

DEVELOPMENT OF AN INTERNATIONAL HARMONIZATION SCHEME
FOR SALT WATER FISH TOXICITY TESTS

150576

E. GOLDSTEIN¹, R. AMAVIS¹, R. CABRIDENC², C. GILLIARD³, and
R. SCHUBERT³

¹ Commission of the European Communities
DG XI/A/2
200, Rue de la Loi
B-1049 Brussels, Belgium

² Institut de Recherche de Chimie Appliquée (IRCHA)
BP 1
F-91710 Vert-le-Petit, France

³ Applied Research Consultants (APRECO)
21, Rue de l'Aurore
B-1050 Brussels, Belgium

ABSTRACT

The proceeding deterioration of the marine environment demonstrates the need and the urgency to take appropriate measures. To obtain significant results all the practical efforts made so far must be sustained by national and international conventions and wherever necessary even by more strict regulations.

One of the major objectives of the administrative and scientific authorities involved, is to establish an harmonized scheme for toxicity testing on marine organisms under controlled laboratory conditions, in order to make a meaningful preliminary hazard assessment for the marine environment.

In this respect, and to screen out those substances which show an immediate adverse effect, a simple, cheap and convenient toxicity test on saltwater fish is urgently needed. The state of the art in this field is, however, not developed as far as it is for freshwater species, for which standards already exist : i.e. ISO, EPA, ASTM, OECD.

This paper attempts to find out which simple methods are more or less commonly used, and if there is a trend in the development of an international harmonized toxicity- testing procedure for saltwater fishes.

KEYWORDS

Marine ecotoxicology, Hazard assessment, Bioassays, Methods, Fish, Standardization, Review.

INTRODUCTION

Marine pollution is a major international preoccupation. Indeed, as a result of many different causes (river input, direct discharge, transportation accidents), the sea receives a considerable amount of mineral or organic pollutants of diverse origin. Such widespread pollution, caused by substances which are often slowly biodegradable and highly toxic, can have very serious consequences on the functioning of the marine ecosystems, with all the resulting ecological, sanitary, and economic implications.

In order to suppress, or at least limit, such harmful effects on the marine environment, it seems indispensable to establish national or international regulations or international conventions. Presently some international conventions are already in application :

- the Oslo Convention, of February 1972, for the prevention of marine pollution resulting from dumping operations carried out by ships and aircrafts ;
- the London Convention, of December 1972, for the prevention of marine pollution resulting from the dumping of wastes and other materials ;
- the Helsinki Convention, of March 1974, for the protection of the marine environment in the Baltic sea zone ;

- the Paris Convention, of June 1974, for the prevention of marine pollution of telluric origin ;
- the Barcelona Convention, of February 1976, for the protection of the Mediterranean sea against pollution.

The following European directives may also be quoted :

- the directive 75/160/CEC of December 8, 1975 concerning the quality of bathing water ;
- the directive 76/464/CEC, of May 4, 1976 concerning the pollution caused by certain dangerous substances discharged into the aquatic environment of the Community ;
- the directive 78/176/CEC of February 20, 1978 concerning waste from the titanium dioxide industry ;
- the directive 79/831/CEC of September 18, 1979 concerning the approximation of laws, regulations, and administrative provisions relating to the classifications, packaging and labelling of dangerous substances.

Moreover, there are in this field a certain number of national regulations resulting, among other sources, from European directives.

The aim of these conventions and regulations is to limit the discharge of certain substances, hence reducing their adverse effect in the marine environment. They rely on the application of laboratory methods, which can forecast the behaviour and effects of these polluting substances.

In this field, it will be necessary to reach an international harmonization scheme and to provide standardized methods leading to reciprocal data recognition. Among the different means to be retained in order to reach an international consensus, the development of a standard acute toxicity test with a marine or estuarine fish is urgently required. This preliminary screening test must be easy to perform and should under arbitrary but fixed conditions, stress the lethal effects for fish, resulting from a short exposure to a toxicant. In a further stage, it would be very useful to examine the effects likely to concern other marine organisms, as well as sublethal effects and longer exposure times.

Working by analogy with the views established in the field of fish toxicity for the freshwater environment, a screening test of this type should be based on the following principles :

- stress of a lethal effect on adults ;
- duration of exposure ranging from 24 to 96 h ;
- the use of a synthetic medium simulating the marine environment ;
- if necessary, the possibility for periodic or continuous renewal of the medium ;
- the choice of a species which is easily available year-round in many countries and obtained from laboratory-bred cultures or fish-farms (guaranteeing the origin, healthy condition, and sensitivity of the test organism) ;
- normally, a species of this kind should be relevant of the marine environment and should also be convenient to maintain in the laboratory (dimensions, temperature, and oxygen requirements).

Moreover, such method should allow sufficient repeatability and reproducibility of the results ; it should be inexpensive and easy to apply, so as to be used on a routine basis by many laboratories.

As it remains difficult to gather all such conditions in one single test, it will be necessary to take among others, the availability of the species likely to be used and the continuous progress made in the field of ecotoxicology with marine fishes into account.

In order to evaluate how much work has to be done to reach a satisfactory solution, it seems appropriate at this stage to take stock of the question by assessing, on the one hand, the plans and proposals made by different national or international organisations, such as ISO, FAO, OECD, CEC, ASTM, EPA, and on the other hand, by examining the methodologies existing in the concerned field.

SCHEMES AND PROPOSALS ISSUED BY NATIONAL AND INTERNATIONAL ORGANIZATIONS

ISO (INTERNATIONAL ORGANIZATION FOR STANDARDIZATION)

In 1981 the subcommittee "Biological Water Tests" recommended the use of standardized toxicity tests concerning marine organisms. A document

established by the Secretariat lists the fish species mostly used for this purpose :

Agonus cataphractus

Platichthys flesus

Pleuronectes platessa

Pomatoschistus microps

Pomatoschistus minutus

It is also suggested to use the rainbow trout, Salmo gairdneri, previously adapted to a salt water environment. A discussion paper on an appropriate method has been introduced by Sweden but, to this day, no discussion has been initiated on the matter and no decision has been made as to how a given proposal should be followed up.

FAO (FOOD AND AGRICULTURE ORGANIZATION OF THE UNITED NATIONS)

The issue was examined in 1977 and the advantages of using toxicity tests for fish were underlined, yet no recommendation was made as to the type of fish species to be used.

OECD (ORGANIZATION FOR ECONOMIC COOPERATION AND DEVELOPMENT)

In 1977 an OECD working group "Ecotoxicology" established a first inventory of laboratory methods susceptible of being used to assess the impact of chemical substances on organisms which are part of water and land ecosystems. Subsequently a list of monographs was published in 1979, of which a small number involved fish. The chosen species were considered to be representative for the marine environment owing to their origin or adaptation capacities. Among the species mentioned, the following can be noted :

Anguilla anguilla

Gadus morhua

Gasterosteus aculeatus

Platichthys flesus

Pleuronectes platessa

Solea solea

No decision has been made as to the advantage and the choice of an ecotoxicity test based on the use of marine species.

Table I. List of fish species recommended in the "Standard methods for the examination of water and waste water" (15th ed.)

Anchoa mitchilli
Brevoortia patronus
Brevoortia tyrannus
Centropristis striata
Clupea harengus
Cyprinodon variegatus
Fundulus heteroclitus
Fundulus parvipinnis
Fundulus similis
Gasterosteus aculeatus
Harengula pensacolatae
Lagodon rhomboides
Leiostomus xanthurus
Menidia berillina
Menidia menidia
Micropogon undulatus
Morone saxatilis
Mugil cephalus
Mugil curema
Pseudopleuronectes americanus
Sardinops sagax

Table II. List of fish species recommended in "Methods for measuring the acute toxicity of effluents to aquatic organisms" (EPA 600/478012, July 78)

Citharichthys stigmatatus
Cymatogaster aggregata
Cyprinodon variegatus
Fundulus heteroclitus
Fundulus similis
Gasterosteus aculeatus
Lagodon rhomboides
Leiostomus xanthurus
Leptocottus armatus
Menidia sp.
Paralichthys dentatus
Paralichthys lethostigma
Parophrys vetulus

EEC (COMMISSION OF THE EUROPEAN COMMUNITY)

The directive 78/166/CEC on titanium dioxide waste recommends the use of toxicity tests with fish species which are commonly found in the discharge areas.

The directive 79/831/CEC on dangerous substances does not exclude ecological studies with marine fishes but does not mention any particular species. The text of the directive does not require explicitly the use of freshwater species, and one can imagine that, in some cases (e.g. for an oil dispersant), the Competent Authorities should require the results of a test with marine species.

The other directives do not impose ecotoxicity tests with marine fishes.

ASTM (AMERICAN SOCIETY FOR TESTING AND MATERIALS)

The 15th edition of "Standard Methods" recommends the use of 21 marine or estuarine fish species, which are mentioned in Table I.

EPA (ENVIRONMENTAL PROTECTION AGENCY)

In 1978 the EPA recommended 13 marine or estuarine species, which are mentioned in Table II. Various proposals have been examined at a national level, but no final decision has been taken to this day.

PRINCIPLES USED FOR THE INVESTIGATION OF EXISTING METHODOLOGIES

To establish the list of publications concerning the toxicity of substances or effluents to marine and estuarine fish or species considered to be representative for such an environment, data base documentation and specific abstracts were consulted. The documents obtained have been filed and used in order to collect data which could be useful in tackling one of the main problems to be solved : namely to designate the one single test species or various suitable species to be retained. The data have been entered into a computer programme and processed in order to obtain answers to the following questions :

1. Which species are used for tests with the following duration of exposure :
 - between 0 and 96 h,
 - between 96 h and 28 days,
 - longer than 28 days ?
2. Which species are retained for tests on a complete life cycle ?
3. Which species are retained for tests based on :
 - lethal effects,
 - effects concerning hatching-reproduction,
 - metabolic modifications ?
4. Which species are likely to be obtained :
 - from fish farms
 - through laboratory breeding, nevertheless making a distinction between tests using eggs and tests using organisms having reached a further stage of development ?
5. Which species are most suitable for tests carried out in an artificial medium ?
6. Which species correspond to the following criteria : available from a fish farm or laboratory, adult, maintainable in an artificial medium, suitable for static or semi-static tests with exposure times of 0 - 96 h, appropriate for the assessment of the LC50 ?

It seemed indeed useful to go beyond our primary objective, so as to be able to obtain additional information concerning toxicity tests, which could be useful in the future to complete the screening procedure.

