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(Article begins on next page)

Impact of DNA repair and stability defects on cortical development

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Abstract

Maintenance of genome stability is a crucial cellular function for normal mammalian development and physiology. However, despite its general relevance, genome stability alteration due to genetic or non-genetic conditions has a particularly profound impact on the developing cerebral cortex. In this review we will analyse the principal pathways involved in maintenance of genome stability, the consequences of their alterations with regard to central nervous system development, as well as the possible molecular and cellular basis of this specificity.

Introduction

Maintenance of genome stability is a complex cellular problem, handled by several interrelated mechanisms that ensure faithful duplication and segregation of genetic material during cell cycle, as well as its repair in proliferating or post-mitotic cells.

Alterations of these mechanisms, due to genetic or non-genetic conditions, have severe consequences, such as cancerous transformation or altered embryonic development.

Among the different tissues, developing human neocortex appears to be particularly vulnerable to alterations of genome stability, as the most common clinical manifestations of these disorders is microcephaly, which is the result of brain volume reduction.

Microcephaly is a common clinical condition that may be evident at birth (primary or congenital microcephaly) or **post-natally** (secondary or progressive microcephaly). In the hereditary forms, microcephaly may be the only defect observed (non-syndromic microcephaly or MCPH) or be associated with other malformations (syndromic microcephaly) [1]. Dissecting the molecular and cellular **mechanisms** implicated in genetic primary microcephaly could also be critical to better understand non-genetic forms, such as intrauterine insults, viral infections [2] [3] or alcohol/drug abuse during pregnancy [4]. In theory, any condition that alters neural progenitor expansion or their survival may induce microcephaly. However, despite **the more or less** specific alteration of brain development that characterize microcephaly syndromes, the identification of causal mutations has revealed that they mostly occur in ubiquitously expressed genes, whose products are tightly involved in cell proliferation and maintenance of genomic stability. Understanding the reasons of this specificity is in most cases an open issue. In this review, we will summarize the main cellular events in the development of the cerebral cortex, with particular emphasis on those that may distinguish cortical progenitors from other developing cell types. We will also underscore the evidence of specific sensitivity of neural

progenitors to genome integrity. Then, we will analyze the genetic causes of microcephaly, grouping the different syndromes on the basis of the main molecular process involved. Finally, we will try to highlight how the interaction between different mechanism may represent a common framework at the basis of the selective vulnerability of neural progenitors.

Milestone events in cortical development.

In vertebrates, the earliest stages of brain development require the segmentation of neural tube along a head-to-tail (anteroposterior) and back-to-front (dorsoventral) axes [5]. The first population of neural progenitors consists of neuroepithelial cells (NECs), which expand their pool by undergoing a series of symmetric proliferative divisions [6]. Then, NECs become more fate-restricted radial glial cells (RGCs). RGCs are bipolar cells that retain an apical-basal orientation and express astroglial markers [7] [8]. RGCs contact the pial surface and the ventricular surface through their basal and apical process, respectively. The onset of neurogenesis is determined by asymmetric division of RGCs cells, i.e. divisions that generate two different daughter cells. RGC asymmetric divisions can be directly neurogenic, when they produce an RGC and a neuron, or may generate an RGC and a more fate-restricted, Basal Progenitor (BP). BPs undergo one or few rounds of symmetric divisions before giving rise to post-mitotic neuroblasts and are one of the major sources of cortical neurons [9]. RGCs may also divide symmetrically to generate two BPs or two neurons [10]. Different daughter cells are spatially segregated: RGCs cell bodies are restricted to the more apical part of the cortical wall, corresponding to the ventricular zone (VZ); BPs bodies form a more basal layer known as the subventricular zone (SVZ). Importantly, additional progenitor populations have been described, such as the basal RG-like cells [11], which are located abventricularly and are found abundantly in brains of gyrencephalic species, such as human, monkey and ferret. Newborn cortical neurons

migrate along the intermediate zone (IZ) to reach the cortical plate (CP), where they accumulate below the basal lamina. Later-born neurons arriving in the CP migrate past earlier born neurons in an inside-out fashion, thus generating multi-layered neocortex.

General impact of genomic stability on brain development and function

Genomic instability (GIN) is defined by an increased mutation rate, due to inactivation of DNA repair pathways or to increase of genotoxic stress from cellular processes that overcome high-fidelity DNA repair. Chromosomal instability (CIN) is a subset of GIN, characterized by altered stability of chromosomes, leading to amplification, deletion or rearrangement of long chromosomal fragments [12] [13]. GIN may result from abnormalities in different steps of cell cycle, including DNA replication [14], abnormal chromosome segregation [15] or abortive cytokinesis [16]. CIN and GIN are especially harmful to cells that continue to proliferate, as they are able to deregulate cell division and prompt senescence, cell death, or uncontrolled proliferation. However, CIN and GIN can also be detrimental in long-lived post-mitotic cells, since they may alter gene expression programs and lead to the production of abnormal proteins [17]. On this basis, it is not surprising that genetic and non-genetic alterations leading to CIN and GIN have profound consequences on CNS development, which is characterized by explosive waves of cell proliferation and generates cells with the longest lifespan in the body. Indeed, CNS is consistently affected in most **conditions** resulting in CIN and GIN. Interestingly, although these conditions should generically affect all proliferating cells in the body, in many cases the severity of CNS alterations is disproportionate if compared to other tissues, indicating that CNS development and function have specific vulnerability factors. In particular, neural progenitors show strong sensitivity to DNA damage [18], which may change at different developmental stage [19].

