effective
246 access of datasets dedicated to the biodeterioration research area, also represent an ongoing
247 challenge (Sterflinger et al. 2018).

248 Whilst there is incontrovertible evidence of mineral disintegration, chemical modification and
249 leaching by lithobionts, other studies have highlighted bioprotective activities, which were also
250 hypothesized many years ago (Krumbein 1968). Such hypotheses were based on early macroscopic
251 observations of differential erosion rates with and without lithobionts, suggesting an umbrella-like
252 protective effect (Mottershead and Lucas 1990; Özvan et al. 2015). Such interpretations of
253 lithobionts as forming a physical barrier against other weathering factors have been supported by
254 the quantification of lower solutional weathering from lithobiont (lichen)-covered limestone slates
255 in comparison with uncolonized controls after 1 year of exposure in the humid climate of Ireland
256 (McIlroy de la Rosa 2014). Ivy (*Hedera helix*) has been found to also have an umbrella-like impact
257 and protect historic limestone walls from pollutants (which can lead to soiling and deterioration)
258 (Sternberg et al. 2011). Additionally, experimental studies illustrate that *Hedera helix* can also
259 protect vulnerable historic stone walls from freeze-thaw damage through modifying microclimatic
260 conditions (Coombes et al. 2018). In drier climates, rock surface microorganisms have also been
261 demonstrated to be chemically involved in the development of rock coatings, which stabilize the
262 rock surfaces and contribute to their long term preservation (Taylor-George et al. 1983; Dorn 2013).

263 Nanometer-scale transport of Fe and Mn accumulated in bacterial sheaths and fixed on clay
264 minerals contributes to the polygenetic formation of rock varnish (Dorn 2013). In arid Jordan,
265 biofilms of cyanobacteria and fungi have been shown to contribute to case-hardening of sandstone
266 by aiding cementation (Viles and Goudie 2004), and in cold arid central Antarctica, endolithic
267 lichens and their EPS have been found to biomineralize iron oxides (Guglielmin et al. 2011). Such
268 effects are also found in wetter climates, for example in the UK where the deposition by epilithic
269 lichens of silica-rich layers within cracks and along mineral boundaries has been observed on
270 granite outcrops alongside biophysical and biochemical weathering impacts (Lee and Parsons
271 1999). Biomineralization of oxalates has also been considered to produce a potential protective
272 shield against abiotic weathering agents, such as wind and runoff (Souza-Egipsy et al. 2004), and
273 Ca-oxalates have often been proposed as producing potentially protective coatings for CHSS (e.g.
274 Rampazzi 2019).

275 Multiple biodeteriorative and bioprotective roles of lithobionts have often been proposed. A notable
276 example is the analysis of roles of the lichen *Verrucaria rubrocincta* on caliche plates in the
277 Sonoran desert (Bungartz et al. 2004; Garvie et al. 2008): simultaneous biodeterioration of the rock
278 substrate and a counterbalancing biomineralization of an upper protecting layer of fine-grained
279 micrite have been recognized and characterized by isotopic analyses. Similarly, calcite dissolution
280 and biomineralization of neoformed calcite around hyphae have been described for non-lichenized
281 fungi (Fomina et al. 2010). Such recognition of both deteriorative and protective effects of
282 lithobionts, and a deep evaluation of the prevailing process, may become crucial in future studies of
283 lithobionts on CHSS (Bartoli et al. 2014). Observations of lithobiont (lichen) communities on tuff
284 churches in Cappadocia showed the deteriorogenic activity of penetrating hyphae, whilst some
285 bioprotective effects were simultaneously envisaged, leaving uncertainty about whether or not the
286 growths should be removed (Casanova-Municchia et al. 2018). Such a case study exemplifies the
287 need for case-specific information on the multiple interactions of lithobionts and CHSS, their spatial
288 patterning and their persistence over time, but also on their actual influence on stone durability,

289 before appropriate management decisions can be made. Indices to evaluate and quantify the
290 potential biodeteriogenic impact of plants (Signorini 1996), lichens (Gazzano et al. 2009) and
291 microorganisms (Gazzano et al. 2011) have been proposed, but are more oriented to a comparative
292 evaluation of the degree of physico-chemical interaction of different species with the substrate,
293 rather than to quantitatively evaluate their impact on the CHSS (Pinna 2014). As a framework
294 proposal, Table 1 summarizes a range of biodeteriorative and bioprotective roles reported for
295 various types of lithobiontic organisms. It is worth remarking that each (potential) role has to be
296 considered species-specific and cannot be generalized for all the members of each group; moreover
297 it also depends on the lithology and the environmental conditions (see the next sub-section).

298

299 *Lithobionts and stone durability*

300 Biodeteriorative and bioprotective processes may or may not balance out, and thus produce
301 complex impacts on the intrinsic stone properties relevant for durability, which can be defined as
302 the capacity of a building material to maintain its size, shape, strength and aesthetic appearance
303 over time (Bell 1993). In particular, knowledge of lithobiont-rock interactions should be combined
304 with quantitative investigation of their impacts on petrophysical properties, including porosity,
305 presence of swelling clay minerals, compactness, water absorption, presence and distribution of
306 anisotropies, compressive strength, and surface hardness (Molina et al. 2013; Wilhelm et al. 2016).
307 Relatively few studies have yet been made of the impacts of lithobionts on durability and resilience
308 of CHSS, although some research has focused on (bio-)geomorphological studies of natural rock
309 outcrops (Viles 2019). The results seem to depend strongly on lithology. For example, lichen
310 colonization has been found to correlate with surface hardening and a reduction of water absorption
311 on tuff (Garcia-Vallès et al. 2003), with a reduction of surface hardness on gneiss (McCarroll and
312 Viles 1995; Favero-Longo et al. 2015), and with variable patterns of surface hardening or softening,
313 and increased water absorption after the scraping of thalli from the surface on limestone (Morando
314 et al. 2017). Such variability suggests the need for tailored investigations to cover a range of

315 lithologies, different lichen species, and other, even more overlooked, lithobionts. Experimental
316 datasets are needed to test conceptual models on the balance between biodeterioration and
317 bioprotective processes and the net impact of lithobionts (lichens, in particular) on stone surfaces
318 (McIlroy de la Rosa et al. 2012). In this regard, one particularly important future research direction
319 is elucidating the impact of lithobionts on their substrate beyond their life-span. Surfaces once
320 covered by lithobionts which have been removed by conservation interventions, or have simply
321 died, may indeed display a long term protection effect due to past biogeochemical processes (e.g.
322 protection by a biomineral deposit, cementation, pore-filling by biogenic silica or varnish).
323 Conversely, such surfaces might also be more prone to disintegration following the loss of
324 biological structures which bound the disentangled mineral fragments. The latter scenario appears
325 particularly critical where stone surface details are a crucial component of the heritage values, as in
326 the case of carved rock-art (Tratebas 2004; Favero-Longo et al. 2019).

327 Developments in laboratory experiments using cultured model lithobiontic organisms, genetically
328 modified lineages, and the combination of closed and open experimental systems provide useful
329 new approaches to disentangling the impacts of lithobionts on CHSS. For example, several studies
330 have cultivated lithobionts on stone coupons and evaluated their impact on stone properties relevant
331 for durability (Favero-Longo et al. 2009; Villa et al. 2015; Pokharel et al. 2017; Seiffert et al. 2016).
332 In this context, it is very important to standardize protocols used in the characterization of
333 biodeteriorative (and bioprotective) processes, using approaches such as microscopic,
334 spectroscopic, culture-based, and ‘omics’ methods, in order to make scientific data truly comparable
335 and offer more reliable support for management decisions (Pinna 2014; Sterflinger et al. 2018).
336 Research on lithobiontic impacts on CHSS should also consider predicted impacts of climate
337 change on lithobiont colonization and growth (Davidson et al. 2018). Changes in temperature and
338 precipitation regimes, beyond their direct impact on CHSS, have been hypothesized to affect
339 lithobiont communities, causing potential shifts from biodeterioration to bioprotective impacts, and
340 vice versa (Viles and Cutler 2012; Fatorić & Seekamp 2017). As an example, Gómez-Bolea et al.

341 (2012) predicted that lithobiotic biomass on CHSS should increase in northern Europe whilst
342 decreasing in southern Europe because of contrasting trends in precipitation. Recent laboratory
343 experiments simulating changing water regimes and increased CO₂ concentrations have confirmed
344 the suggested impacts on biofilm composition and a shift towards biodeterioration effects (Prieto et
345 al. 2020). From a different point of view, subaerial biofilms have also been noted as potentially
346 important climate regulators, as agents of carbon sequestration, biogeochemical cycles and
347 elemental transformations (Villa et al. 2016).