Quite evidently a certain number of documents escaped our investigation, in particular publications from Japan and the USSR. Moreover, scientific literature does not always give a faithful description of the situation, as it does not always mention routine tests. Certain publications lack precision regarding the origin and age of the fish, as well as the test conditions, etc. In addition, a certain confusion seems to exist as to the nomenclature. It should also be noted that certain countries, such as the United States and the Scandinavian countries, have more experience in the field of marine exotoxicology. Consequently, the species used in these countries take on considerable importance in the light of our investigation, although their availability, at an international level does not match this in any way.

Table III. Occurrence frequency of species used in 261 toxicity tests described in the literature (total number of species : 80)

Species	Frequency	References
<u>Agonus cataphractus</u>	1	92
<u>Alburnus alburnus</u>	2	8, 70
<u>Alosa aestivalis</u>	3	6, 88, 89
<u>Alosa pseudoharengus</u>	2	6, 111
<u>Alosa sapidissima</u>	1	6
<u>Anguilla anguilla</u>	2	66, 92
<u>Anguilla rostrata</u>	1	28
<u>Aphanius dispar</u>	2	55, 56
<u>Archosargus probatocephalus</u>	1	106
<u>Arius felis</u>	2	106, 133
<u>Belone belone</u>	1	125
<u>Blenius pavo</u>	1	65
<u>Brevoortia tyrannus</u>	2	13, 103
<u>Chromis punctipinnis</u>	1	
<u>Citharichthys stigmaeus</u>	2	35, 85
<u>Clupea harengus</u>	7	62, 124, 126, 138, 139, 140
<u>Clupea harengus membras</u>	3	73, 74, 75
<u>Clupea harengus pallasii</u>	4	14, 32, 112
<u>Clupea pallasii</u>	2	2, 3
<u>Cymatogaster aggregata</u>	3	35, 115, 119
<u>Cynoscion nebulosus</u>	1	59
<u>Cyprinodon variegatus</u>	19	5, 21, 29, 34, 35, 43, 44, 46, 48, 52, 96, 97, 107, 108, 109, 110, 131, 132
<u>Dicentrarchus labrax</u>	1	100
<u>Engraulis mordax</u>	1	102
<u>Fundulus grandis</u>	1	36
<u>Fundulus heteroclitus</u>	17	16, 17, 24, 25, 28, 29, 30, 31, 35, 37, 76, 77, 83, 127, 134, 135, 136
<u>Fundulus majalis</u>	1	29
<u>Fundulus similis</u>	4	5, 35, 91, 109
<u>Gadus morhua</u>	7	38, 64, 78, 79, 92, 94, 98
<u>Gasterosteus aculeatus</u>	4	35, 60, 90, 92
<u>Gillichthys mirabilis</u>	1	114
<u>Gobius microps</u>	1	1
<u>Gobius minutus</u>	1	1
<u>Heteropneustes fossilis</u>	1	104
<u>Labrus bergylta</u>	1	26
<u>Lagodon rhomboides</u>	8	22, 35, 47, 91, 106, 107, 109, 117
<u>Lebistes reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	8	13, 33, 35, 80, 87, 91, 103, 110
<u>Leptocottus armatus</u>	1	35
<u>Leuresthes tenuis</u>	1	27
<u>Liza macrolepis</u>	1	53
<u>Melangorammus aeglefinus</u>	2	140

<u>Menidia beryllina</u>	1	5
<u>Menidia menidia</u>	5	28, 33, 89, 103, 129
<u>Menidia sp.</u>	1	35
<u>Micropogon undulatus</u>	1	106
<u>Microstomus kitt</u>	2	140
<u>Morone americana</u>	3	6, 88, 89
<u>Morone labrax</u>	1	82
<u>Morone saxatilis</u>	5	6, 45, 86, 88, 89
<u>Mugil cephalus</u>	4	18, 30, 101, 120
<u>Myoxocephalus octodecemspinosus</u>	1	98
<u>Notropis hudsonius</u>	1	111
<u>Oncorhynchus gorbuscha</u>	3	16, 63, 116
<u>Oncorhynchus kisutch</u>	12	11, 12, 20, 41, 42, 84, 111, 115, 118, 122
<u>Oncorhynchus tshawytscha</u>	6	17, 19, 23, 51, 54, 116
<u>Ophiocephalus punctatus</u>	1	104
<u>Osmerus mordax</u>	1	111
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Parophrys vetulus</u>	1	35
<u>Peltorhamphus latus</u>	1	95
<u>Perca flavescens</u>	1	6
<u>Perca fluviatilis</u>	4	66, 70, 71, 72,
<u>Phoxinus phoxinus</u>	4	7, 9, 90, 93
<u>Platichthys flesus</u>	10	10, 49, 50, 67, 68, 69, 78, 79, 92
<u>Pleuronectes flesus</u>	2	57, 66
<u>Pleuronectes platessa</u>	10	1, 4, 78, 79, 81, 92, 105, 139, 140
<u>Poecilia reticulata</u>	1	1
<u>Pseudopleuronectes americanus</u>	6	17, 28, 98, 113, 128, 130
<u>Roccus saxatilis</u>	1	30
<u>Salmo gairdneri</u>	3	19, 20, 58
<u>Salmo salar</u>	3	38, 137, 141
<u>Sardina pilchardus</u>	2	140
<u>Scyliorhinus canicula</u>	1	40
<u>Siganus canaliculatus</u>	1	121
<u>Solea solea</u>	4	4, 139, 140
<u>Stenotomus chrysops</u>	1	30
<u>Stenotomus versicolor</u>	1	17
<u>Tautoglabrus adspersus</u>	4	61, 39, 99, 123

Table IV. Occurrence frequency of species used in tests with an exposure period between 0 and 96 h (total number of species : 62)

Species	Frequency	References
<u>Alosa aestivalis</u>	2	6, 89
<u>Alosa pseudoharengus</u>	2	6, 111
<u>Alosa sapidissima</u>	1	6
<u>Anguilla anguilla</u>	1	92
<u>Anguilla rostrata</u>	1	28
<u>Aphanius dispar</u>	2	55, 56
<u>Archosargus probatocephalus</u>	1	106
<u>Arius felis</u>	1	106
<u>Brevoortia tyrannus</u>	2	13, 103
<u>Chromis punctipinnis</u>	1	
<u>Citharichthys stigmatæus</u>	2	35, 85
<u>Clupea harengus</u>	2	138, 139
<u>Clupea harengus membras</u>	1	75
<u>Clupea harengus pallasii</u>	2	32, 112
<u>Clupea pallasii</u>	1	2
<u>Cymatogaster aggregata</u>	2	35, 115
<u>Cynoscion nebulosus</u>	1	59
<u>Cyprinodon variegatus</u>	13	5, 21, 29, 35, 43, 48, 52, 96, 97, 107, 109, 110
<u>Dicentrarchus labrax</u>	1	100
<u>Engraulis mordax</u>	1	102
<u>Fundulus heteroclitus</u>	9	16, 17, 24, 25, 29, 35, 83, 127
<u>Fundulus majalis</u>	1	29
<u>Fundulus similis</u>	3	5, 35, 109
<u>Gadus morhua</u>	4	38, 64, 78, 92
<u>Gasterosteus aculeatus</u>	4	35, 60, 90, 92
<u>Gobius microps</u>	1	1
<u>Gobius minutus</u>	1	1
<u>Labrus bergylta</u>	1	26
<u>Lagodon rhomboides</u>	5	22, 35, 106, 107, 109
<u>Lebistes reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	5	13, 35, 87, 103, 110
<u>Leptocottus armatus</u>	1	35
<u>Liza macrolepis</u>	1	53
<u>Menidia beryllina</u>	1	5
<u>Menidia menidia</u>	3	28, 89, 103, 129
<u>Menidia sp.</u>	1	35
<u>Micropogon undulatus</u>	1	106
<u>Microstomus kitt</u>	2	139
<u>Morone americana</u>	2	6, 89
<u>Morone labrax</u>	1	82
<u>Morone saxatilis</u>	4	6, 45, 86, 89
<u>Mugil cephalus</u>	3	28, 101, 120
<u>Notropis hudsonius</u>	1	111
<u>Oncorhynchus gorbusha</u>	3	16, 63, 116
<u>Oncorhynchus kisutch</u>	5	12, 20, 84, 111, 115
<u>Oncorhynchus tshawytscha</u>	4	16, 19, 23, 116

<u>Osmerus mordax</u>	1	111
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Parophrys vetulus</u>	1	35
<u>Perca flavescens</u>	1	6
<u>Phoxinus phoxinus</u>	2	90, 93
<u>Platichthys flesus</u>	2	78, 92
<u>Pleuronectes platessa</u>	5	1, 4, 78, 81, 92
<u>Poecilia reticulata</u>	1	1
<u>Pseudopleuronectes americanus</u>	2	17, 28
<u>Salmo gairdneri</u>	2	19, 20
<u>Salmo salar</u>	2	38, 141
<u>Scylliorhinus canicula</u>	1	40
<u>Solea solea</u>	1	4
<u>Stenotomus versicolor</u>	1	17
<u>Tautoglabrus adspersus</u>	2	61, 123

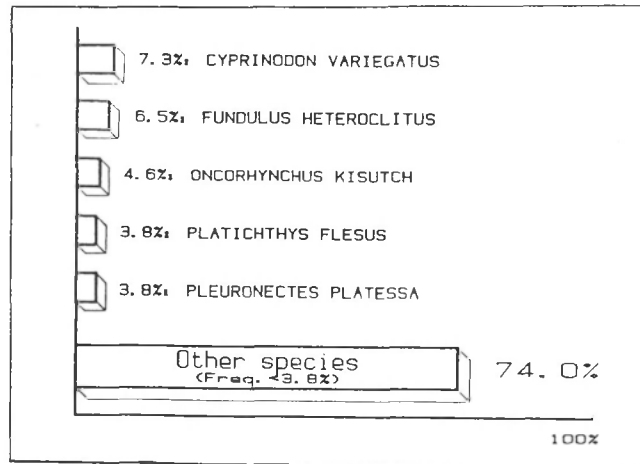


Fig. 1. Frequency (%) of species used in 261 toxicity tests described in the literature.

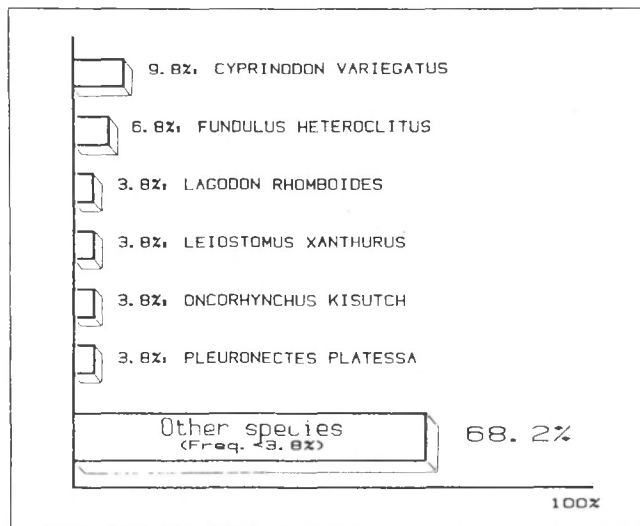


Fig. 2. Frequency (%) of species used in tests with an exposure period between 0 and 96 h. On 261 toxicity tests, 132 fulfill the condition, using 62 different species.