One of the most dramatic evidences of this sensitivity has been provided by follow-up studies of atomic bomb survivors in Hiroshima and Nagasaki, showing that the frequency of microcephaly in **new-borns** exposed to radiations in utero increased ten times more than the incidence of radiation-induced tumors [20]. In addition, a huge array of human syndromes characterized by neurodevelopmental defects, neurodegeneration or brain tumors display defects in DNA damage signalling [21].

This peculiar sensitivity has been correlated with the relatively low capability of different neural progenitors to cope with DNA damage. It has been shown that neuronal progenitors process IR induced DNA damage more slowly than neurons and, as a consequence, are also more susceptible to IR induced apoptosis [22] [23]. A second factor that may justify the specific susceptibility of different neural progenitors to genotoxic insults is the different duration of their cell cycle phases. G1 phase lengthening is associated with differentiating progenitors [24]: neural progenitors undergoing rapid expansion have longer S phase, compared with progenitors committed to the neurogenic lineage [25]. Longer replicative phase during early progenitor expansion may facilitate accurate DNA repair through homologous recombination (HRR), which requires more time to be completed than non-homologous end joining (NHEJ) [26], thus avoiding transmission of genetic errors to a large population of brain cells. On the other hand, differentiating progenitors rely more on NHEJ to repair DSBs [27], which can be more effective for coping with the high replicative stress associated with rapid proliferation of late neural progenitors [28] [29] [30].

Many knock out mouse models of DNA repair genes, such as *Polb* mutants [31], show lethal phenotypes due to neuronal defects.

Differential usage of DNA repair pathways at different developmental stages, associated with a different apoptotic threshold, may justify most of the specific effects of GIN. For instance, mice defective for NHEJ because of DNA ligase IV (*Lig4*) inactivation show DNA

damage-induced apoptosis in **postmitotic differentiating immature neurons**, while HRR-deficient **Xrcc2** knockout mice show apoptosis only in neural precursors cells, thus suggesting that during early development DSB are repaired mainly using HRR [27]. Another example of differential requirement of **DNA damage response (DDR)** factors during development is the conditional ablation of **Topbp1**, which is an activator of ATR during DDR and an important factor to maintain DNA integrity during replication. Deletion of **Topbp1** in early cortical progenitors, by using an **Emx1-Cre** driver, leads to microcephaly and strong apoptosis during development. In contrast, deletion at later stages using **Nes-Cre** results in similar levels of DNA damage but attenuated apoptosis. In addition to the temporally-restricted dependence on different DNA repair pathways, these data suggest that neural progenitors have different threshold for apoptosis at different developmental stages and support the idea that apoptosis in early progenitors might be a crucial mechanism to ensure genome integrity in CNS [32] [29] [28].

Neurons display high sensitivity to GIN even after the end of development. The high levels of oxidative metabolism that characterize brain produces a high burden of reactive oxygen species, which are the main cause of SSB. Transcription associated breaks [33] and neurotransmission-associated stress are additional SSB sources [34]. Accordingly, most patients bearing mutations in gene responsible to sense or repair SSB display neurological syndromes [32].

DNA damage sensing pathways and cortical development.

In all tissues, DNA damage is sensed by genome surveillance pathways that may prevent cell cycle progression, or initiate apoptosis, thus allowing DNA repair or preventing the potential danger posed by damaged cells [35]. Genome surveillance is ensured by three PI3K-like protein kinases: ATM (ataxia-telangiectasia, mutated), ATR (ATM and Rad3-related) and **PRKDC** (the catalytic subunit of the DNA-dependent **protein** kinase) [23]. The