348

349 **Approaches to managing lithobionts on CHSS**

350 *Factors affecting the opportunity to remove (or preserve) lithobionts*

351 Aspects of culture and tradition have shaped and continue to shape perceptions of the growth of
352 lithobionts on CHSS. Ruins covered by plants and other lithobionts, recognized as evidence of
353 natural decay, are a symbol of the European Romantic taste of the 19th century (Stanford 2000;
354 Huyssen 2006). Similarly, an abundance of lithobionts on some types of heritage sites, such as a
355 Celtic graveyard or a Shinto shrine, can be seen in a positive light today, especially in places where
356 local tradition tends to appreciate, rather than worry about, the merging of CHSS and nature,
357 sharing a common acceptance of human impermanence (Fitzsimons et al. 2012). In most cases,
358 however, alterations in CHSS caused by both biotic and abiotic drivers are considered to be
359 negative, reducing heritage values (Brimblecombe and Grossi 2005; Prieto et al. 2007). As a
360 consequence, removal of lithobionts is usually recognized as a necessary part of conservation
361 interventions (Pinna 2017). However, scientific evidence justifying the removal of lithobionts is
362 often missing.

363 In order to make more informed decisions about whether to remove (or preserve, or even encourage
364 further) lithobionts, aesthetic evaluations should be combined with data on the impacts of
365 lithobionts on stone durability, based on an analysis of the balance between biodeteriorative and

366 bioprotective processes. Such an approach is not in conflict with the priority often given (at least in
367 some countries) to aesthetic evaluations as the basis of management strategies for heritage sites, but
368 enables a broader, more balanced assessments of risks vs benefits. By way of an example, in cases
369 where lithobionts are shown to be predominantly bioprotective, their removal might cause a
370 decrease in durability of CHSS: if the aesthetic damage they cause is more severe, however, and
371 they are removed, strategies to substitute their bioprotective function should be considered. In cases
372 where the aesthetic damage caused by lithobionts is minor, and there is evidence of a bioprotective
373 effect, or of only minor biodeterioration impact, a recommendation could be made to preserve the
374 lithobionts and promote their biodiversity as an additional cultural value of the heritage site (Nimis
375 et al. 1992; Steinbauer et al. 2013). Such an approach may be particularly valuable in the case of
376 CHSS immersed in the natural environment, where the removal of a mature-climactic lithobiontic
377 community, similar to that found on surrounding natural outcrops, is often followed by a rapid
378 recolonization by a simplified community of ‘banal’ species (Nascimbene et al. 2009). The
379 inclusion of biodiversity in the concept of cultural value of a heritage site agrees with official
380 measures addressing parallel conservation of cultural and natural heritage (e.g. Italian Ministry for
381 Cultural Heritage and Activities 2004: the Italian code of cultural heritage and landscape; UNESCO
382 2014: Florence Declaration). This may be particularly important for cemeteries, monumental or
383 archaeological sites which are hotspots of lithology or microhabitat diversity and consequently host
384 strongly heterogeneous lithobiontic and plant communities, making them easily accessible to the
385 public (e.g. Nimis et al. 1987; Cicinelli et al. 2018; Löki et al. 2019). In other cases, cultural
386 heritage sites have been clearly identified as preferential sites for the preservation of threatened
387 species or even their re-introduction (Valkó et al. 2018; BLS 2020).

388 CHSS management should consider all facets of lithobiontic growths before automatically
389 removing lithobionts to conserve and present heritage sites. In all cases, once the decision has been
390 taken to remove lithobionts, the adoption of an effective strategy to clean the CHSS is an absolute
391 priority (Pinna 2017). A major issue for CHSS management is indeed the frequent need to repeat

392 treatments to control lithobionts, which in most cases are stress-tolerant (micro-)organisms well-
393 equipped to thwart human attempts to remove them and prevent their regrowth.

394

395 *Direct and indirect strategies to control lithobionts*

396 Two main strategies to control lithobionts on CHSS can be distinguished, although their choice
397 should not be considered exclusive and they may be used in combination: first, an indirect approach
398 focused on the control of microenvironmental parameters which favour/allow their growth, and
399 second, direct intervention to remove them from CHSS by physical and/or mechanical and/or
400 chemical methods (Pinna 2017). The first approach is based on the fact that the presence of
401 lithobiontic communities, and, in particular, of certain biodeteriogenic species, depends on the
402 availability of suitable microenvironmental conditions, allowing their establishment, expansion, and
403 reproduction. At any one heritage site, different CHSS often represent diverse ecological micro-
404 niches, hosting different lithobiontic communities which interact with the substrate and influence its
405 conservation in different ways (Marques et al. 2016; Tonon et al. 2019). Knowledge and monitoring
406 of (micro-)environmental parameters favouring or discouraging lithobiontic communities, and in
407 particular, deteriogenic species, can help recognise and achieve the best conservation solutions
408 (Caneva et al. 2016; Schumacher and Gorbushina 2020). For example, the development of
409 phototrophic biofilms on hypogean CHSS, and even on outdoor surfaces lit by artificial lighting,
410 may be controlled through careful illumination strategies, based on modifying the spectral emission
411 of lamps to affect microbial biomass, colour and EPS production (Albertano and Bruno 2003;
412 Sanmartín et al. 2017). Careful management of higher plants in heritage sites may limit their
413 shading of CHSS, with the consequent increase in humidity, and their supply of nutrients, both of
414 which favour lithobiontic growth (Salvadori and Charola 2011). Such ecologically-based
415 approaches may also be used to choose the most appropriate contemporary building materials
416 (Caneva et al. 2008).

417 Direct intervention to clean CHSS should satisfy the main aims of removing the lithobiotic
418 communities and preventing (re-)colonization processes by limiting surface bioreceptivity. These
419 objectives have been variously pursued by combining several mechanical, physical and chemical
420 approaches, already reviewed in detail elsewhere (Pinna 2017). The crucial point is that lithobiotic
421 colonization usually extends well beneath the rock surface and microbial structures generally show
422 a strong capacity to recover from disturbance, and so many cleaning interventions without
423 subsequent biocide treatment are usually followed by rapid recolonization (Sohrabi et al. 2016).
424 Removal by (pressurized) water and brushing may be effective in the case of superficial, subaerial
425 biofilms (Sanmartín et al. 2020), but in other situations they may spread microbial structures,
426 pushing them deeper within the substrate (Pinna 2017). Valuable protocols for the physical removal
427 of lithobionts by laser ablation have been drawn up in the last decade (Sanz et al. 2017), but even
428 the combination of scalpel and laser may sometimes fail to eliminate lithobiotic structures within
429 fissures (Rivas et al. 2018). Similarly, some laser ablation can cause damage to the mineral surface
430 whilst not completely removing the lichens (Pozo-Antonio et al. 2019). As a result of these issues,
431 biocide treatments often precede mechanical removal (Kakakhel et al. 2019). Although many of the
432 more toxic biocides have been banned, safer compounds may have some drawbacks. For example,
433 the effectiveness of quaternary ammonium salts (e.g benzalkonium chloride) to disrupt cell
434 membranes and kill lithobionts has been well demonstrated (Wessels and Ingmer 2013; Vannini et
435 al. 2018; Sanmartín et al. 2020), but their use can promote bacterial adaptation and antibiotic
436 resistance (Kampf 2018; Kim et al. 2018; Poursat et al. 2019). Moreover, their degradation is
437 suspected to contribute to nitrogen supply (Scheerer et al. 2009), thus favouring, rather than
438 preventing, recolonization by aggressive nitrophytic species. Such alarming findings call for caution
439 in biocide application (Stupar et al. 2014) and the need for non-chemical alternatives. As mentioned
440 above, electromagnetic methods, which include laser irradiation as well as UV, microwave and
441 gamma rays (Riminesi and Olmi 2016), are useful but still have limitations for large-scale
442 applications to outdoor CHSS. Alternatively, heat shock treatments have been shown to provide a

443 sustainable approach to killing lithobionts such as lichens and mosses (Tretiach et al. 2012; Bertuzzi
444 et al. 2013). These poikilohydric organisms tolerate thermal stress well whilst dehydrated, but do
445 not survive heating to 40-60°C for 6 hours when hydrated. Unfortunately, such an approach was
446 less effective against microalgae (Bertuzzi et al. 2017) and the original proposal to exploit solar
447 irradiation to provide the heating may be only suitable under stable weather conditions in warm
448 countries, which limits its applicability. Nevertheless, the combination and improvement of all the
449 cited and other approaches may be expected to shortly produce valuable strategies to moderate the
450 use of biocides.