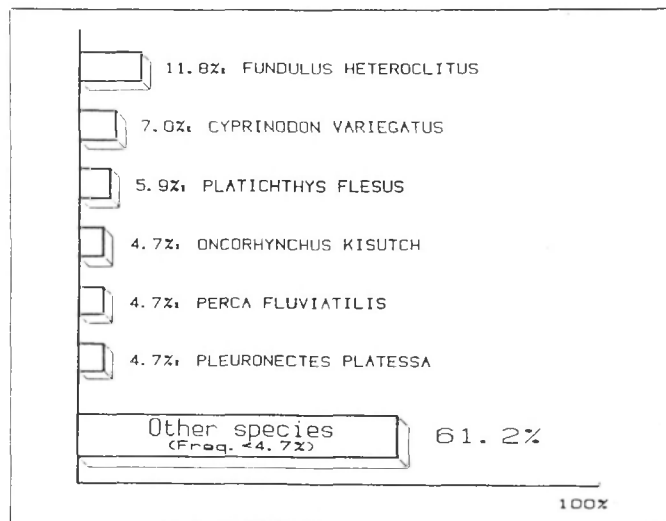


Fig. 3. Frequency (%) of species used in tests with an exposure period between 96 h and 28 days. On 261 toxicity tests, 85 fulfill the condition, using 46 different species.

Table V. Occurrence frequency of species used in tests with an exposure period between 96 h and 28 days (total number of species : 46)

Species	Frequency	References
<u>Alburnus alburnus</u>	1	70
<u>Alosa aestivalis</u>	1	6
<u>Alosa pseudoharengus</u>	1	6
<u>Alosa sapidissima</u>	1	6
<u>Anguilla anguilla</u>	2	66, 92
<u>Anguilla rostrata</u>	1	28
<u>Aphanius dispar</u>	1	56
<u>Arius felis</u>	1	133
<u>Citharichthys stigmaeus</u>	1	85
<u>Clupea harengus</u>	2	139, 140
<u>Clupea harengus membras</u>	2	73, 75
<u>Clupea harengus pallasii</u>	2	14, 112
<u>Cyprinodon variegatus</u>	6	21, 43, 46, 108, 132
<u>Fundulus heteroclitus</u>	10	24, 25, 28, 29, 30, 31, 37, 77, 83, 135
<u>Fundulus similis</u>	2	91, 107
<u>Gadus morhua</u>	2	38, 92
<u>Gasterosteus aculeatus</u>	1	92
<u>Gobius microps</u>	1	1
<u>Gobius minutus</u>	1	1
<u>Heteropneustes fossilis</u>	1	104
<u>Lagodon rhomboides</u>	2	91, 117
<u>Leiostomus xanthurus</u>	2	33, 91
<u>Melanogrammus aeglefinus</u>	1	140
<u>Menidia menidia</u>	1	33
<u>Microstomus kitt</u>	1	140
<u>Morone americana</u>	1	6
<u>Morone saxatilis</u>	1	6
<u>Mugil cephalus</u>	1	118
<u>Oncorhynchus kisutch</u>	4	41, 42, 118, 122
<u>Oncorhynchus tshawytscha</u>	2	51, 54
<u>Ophiocephalus punctatus</u>	1	104
<u>Peltorhamphus latus</u>	1	95
<u>Perca flacescens</u>	1	6
<u>Perca fluviatilis</u>	4	66, 70, 71, 72
<u>Platichthys flesus</u>	5	10, 49, 50, 67, 69
<u>Pleuronectes flesus</u>	1	66
<u>Pleuronectes platessa</u>	4	1, 4, 139, 140
<u>Poecilia reticulata</u>	1	1
<u>Pseudopleuronectes americanus</u>	1	130
<u>Roccus saxatilis</u>	1	28
<u>Salmo salar</u>	2	38, 137
<u>Sardina pilchardus</u>	1	140
<u>Scyliorhinus canicula</u>	1	40
<u>Siganus canaliculatus</u>	1	121
<u>Solea solea</u>	3	4, 139, 140
<u>Stenotomus chrysops</u>	1	28

RESULTS

The investigated papers cover 261 tests using 80 different fish species listed in Table III and Fig. 1.

EXPOSURE PERIOD

Short-term tests : from 0 to 96 h

This period of exposure is used for acute toxicity tests. On 261 tests examined, 132 use this period of exposure for 62 different species (Table IV, Fig. 2). Cyprinodon variegatus and Fundulus heteroclitus are the species most commonly used.

Medium-term tests : from 96 h to 28 days

Out of 261 tests, 85 use this period of exposure for 46 different species (Table V, Fig. 3). Fundulus heteroclitus and Cyprinodon variegatus are the species most commonly used.

Long-term tests

1. Longer than 28 days

An exposure period longer than 28 days is used in 47 tests, in which 25 different species are involved (Table VI, Fig. 4). Cyprinodon variegatus and Platichthys flesus are the species most commonly used.

2. Full life cycle

Only five tests are full life-cycle studies with two species, Cyprinodon variegatus and Oncorhynchus tshawitscha (Table VII).

It is clear from the examination of the exposure period criteria that Cyprinodon variegatus can be used in all cases, whereas Fundulus heteroclitus are used for short- and medium-term tests.

Table VI. Occurrence frequency of species used in test with an exposure period exceeding 28 days (total number of species :25)

Species	Frequency	References
<u>Alburnus alburnus</u>	2	8, 70
<u>Anguilla anguilla</u>	1	92
<u>Alphanius dispar</u>	2	55, 56
<u>Blencius pavo</u>	1	65
<u>Cymatogaster aggregata</u>	1	119
<u>Cyprinodon variegatus</u>	8	34, 43, 44, 46, 48, 96, 97
<u>Fundulus grandis</u>	1	36
<u>Fundulus heteroclitus</u>	2	28, 76
<u>Gadus morhua</u>	3	92, 94, 98
<u>Gasterosteus aculeatus</u>	1	92
<u>Gillichthys mirabilis</u>	1	114
<u>Lagodon rhomboides</u>	1	46
<u>Lebistes Reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	1	80
<u>Mugil cephalus</u>	1	18
<u>Myoxocephalus octodecemspinosus</u>	1	98
<u>Oncorhynchus kisutch</u>	2	11, 42
<u>Phoxinus phoxinus</u>	2	7, 9
<u>Platichthys flesus</u>	5	49, 67, 68, 92
<u>Pleuronectes flesus</u>	2	57, 66
<u>Pleuronectes platessa</u>	3	1, 92, 105
<u>Poecilia reticulata</u>	1	1
<u>Pseudopleuronectes americanus</u>	1	98
<u>Salmo gairdneri</u>	1	58
<u>Tautoglabrus adspersus</u>	2	39, 99

Table VII. Occurrence frequency of species used in test lasting a complete life cycle out of 261 tests, only five fulfill the condition, using two species

Species	Frequency	References
<u>Cyprinodon variegatus</u>	4	34, 44, 48, 108
<u>Oncorhynchus tshawytscha</u>	1	54

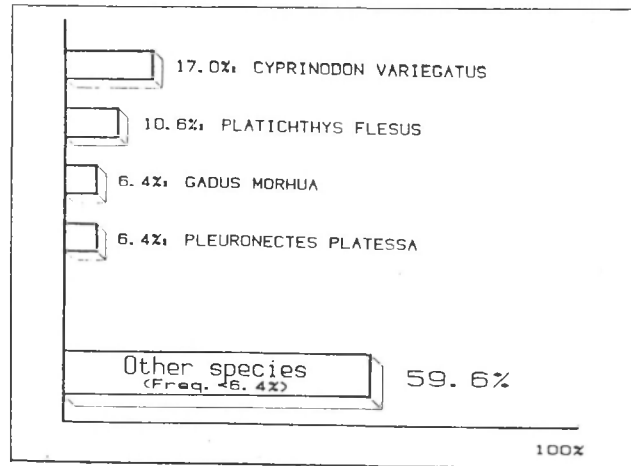


Fig. 4. Frequency (%) of fish species used in tests with an exposure period longer than 28 days. On 261 toxicity tests, 47 fulfill the condition, using 25 different species.

Table VIII. Occurrence frequency of species used in lethal effect tests (total number of species : 62)

Species	Frequency	References
<u>Agonus cataphractus</u>	1	92
<u>Alosa aestivalis</u>	1	89
<u>Alosa pseudoharengus</u>	1	111
<u>Anguilla anguilla</u>	1	92
<u>Anguilla rostrata</u>	1	30
<u>Aphanius dispar</u>	1	56
<u>Arius felis</u>	1	133
<u>Belone belone</u>	1	125
<u>Brevoortia tyrannus</u>	2	13,103
<u>Citharichthys stigmaeus</u>	2	35,85
<u>Clupea harengus</u>	5	62,124,126, 139,140
<u>Clupea harengus membras</u>	3	73,74,75
<u>Clupea harengus pallasii</u>	1	112
<u>Cymatogaster aggregata</u>	2	35,115
<u>Cynoscion nebulosus</u>	1	59
<u>Cyprinodon variegatus</u>	19	5,21,29,34,35,43,44,46,48, 52,96,97,107,108,109,110, 131,132

Table VIII. (cont'd)