relative importance of individual kinases in CNS development is dependent on the type of DNA lesion and cells [36]. ATR exerts a prominent role in protecting cells from DNA damage arising during S-phase in consequence of DNA stressors like ultraviolet radiation, DNA polymerase inhibitors, nucleotide depletion or DNA crosslinkers, leading to DNA polymerase slowing or stalling [23]. These agents converge on formation of single stranded DNA (ssDNA) stretches at the replication fork, which may produce replication intermediates leading to high risks of mutation and/or genomic rearrangements. Canonical ATR pathway activation is triggered when the replication protein A complex (RPA) binds to ssDNA, generating a recruitment platform for different proteins, including ATR-interacting protein (ATRIP), which in turn recruits ATR. In addition to ATR-ATRIP complex formation, full ATR activation requires the simultaneous presence on ssDNA of the heterotrimeric ring-shaped complex RAD9–RAD1–HUS1 [37]. This complex locally recruits TOPBP1 that binds to and activates ATR, leading to phosphorylation of checkpoint kinase-1 (CHK1) and other downstream ATR effectors, which slow origin firing, arrest cell-cycle, stabilize and restart stalled replication forks [38]. In contrast, ATM and PRKDC are activated by DNA double strand breaks (DSBs) and exert a pivotal function in DSB recognition [23]. When DSB are recognized by the MRN complex (MRE11-RAD50-NBS1), ATM is recruited at DNA damage sites, leading to phosphorylation of H2AX histone variant at S139 (γ H2Ax) in proximity of the break. γ H2Ax binds to MDC1, which further amplifies the signal by recruiting more MRN molecules and TP53BP1 [39]. ATM activation is crucial in preventing DNA-damaged cells from entering S-phase. ATM also activates TP53, which in turn induces cell cycle blocker CDKN1A (p21CIP1) as well as apoptosis-initiating genes. Most DSB occurring outside S phase are repaired by NHEJ independently of ATM, but a minority of DSBs located in heterochromatic regions or with blocked ends, require processing in a ATM dependent fashion. PRKDC is engaged to DSBs by the heterodimer

complex **KU70/80** to form the **DNA-PK holoenzyme** and facilitate DNA repair via NHEJ [40]. However, NHEJ core components inactivation leads to extensive neural apoptosis and embryonic lethality, while mice lacking **PRKDC** are viable [41] [42]. How these three kinases collectively organise the responses to DNA damage has been elegantly demonstrated by Enriquez-Rios and colleagues: mice bearing single or combined mutations of *Atm*, *Atr* or *Prkdc* showed differential cellular sensitivity in response to IR [36]. Authors confirmed that ATM transduces pro-apoptotic signals in non-cycling cells, while it turned out to be less important in replicating progenitors in VZ. In contrast, ATR deletion did not affect the increased apoptosis induced by IR in presence of **PRKDC** loss. Interestingly, apoptosis levels were increased by **PRKDC** loss even when both ATM and ATR were deleted (triple knockout mice), implying the existence of an additional, non-defined pathway converging on **TP53** activation. In humans, *PRKDC* mutations have been associated to a rare syndrome characterized by immune-deficiency and microcephaly [43] [44]. *ATM* loss is responsible for the neurodegenerative syndrome ataxia telangiectasia, while hypomorphic *ATR* mutations are associated with one of the complementation groups of Seckel syndrome (SCKS) [45] [29] (see below).

In summary, the DNA-damage sensors ATR and PRKDC appear to be crucial for coping with endogenous and exogenous replication-related stress arising during embryonic development. Neural tissue could be affected more than other districts because of the stress imposed by the explosive proliferation of neural progenitors, that especially characterize late-born neurons [46]. On the other hand, ATM is much more important for post-mitotic neurons, but in this case the reasons for specificity remain undefined.

DNA repair pathways in cortical development.

In human gestation, cortical neurogenesis lasts for a long period, spanning from 7th till 28th week [47]. Neural progenitors, committed progenitors and differentiated cells are continually under threat by lesions from endogenous or exogenous sources. Base **modifications**, similar to those induced by ultraviolet radiation, may distort the helical structure of the DNA and are repaired by the nucleotide excision repair pathway (NER), while DNA base modifications caused by alkylation, deamination or oxidation, such as **those** induced by reactive oxygen species (ROS), are mainly repaired by the base excision repair pathway (BER). DNA double-strand breaks (DSB) are the most harmful type of DNA lesion, which if left unrepaired or not correctly repaired may lead to apoptosis or mutagenic rearrangements, such as translocations [35]. DSBs may be generated exogenously, by Ionizing Radiations (IR) or different chemical agents (such as chemotherapeutic drugs), or endogenously, by prolonged replication fork stalling or ROS [48]. DSBs can be managed by cells through two different core pathways: HRR and NHEJ [49].

Base Excision Repair

The most common DNA lesion, DNA single strand break (SSB), **may be produced by ROS** reaction with the deoxyribose sugar of DNA, as normal enzymatic **intermediate** of Base Excision Repair (BER) pathway, **from** catalytic intermediates generated by topoisomerase 1 (TOP1) to resolve supercoiled structures during **replication, transcription or other events**. Given the different sources of SSBs, chemical composition of SSB ends can be highly heterogeneous and a wide variety of sub-pathways of SSB repair **have** evolved. Persistent SSBs may lead to replication fork collapse during chromosome duplication with consequent formation of DSBs, as well as transcription block in non-proliferating cells [32]. To deal with oxidation, deamination and spontaneous hydrolysis of bases, specific subtypes of BER operate during all stages of the cell cycle and exert a key function in both