451 Whatever the chosen approach to kill lithobionts, it is necessary to monitor its efficacy on the target
452 organisms and the examined CHSS: widely used biocides applied to lichens showed species- and
453 site-specific effectiveness, likely depending on differences in the resistance of symbionts and the
454 influence of different stone surfaces and climate/meteorological conditions (Favero-Longo et al.
455 2017). Biocides may have different effectiveness against epilithic and endolithic lithobionts (de los
456 Ríos et al. 2012). Moreover, the method chosen to apply biocides, often defined for economic
457 reasons, may strongly influence the final treatment success (Favero-Longo et al. 2017; Matteucci et
458 al. 2019). A preliminary assessment of the viability of targeted microorganisms before and after
459 treatment should be routinely performed by in situ methods, such as fluorimetry (Tretiach et al.
460 2010).

461 Although the effective devitalization of lithobionts contributes to keeping CHSS in a “clean” state
462 for longer, (re-)colonization is inevitable (Fig. 3). However, longer term cleaning can be achieved
463 by avoiding the application of compounds after cleaning which may increase bioreceptivity and
464 promote recolonization, as demonstrated in the case of certain consolidants and other restoration
465 products (Barriuso et al. 2017; Favero-Longo et al. 2018). In contrast, some other lithobiont control
466 strategies may contribute to delay recolonization, including the application of photocatalytic
467 nanoparticles (Fonseca et al. 2010; Sierra-Fernandez et al. 2017), and eco-friendly products, as
468 essential oils and other biological substances exerting allelopathic effects (Fidanza and Caneva

469 2019). Monitoring programs are increasingly needed to monitor the long-term effectiveness of such
470 approaches, in comparison with the traditional application of biocides following mechanical
471 removal (Bruno et al. 2019; Favero-Longo et al. 2019; Sanmartín et al. 2020). Under certain climate
472 conditions, however, even the most effective strategies may not fully prevent recolonization (Pinna
473 et al. 2018), confirming the necessity of an integrated, site-specific approach (including
474 microclimate control, where possible) to improve the success of management strategies for the
475 conservation of CHSS.

476

477 *Concluding remarks*

478 Advances in technology and medical research have provided great tools to diagnose, control and
479 prevent human diseases, and there is a clear integration between scientific research and managing
480 health. This review has outlined that similarly integrated scientific approaches are available to
481 diagnose the role of lithobionts on CHSS and inform management strategies. Lithobiont-CHSS
482 systems display remarkable complexity, encompassing equilibria between lithosphere (i.e. the stone
483 substrate) and biosphere (i.e. the lithobionts), as well as atmosphere/hydrosphere and
484 anthroposphere, which are additional components affecting colonization and weathering dynamics.
485 As in the case of human health, management decisions about the conservation of stone cultural
486 heritage should increasingly take into consideration all the levels of knowledge available (or
487 implementable) to decipher this complexity, and should be based on accurate diagnosis of each
488 situation. Figure 4 presents a framework to include the above-reviewed spheres of investigation
489 required for decision-making on lithobionts and CHSS, with a focus on whether to remove or
490 preserve lithobionts and, if removal is chosen how to choose a suitable control strategy. As Figure 4
491 demonstrates, there is no ‘one size fits all’ approach because durability of different CHSS depends
492 on the balance between biodeterioration and bioprotection effects driven by lithobiontic
493 communities, which in turn depends on the species and lithology involved and the (micro)-
494 environmental context. Moreover, these same factors (species-lithology-environment) also

495 determine the effectiveness of control strategies. Thus, whatever the protocol used (innovative or
496 based on traditional techniques) an experimental assessment of its suitability for the CHSS of
497 interest is vital. Funding limitations may discourage scientific investigations on the role of
498 lithobionts and the efficacy of control strategies, but such investigations are vital for cost-effective
499 and environmentally-sustainable management of CHSS in future.

500

501 **References**

- 502 Adamo P, Violante P (2000). Weathering of rocks and neogenesis of minerals associated with
503 lichen activity. *Appl Clay Sci* 16: 229-256
- 504 Albertano P (2012). Cyanobacterial biofilms in monuments and caves. In: Whitton BA (ed)
505 Ecology of cyanobacteria II. Springer, Dordrecht, pp. 317-343
- 506 Albertano P, Bruno L (2003). The importance of light in the conservation of hypogean monuments.
507 In: Saiz-Jimenez C (ed) *Molecular Biology and Cultural Heritage*. Balkema, Lisse, pp. 171-177
- 508 Ascaso C, Galván J, Rodríguez-Pascual C (1976). Studies on the pedogenetic action of lichen acids.
509 *Pedobiologia* 16: 321-331
- 510 Banfield JF, Barker WW, Welch SA, Taunton A (1999). Biological impact on mineral dissolution:
511 application of the lichen model to understanding mineral weathering in the rhizosphere. *Proc Natl*
512 *Acad Sci USA* 96: 3404-3411
- 513 Barriuso BC, Botticelli G, Cuzman OA, Osticioli I, Tiano P, Matteini M (2017). Conservation of
514 calcareous stone monuments: Screening different diammonium phosphate based formulations for
515 countering phototrophic colonization. *J Cult Herit* 27: 97-106
- 516 Bartoli F, Casanova-Municchia A, Futagami Y, Kashiwadani H, Moon KH, Caneva G (2014).
517 Biological colonization patterns on the ruins of Angkor temples (Cambodia) in the
518 biodeterioration vs bioprotection debate. *Int Biodeterior Biodegrad* 96: 157–165.

519 Bartoli F, Romiti F, Caneva G (2017). Aggressiveness of *Hedera helix* L. growing on monuments:
520 Evaluation in Roman archaeological sites and guidelines for a general methodological approach.
521 Plant Biosyst 151: 866-877

522 Bell FG (1993). Durability of carbonate rock as building stone with comments on its preservation.
523 Environ Geol 21: 187-200

524 Bellezza S, Albertano P, de Philippis R, Paradossi G (2006). Exopolysaccharides of two
525 cyanobacterial strains from Roman hypogea. Geomicrobiol J 23: 301-310

526 Bertuzzi S, Candotto Carniel F, Pipan G, Tretiach M (2013). Devitalization of poikilohydric
527 lithobionts of open-air monuments by heat shock treatments: a new case study centred on
528 bryophytes. Int Biodeterior Biodegrad 84: 44-53.

529 Bertuzzi S, Gustavs L, Pandolfini G, Tretiach M (2017). Heat shock treatments for the control of
530 lithobionts: A case study with epilithic green microalgae. Int Biodeterior Biodegrad 123: 236-243

531 Bjelland T, Grube M, Hoem S, Jorgensen SL, Daae FL, Thorseth IH, Øvreås L (2011). Microbial
532 metacommunities in the lichen–rock habitat. Environ Microbiol Rep 3: 434-442

533 BLS (British Lichen Society) (2020). Churchyard lichen conservation.
534 [https://www.britishlichensociety.org.uk/activities/churchyard-survey/churchyard-lichen-](https://www.britishlichensociety.org.uk/activities/churchyard-survey/churchyard-lichen-conservation)
535 conservation. Accessed 02 April 2020

536 Bolívar FC, Sánchez-Castillo PM (1997). Biomineralization processes in the fountains of the
537 Alhambra, Granada, Spain. Int Biodeterior Biodegrad 40: 205-215

538 Breitenbach R, Silbernagl D, Toepel J, Sturm H, Broughton WJ, Sasaki GL, Gorbushina AA
539 (2018). Corrosive extracellular polysaccharides of the rock-inhabiting model fungus *Knufia*
540 *petricola*. Extremophiles 22: 165-175

541 Brimblecombe P, Grossi CM (2005). Aesthetic thresholds and blackening of stone buildings. Sci
542 Total Environ 349: 175-189

543 Bruno L, Rugnini L, Spizzichino V, Caneve L, Canini A, Ellwood NTW (2019). Biodeterioration of
544 Roman hypogea: the case study of the Catacombs of SS. Marcellino and Pietro (Rome, Italy). *Ann*
545 *Microbiol* 69: 1023-1032

546 Bungartz F, Garvie LA (2004). Anatomy of the endolithic Sonoran Desert lichen *Verrucaria*
547 *rubrocincta* Breuss: implications for biodeterioration and biomineralization. *Lichenol* 36: 55-73

548 Caneva G (1993). Ecological approach to the genesis of calcium oxalate patinas on stone
549 monuments. *Aerobiol* 9: 149-156

550 Caneva G, Bartoli F, Savo V, Futagami Y, Strona G (2016). Combining statistical tools and
551 ecological assessments in the study of biodeterioration patterns of stone temples in Angkor
552 (Cambodia). *Sci Rep* 6: 32601

553 Caneva G, Galotta G, Cancellieri L, Savo V (2009). Tree roots and damages in the Jewish
554 catacombs of Villa Torlonia (Roma). *J Cult Herit* 10: 53-62

555 Caneva G, Nugari MP, Salvadori O. (2008). Plant biology for cultural heritage: biodeterioration and
556 conservation. Getty Conservation Institute, Los Angeles

557 Carter NEA, Viles HA (2004). Lichen hotspots: raised rock temperatures beneath *Verrucaria*
558 *nigrescens* on limestone. *Geomorphol* 62: 1-16