Species	Frequency	References
<u>Dicentrarchus labrax</u>	1	100
<u>Engraulis mordax</u>	1	102
<u>Fundulus grandis</u>	1	36
<u>Fundulus heteroclitus</u>	11	16, 17, 24, 28, 29, 30, 31, 35, 37, 77, 127
<u>Fundulus majalis</u>	1	29
<u>Fundulus similis</u>	3	5, 35, 109
<u>Gadus morhua</u>	4	38, 64, 79, 92
<u>Gasterosteus aculeatus</u>	4	35, 60, 90, 92
<u>Gobius microps</u>	1	1
<u>Gobius minutus</u>	1	1
<u>Labrus bergylta</u>	1	26
<u>Lagodon rhomboides</u>	5	22, 35, 47, 107, 109
<u>Lebistes reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	6	13, 35, 80, 87, 103, 110
<u>Leptocottus armatus</u>	1	35
<u>Melanogrammus aeglefinus</u>	1	139
<u>Menidia beryllina</u>	1	5
<u>Menidia menidia</u>	4	28, 89, 103, 129
<u>Menidia sp.</u>	1	35
<u>Microstomus kitt</u>	1	140
<u>Morone americana</u>	1	89
<u>Morone labrax</u>	1	82
<u>Morone saxatilis</u>	3	45, 86, 89
<u>Mugil cephalus</u>	2	28, 101
<u>Notropis hudsonius</u>	1	111
<u>Oncorhynchus grobuscha</u>	3	16, 63, 116
<u>Oncorhynchus kisutch</u>	6	20, 84, 111, 115, 118, 122
<u>Oncorhynchus tshawytscha</u>	5	16, 19, 23, 54, 116
<u>Osmerus mordax</u>	1	111
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Parophrys vetulus</u>	1	35
<u>Peltorhamphus latus</u>	1	95
<u>Phoxinus phoxinus</u>	4	7, 9, 90, 93
<u>Platichthys flesus</u>	3	67, 79, 92
<u>Pleuronectes platessa</u>	6	1, 4, 81, 92, 139, 140
<u>Poecilia reticulata</u>	1	1
<u>Pseudopleuronectes americanus</u>	2	17, 28
<u>Roccus saxatilis</u>	1	30
<u>Salmo gairdneri</u>	2	19, 20
<u>Salmo salar</u>	3	38, 137, 141
<u>Sardina pilchardus</u>	1	140
<u>Siganus canaliculatus</u>	1	121
<u>Solea solea</u>	3	4, 139, 140
<u>Stenotomus chrysops</u>	1	30
<u>Stenotomus versicolor</u>	1	17

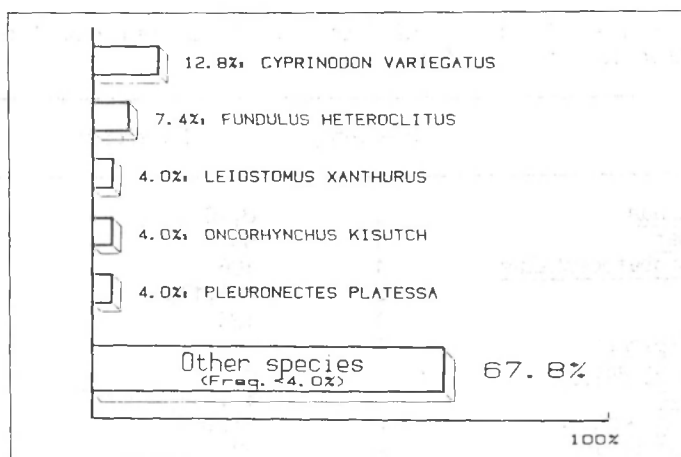


Fig. 5. Frequency (%) of species used in lethal effect tests. On 261 tests, 149 fulfill the condition, using 62 different species.

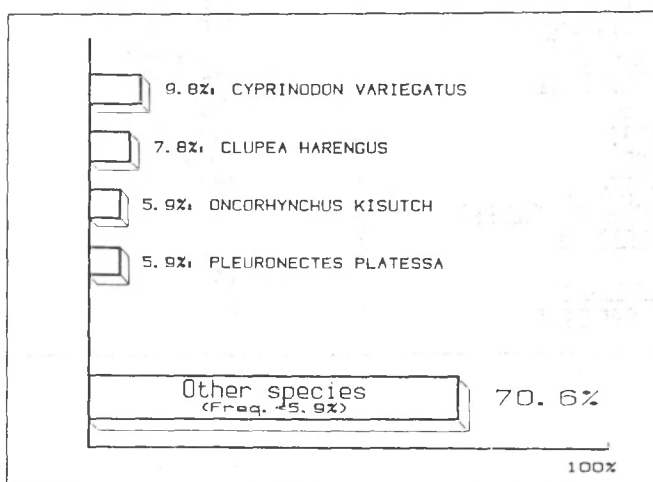


Fig. 6. Frequency (%) of species used in behaviour tests. On 261 tests, 51 fulfill the condition, using 31 different species.

Table IX. Occurrence frequency of species used in behavioral tests (total number of species : 31)

Species	Frequency	References
<u>Alburnus alburnus</u>	2	8,70
<u>Aphanius dispar</u>	1	56
<u>Archosargus probatocephalus</u>	1	106
<u>Arius felis</u>	2	106,133
<u>Belone belone</u>	1	125
<u>Chromis punctipinnis</u>	1	
<u>Citharichthys stigmaeus</u>	1	85
<u>Clupea harengus</u>	4	62,126,138,139
<u>Clupea harengus membras</u>	2	73,74
<u>Clupea harengus pallasii</u>	1	32
<u>Cymatogaster aggregata</u>	2	115,119
<u>Cyprinodon variegatus</u>	5	34,43,46,48
<u>Fundulus heteroclitus</u>	2	16,136
<u>Gadus morhua</u>	2	92,94
<u>Gasterosteus aculeatus</u>	1	92
<u>Gillichthys mirabilis</u>	1	114
<u>Lagodon rhomboides</u>	2	106,117
<u>Leiostomus xanthurus</u>	1	87
<u>Micropogon undulatus</u>	1	106
<u>Oncorhynchus gorbuscha</u>	1	116
<u>Oncorhynchus kisutch</u>	3	41,42,115
<u>Oncorhynchus tshawytscha</u>	1	116
<u>Perca fluviatilis</u>	2	70,72
<u>Phoxinus phoxinus</u>	2	7,93
<u>Platichthys flesus</u>	3	10,92
<u>Pleuronectes platessa</u>	1	139
<u>Pseudopleuronectes americanus</u>	1	17
<u>Scyliorhinus canicula</u>	1	40
<u>Solea solea</u>	1	139
<u>Stenotomus versicolor</u>	1	17
<u>Tautoglabrus adspersus</u>	1	61

TOXIC EFFECTS

Lethal effect

Out of 261 tests, 149 measure the lethal effect on 62 different fish species. Cyprinodon variegatus and Fundulus heteroclitus are the species most commonly used (Table VIII, Fig. 5).

Behavioral modifications

Changes in the behaviour of the fish were studied in 51 tests, in 31 different species. Cyprinodon variegatus is the species most commonly used (Table IX, Fig. 6).

Effects on hatching and/or reproduction

Changes in the hatching or reproduction rate and the measure of morphological abnormalities in early life stages were encoded under the same topic in the data bank. Changes in the reproduction of the fish were observed in 53 test in 23 different species. Cyprinodon variegatus is the species most commonly used (Table X, Fig. 7).

Modification in metabolic activity

Physiological studies were performed in 47 tests with 28 different species. Platichthys flesus is the species most commonly used (Table XI, Fig. 8).

ORIGIN OF THE TEST ORGANISMS

Test organisms obtained from fish farms

Out of 261 tests, 24 tests were performed using 11 different commercially available fish species (Table XII, Fig. 9). Oncorhynchus kisutch is the species most commonly used. It should be pointed out however that this species is rather cumbersome to manipulate due to its size.

Table X. Occurrence frequency of species used in hatching and reproduction tests (total number of species : 23)

Species	Frequency	References
<u>Alosa aestivalis</u>	3	6,88,89
<u>Alosa pseudoharengus</u>	1	6
<u>Alosa sapidissima</u>	1	6
<u>Belone belone</u>	1	125
<u>Clupea harengus</u>	4	62,124,126,139
<u>Clupea harengus membras</u>	2	74,75
<u>Clupea harengus pallasii</u>	1	112
<u>Cyprinodon variegatus</u>	9	34,43,46,48, 96,97,108,132
<u>Fundulus grandis</u>	1	36
<u>Fundulus heteroclitus</u>	3	4,77,135
<u>Gadus morhua</u>	4	64,78,79,92
<u>Gasterosteus aculeatus</u>	1	92
<u>Leiostomus xanthurus</u>	1	33
<u>Leuresthes tenuis</u>	1	27
<u>Menidia menidia</u>	3	33,89,129
<u>Morone americana</u>	2	6,88
<u>Morone saxatilis</u>	2	6,88
<u>Perca flavescens</u>	1	6
<u>Phoxinus phoxinus</u>	1	7
<u>Platichthys flesus</u>	3	78,79,92
<u>Pleuronectes platessa</u>	4	1,78,79,139
<u>Pseudopleuronectes americanus</u>	3	113,128,130
<u>Solea solea</u>	1	139

Table XI. Occurrence frequency of species used in physiological tests (total number of species : 28)

Species	Frequency	References
<u>Anguilla anguilla</u>	1	66
<u>Aphanius dispar</u>	1	55
<u>Blenius pavo</u>	1	65
<u>Clupea harengus</u>	1	139
<u>Clupea harengus membras</u>	1	75
<u>Clupea harengus pallasii</u>	1	32
<u>Clupea pallasii</u>	1	2
<u>Cyprinodon variegatus</u>	2	21,43
<u>Fundulus heteroclitus</u>	3	24,25,83
<u>Gadus morhua</u>	2	92,98
<u>Gillichthys mirabilis</u>	1	114
<u>Heteronneustes fossilis</u>	1	104

<u>Lagodon rhomboides</u>	2	22,117
<u>Leiostomus xanthurus</u>	1	87
<u>Liza macrolepis</u>	1	53
<u>Mugil cephalus</u>	2	18,120
<u>Myoxocephalus octodecemspinosus</u>	1	98
<u>Oncorhynchus kisutch</u>	3	11,12,118
<u>Oncorhynchus tshawytscha</u>	1	23
<u>Ophiocephalus punctatus</u>	1	104
<u>Perca fluviatilis</u>	1	66
<u>Platichthys flesus</u>	7	49,50,67,68,69,92
<u>Pleuronectes flesus</u>	2	57,66
<u>Pleuronectes platessa</u>	3	81,92,139
<u>Pseudopleuronectes americanus</u>	1	98
<u>Salmo gairdneri</u>	1	58
<u>Solea solea</u>	1	139
<u>Tautoglabrus adspersus</u>	3	39,99,123

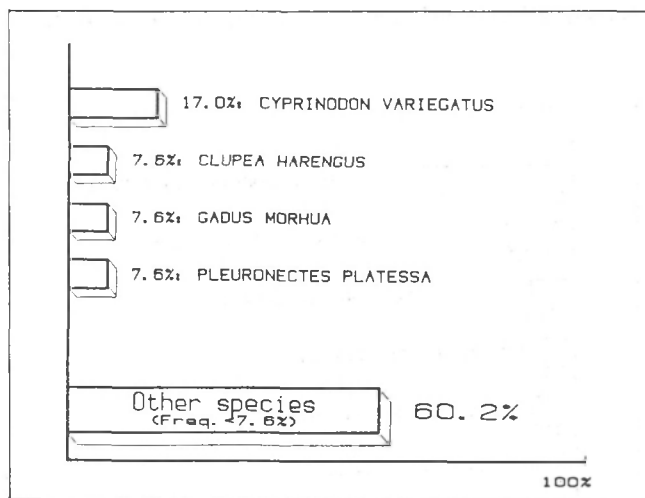


Fig. 7. Frequency (%) of species used in reproduction and hatching tests. On 261 tests, 53 fulfill the condition, using 23 different species.