dividing and non-dividing cells. The sequence of BER is started by a lesion-specific DNA glycosylase, which recognizes and hydrolyzes the N-glycosidic bond of a substrate base, generating an **apurinic/aprimidinic** site intermediate. Apurinic sites, which can be formed frequently by spontaneous or damage-induced hydrolysis of the N-glycosidic bond, are cut by an endonuclease, APEX1. This protein works in coordination with PNKP to generate 5'-P and 3'-OH priming **group**, for repair synthesis and ligation. BER then engages Pol β , to replace the missing nucleotide (shortpatch repair) or Pol δ/ϵ in collaboration with PCNA, to **perform** displacing synthesis during S phase or when ATP concentrations are low (longpatch repair). Finally, the repair factor complex XRCC1 and XRCC1 and **DNA ligases** **LIG1 or LIG3** seal the remaining nick [50]. **Mutations** in genes encoding for BER components such as **APTX, TDP1 or PNKP** may produce different CNS phenotypes. *TDP1* mutations cause a neurodegenerative disorder, Spinocerebellar ataxia with axonal neuropathy (SCAN1) [51], while mutations in *APTX* are responsible for Ataxia-ocular motor apraxia 1 (AOA1) [52]. *PNKP* mutations may result in neurodegeneration (AOA4) [53], but may also cause a neurodevelopmental disorder, Microcephaly with Seizures (MCSZ) [54]. This heterogeneity could be explained by the dual role played by PNKP, which is a key factor in BER, but is also involved in NHEJ **through its interaction with** XRCC4. PNKP is thus involved in repair of both SSBR and DSBR [55] [32], but it is likely to play a role only in a fraction of **breaks** repaired by NHEJ after IR damage [56].

Nucleotide Excision Repair

Nucleotide excision repair (NER) is a repair system capable of removing a wide variety of DNA helix-distorting lesions, such as base modifications or UV-induced photolesions and DNA adducts induced by chemicals like N-acetoxy-2-acetylaminofluorene or aflatoxinB1. Patients bearing congenital defects in NER components are peculiarly sensitive to sunlight, and suffer for cancer predisposition and/or premature aging [57].

NER pathway is very important during neural progenitor proliferation, as NER can cope with stalled RNA polymerase or abortive transcripts, monitor DNA integrity and activate DNA damage signaling. In this repair system, a central role is exerted by components of the basal TFIIH transcription and repair complex, required for RNA pol II function. This system is responsible for transcription coupled repair (TCR), as well as for transcription-independent global genomic repair (GGR). It is worth noticing that mutations involving TCR lead to neuropathology, while those affecting GGR have much milder neurological implications. The two pathways share a high number of repair factors but differ during the DNA damage recognition phase [48]. In TCR, DNA lesions affect a transcribed gene and are sensed in consequence of RNA Pol II stalling. In this case, the repair process is initiated by **ERCC6** that, in turn, recruits **ERCC8**. Instead, the proteins required to recognize DNA damage in GGR are XPC and XPE. For both pathways DNA damage verification and unwinding is carried out by TFIIH, a DNA-dependent ATPase and helicase activity that, in complex with **ERCC3** or **ERCC2**, opens up the DNA helix around the lesion. Finally, the DNA strand that contain the lesion is cut at the single- to double-strand DNA transitions by the structure-specific endonucleases **ERCC5** and **ERCC1**, gap filling is carried out by DNA **Pol δ/ε** using undamaged strand as a template and the nick is sealed by DNA ligase I or III [58]. Mutations in several components of the NER pathway can lead to the human syndromes Xeroderma Pigmentosum (XP), Cockayne Syndrome (CS), and Trichothiodystrophy (TTD). Defects in NER components may lead to a complex relationship between genetic alterations and clinical phenotype [59]. While mutations in different **genes**, grouped in eight XP complementation groups, lead to very similar phenotypes, different mutations in just one gene (*ERCC2*) may lead to 6 different clinical disorders: XP, XP with neurological disease, TTD, XP/CS complex, XP/TTD complex or

Cerebro-oculo-facio-skeletal syndrome. XP is rare disease affecting the skin and leading to neurodegeneration (that results in brain atrophy). Exposure of these patients to sunlight results in a high incidence of skin and mucous membrane cancer, squamous and basal cell carcinomas and melanomas [60]. However, mutations in *ERCC8* or *ERCC6*, lead to CS. CS is a developmental and neurological disorder, associated with reduced lifespan without an increased incidence of cancer. In this case, neuropathology is profoundly different from that associated with XP, as CS is characterized by microcephaly with intellectual disability. Microcephaly in CS is likely due to combination of defective cortical development and progressive brain atrophy. In addition, vasculopathy and calcification of the basal ganglia may also be observed. To complicate even further the situation, CS may be caused by mutations in *ERCC2*, *ERCC3* or *ERCC5* [61]. TTD is a rare autosomal recessive disorder characterized by sulphur deficient brittle hair, sometimes accompanied by facial dysmorphism and intellectual disability. TTD arises from mutations in the gene encoding ERCC2 helicase or, less frequently, *ERCC3* or *GTF2H5 proteins*. The neurological abnormalities observed in TTD include developmental microcephaly associated with intellectual disability, deafness and ataxia [62]. Similar to CS, TTD may further affect cortical development by altering myelin formation. Moreover, neurological defects observed in TTD patients may be due to the deregulation of thyroid hormone target genes in the brain [63].

Homologous Recombination Repair and Fanconi Anemia pathways.