559 Carter NEA, Viles HA (2005). Bioprotection explored: the story of a little known earth surface
560 process. *Geomorphol* 67: 273-281

561 Casanova-Municchia A, Bartoli F, Taniguchi Y, Giordani P, Caneva G (2018). Evaluation of the
562 biodeterioration activity of lichens in the Cave Church of Üzümlü (Cappadocia, Turkey). *Int*
563 *Biodeterior Biodegrad* 127: 160-169

564 Casanova Municchia A, Percario Z, Caneva G (2014). Detection of endolithic spatial distribution in
565 marble stone. *J Microsc* 256: 37-45

566 Charter M, Tischner U (2017). Sustainable solutions: developing products and services for the
567 future. Routledge, London

568 Chen J, Blume HP, Beyer L (2000). Weathering of rocks induced by lichen colonization - a review.
569 Catena 39: 121-146

570 Cicinelli E, Salerno G, Caneva G (2018). An assessment methodology to combine the preservation
571 of biodiversity and cultural heritage: the San Vincenzo al Volturno historical site (Molise, Italy).
572 Biodivers Conserv 27: 1073-1093

573 Convention Concerning the Protection of the World Cultural and Natural Heritage, 1972.
574 <https://whc.unesco.org/en/conventiontext/>. Accessed 02 April 2020

575 Coombes MA, Viles HA, Zhang H (2018) Thermal blanketing by ivy (*Hedera helix* L.) can protect
576 building stone from damaging frosts. Sci Rep 8: 9834

577 Costantini I, Castro K, Madariaga JM (2018). Portable and laboratory analytical instruments for the
578 study of materials, techniques and environmental impacts in mediaeval mural paintings. Anal
579 Methods 10: 4854-4870

580 Crispim CA, Gaylarde CC (2005). Cyanobacteria and biodeterioration of cultural heritage: a
581 review. Microb Ecol 49: 1-9

582 Cuadros J (2017). Clay minerals interaction with microorganisms: a review. Clay Min 52: 235-262

583 Davidson TM, Altieri AH, Ruiz GM, Torchin ME (2018). Bioerosion in a changing world: a
584 conceptual framework. Ecol Lett 21: 422–438

585 De Leo F, Antonelli F, Pietrini AM, Ricci S, Urzì C (2019). Study of the euendolithic activity of
586 black meristematic fungi isolated from a marble statue in the Quirinale Palace's Gardens in Rome,
587 Italy. Facies 65: 18

588 de los Rios A, Ascaso C (2005). Contributions of in situ microscopy to the current understanding of
589 stone biodeterioration. Int Microbiol 8: 181-188

590 de los Ríos A, Pérez-Ortega S, Wierzos J, Ascaso C (2012). Differential effects of biocide
591 treatments on saxicolous communities: case study of the Segovia cathedral cloister (Spain). Int
592 Biodeterior Biodegrad 67: 64-72

593 Dorn R (2013). Rock coatings. In: Shroder JF (ed) Treatise on geomorphology. Academic Press,
594 San Diego, pp. 70-97

595 Edwards HG, Seaward MR, Attwood SJ, Little SJ, de Oliveira LF, Tretiach M (2003). FT-Raman
596 spectroscopy of lichens on dolomitic rocks: an assessment of metal oxalate formation. *Analyst*
597 128: 1218-1221

598 Fatorić S, Seekamp E (2017). Are cultural heritage and resources threatened by climate change? A
599 systematic literature review. *Clim Chang* 142: 227-254

600 Favero-Longo SE, Accattino E, Matteucci E, Borghi A, Piervittori R (2015). Weakening of gneiss
601 surfaces colonized by endolithic lichens in the temperate climate area of northwest Italy. *Earth*
602 *Surf Process and Landf* 40: 2000-2012

603 Favero-Longo SE, Benesperi R, Bertuzzi S, Bianchi E, Buffa G, Giordani P, Loppi S, Malaspina P,
604 Matteucci E, Paoli L, Ravera S, Roccardi A, Segimiro A, Vannini A (2017). Species-and site-
605 specific efficacy of commercial biocides and application solvents against lichens. *Int Biodeterior*
606 *Biodegrad* 123: 127-137

607 Favero-Longo SE, Borghi A, Tretiach M, Piervittori R (2009). In vitro receptivity of carbonate
608 rocks to endolithic lichen-forming aposymbionts. *Mycol Res* 113: 1216-1227

609 Favero-Longo SE, Brigadeci F, Segimiro A, Voyron S, Cardinali M, Girlanda M, Piervittori R
610 (2018). Biocide efficacy and consolidant effect on the mycoflora of historical stuccos in indoor
611 environment. *J Cult Herit* 34: 33-42

612 Favero-Longo SE, Castelli D, Salvadori O, Belluso E, Piervittori R (2005). Pedogenetic action of
613 the lichens *Lecidea atrobrunnea*, *Rhizocarpon geographicum* gr. and *Sporastatia testudinea* on
614 serpentized ultramafic rocks in an alpine environment. *Int Biodeterior Biodegrad* 56: 17-27

615 Favero-Longo SE, Gazzano C, Girlanda M, Castelli D, Tretiach M, Baiocchi C, Piervittori R
616 (2011). Physical and chemical deterioration of silicate and carbonate rocks by meristematic
617 microcolonial fungi and endolithic lichens (Chaetothyriomycetidae). *Geomicrobiol J* 28: 732-744

618 Favero-Longo SE, Matteucci E, Ruggiero MG (2019). Caratterizzazione di licheni e patine
619 microbiologiche sulle rocce istoriate del Parco Nazionale delle Incisioni Rupestri di Naquane
620 (Valle Camonica) e metodiche per il loro controllo. *Not Soc Lichenol Ital* 32: 31

621 Favero-Longo SE, Turci F, Fubini B, Castelli D, Piervittori R (2013). Lichen deterioration of
622 asbestos and asbestiform minerals of serpentinite rocks in Western Alps. *Int Biodeterior*
623 *Biodegrad* 84: 342-350

624 Fernandes P (2006). Applied microbiology and biotechnology in the conservation of stone cultural
625 heritage materials. *Appl Microbiol Biotechnol* 73: 291-296

626 Ferretti V, Comino E (2015). An integrated framework to assess complex cultural and natural
627 heritage systems with Multi-Attribute Value Theory. *J Cult Herit* 16: 688-697

628 Fidanza MR, Caneva G (2019). Natural biocides for the conservation of stone cultural heritage: A
629 review. *J Cult Herit* 38: 271-286

630 Fitzsimons A, Morrissey S, Woods J (2012). Between East and West. In: De Angelis I. (ed) *The*
631 *Japanese effect in contemporary Irish poetry*. Springer, Berlin, pp. 138-158

632 Flemming HC, Neu TR, Wingender J (2017). *The perfect slime: microbial extracellular polymeric*
633 *substances (EPS)*. IWA publishing, London

634 Flemming HC, Neu TR, Wozniak DJ (2007). The EPS matrix: the “house of biofilm cells”. *J of*
635 *Bacteriol* 189: 7945-7947

636 Fomina M, Burford EP, Hillier S, Kierans M, Gadd GM (2010). Rock-building fungi. *Geomicrobiol*
637 *J* 27: 624-629

638 Fonseca AJ, Pina F, Macedo MF, Leal N, Romanowska-Deskins A, Laiz L, Gómez-Bolea A, Saiz-
639 Jimenez C (2010). Anatase as an alternative application for preventing biodeterioration of mortars:
640 evaluation and comparison with other biocides. *Int Biodeterior Biodegrad* 64: 388-396

641 Fry EJ (1924). A suggested explanation of the mechanical action of lithophytic lichens on rocks
642 (shale). *Ann Bot* 38: 175-196

643 Fry, EJ (1927). The mechanical action of crustaceous lichens on substrata of shale, schist, gneiss,
644 limestone, and obsidian. *Ann Bot* 41: 437-460

645 Gadd GM (2017). Fungi, rocks, and minerals. *Elements* 13: 171-176

646 Gadd GM, Bahri-Esfahani J, Li Q, Rhee YJ, Wei Z, Fomina M, Liang X. (2014). Oxalate
647 production by fungi: significance in geomycology, biodeterioration and bioremediation. *Fungal*
648 *Biol Rev* 28: 36-55

649 Gadd GM, Dyer TD (2017). Bioprotection of the built environment and cultural heritage. *Microb*
650 *Biotechnol* 10: 1152-1156

651 Garcia-Vallès M, Topal T, Vendrell-Saz M (2003). Lichenic growth as a factor in the physical
652 deterioration or protection of Cappadocian monuments. *Environ Geol* 43: 776-781

653 Garty J (1990). Influence of epilithic microorganisms on the surface temperature of building walls.
654 *Can J Bot* 68: 1349-1353