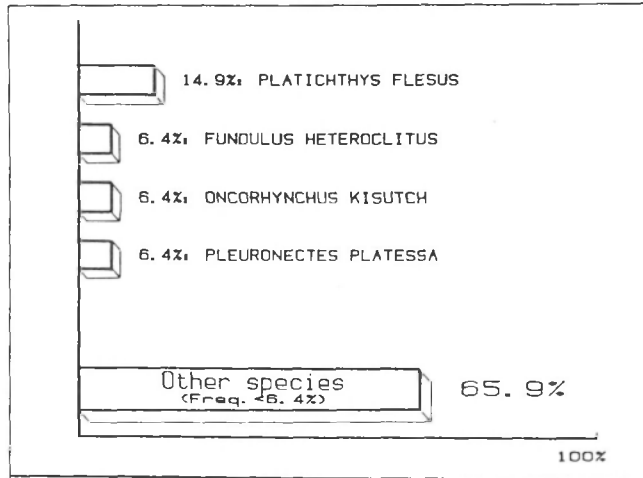


Fig. 8. Frequency (%) of species used in physiological tests. On 261 tests, 47 fulfill the condition, using 28 different species.

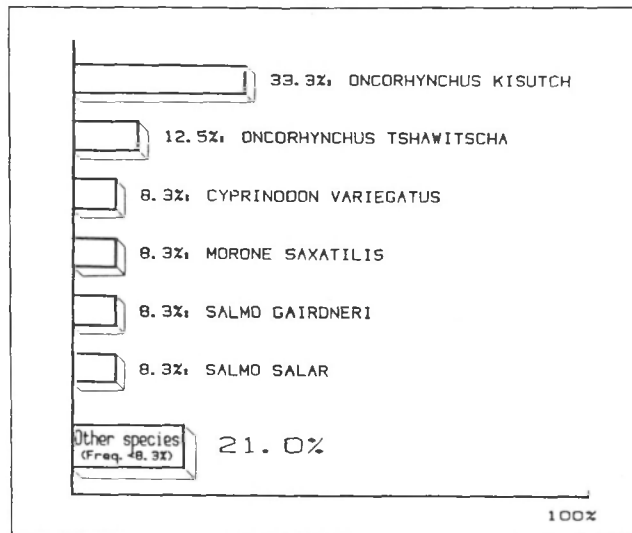


Fig. 9. Frequency (%) of species obtained from fish farms. On 261 tests, 24 fulfill the condition, using 11 different species.

Table XII. Occurrence frequency of species obtained from fish farms (total number of species : 11)

Species	Frequency	References
<u>Cymatogaster aggregata</u>	1	115
<u>Cyprinodon variegatus</u>	2	44,52
<u>Dicentrarchus labrax</u>	1	100
<u>Fundulus heteroclitus</u>	1	76
<u>Gillichthys mirabilis</u>	1	114
<u>Microstomus kitt</u>	1	140
<u>Morone saxatilis</u>	2	45,86
<u>Oncorhynchus kisutch</u>	8	11,12,41,42,84,111,115,118
<u>Oncorhynchus tshawytscha</u>	3	19,23,116
<u>Salmo gairdneri</u>	2	19,58
<u>Salmo salar</u>	2	38,137

Table XIII. Occurrence frequency of species cultivated in laboratories (total number of species : 18)

Species	Frequency	References
<u>Citharichthys stigmaeus</u>	1	35
<u>Cymatogaster aggregata</u>	1	35
<u>Cyprinodon variegatus</u>	8	34,35,43,46,48,96,132
<u>Fundulus heteroclitus</u>	2	24,35
<u>Fundulus similis</u>	1	35
<u>Gasterosteus aculeatus</u>	1	35
<u>Labrus bergylta</u>	1	26
<u>Lagodon rhomboides</u>	1	35
<u>Lebistes reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	1	35
<u>Leptocottus armatus</u>	1	35
<u>Menidia sp.</u>	1	35
<u>Oncorhynchus tshawytscha</u>	1	54
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Paraphrys vetulus</u>	1	35
<u>Pleuronectes platessa</u>	2	4,92
<u>Solea solea</u>	1	4

Test organisms cultivated in laboratories

In 27 tests, fishes cultivated in laboratories were used. The 18 different species are listed in Table XIII and Fig. 10, and Cyprinodon variegatus is most commonly used.

TEST CARRIED OUT IN AN ARTIFICIAL MEDIUM

Out of 261 tests, 36 tests were carried out with different kinds of artificial or reconstituted water, and this with 25 different species. Cyprinodon variegatus and Fundulus heteroclitus were the species most commonly used (Table XIV, Fig. 11).

TESTS CORRESPONDING TO VARIOUS CRITERIA SIMULTANEOUSLY

Toxicity tests were selected in which :

- the species is obtained from fish farms and/or laboratory breeding ;
- the species is used in the adult stage ;
- an artificial medium is used ;
- a static or semistatic procedure is used ;
- the exposure period is from 0 to 96 h ;
- the test criterion is the lethal effect.

From the 261 tests studied, 13 tests meet these prerequisites, using 13 different species. None of these 13 species has been used twice with the same criteria (Table XV).

The variability of species used by laboratories during toxicity tests with marine fish can be explained mainly by the objectives pursued. One species, namely Cyprinodon variegatus, has been used several times in slightly different conditions (duration of exposure, cultivation medium etc.). Sometimes, several species have been used in similar conditions, in order to test the sensitivity of the different species. This method was used to assess the impact of a substance (*i.e.* oil dispersant, pesticide, etc.) on an entire community.

When all prerequisites for a routine acute toxicity test with marine fish are applied simultaneously, it becomes obvious that there is no consensus among scientists to use a common test organism.

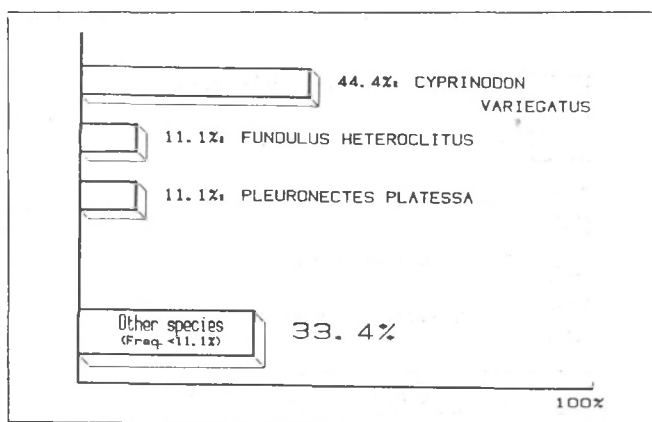


Fig. 10. Frequency (%) of species cultivated in laboratories (tests using only early life stages excluded). On 261 tests, 27 fulfill the condition, using 18 different species.

Table XIV. Occurrence frequency of species used in tests with artificial or reconstituted water (total number of species : 25)

Species	Frequency	References
<u>Archosargus probatocephalus</u>	1	106
<u>Arius felis</u>	1	106
<u>Cirrharchichthys stigmæus</u>	1	35
<u>Cymatogaster aggregata</u>	1	35
<u>Cyprinodon variegatus</u>	6	5, 21, 29, 34, 35, 97
<u>Dicentrarchus labrax</u>	1	100
<u>Fundulus grandis</u>	1	36
<u>Fundulus heteroclitus</u>	4	25, 29, 35, 77
<u>Fundulus majalis</u>	1	29
<u>Fundulus similis</u>	2	5, 35
<u>Gadus morhua</u>	1	92
<u>Gasterosteus stigmæus</u>	1	35
<u>Lagodon rhomboides</u>	2	35, 106
<u>Lebistes reticulatus</u>	1	15
<u>Leiostomus xanthurus</u>	1	35
<u>Leptocottus armatus</u>	1	35
<u>Menidia beryllina</u>	1	5
<u>Menidia sp.</u>	1	35
<u>Micropogon undulatus</u>	1	106
<u>Mugil cephalus</u>	1	120
<u>Oncorhynchus tshawytscha</u>	1	23
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Parophrys vetulus</u>	1	35
<u>Pleuronectes platessa</u>	2	1, 92

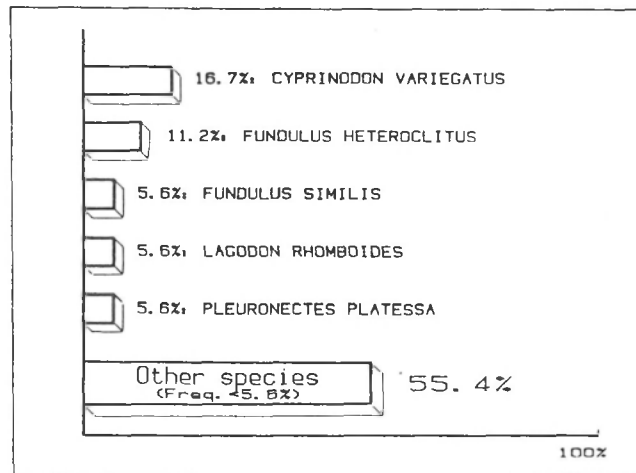


Fig. 11. Frequency (%) of species used in tests using artificial or reconstituted water. On 261 toxicity tests, 36 fulfill the condition, using 25 different species.

Table XV. Occurrence frequency of species used in tests which fulfill the following criteria: obtained from fish farms or laboratories, adults used maintained in artificial medium in static or semi-static conditions, exposure time between 0 and 96 h, lethal effect used as test criterion. Out of 261 tests, 13 fulfill the conditions using 13 different species

Species	Frequency	References
<u>Citharichthys stigmaeus</u>	1	35
<u>Cymatogaster aggregata</u>	1	35
<u>Cyprinodon variegatus</u>	1	35
<u>Fundulus heteroclitus</u>	1	35
<u>Fundulus similis</u>	1	35
<u>Gasterosteus aculeatus</u>	1	35
<u>Lagodon rhomboides</u>	1	35
<u>Leiostomus xanthurus</u>	1	35
<u>Leptocottus armatus</u>	1	35
<u>Medinia sp.</u>	1	35
<u>Paralichthys dentatus</u>	1	35
<u>Paralichthys lethostigma</u>	1	35
<u>Parophrys vetulus</u>	1	35

CONCLUSIONS

In the present state of the art of ecotoxicological testing with marine fish, the information collected has led to the following conclusions :

1. Many methods have been described, which are based on different toxicity criteria, as well as different durations of exposure and involve a large number of species.
2. These methods are usually used to study certain groups of chemical products (pesticides, crude oil dispersing agents, etc.) and are not easily adaptable to investigate all chemical substances.
3. At the moment there is no method which meets the aims pursued for a routine preliminary screening test, suitable for the implementation of international regulations and conventions.
4. Attempts towards international harmonization (ISO, OECD) have not yet led to an agreement on the general principles and more particularly on the choice of a single test species which would satisfy everyone by meeting the criteria retained.