In cycling neural progenitors, genomic fidelity during replication is ensured by HRR. HRR is considered an error free mechanism, occurring during S or G2 phase of the cell cycle. During the initial stages of HRR, ATM and ATR kinases recognizes double-strand DNA break and phosphorylate downstream targets such as H2AX, BRCA1, CHEK2 and *TP53*. BRCA1, along with BARD1 and BRIP1, acts as a scaffold that coordinates proteins at the

damage site [64]. HRR requires a sister chromatid as a template to repair DNA and is initiated by the MRE11-RAD50-NBS1 complex, in collaboration with **RBBP8**, which generates 3' ssDNA overhang for strand exchange [65]. MRN complex participates in both HR and NHEJ (see below); in contrast RBBP facilitates HR, while suppressing NHEJ [66] [67]. RAD51 is recruited to 3' ssDNA overhangs through a complex mechanism involving the RPA, BRCA2, BRCA1 and XRCC2 proteins, to form RAD51 nucleoprotein filaments, which are essential for strand invasion of the intact homologous DNA region used as a template for repair. **Studies in knockout mouse models have shown that HRR genes are fundamental for brain development** [68–70]. In humans, mutations in these genes have been associated with heterogeneous clinical manifestations, which may (**BRCA1**) [71] [72] or may not (**BRCA2**) include microcephaly [73] [74]. One of the peculiar phenotypes that may derive from HRR pathway mutations is Fanconi Anemia (FA), a clinically and genetically heterogeneous disorder mainly characterized by bone marrow failure and cancer predisposition. However, some patients also display microcephaly and dwarfism features, reminiscent of **Seckel syndrome** [75]. To date, at least eighteen FANC genes are associated with patient mutations. The FA nuclear complex, composed of **FANCA**, **FANCC**, **FANCG** and **FANCF** proteins, is essential for protection against chromosome breakage. The core complex of the FA pathway is necessary to monoubiquitylate a specific site in each protein of the FANCI/FANCD2 (ID2) heterodimer. **Monoubiquitination** of ID2 triggers nucleases **SLX4** and **ERCC4**, **as well as** downstream repair factors that include **BRIP**, **BRCA2**, **PALB2**, and **RAD51C** [76]. The latter two genes are in common with the HRR pathway. FA proteins have a prominent role in promoting the repair of DNA interstrand crosslinks (ICLs) [77]. Unrepaired ICLs are very harmful, especially in dividing cells, as they prevent strand separation, thereby stalling DNA replication and leading to chromosomal instability [78]. FA pathway is closely dependent on ATR and **CHEK1**

function. Indeed, ATR deficient cells are hypersensitive to ICLs, and many FA proteins , such as FANCD2, FANCA, FANCI and FANCE, are direct substrates of ATR or CHEK1 [79].

Non-Homologous End Joining

NHEJ, which works in both non-replicating and replicating cells, is the predominant DSB repair system in higher eukaryotes. NHEJ involves ligation of two broken DNA ends without needing a repair template. NHEJ core factors, including XRCC4, LIG4 and NHEJ1, associate with DSB after PRKDC recruits Ku proteins to the broken ends, which are subsequently closely aligned and ligated. The repair of some DSBs by NHEJ may require additional accessory proteins, such as DNA polymerases and nucleases. One of such factors is DCLRE1C, an endonuclease interacting with PRKDC to promote repair of a subset of DSBs [80] [81] [82]. Indeed, PRKDC recruits DCLRE1C to promote DNA-end processing [83] [84].

Although NHEJ is often described as an error prone and mutagenic process, it is more efficient and accurate than previously thought [85]. Only when classical NHEJ (cNHEJ) fails, cells use an alternative end-joining pathway mediated by DNA polymerase theta, the so called theta-mediated end-joining (TMEJ), which can easily introduce mutations ([86], [87]). The study of mice mutated for genes important in NHEJ [88] showed elevated apoptosis and enhanced IR sensitivity in cortical progenitors. NHEJ is the main repair pathway in differentiating and post-mitotic neurons [89]. NHEJ is also required during immune system development for V(D)J and Class Switch Recombination mechanisms [90]. For these reasons, congenital defects in NHEJ pathway components such PRKDC [44], LIG4 [91] and genes encoding XRCC4 and NHEJ1 [92] [74] are associated with developmental delay, malignancy, variable degree of immunodeficiency and microcephaly.

The MCPH-SCKS spectrum: many roads leading from genomic instability to primary microcephaly.

Severe congenital microcephalies are characterized by reduced head circumference at birth, to at least 3 standard deviations below the mean [1] [93]. The two main clinical phenotypes are referred to as primary hereditary microcephaly (MCPH) and Seckel syndrome (SCKS) [1] [93]. In MCPH, brain size reduction is the main clinical feature, associated with conserved brain architecture and mild to moderate intellectual disability.