655 Garvie LA, Knauth LP, Bungartz F, Klonowski S, Nash TH (2008). Life in extreme environments:
656 survival strategy of the endolithic desert lichen *Verrucaria rubrocincta*. *Naturwissenschaften* 95:
657 705-712

658 Gazzano C, Favero-Longo SE, Matteucci E, Piervittori R (2011) Index of Lichen Potential
659 Biodeteriogenic Activity: towards a statistical validation and the extension to other biodeteriogens.
660 In: Book of abstract of the XV International Biodeterioration & Biodegradation Symposium
661 (IBBS-15), 20-23 September 2011, Vienna (Austria). p. 104

662 Gazzano C, Favero-Longo SE, Matteucci E, Roccardi A, Piervittori R (2009). Index of Lichen
663 Potential Biodeteriogenic Activity (LPBA): a tentative tool to evaluate the lichen impact on
664 stonework. *Intl Biodeterior Biodegrad* 63: 836-843

665 Gómez-Bolea A, Llop E, Ariño X, Saiz-Jimenez C, Bonazza A, Messina P, Sabbioni C (2012).
666 Mapping the impact of climate change on biomass accumulation on stone. *J Cult Herit* 13: 254-
667 258

668 Gorbushina AA, Broughton WJ (2009). Microbiology of the atmosphere-rock interface: how
669 biological interactions and physical stresses modulate a sophisticated microbial ecosystem. *Ann*
670 *Rev Microbiol* 63: 431-450

671 Guglielmin M, Favero-Longo SE, Cannone N, Piervittori R, & Strini A (2011). Role of lichens in
672 granite weathering in cold and arid environments of continental Antarctica. *Geol Soc Spec Publ*
673 354: 195-204

674 Guillitte O (1995). Bioreceptivity: a new concept for building ecology studies. *Sci Total Environ*
675 167: 215-220

676 Gumbel CW (1856) Mittheilungen über die neue Färberflechte *Lecanora ventosa* Ach., nebst
677 Beitrag zur Entwicklungsgeschichte der Flechten. Denkschriften der Mathematisch-
678 Naturwissenschaftlichen Classe Der Kaiserlichen Akademie der Wissenschaften Besonders
679 Abgedruckt 11. Kaiserlich-Königlichen Hof- und Staatsdruckerei, Wien

680 Haas JR, Purvis OW (2006). Lichen biogeochemistry. In: Gadd GM (ed) *Fungi in biogeochemical*
681 *cycles*. Cambridge University Press, Cambridge, pp. 344-376

682 Hueck HJ (1965). The biodeterioration of materials as a part of hylobiology. *Material und*
683 *Organismen* 1: 5-34

684 Huysen A (2006). Nostalgia for ruins. *Grey Room* 23: 6-21

685 Italian Ministry for Cultural Heritage and Activities (2004) Legislative Decree no. 42 of 22 January
686 2004 - Code of the Cultural and Landscape Heritage, pursuant to article 10 of law no. 137 of 6
687 July 2002. <https://whc.unesco.org/document/155711>. Accessed 3 April 2020

688 Kakakhel MA, Wu F, Gu JD, Feng H, Shah K, Wang W (2019). Controlling biodeterioration of
689 cultural heritage objects with biocides: A review. *Int Biodeterior Biodegrad* 143: 104721

690 Kampf G (2018). Adaptive microbial response to low-level benzalkonium chloride exposure. *J of*
691 *Hosp Infect* 100, e1-e22

692 Kim M, Weigand MR, Oh S, Hatt JK, Krishnan R, Tezel U, Pavlostathis SG, Konstantinidis KT
693 (2018). Widely used benzalkonium chloride disinfectants can promote antibiotic resistance. *Appl*
694 *Environ Microbiol* 84: e01201-18

695 Krumbein WE (1968). Zur Frage der biologischen Verwitterung: Einfluß der Mikroflora auf die
696 Bausteinverwitterung und ihre Abhängigkeit von edaphischen Faktoren. *Z allg Mikrobiol* 8: 107-
697 117

698 Lee MR, Parsons I (1999). Biomechanical and biochemical weathering of lichen-encrusted granite:
699 textural controls on organic–mineral interactions and deposition of silica-rich layers. *Chem Geol*
700 161: 385-397

701 Löki V, Deák B, Lukács AB, Molnár VA (2019). Biodiversity potential of burial places—a review
702 on the flora and fauna of cemeteries and churchyards. *Glob Ecol Conserv*: e00614

703 Maguregui M, Knuutinen U, Trebolazabala J, Morillas H, Castro K, Martinez-Arkarazo I,
704 Madariaga JM (2012). Use of in situ and confocal Raman spectroscopy to study the nature and
705 distribution of carotenoids in brown patinas from a deteriorated wall painting in Marcus Lucretius
706 House (Pompeii). *Anal Bioanal Chem* 402: 1529-1539

707 Mandrioli P, Caneva G, Sabbioni C (2003). Cultural heritage and aerobiology. Methods and
708 measurement techniques for biodeterioration monitoring. Kluwer Academic Publishers, Dordrecht

709 Mapelli F, Marasco R, Balloi A, Rolli E, Cappitelli F, Daffonchio D, Borin S (2012). Mineral–
710 microbe interactions: biotechnological potential of bioweathering. *J Biotechnol* 157: 473-481

711 Marques J, Gonçalves J, Oliveira C, Favero-Longo SE, Paz-Bermúdez G, Almeida R, Prieto B
712 (2016). On the dual nature of lichen-induced rock surface weathering in contrasting micro-
713 environments. *Ecol* 97: 2844-2857

714 Marvasi M, Cavalieri D, Mastromei G, Casaccia A, Perito B (2019). Omics technologies for an in-
715 depth investigation of biodeterioration of cultural heritage. *Int Biodeterior Biodegrad* 144: 104736

716 Matteucci E, Scarcella AV, Croveri P, Marengo A, Borghi A, Benelli C, Hamdan O, Favero-Longo
717 SE (2019). Lichens and other lithobionts on the carbonate rock surfaces of the heritage site of the

718 tomb of Lazarus (Palestinian territories): diversity, biodeterioration, and control issues in a semi-
719 arid environment. *Ann Microbiol* 69: 1033-1046

720 McCarroll D, Viles H (1995). Rock-weathering by the lichen *Lecidea auriculata* in an arctic alpine
721 environment. *Earth Surf Process Landf* 20: 199-206

722 McIlroy de la Rosa JP, Warke PA, Smith BJ (2012). Lichen-induced biomodification of calcareous
723 surfaces: bioprotection versus biodeterioration. *Prog Phys Geogr* 37: 325-351

724 McIlroy de la Rosa JP, Warke PA, Smith BJ (2014). The effects of lichen cover upon the rate of
725 solutional weathering of limestone. *Geomorphol* 220: 81-92

726 Miller AZ, Sanmartín P, Pereira-Pardo L, Dionísio A, Sáiz-Jiménez C, Macedo MF, Prieto B
727 (2012). Bioreceptivity of building stones: a review. *Sci Total Environ* 426: 1-12

728 Molina E, Cultrone G, Sebastian E, Alonso FJ (2013). Evaluation of stone durability using a
729 combination of ultrasound, mechanical and accelerated aging tests. *J Geophys Eng* 10: 035003

730 Morando M, Wilhelm K, Matteucci E, Martire L, Piervittori R, Viles HA, Favero-Longo SE (2017).
731 The influence of structural organization of epilithic and endolithic lichens on limestone
732 weathering. *Earth Surf Process Landf* 42: 1666-1679

733 Mottershead D, Lucas G (2000). The role of lichens in inhibiting erosion of a soluble rock. *Lichenol*
734 32: 601-609

735 Nascimbene J, Salvadori O, Nimis PL (2009). Monitoring lichen recolonization on a restored
736 calcareous statue. *Sci Total Environ* 407: 2420-2426

737 Negi A, Sarethy IP (2019). Microbial biodeterioration of Cultural Heritage: Events, colonization,
738 and analyses. *Microb Ecol* 78: 1014-1029

739 Nimis PL, Monte M, Tretiach M (1987). Flora e vegetazione lichenica di aree archeologiche del
740 Lazio. *Studia Geobotanica* 7: 3-161

741 Nimis PL, Pinna D, Salvadori O (1992). *Licheni e Conservazione dei Monumenti*. Cooperativa
742 Libreria Universitaria Editrice, Bologna