With regard to the choice of the various species, which have been used in the 261 papers that were investigated, the following observations can be made :

1. The species mentioned are very numerous ;
2. A small number of species are mentioned more often (Fig. 12) but for various reasons do not meet the required criteria because of their :
 - geographic specificity ;
 - limited relevance to the marine environment in general ;
 - limited availability ;
 - uncertain resistance to handling in the laboratory.

Under such conditions and owing to the difficulties encountered, it is necessary to reach an international agreement without delay on the choice of one or several species. This would be a temporary compromise which may, if necessary, deviate from the principles preciously agreed upon (i.e. choice of an aquarium fish which is not relevant of the marine environment, choice of a freshwater species adapted to a saltwater environment).

Having reached an international agreement, the expert should introduce proposals for methodologies which would lay the foundations for inter-laboratory tests, prior to the international standardization of a first routine toxicity test.

	CYPRINODON VARIEGATUS	FUNDULUS HETEROCLITUS	ONCORHYNCHUS KISUTCH	PLATICHTHYS FLESUS	PLEURONECTES PLATESSA
ORIGIN	FISHFARMING LABORATORY NATUR. ENVIR.	FISHFARMING LABORATORY NATUR. ENVIR.	FISHFARMING NATUR. ENVIR.	NATUR. ENVIR.	NATUR. ENVIR. LABORATORY
GEOGRAPHICAL DISTRIBUTION	EASTERN USA	N-E ATLANTIC	NORTH PACIFIC	NORTH SEA COASTAL WATER	NORTH SEA BRITISH ISLES ICELAND
MORPHOLOGY	+/- 8 cm	+/- 12 cm	+/- 90 cm 4-5 kg	+/- 51 cm	+/- 50 cm
PHYSIOLOGY	BRACKISH WATER +/- 22 °C	BRACKISH and ESTUARINE WATER -	SALT and FRESH WATER -	ESTUARINE and COASTAL WATER 10 - 20 °C	COASTAL WATER 10 - 20 °C
ADAPTATION TO LAB. TESTS	GOOD	GOOD	LIMITED (adult)	LIMITED (adult)	LIMITED (adult)
RELEVANCE TO MARINE ENVIRON.	LIMITED	LIMITED	LIMITED	LIMITED	LIMITED
AVAILABILITY	USA ONLY	CANADA, USA (easily transportable)	WESTERN USA	EUROPE	EUROPE

Fig. 12. Characteristics of the fish species most commonly used in toxicity tests.

Having taken this first indispensable step, investigations in the field of marine ecotoxicology with fish should be pursued in order to be able to make use of newly developed toxicity tests as soon as possible. These tests would be more relevant for the marine environment and allow a better prediction of the real impact of chemical substances on the marine environment.

LITERATURE CITED

1. Adema D.M.M. 1981.
Ecotoxicological testing in relation to the prior consultation procedure. *Chemosphere* 10(6):515-532.
2. Alberdige D.F, T. Rao, and H. Rosenthal. 1979.
Osmotic responses of eggs and larvae of the Pacific herring to salinity and Cd. *Helgoländer wiss. Meeresunters.* 32:508-538.
3. Alberdige D.F., H. Rosenthal, and F.P. Velsen. 1979.
Influence of salinity and Cadmium on the volume of Pacific herring eggs. *Helgoländer wiss. Meeresunters.* 32:163-178.
4. Alderson R. 1974.
Seawater chlorination and the survival stages of plaice, Pleuronectes platessa L. and dover sole, Solea solea L. *Aquaculture* 4:41-53.
5. Andersen J.W., J.M. Neff, B.A. Cox, H.A. Tatem, and G.M. Hightower. 1974.
Characteristics of dispersions and water soluble extracts of crude and refined oils and their toxicity to estuarine crustaceans and fish. *Mar. Biol.* 27:75-88.
6. Auld A.M. and J.R. Schubel. 1978.
Effects of suspended sediments on fish, eggs and larvae : a laboratory assessment. *Estuar. Coastal Mar. Sc.* 6:153-164.
7. Bengtsson B.-E. 1980.
Long-term effects of PCB (Clophen A 50) on growth, reproduction and swimming performance in the minnow, Phoxinus phoxinus. *Water Res.* 14:681-687.
8. Bengtsson B.-E. 1982.
Long-term studies on uptake and elimination of some chlorinated paraffins in the bleak, Alburnus alburnus. *Ambio* 11(1):38-40.
9. Bengtsson B.-E., C.M. Carlin, A. Larsson, and O. Suanberg. 1975.
Vertebral damage in minnows Phoxinus phoxinus L, exposed to Cd. *Ambio* 4:166-168.
10. Bengtsson B.-E. and A. Larsson. 1981.
Hyperactivity and changed diurnal activity in flounders, Platichthys flesus, exposed to DDT. *Mar. Pollut. Bull.* 12(3):100-102.

11. Buckley J.A., C.M. Whitmore, and R.I. Matsuda. 1976.
Change in blood chemistry and blood cell morphology in coho salmon (Onchorhynchus kisutch) following exposure to sublethal levels of total residual chlorine in municipal wastewater. *J. Fish. Res. Bd. Can.* 33(4):776-782.
12. Buckley J.A. 1976.
Acute toxicity of residual chlorine in waste water to coho salmon (Onchorhynchus kisutch) and some resultant hematologic changes. *J. Fish. Res. Bd. Can.* 33:2854-2856.
13. Burton D.T. 1980.
Evaluation of Pentron D-90 toxicity to juvenile estuarine fish and blue crabs. *Bull. Environ. Contam. Toxicol.* 25(3):470-476.
14. Cameron J.A. and R.L. Smith. 1980.
Ultrastructural effects of crude oil on early life stages of Pacific herring. *Trans. Amer. Fish. Soc.* 109(2):224-226.
15. Canton J.H., R.C.C. Wegman, T.J.A. Vulto, C.H. Verhoef, and G.J. Van Esch. 1978.
Toxicity, accumulation and elimination studies of - hexachlorocyclohexane (HCH) with saltwater organisms of different trophic levels. *Water Res.* 12:687-690.
16. Capuzzo J.M. 1979.
The effect of temperature on the toxicity of chlorinated cooling waters to marine animals - A preliminary review. *Mar. Pollut. Bull.* 10:45-47.
17. Capuzzo J.M., J.A. Davidson, S.A. Lawrence, and M. Libni. 1977.
The differential effects of free and combined chlorine on juvenile marine fish. *Estuar. Coastal Mar. Sci.* 5(6):733-741.
18. Chambers J. 1979.
Enzyme activities following chronic exposure to crude oil in an simulated ecosystem. *Environ. Res.* 20:140-147.
19. Chapman G.A. 1978.
Toxicities of cadmium, copper and zinc to four juvenile stages of chinook salmon and sheepshead. *Trans. Amer. Fish. Soc.* 107(6):841-847.
20. Chapman G.A. and D.G. Stevens. 1978.
Acutely lethal levels of cadmium, copper and zinc to adult male coho salmon and steelhead. *Trans. Amer. Fish. Soc.* 107(6):837-840.

21. Coppage D.L. 1972.
Organophosphate pesticides : specific level of brain Ach E inhibition related to death in sheepshead minnows. *Trans. Amer. Fish. Soc.* 101:534-536.
22. Coppage D. and E. Matthews. 1975.
Brain acetylcholinesterase inhibition in a marine teleost during lethal and sublethal exposures to 1,2-dibromo-2,2-dichloroethyl dimethyl phosphate (Naled) in seawater. *Toxicol. Appl. Pharmacol.* 31:128-133.
23. Crawford R.E. and G.H. Allen. 1977.
Seawater inhibition of nitrite toxicity to chinook salmon. *Trans. Amer. Fish. Soc.* 106(1):105-109.
24. Crawford R.B. and A.M. Guarino. 1976.
Effects of DDT in Fundulus : studies on toxicity, fate and reproduction. *Arch. Environ. Contam. Toxicol.* 4(3):334-348.
25. Dimichele L. and M.H. Taylor. 1978.
Histopathological and physiological responses of Fundulus heteroclitus to naphthalene exposure. *J. Fish. Res. Bd Can.* 35(8):1060-1066.
26. Donnier B. 1972.
Etude de la toxicité d'effluents de papeterie en milieu marin. *Rev. int. Océanogr. méd.* 28:53-93.
27. Ehrlich K. 1977.
Inhibited hatching success of marine fish eggs by power plant effluent. *Mar. Pollut. Bull.* 8(10):228-229.
28. Eisler R. 1965.
Some effects of a synthetic detergent on estuarine fishes. *Trans. Amer. Fish. Soc.* 94:26-31.
29. Eisler R. 1971.
Cadmium poisoning in Fundulus heteroclitus and other marine organisms. *J. Fish. Res. Bd Can.* 28:1225-1234.
30. Eisler R., G.R. Gardner, R.J. Hennekey, G. LaRoche, D.F. Walsh, and P.P. Yevich. 1972.
Acute toxicology of sodium nitrilotriacetic acid (NTA) and NTA containing detergents to marine organisms. *Water Res.* 6(9):1009-1027.
31. Eisler R. and R.J. Hennekey. 1977.
Acute toxicities of Cd^{2+} , Cr^{+6} , Mg^{2+} , Ni^{2+} and Zn^{2+} to estuarine macrofauna. *Arch. Environ. Contam. Toxicol.* 6(23) 315-323.