MCPH is a genetically heterogeneous condition, with at least 18 complementation groups. Most of the genetic defects identified in MCPH involve genes that play fundamental roles in various processes that collectively enable cells to faithfully segregate chromosomes and allow correct progression of mitotic division [94]. These defects may range from abnormal microtubule formation, stabilization and polymerization, to alteration of spindle structure, defective centrosome function, along with impaired cell cycle checkpoint function. Although these disfunctions may all indirectly lead to CIN and GIN (see below), some of MCPH genes have a more direct involvement in DNA repair and chromosome stability. Under this point of view, the case of MCPH1 is particularly striking, because this multifunctional gene may affect diverse aspects of cell cycle regulation and integrity. **MCPH1** [95] [96] encodes for a centrosomal protein containing three BRCT (BRCA1 C-terminal) domains. Various studies demonstrated that **MCPH1** plays an important role in G1/S and G2/M checkpoint regulation and in maintaining telomere integrity [97]. MCPH1-deficient cells exhibit mitotic defects, such as a prophase-like state and premature chromosome condensation [98] [99]. MCPH1 plays a crucial role in different aspects of DNA damage response downstream of both ATM and ATR pathways. Indeed, MCPH1 is concentrated at DNA damage foci in response to UV or IR treatment, colocalizing with γ H2AX, **TP53BP1**, pATM, ATR and RPA. In MCPH1-deficient cells, the IR-induced formation of **TP53BP1** and pATM foci or RPA

recruitment to ssDNA are impaired. ATR signaling amplification under replication stress requires MCPH1 to recruit **TOPBP1** [100]. Moreover, during HRR, MCPH1 binds to BRCA2 and regulates the localization of BRCA2 and RAD51 at DNA damage sites [101]. In addition, MCPH1 transcriptionally regulates CHK1 and BRCA1 expression, by interacting with E2F1 on their promoters [102]. Finally, MCPH1 directly acts on chromatin structure, facilitating the access of repair proteins to DNA damage sites, by interacting with the chromatin remodeling complex SWI/SNF in an ATR-dependent manner [103]. Chromatin remodeling is also regulated by PHC1, a human homologue of the *Drosophila* Polyhomeotic gene, which takes part to the Polycomb Repressive Complex 1 and is mutated in MCPH11. Cells defective for PHC1 show enhanced levels of the DNA replication inhibitor **GMNN**, accumulate DNA damage and are defective in DNA repair and proliferation after IR treatment [104]. A direct involvement in GIN has recently been proposed for ASPM, the most frequently mutated MCPH gene [105]. ASPM-dependent microcephaly (MCPH5) is commonly believed to result from abnormal spindle function and positioning, leading to the alteration of symmetric to asymmetric cell division [106]. However, ASPM levels are influenced by IR and ASPM affects efficient DSB repair and reduces DNA damage during cerebellar granule neuron progenitors cell proliferation [107] [108]. Another multifunctional gene causing MCPH is Citron Rho-Interacting Kinase (**CIT**), encoding Citron kinase protein (**CITK**). **CITK** localizes at the centrosomes and midbody, where it plays a major role in regulating the stability of the microtubule cytoskeleton. Mutation of **CIT** lead to apoptosis in neural progenitors, newborn neurons and in male germ cells [109] [110] [111], which was initially proposed to result from cytokinesis failure [112]. In addition, **mouse Cit**-mutant neural progenitors show aberrant cleavage plane orientation [113] and accumulate DNA double strand breaks [114]. The latter phenotype correlates with defective RAD51 localization to DNA damage foci and is probably

responsible of the apoptotic phenotype through TP53 activation [114]. Altogether, these studies underscore that maintenance of genome stability is a critical functional aspect common to many of MCPH genes.

On the other hand, SCKS is characterized by intrauterine growth retardation, severe proportionate short stature, profound microcephaly with intellectual disability and characteristic craniofacial features, in the absence of visceral malformations.

In addition to ATR mutation, SCKS can be caused by mutation of RBBP8, CENPJ, CEP152, CEP63, NIN, DNA2, TRAIP and NSMCE2. However, patients carrying mutations in some of these genes may show phenotypes intermediate between SCKS and MCPH, or even pure MCPH. These genes include RBBP8, CEP152 (involved in MCPH9), CENPJ (involved in MCPH6), CEP63, and PHC1 (involved in MCPH11). Therefore, while in the past SCKS and MCPH were discriminated by height, the most recent view is that the two phenotypes define the extremes of a spectrum of phenotypes in which genomic stability of neural progenitors is mostly compromised.

Centrosomes and chromosome segregation at the core of genome stability and neurologic disease

Centrosome is the main microtubule-organizing center (MTOC), essential for coordinating cell cycle progression, mitotic cell division, cilia formation and DNA damage response [115]. Moreover, in cortical neural progenitors, centrosome plays a pivotal role in regulating cleavage plane orientation [116], cell fate decision and neuronal migration [117]. During mitosis, an intact centrosome number is pivotal for correct chromosome segregation and to maintain genomic integrity. Alteration of centrosomes' number and function can impact on genome stability in different ways. Increased centrosome number may lead to multipolar mitoses, which may result in mitotic catastrophe or undergo bipolar

resolution [118]. Even in this case, there is a high frequency of abnormal microtubule to kinetochore attachments, resulting in chromosome missegregation [118]. The latter event may lead to aneuploidy, DNA damage through micronuclei formation, or even cytokinesis failure ending up with polyploidy [119]. These defects can result from either complete loss of centrosomes, disruption of normal centrosome activity, or from defects in centrosome separation [120]. All these events may have a profound importance on cortical development [121–123].