743 Ortega-Morales O, Guezennec J, Hernandez-Duque G, Gaylarde CC, Gaylarde PM (2000).
744 Phototrophic biofilms on ancient Mayan buildings in Yucatan, Mexico. *Curr Microbiol* 40: 81-85
745 Özvan A, Dinçer İ, Akın M, Oyan V, Tapan M (2015). Experimental studies on ignimbrite and the
746 effect of lichens and capillarity on the deterioration of Seljuk Gravestones. *Eng Geol* 185: 81-95
747 Pawlik Ł, Phillips JD, Šamonil P (2016). Roots, rock, and regolith: Biomechanical and biochemical
748 weathering by trees and its impact on hillslopes - A critical literature review. *Earth Sci Rev* 159:
749 142-159
750 Pena-Poza J, Ascaso C, Sanz M, Pérez-Ortega S, Oujja M, Wierzchos J, Souza-Egipsy V,
751 Cañamares MV, Urizal M, Castillejo M, García-Heras M (2018). Effect of biological colonization
752 on ceramic roofing tiles by lichens and a combined laser and biocide procedure for its removal. *Int*
753 *Biodeterior Biodegrad* 126: 86-94
754 Piervittori R, Favero-Longo SE, Gazzano C (2009). Lichens and biodeterioration of stonework: a
755 review. *Chimica oggi-Chem Today* 27: 8-11
756 Piñar G, Sterflinger K (2018). Two decades using molecular techniques to study biodeterioration of
757 cultural heritage: An amazing biotechnological development. In: Mosquera MJ, Almoraima Gil
758 ML (eds) *Conserving Cultural Heritage: Proceedings of the 3rd International Congress on Science*
759 *and Technology for the Conservation of Cultural Heritage (TechnoHeritage 2017)*, May 21-24,
760 2017, Cadiz, Spain. CRC Press, Boca Raton, pp. 299-302
761 Pinheiro AC, Mesquita N, Trovão J, Soares F, Tiago I, Coelho C, Paiva de Carvalho H, Gil F,
762 Catarino L, Piñar G, Portugal A (2019). Limestone biodeterioration: A review on the Portuguese
763 cultural heritage scenario. *J Cult Herit* 36: 275-285
764 Pinna D (2014) Biofilms and lichens on stone monuments: Do they damage or protect? *Front*
765 *Microbiol* 5: 1–3
766 Pinna D (2017) *Coping with biological growth on stone heritage objects: methods, products,*
767 *applications, and perspectives.* Apple Academic Press, Oakville

768 Pinna D, Galeotti M, Perito B, Daly G, Salvadori B (2018). In situ long-term monitoring of
769 recolonization by fungi and lichens after innovative and traditional conservative treatments of
770 archaeological stones in Fiesole (Italy). *Int Biodeterior Biodegrad* 132: 49-58

771 Pinna D, Salvadori O, Tretiach M (1998). An anatomical investigation of calcicolous endolithic
772 lichens from the Trieste karst (NE Italy). *Plant Biosyst* 132: 183-195

773 Pokharel R, Gerrits R, Schuessler JA, Floor GH, Gorbushina AA., von Blanckenburg F (2017). Mg
774 isotope fractionation during uptake by a rock-inhabiting, model microcolonial fungus *Knufia*
775 *petricola* at acidic and neutral pH. *Environ Sci Technol* 51: 9691-9699

776 Poursat BA, van Spanning RJ, de Voogt P, Parsons JR (2019). Implications of microbial adaptation
777 for the assessment of environmental persistence of chemicals. *Crit Rev Environ Sci Technol* 49:
778 2220-2255

779 Pozo-Antonio JS, Barreiro P, González P, Paz-Bermúdez G (2019). Nd: YAG and Er: YAG laser
780 cleaning to remove *Circinaria hoffmanniana* (Lichenes, Ascomycota) from schist located in the
781 Côa Valley Archaeological Park. *Int Biodeterior Biodegrad* 144: 104748

782 Prieto B, Aira N, Silva B (2007). Comparative study of dark patinas on granitic outcrops and
783 buildings. *Sci Total Environ* 381: 280-289

784 Prieto B, Vázquez-Nion D, Fuentes E, Durán-Román AG (2020). Response of subaerial biofilms
785 growing on stone-built cultural heritage to changing water regime and CO₂ conditions. *Int*
786 *Biodeterior Biodegrad* 148: 104882

787 Rampazzi L (2019). Calcium oxalate films on works of art: A review. *J Cult Herit* 40: 195-214

788 Ricci S, Altieri A (2008). Il ruolo delle briofite nel deterioramento dei Beni Culturali. In: Aleffi M
789 (ed) *Biologia ed ecologia delle Briofite*. Antonio Delfino, Roma, pp. 417-434

790 Riminesi C, Olmi R (2016). Localized microwave heating for controlling biodeteriogens on cultural
791 heritage assets. *Int J Conserv Sci* 7(Special Issue 1): 281-294

792 Rivas T, Pozo-Antonio JS, de Silanes ML, Ramil A, López AJ (2018). Laser versus scalpel
793 cleaning of crustose lichens on granite. *Appl Surf Sci* 440: 467-476

794 Sáiz-Jiménez C, García Rowe J, Rodríguez Hidalgo JM (1991). Biodeterioration of polychrome
795 Roman mosaics. *Int Biodeterior Biodegrad* 28: 65- 79

796 Salvadori O, Charola AE (2011). Methods to prevent biocolonization and recolonization: an
797 overview of current research for architectural and archaeological heritage. In: Charola AE,
798 McNamara C, Koestler RJ (eds) *Biocolonization of stone: Control and preventive methods*.
799 *Proceedings from the MCI Workshop Series. Smithsonian Contributions to Museum Conservation*
800 (Vol 2). Smithsonian Inst. Press, Washington, pp. 37-50

801 Salvadori O, Casanova-Municchia A (2016). The role of fungi and lichens in the biodeterioration of
802 stone monuments. *Open Conf Proc J* 7 (suppl. 1: M4): 39–54

803 Sanmartín P, DeAraujo A, Vasanthakumar A (2018). Melding the old with the new: trends in
804 methods used to identify, monitor, and control microorganisms on cultural heritage materials.
805 *Microb Ecol* 76: 64-80.

806 Sanmartín P, Rodríguez A, Aguiar U (2020). Medium-term field evaluation of several widely used
807 cleaning-restoration techniques applied to algal biofilm formed on a granite-built historical
808 monument. *Int Biodeterior Biodegrad* 147: 104870

809 Sanmartín P, Vázquez-Nion D, Arines J, Cabo-Domínguez L, Prieto B (2017). Controlling growth
810 and colour of phototrophs by using simple and inexpensive coloured lighting: A preliminary study
811 in the Light4Heritage project towards future strategies for outdoor illumination. *Int Biodeterior*
812 *Biodegrad* 122: 107-115

813 Sanz M, Oujja M, Ascaso C, Pérez-Ortega S, Souza-Egipsy V, Fort R, de los Ríos A, Wierzechos J,
814 Cañamares MV, Castillejo M (2017). Influence of wavelength on the laser removal of lichens
815 colonizing heritage stone. *Appl Surf Sci* 399: 758-768

816 Sazanova KV, Vlasov DY, Osmolovskay NG, Schiparev SM, Rusakov AV (2016). Significance
817 and regulation of acids production by rock-inhabited fungi. In: Frank-Kamenetskaya OV, Panova
818 EG, Vlasov DY (eds) *Biogenic - abiogenic interactions in natural and anthropogenic systems*.
819 Springer, Cham, pp. 379-392

820 Scarciglia F, Saporito N, La Russa MF, Le Pera E, Macchione M, Puntillo D, Crisci GM, Pezzino A
821 (2012). Role of lichens in weathering of granodiorite in the Sila uplands (Calabria, southern Italy).
822 *Sediment Geol* 280: 119-134

823 Scheerer S, Ortega-Morales O, Gaylarde C (2009). Microbial deterioration of stone monuments - an
824 updated overview. *Adv Appl Microbiol* 66: 97-139

825 Schröder V, Turcanu-Carutiu D, Honcea A, Ion RM (2019). Microscopical methods for the in situ
826 investigation of biodegradation on Cultural Heritage. In: Turcanu-Carutiu D, Ion RM, Hmood K
827 (eds) *Advanced methods and new materials for Cultural Heritage preservation*. IntechOpen,
828 10.5772/intechopen.77505

829 Schumacher J, Gorbushina A (2020). Light sensing in plant-and rock-associated black fungi. *Fungal*
830 *Biol*. In press (<https://doi.org/10.1016/j.funbio.2020.01.004>)

831 Seaward MRD(2015). Lichens as agents of biodeterioration. In: Upreti DK, Divakar PK, Shukla V,
832 Bajpai R (eds), *Recent advances in Lichenology. Modern methods and approaches in*
833 *biomonitoring and bioprospection (Vol 1)*. Springer India, New Delhi, pp. 189–211

834 Seiffert F, Bandow N, Kalbe U, Milke R, Gorbushina AA (2016). Laboratory tools to quantify
835 biogenic dissolution of rocks and minerals: a model rock biofilm growing in percolation columns.
836 *Front Earth Sci* 4: 31