32. Eldridge M.B., T. Echeverria, and J.A. Whipple. 1977.
Energetics of pacific herring (Clupea harengus pallasi) embryos and larvae exposed to low concentrations of benzene, a monaromatic component of crude oil. *Trans. Amer. Fish. Soc.* 106(5):452-461.
33. Engel D.W. and W.G. Sunda. 1979.
Toxicity of cupric ion to eggs of the spot Leiostomus xanthurus and the atlantic silverside Menidia menidia. *Mar. Biol.* 50(2):121-126.
34. EPA. 1980.
Bioassays for surface waters, wastes and sediments. Bioassay Subcomm. Biolog. Adisory Comm. Cincinnati, Ohio p. 229-236.
35. EPA. (Peltier W.). 1978.
Methods for measuring the acute toxicity of effluents to aquatic organisms. *Environm. Monitor. Suport Lab, Cincinnati, Ohio.* EPA 600/4-7812. 52 p.
36. Ernst V.V. and J.M. Neff. 1977.
The effects of the water-soluble fractions of No. 2 fuel-oil on the early development of the estuarine fish, Fundulus grandis. *Environ. Pollut.* 14(12):25-35.
37. Ferraro L.A., R.E. Wolke, and P.P. Yevich. 1977.
Acute toxicity of water-borne dimethyl nitrosamine (DMN) to Fundulus heteroclitus (L.). *J. Fish. Biol.* 10(3):203-209.
38. Fletcher G.L. and R.J. Hoyle. 1972.
Acute toxicity of yellow phosphorus to Atlantic cod (Gadus morhua) and Atlantic salmon (Salmon salar) smolts. *J. Fish Res. Bd Can.* 29(9):1295-1301.
39. Fletcher G.L., J.W. Kiceniuk, M.J. King, and J.F. Payne, 1979.
Reduction of blood plasma copper concentrations in a marine fish following a six month exposure to crude oil. *Bull. Environ. Contam. Toxicol.* 22:548-551.
40. Flos R. 1979.
Zinc content in organs of dogfish (Scyliorhinus canicula L.) subject to sublethal experimental aquatic zinc pollution. *Comp. Biochem. Physiol.* 64(C)1:77-81.
41. Folmar L.C., D.R. Craddock, J.W. Blackwell. G.J. Joyce, and H.O. Hodgings. 1981.
Effects of petroleum exposure on predatory behaviour of coho salmon (Oncorhynchus kisutch). *Bull. Environ. Contam. Toxicol.* 27:458-462.

42. Folmar L. and H. Hodgins. 1982.
Effects of Arocolor 1254 and No. 2 fuel oil, singly and in combination on predator-prey interactions in coho salmon (Oncorhynchus kisutch). Bull. Environ. Contam. Toxicol. 29:24-28.
43. Goodman L, D. Hansen, and D. Copping. 1979.
Diazinon (trade name) chronic toxicity to and brain acetylcholinesterase inhibition in the sheepshead minnow (Cyprinodon variegatus). Trans. Amer. Fish. Soc. 108:479-488.
44. Graves W.C., D.T. Burton, L.B. Richardson, and S.L. Meyrey. 1981.
The interaction of treated bleached kraft mill effluent and dissolved oxygen concentration on the survival of the development stages of the sheepshead minnow (Cyprinodon variegatus). Water Res. 15:1005-1011.
45. Hall L.W., Jr, D.T. Burton, S.T. Margrey, and W.C. Graves. 1983.
The effect of acclimation temperature on the interaction of chlorine, T and exposure duration to eggs, prolarvae and larvae of striped bass Morone saxatilis. Water Res. 17(3):309-317.
46. Hansen D.J., L.L. Goodman, and A.J. Wilson, Jr. 1977.
Kepone(R) chronic effects on embryo, fry, juvenile and adult sheepshead minnows (Cyprinodon variegatus). EPA Rep. No. 600/J-77-031.
47. Hansen D.J., P.R. Parrish, and J. Forester. 1974.
Arocolor 1016 : Toxicity to and uptake by estuarine animals. Environ. Res. 7:363-373.
48. Hansen D.J., S. Schimmel, and J. Forester. 1977
Endrin : Effects on the entire life cycle of saltwater fish. J. Toxicol. Environ. Health 3:721-733.
49. Haux C. and A. Larsson. 1979.
Effects of DDT on blood plasma electrolytes in the flounder, Platichthys flesus L., in hypotonic brackish water. Ambio 8(4):171-172.
50. Haux C., Å. Larsson, U. Lidman, L. Förlin, T. Hansson, and M.-L. Johansson-Sjöbeck. 1982.
Sublethal physiological effects of chlorinated paraffins on the flounder, Platichthys flesus L. Ecotoxicol. Environ. Safety 6:49-59.
51. Hawkes J.W., E.H. Gruger, and O. Olson. 1980.
Effect of petroleum hydrocarbons and chlorinated biphenyls on the morphology of the intestine of chinook salmon (Oncorhynchus tshawytscha). Environ. Res. 23:141-161.

52. Heitmuller P.T., T.A. Hollister, and P.R. Parrish, 1981.
Acute toxicity of 54 industrial chemicals to sheepshead minnows.
Bull. Environ. Contam. Toxicol. 27(5):596-604.
53. Helmy M.M., A.E. Lemke, P.G. Jacob, and Y.Y. Al-Sultan. 1979.
Haematological changes in Kuwait mullet, Liza macrolepis (Smith),
induced by heavy metals. Indian Mar. Sci. 8: 278-281.
54. Hershberger W.K., K. Bonham, and L.R. Donaldson, 1978.
Chronic exposure of chinook salmon eggs and alevins to gamma
irradiation. Effects on their return to freshwater as adults.
Trans. Amer. Fish. Soc. 107(4):622-631.
55. Hilmy A.M. and M.B. Shabana. 1980.
Blood chemistry levels after acute and chronic exposure to HgCl₂
in the killifish Aphanius dispar (Rupp). Water Air Soil Pollut.
14: 409-417.
56. Hilmy A.M., M.B. Shabana, and M.M. Sajed. 1981.
Comparative study of mercury poisoning on Aphanius dispar
(Teleostei), Sergestes lucens (Crustacea) and Modiolus modiolus
(Mollusca) of the Red Sea. Comp. Biochem. Physiol. 68(C):199-204.
57. Johansson-Sjöbeck M.-L. and A. Larsson. 1978.
The effect of Cd on the hematology and on the activity of F.
aminolevulinic acid dehydratase (ALA-D) in blood and hematopoietic
tissues of the flounder Pleuronectes flesus L. Environ. Res.
17:191-204.
58. Johansson-Sjöbeck M.-L. and A. Larsson. 1979.
Effects of inorganic lead on delta-aminolevulinic acid dehydratase
activity and hematological variables in the rainbow trout Salmo
gairdnerii. Arch. Environ. Contam. Toxicol. (8):419-431.
59. Johnson A.G., T.O. Williams, and C.R. Arnold. 1977.
Chlorine induced mortality of eggs and larvae of spotted seatrout
(Cynoscion nebulosus). Trans. Amer. Fish. Soc. 106(5):466-469.
60. Katz M. and G. Chadwick. 1961.
Toxicity of Endrin to some Pacific northwest fishes. Trans. Amer.
Fish. Soc. 90(4):394-397.
61. Kiceniuk J.W. 1978.
Oil spill dispersant cause bradycardia in a marine fish. Mar.
Pollut. Bull. 9(2):42-45.
62. Kinne O. and H. Rosenthal. 1967.
Effects of sulfuric water pollutants on fertilization, embryonic
development and larvae of the herring Clupea harengus. Mar. Biol.
1:65-83.

63. Korn S., D.A. Moles, and S.D. Rice. 1979.
Effects of T on the median tolerance limit of pink salmon and shrimp exposed to toluene, naphthalene and Cook Inlet crude oil. Bull. Environ. Contam. Toxicol. 21(4-5):521-525.
64. Kühnhold W.W. 1974.
Investigations on the toxicity of seawater extracts of three crude oils on eggs of cod (Gadus morhua L.). Ber. Deutsche Wiss. Komm. Meeresforsch. 23:165-180.
65. Kurelec B., S. Britvic, M. Rijavec, W.E.G. Mueller, and R.K. Zahn. 1977.
Benzo (A) pyrene monooxygenase induction in marine fish-molecular response to oil pollution. Mar. Biol. 44(3):211-216.
66. Larsson A. 1975.
Some biochemical effects of cadmium on fish p. 3-13. In: Sublethal effects of toxic chemicals on aquatic animals. Koeman J.H. and J.J.T.W.A. Strik (Eds). Elsevier, Scient. Publ. Comp., Amsterdam. 234 p.
67. Larsson A., B.-E. Bengtsson, and C. Haux. 1980.
Disturbed ion balance in flounder Platichthys flesus L. exposed to sublethal levels of Cd. Aquat. Toxicol. 1:13-19.
68. Larsson A. and C. Haux. 1982.
Altered carbohydrate metabolism in fish exposed to sublethal levels of Cd. J. Environ. Biol. 3 (2):71-81.
69. Larsson A., K.J. Lehtinen, and C. Haux. 1980.
Biochemical and hematological effects of a titanium dioxide industrial effluent on fish. Bull. Environ. Contam. Toxicol. 25:427-435.
70. Lehtinen K.J. 1980.
Effects on fish exposed to effluent from a titanium dioxide industry and tested with rotary-flow technique. Ambio 9(1):31-33.
71. Lehtinen K.J. and G. Klingstedt. 1983.
X-ray microanalysis in the scanning electron microscope on fish gills effected by acidic, heavy metal containing industrial effluents. Aquat. Toxicol. 3:93-102.
72. Lehtinen K.J. and A. Oikari. 1980.
Sublethal effects of kraft pulp mill waste water on the perch, Perca fluviatilis, studied by rotary-flow and histological techniques. Ann. Zool. Fennici 17(4):255-259.

73. Linden O. 1975.
Acute effect of oil and oil/dispersant mixture on larvae of Baltic herring. *Ambio* 4(3):130-133.
74. Linden O. 1976.
The influence of crude oil and mixtures of crude oil dispersants on the ontogenic development of the Baltic herring Clupea harengus membras L. *Ambio* 5(3):136-140.
75. Linden O. 1978.
Biological effects of oil on early development of the Baltic herring Clupea harengus membras. *Mar. Biol.* 45(3):273-283.
76. Linden O., R. Laughlin, Jr., J.R. Sharp, and J.M. Neff. 1980.
The combined effects of salinity, temperature and oil on the growth pattern of embryos of the killifish, Fundulus heteroclitus Walbaum. *Mar. Environ. Res.* 3:129-144.
77. Linden O., J.R. Sharp, R. Laughlin, and J.M. Neff. 1979.
Interactive effects of salinity, temperature and chronic exposure to oil on the survival and development rate of embryos of the estuarine killifish Fundulus heteroclitus. *Mar. Biol.* 51(2):101-109.
78. Lonning S. 1977.
The effects of crude Ekofisk oil and oil products on marine fish larvae. *Astarte* 10:37-47.
79. Lonning S. and B.E. Hagström. 1976.
Deleterious effects of Corexit 9527 on fertilization and development. *Mar. Pollut. Bull.* 7:124-127.
80. Lowe J.I. 1964.
Chronic exposure of spot Leiostomus xanthurus to sublethal concentrations of toxaphene in seawater. *Trans. Amer. Fish. Soc.* 93:396-399.
81. Mackie A.M., H.T. Singh, and T.C. Fletcher. 1975.
Studies on the cytotoxic effects of sun star (Marthasterias glacialis) saponins and synthetic surfactant in the plaice (Pleuronectes platessa). *Mar. Biol.* 29(4):307-314.
82. Marchetti R. 1978.
Acute toxicity of alkyl leads to some marine organisms. *Mar. Pollut. Bull.* 9:206-207.
83. Mc Kee M.J., A.C. Hendricks, and R.E. Ebel. 1983.
Effects of naphthalene on benzo(a) pyrene hydroxylase and cytochrome P-450 in Fundulus heteroclitus. *Aquat. Toxicol.* 3:103-104.