Therefore, it is not surprising that mutations in genes encoding structural centrosome components or centrosome-associated proteins are the most frequent cause of MCPH (CEP152, CEP63, SASS6, STIL, CENPJ, CEP135, CDK5RAP2, CDK6, ASPM and WDR62), and Microcephalic primordial dwarfism (MPD) [124] [1] [125]. The importance of correct chromosome segregation is further suggested by the occurrence of microcephaly in the so called "condensinopathies", i.e. genetic syndromes deriving from mutations in components of condensin complexes (NCAPD2, NCAPH or NCAPD3) [126].

Condensinopathies are characterized by inappropriate chromosome compaction and decatenation, chromosome segregation errors and micronucleus formation, with consequent increased frequency of aneuploidy [126]. Of note, the function of Condensin II complex is regulated by MCPH1, explaining the premature chromosome condensation (PCC) defect observed in cells derived from MCPH1 patients [127]. It is also emerging that centrosomes could be important integration points for DNA repair pathways and control cell cycle progression after DNA damage. Indeed, some DDR players such as ATM, ATR, CHEK2, CHEK1, BRCA1, BRCA2 and PARPs may localize to the centrosome.

Centrosomal localization of CHEK1 is important to inactivate the phosphatase CDC25, which is an activator of CDK1-cyclinB, the M phase promoting factor [128]. MCPH1 controls the localization of CHEK1 to centrosomes and its loss causes premature CHEK1

activation and early mitotic entry, thus uncoupling mitosis and the centrosome cycle [129]. Moreover **TP53**, a common effector of the DNA damage response implicated in cell cycle progression, DNA repair and apoptosis, can localize to centrosome after DNA damage, in an ATM dependent manner [130]. Another important link between DDR and centrosome is the observation that nuclear-cytoplasmic trafficking of DDR proteins occurs on microtubules and is dependent on dynein. Interestingly, microtubule-targeting agents (e.g. taxanes), can **alter** the intracellular distribution of several DDR proteins (ATM, ATR, **PRKDC, MRE11, RAD50, TP53, NBS1, TP53BP1, and TP63**) [131]. Finally, microtubules are required to control chromatin mobility, in particular in case of DNA break, where an enhanced chromatin moving seems to be important to "expose" the damage and allow DDR factors to reach and repair the lesion [132]. An additional link between centrosome and DNA damage response is represented by **SCKS and FA**, which display centrosomal dysfunction and DDR impairment. **SCKS** may arise from mutations in centrosomal proteins CEP152 or PLK4 [133] [134] [135], leading to impaired centrosome biogenesis, supernumerary centrosomes and impaired cell cycle progression, together with genomic instability and defective DDR. Loss of FA signaling can weaken the spindle assembly checkpoint (SAC), which in normal conditions prevents separation of the duplicated chromosomes until each chromosome is properly attached to the spindle apparatus. FA deficient cells display chromosome instability caused by abnormal cell cycle progression, cytokinesis failure and accumulation of ultrafine bridges. Moreover FA deficient cells accumulate DNA damage since they are hypersensitive to some metabolic by-products such as aldehydes [75].

Perspectives and conclusions

Knowledge of the fundamental processes underlying brain cortex development has grown exponentially during the last few years. These studies have underscored the crucial

relevance for brain development and physiology of the different pathways that have evolved to ensure faithful duplication, transmission and maintenance of the genomic information. At the same time, these studies have started to unravel the complex interrelation between the molecular details of these pathways and the phenotypic spectrum of CNS abnormalities produced by their alterations. Of particular relevance, the centrosome is progressively emerging **as a possible integration hub** for molecular events specifically important for neurogenesis. It is therefore easy to foresee that the study of genome stability maintenance in CNS will be a very interesting topic for future studies. Among the most interesting challenges, it will be important to determine how centrosome functions and DDR are finely orchestrated and why neurogenesis is so sensitive to their alterations. Moreover, it appears very important to better define the role of microtubules in this context. A deeper knowledge of these phenomena could be critical to better understand why alterations of apparently different pathways produce similar phenotypes, as well as how different mutations of the same gene may result in completely different pathologies.

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Figure legends

Figure 1

Multiple kind of genome lesions can impact the nervous system at all stages of development and maturity. Replication stress primarily affects proliferating neural progenitors that may require DNA DSB repair, which involves either HR or NHEJ. HR is dependent on using replicated sister chromatids as an error-free repair templates and is not active in non-replicating or differentiated cells. In this phase, ATR exerts a prominent role in protecting cells from DNA damage arising during S-phase in consequence of DNA stressors. Replicating cells may also experience other types of damage, including inter-strand crosslinks or strand breaks from other sources, including oxidative lesions and transcription-associated damage. ATM and DNA-PK are activated by DNA DSBs and exert a pivotal function in DSB recognition. In non-cycling cells, NHEJ repairs DNA DSBs, whereas other types of DNA damage require the BER/ single-stranded break repair (SSBR) pathway or NER. Oxidative damage can also impact immature cells but will be an ongoing threat to the mature nervous system.