837 Shraddha G, Darshan M (2019). Microbially induced calcite precipitation through urolytic
838 organisms - A review. *Int J Life Sci* 7: 133-139

839 Sierra-Fernandez A, Gomez-Villalba LS, Rabanal ME, Fort R (2017). New nanomaterials for
840 applications in conservation and restoration of stony materials: A review. *Materiales de*
841 *Construcción* 67: 107

842 Signorini MA (1996) L'Indice di Pericolosità: un contributo del botanico al controllo della
843 vegetazione infestante nelle aree monumentali. *Inf Bot Ital* 28: 7-14

844 Silverman MP (1979). Biological and organic chemical decomposition of silicates. *Stud Environl*
845 *Sci* 3: 445-465

846 Slater JW (1856) On some reactions of oxalic acid. Chem Gaz 14: 130-131

847 Smith AL (1921). Lichens. Cambridge University Press, Cambridge

848 Sohrabi M, Favero-Longo SE, Pérez-Ortega S, Ascaso C, Haghghat Z, Talebian MH, Fadaei H, de
849 los Ríos A (2017). Lichen colonization and associated deterioration processes in Pasargadae,
850 UNESCO world heritage site, Iran. Int Biodeterior Biodegrad 117: 171-182

851 Sollas WJ (1880) On the activity of a lichen on limestone. Report of the British Association for the
852 Advancement of Science, 586

853 Souza-Egipsy V, Wierzchos J, Sancho C, Belmonte A, Ascaso C (2004). Role of biological soil
854 crust cover in bioweathering and protection of sandstones in a semi-arid landscape (Torrollones de
855 Gabarda, Huesca, Spain). Earth Surf Process Landf 29: 1651-1661

856 Stanford C (2000) On preserving our ruins. J Archit Conserv 6: 28-43

857 Steinbauer MJ, Gohlke A, Mahler C, Schmiedinger A, Beierkuhnlein C (2013) Quantification of
858 wall surface heterogeneity and its influence on species diversity at medieval castles—implications
859 for the environmentally friendly preservation of cultural heritage. J Cult Herit 14: 219-228

860 Sterflinger K (2010). Fungi: their role in deterioration of cultural heritage. Fungal Biol Rev 24: 47-
861 55

862 Sterflinger K, Krumbein WE (1997). Dematiaceous fungi as a major agent for biopitting on
863 Mediterranean marbles and limestones. Geomicrobiol J 14: 219-230

864 Sterflinger K, Little B, Pinar G, Pinzari F, de los Ríos A, Gu JD (2018). Future directions and
865 challenges in biodeterioration research on historic materials and cultural properties. Int Biodeterior
866 Biodegrad 129: 10-12

867 Sternberg T, Viles HA, Cathersides A, Edwards M (2010). Dust particulate absorption by ivy
868 (*Hedera helix* L) on historic walls in urban environments. Sci Total Environ 409: 162-168

869 Stupar M, Grbić ML, Džamić A, Unković N, Ristić M, Jelikić A, Vukojević J (2014). Antifungal
870 activity of selected essential oils and biocide benzalkonium chloride against the fungi isolated
871 from cultural heritage objects. S Afr J Bot 93: 118-124

872 Taylor-George S, Palmer F, Staley JT, Borns DJ, Curtiss B, Adams JB (1983). Fungi and bacteria
873 involved in desert varnish formation. *Microb Ecol* 9: 227-245

874 Tonon C, Favero-Longo SE, Matteucci E, Piervittori R, Croveri P, Appolonia L., Meirano V, Serina
875 M, Elia D (2019). Microenvironmental features drive the distribution of lichens in the House of
876 the Ancient Hunt, Pompeii, Italy. *Int Biodeterior Biodegrad* 136: 71-81

877 Totsche KU, Rennert T, Gerzabek MH, Kögel-Knabner I, Smalla K, Spiteller M, Vogel HJ (2010).
878 Biogeochemical interfaces in soil: The interdisciplinary challenge for soil science. *J Plant Nutr*
879 *Soil Sci* 173: 88-99

880 Tratebas AM (2004) Biodeterioration of Prehistoric rock art and issues in site preservation. In:
881 Seaward MRD, St. Clair LL (eds) *Biodeterioration of stone surfaces*. Springer, Dordrecht, pp.
882 195-228

883 Tretiach M, Bertuzzi S, Candotto Carniel F (2012). Heat shock treatments: a new safe approach
884 against lichen growth on outdoor stone surfaces. *Environ Sci Technol* 46: 6851-6859

885 Tretiach M, Bertuzzi S, Salvadori O (2010). Chlorophyll a fluorescence as a practical tool for
886 checking the effects of biocide treatments on endolithic lichens. *Int Biodeterior Biodegrad* 64:
887 452-460

888 Trovão J, Portugal A, Soares F, Paiva DS, Mesquita N, Coelho C, Pinheiro C, Catarino L, Gil F,
889 Tiago I (2019). Fungal diversity and distribution across distinct biodeterioration phenomena in
890 limestone walls of the old cathedral of Coimbra, UNESCO World Heritage Site. *Int Biodeterior*
891 *Biodegrad* 142: 91-102

892 Uloth W (1861) Beiträge zur Flora der Laubmoose und Flechten von Kurhessen. *Flora* 44: 565-576

893 UNESCO (2014). Culture, creativity and sustainable development. Research, innovation,
894 opportunities. Florence declaration - 4 October 2014.

895 <https://unesdoc.unesco.org/ark:/48223/pf0000230394>. Accessed 03 April 2020

896 Valkó O, Tóth K, Kelemen A, Miglécz T, Radócz S, Sonkoly J, Tóthmérész B, Török P, Deák B
897 (2018). Cultural heritage and biodiversity conservation—plant introduction and practical restoration
898 on ancient burial mounds. *Nat Conserv* 24: 65-80

899 Vannini A, Contardo T, Paoli L, Scattoni M, Favero-Longo SE, Loppi S (2018). Application of
900 commercial biocides to lichens: Does a physiological recovery occur over time? *Int Biodeterior*
901 *Biodegrad* 129: 189-194

902 Vázquez-Nion D, Silva B, Prieto B (2018). Bioreceptivity index for granitic rocks used as
903 construction material. *Sci Total Environ* 633: 112-121

904 Viles H (2019). Biogeomorphology: Past, present and future. *Geomorphol.* In press
905 (<https://doi.org/10.1016/j.geomorph.2019.06.022>)

906 Viles HA, Cutler NA (2012). Global environmental change and the biology of heritage structures.
907 *Glob Chang Biol* 18: 2406-2418

908 Viles HA, Goudie AS (2004). Biofilms and case hardening on sandstones from Al-Quwayra,
909 Jordan. *Earth Surf Process Landf* 29: 1473-1485

910 Villa F, Pitts B, Lauchnor E, Cappitelli F, Stewart PS (2015). Development of a laboratory model of
911 a phototroph-heterotroph mixed-species biofilm at the stone/air interface. *Front Microbiol* 6: 1251

912 Villa F, Stewart PS, Klapper I, Jacob JM, Cappitelli F (2016). Subaerial biofilms on outdoor stone
913 monuments: changing the perspective toward an ecological framework. *Bioscience* 66: 285-294

914 Wang J, Ersan YC, Boon N, De Belie N (2016). Application of microorganisms in concrete: a
915 promising sustainable strategy to improve concrete durability. *Appl Microbiol Biotechnol* 100:
916 2993-3007

917 Warscheid T, Braams J (2000). Biodeterioration of stone: a review. *Int Biodeterior Biodegrad* 46:
918 343-368

919 Weber B, Scherr C, Bicker F, Friedl T, Büdel B (2011). Respiration-induced weathering patterns of
920 two endolithically growing lichens. *Geobiol* 9: 34-43

- 921 Wei Z, Kierans M, Gadd GM (2012). A model sheet mineral system to study fungal bioweathering
922 of mica. *Geomicrobiol J* 29: 323-331
- 923 Wessels S, Ingmer H (2013). Modes of action of three disinfectant active substances: a review.
924 *Regul Toxicol Pharmacol* 67: 456-467
- 925 Wilhelm K, Viles H, Burke O, Mayaud J (2016). Surface hardness as a proxy for weathering
926 behaviour of limestone heritage: a case study on dated headstones on the Isle of Portland, UK.
927 *Environ Earth Sci* 75: 931
- 928 Zhang G, Gong C, Gu J, Katayama Y, Someya T, Gu JD (2019). Biochemical reactions and
929 mechanisms involved in the biodeterioration of stone world cultural heritage under the tropical
930 climate conditions. *Int Biodeterior Biodegrad* 143: 104723
- 931

932 **Table**

933 Table 1 - Biodeteriorative and bioprotective roles often reported (●) or hypothesized/debated (?) for
 934 various types of lithobiontic organisms. It is worth noting that each potential role has to be
 935 considered species-specific and cannot be generalized for all the members of each group.