Figure 2.

Cellular defects that impair the fidelity of mitosis promote chromosome mis-segregation (Chromosome Instability CIN) and aneuploidy. Increasing evidence reveals that errors in mitosis can also promote the direct and/or indirect acquisition of DNA damage and chromosome breaks. Unrepaired DNA damage is also deemed a contributor to aneuploidy, which might be especially relevant in the central nervous system (CNS) due to evidence for a low repair capacity of neuronal precursors and differentiated neurons. The induction of DNA damage may lead to apoptosis, which occurs in the majority of conditions described in this review. In contrast, some MCPH, Seckel syndrome and FA are characterized by cell cycle arrest due to mitotic alterations. Color-coded bars indicate the relative contribution to the cortical phenotype from cell cycle arrest (green) and apoptosis (blue). MCPH1: all the genes mutated in MCPH that show mitotic alterations. MCPH2: MCPH1, ASPM, CITK

Figure 1

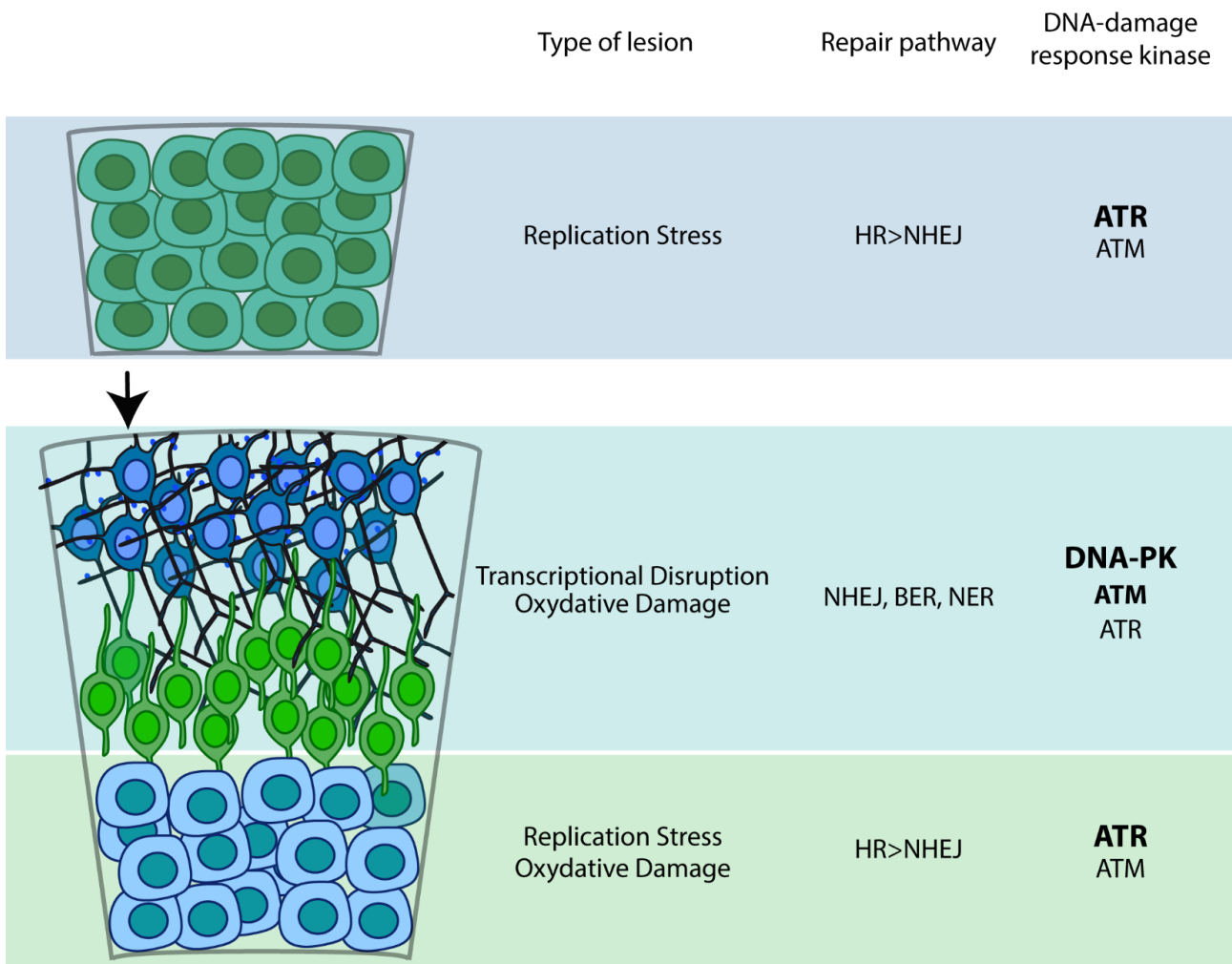


Figure 2