				Higher plants	Bryophytes	Lichens	Microbial biofilms		
							Algae	Cyanobacteria	Microcolonial fungi
Biodeteriorative roles	During life	Biogeophysical	"Rooting"	●	●	●			●
			Wetting/ drying			●	●	●	
			Enhanced thermal stresses		?	●	●	●	●
		Biogeochemical	Biom mineralization		●	●		●	
			Acid/ complex dissolution	●	●	●	●	●	?
	Aesthetic loss	Surface coverage/ soiling	●	●	●	●	●	●	
	After death	Remnant biocrusts	Disfiguring patina	?	●	●	●	●	●
Bioprotective roles	During life	Shielding	Umbrella effect	●	●	●	?	?	
			Reduced thermal stress	●	●	●	●	?	
			From pollutants	●	?	?	?	?	
		Biogeochemical	Rock varnish & case hardening			●		●	●
			Biom mineralization		●	●		●	
	Aesthetic/ biodiversity enhancement	'Greening' walls	●	●	●	●	●		
	After death	Remnant biocrusts	Protective patina	?	●	●	●	●	●

936

937

938 **Figure captions**

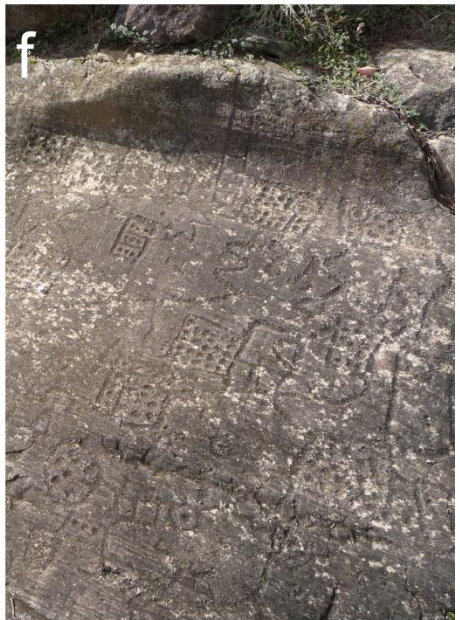
939 Fig. 1. Lithobionts on stone cultural heritage. **a** Residence of the Royal House of Savoy in Govone
940 (Italy; year 2010); **b** Fushimi Inari-Taisha-Shrine in Kyoto (Japan; 2019); **c** Sanctuaire de Notre-
941 Dame de Laghet in La Trinité (France; 2019); **d** Roman underground cistern down to Lithostrotos
942 in Jerusalem Old Town (2017); **e** 16th century Kelmscott Manor near Oxford (UK; 2018); **f**
943 Engravings of the ‘Rock of the Map’ in Valle Camonica (Italy; 2016); **g** St Andrew’s Churchyard in
944 the Isle of Portland (UK; 2018); **h** North Grotto Temple in the Gansu province (China; 2018); **i** The
945 Mostaccini Fountain in the Boboli Gardens of Florence (Italy; 2016).

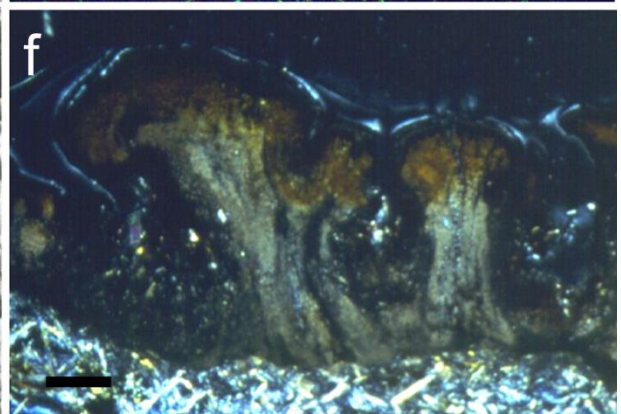
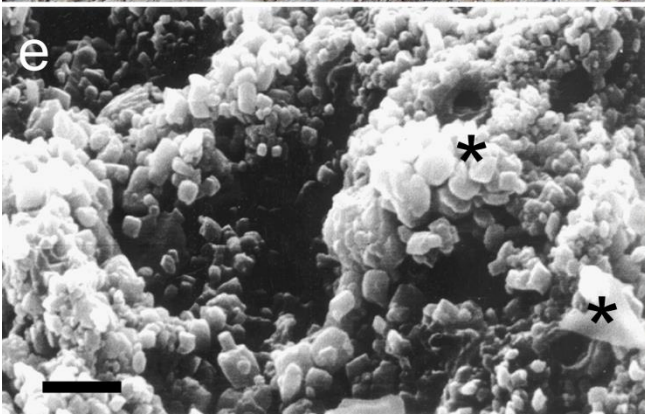
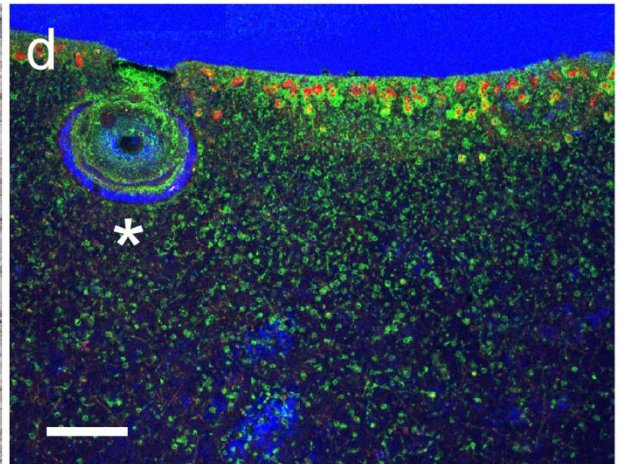
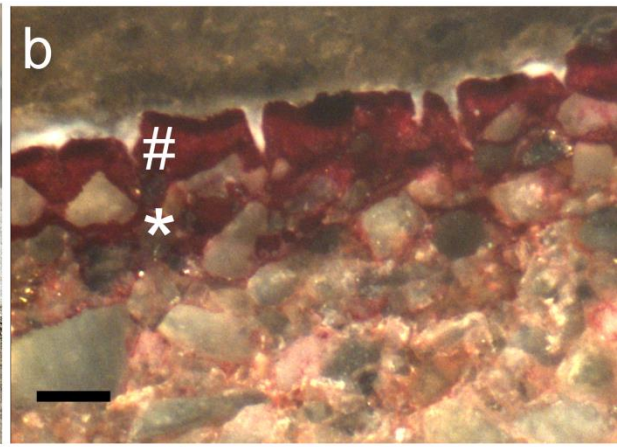
946 Fig. 2. Lithobiont-related processes and spatial patterning on CHSS: the example of lichen-forming
947 fungi. **a** rock flaking associated with lichen colonization of a sandstone (Pietra Serena) sculpted
948 surface. **b** Pietra Serena colonized by the lichen *Aspicilia cinerea* (L.) Körb., displaying the thalline
949 component (#) at the sandstone surface, and the hyphal penetration component (*) developing along
950 the grain borders (polished cross section stained by PAS and observed under reflected light
951 microscopy; bar: 500 µm); **c** pitting at the surface of a limestone previously colonized by an
952 endolithic lichen; **d** Aurisina limestone colonized by the endolithic lichen *Bagliettoa baldensis* (A.
953 Massal.) Vězda, displaying both the photobiont (red) and the mycobiont (green) developing within
954 the substrate, and a pit occupied by a fruiting body (*) (polished cross section stained with FITC-
955 Concanavalin A and observed under confocal laser scanning microscopy; bar: 500 µm); **e**
956 biomineralization of calcium oxalates at the interface between a lichen thallus and a serpentinite
957 rock (*, cross sectioned hyphae submerged by the oxalates; scanning electron microscopy; bar: 5
958 µm); **f** serpentinite colonized by the lichen *Lecidea atrobrunnea* (DC.) Schaer., displaying oxalates
959 (milk-like high birefringence colours) in the medulla layer and at the rock interface (thin cross
960 section observed under cross polarized transmitted light; bar: 500 µm).

961 Fig. 3. Colonization dynamics on CHSS. **a** lichens on a granite gravestone in the Walser graveyard
962 of the alpine village Gressoney-la-Trinité (Italy; year 2010), **b** the same surface after cleaning by
963 professional restorers (year 2011); **c** lichen recolonization after eight years (year 2019).

964 Fig. 4. Framework summarizing the spheres of knowledge dealing with the lithobiotic colonization
965 of CHSS (white boxes) and the potential control strategies (green), with the related diagnostic
966 approaches (blue). Such knowledge may properly support management decisions (orange) to
967 improve the conservation of stone cultural heritage.

968







971