84. Mc Leay D.L., C.L. Walden, and J.R. Munro. 1979.
Effect of pH on toxicity of kraft pulp and paper mill effluent to salmonid fish in fresh and seawater. *Water Res.* 13:249-254.
85. Mearns A.J., P.S. Oshida, M.J. Sherwood, D.R. Young, and D.J. Reish. 1976.
Chromium effects on coastal organisms. *Water Pollut. Control Fed.* 48(3):1929-1939.
86. Meyerhoff R.D. 1975.
Acute toxicity of benzene, a component of crude oil to juvenile striped bass Morone saxatilis. *J. Fish. Res. Bd Can.* 32:1864-1866.
87. Middaugh O.P., L.E. Burnett, and J.A. Couch. 1980.
Toxicological and physiological responses of the fish Leiostomus xanthurus, exposed to chlorine produced oxidants. *Estuaries* 3(2):132-141.
88. Morgan P.P. and R.D. Prince. 1978.
Chlorine effects on larval development of striped bass (Morone saxatilis) white perch (M. americana) and blue back herring (Alosa aestivalis). *Trans. Amer. Fish Soc.* 107(4):636-641.
89. Morgan P.P. and R.D. Prince. 1979.
Chlorine toxicity to eggs and larvae of five Chesapeake Bay fishes. *Trans Amer. Fish. Soc.* 106:380-385.
90. Nagell B., M. Notini, and O. Grahn. 1974.
Toxicity of four oil dispersants to some animals from the Baltic Sea. *Mar. Biol.* 28(4):237-243.
91. Nimmo D. 1975.
Toxicity of Aroclor[®] 1254 and its physiological activity in several estuarine organisms. *Arch. Environ. Contam. Toxicol.* 3(1):22-39.
92. OECD. 1980.
Report on the assessment of potential environmental effects of chemicals. The effects on organisms other than man and on ecosystems. Vol. 3. TNO Dep. Biology Study and Informat. Centre, Delft, the Netherlands, February 1980. 28 p. + 9 annexes.
93. Oksama M. 1979.
The toxicity of phenol to Phoxinus phoxinus, Gammarus duebeni and Mesidotea entomon in brackish water. *Ann. Zool. Fennici* 16(3):209-216.
94. Olofsson S. and P.E. Lindahl. 1979.
Decreased fitness of cod (Gadus morhua L.) from polluted water. *Mar. Environ. Res.* 2:33-45.

95. Pankhurst N., G. Boyden, and J.B. Wilson. 1980.
The effect of a fluoride effluent on marine organisms. *Environ. Pollut., Ser. A*(23):299-312.
96. Parrish P.R., E.E. Dyar, J.M. Enos, and W.G. Wilson. 1978.
Chronic toxicity of chlordane trifluralin and pentachlorophenol to sheepshead minnows, etc. *Environ. Res. Lab. Gulf Breeze, Florida. EPA/600/3-78/010.* 53 p.
97. Parrish P. R., E.E. Dyar, M.A. Lindberg, C.M. Shanika, and J.M. Enos 1977.
Chronic toxicity of methoxychlor, malathion and carbofuran to sheepshead minnows (*Cyprinodon variegatus*). *Environ. Res. Lab. Gulf Breeze, Florida. EPA 600/3-77-059.* 36 p.
98. Payne J. and L. Fancey. 1982.
Effect of long-term exposure to petroleum on mixed function oxygenases in fish : further support for use of the enzyme system in biological monitoring. *Chemosphere* 11(2):207-213.
99. Payne J.E., J.W. Kiceniuk, and W.R. Squires. 1978.
Pathological changes in a marine fish after a 6 month exposure to petroleum. *J. Fish. Res. Bd Can.* 35(5):665-667.
100. Pérès G. and M.H. Brichon. 1979.
Le loup (*Dicentrarchus labrax*) poisson de laboratoire : intérêt et difficultés de son utilisation en écotoxicologie. *Bull. Soc. Sc. Vét. Méd. Comp.* 81(4):219-222.
101. Powell J.H. and D.R. Fielder. 1982.
Temperature and toxicity of DDT to sea mullet (*Mugil cephalus* L.). *Mar. Pollut. Bull.* 13(7):228-230.
102. Rice D.W. and F.L. Harrison. 1980.
Effects of copper on early life history stages of northern anchovy, *Engraulis mordax*. *Fish. Bull.* 78(3):675-684.
103. Roberts M., Jr. and R. Gleeson. 1978.
Acute toxicity of bromo-chlorinated seawater to selected estuarine species with a comparison to chlorinated seawater toxicity. *Mar. Environ. Res.* 1:19-30.
104. Saspry K.V. and P.K. Gupta. 1980.
Alterations in the activities of a few dehydrogenases in the digestive system of two teleost fishes exposed to lead nitrate. *Ecotoxicol. Environ. Safety* 4:232-239.
105. Saward D., A. Stirling, and G. Topping. 1975.
Experimental studies on the effects of copper on a marine food chain. *Mar. Biol.* 29(4):351-361.

106. Scarfe A.D., K.A. Jones, C.W. Steele, H. Kleerekoper, and M. Corbett. 1982.
Locomotor behaviour of four marine teleosts in response to sublethal copper exposure. *Aquat. Toxicol.* 2:335-353.
107. Schimmel S.C. 1977.
Uptake and toxicity of toxaphene in several estuarine organisms. *Arch. Environ. Contam. Toxicol.* 5(3):353-367.
108. Schimmel S.C., D.J. Hansen, and J. Forester. 1974.
Effects of Arocolor 1254 on laboratory-reared embryos and fry of sheepshead minnows (Cyprinodon variegatus). *Trans. Amer. Fish. Soc.* 103(3):582-586.
109. Schimmel S.C., J. Patrick, and J. Forester. 1977.
Toxicity and bioconcentration of BHC and Lindane in selected estuarine animals. *Arch. Environ. Contam. Toxicol.* 6:355-363.
110. Schimmel S.C. and A.J. Wilson, Jr. 1977.
Acute toxicity of kepone (Trademark) to four estuarine animals. *Chesapeake Sci.* 18(2):224-227.
111. Seegert G.L. and A.S. Brooks. 1978.
The effects of intermittent chlorination on coho salmon, alewife, spottail shiner, and rainbow smelt. *Trans. Amer. Fish. Soc.* 107(2):346-353.
112. Smith R.L. and J.A. Cameron. 1979.
Effect of water soluble fraction of Prudhoe Bay crude oil on embryonic development of Pacific herring. *Trans. Amer. Fish. Soc.* 108(1):70-75.
113. Smith R.M. and C.F. Cole. 1973.
Effects of egg concentrations of DDT and dieldrin on development in winter flounder (Pseudopleuronectes americanus). *J. Fish. Res. Bd Can.* 30(12):1894-1898.
114. Somero G.N., P.H. Yancey, T.J. Chow, and C.E. Snyder. 1977.
Lead effects on tissue and whole organism respiration of the estuarine teleost fish, Gillichthys mirabilis. *Arch. Environ. Contam. Toxicol.* 6(2-3):349-354.
115. Stober Q.J., P.A. Dinnel, E.F. Huriburt, and H. DiJulio. 1980.
Acute toxicity and behavioural responses of coho salmon (Onchorhynchus kisutch) and shiner perch (Cymatogaster aggregata) to chlorine in heated sea-water. *Water Res.* 14(4):347-354.

116. Stober Q. and C.H. Hanson. 1974.
Toxicity of chlorine and lead to pink (*Oncorhynchus gorbuscha*) and chinook salmon (*O. tshawytscha*). *Trans. Amer. Fish. Soc.* 103(3):569-576.
117. Stoner A.W. and R.J. Livingston. 1978.
Respiration, growth and food conversion efficiency of pin fish (*Lagodon rhomboides*) exposed to sublethal concentrations of bleached kraft mill effluent. *Environ. Pollut.* 17(3):207-217.
118. Suggat R.H. 1980.
Effects of sublethal sodium dichromate exposure in freshwater on the salinity tolerance and serum osmolality of juvenile coho salmon, *Oncorhynchus kisutch* in seawater. *Arch. Environ. Contam. Toxicol.* 9(1):41-52.
119. Thatcher T. 1979.
A morphological defect in shiner perch resulting from chronic exposure to chlorinated seawater. *Bull. Environ. Contam. Toxicol.* 21(4-5):433-439.
120. Thomas P., B.R. Woodin, and J.M. Nefe. 1980.
Biochemical responses of the striped mullet *Mugil cephalus* to oil exposure. I. Acute responses interrenal activations and secondary stress responses. *Mar. Biol.* 59:141-149.
121. Thompson G.B. and R.R.S.S. Wy. 1981.
Toxicity testing of oil slick dispersants in Hong Kong. *Mar. Pollut. Bull.* 12(7):233-237.
122. Thompson J.A.J., J.C. Davis, and R.E. Drew. 1976.
Toxicity, uptake and survey studies of boron in the marine environment. *Water Res.* 10(10):869-875.
123. Thurberg F. and R. Collier. 1977.
Respiratory response of cunners to silver. *Mar. Pollut. Bull.* 8(2):40-41.
124. von Westerhagen H. 1979.
Combined effects of Cd, Cu and Pb on developing herring eggs and larvae. *Helgoländer wiss. Meeresunters.* 32(3):257-278.
125. von Westernhagen H., V. Dethlefsen, and H. Rosenthal. 1975.
Combined effects of Cd and salinity on development and survival of garpike eggs. *Helgoländer wiss. Meeresunters.* 27:268-282.
126. von Westernhagen H., H. Rosenthal, and K.R. Sperling. 1974.
Combined effects of Cd and salinity on development and survival of herring eggs. *Helgoländer wiss. Meeresunters.* 26:(3-4):416-433.

127. Voyer R.A. 1975.
Effect of dissolved oxygen concentration on the acute toxicity of Cd to the mummichog Fundulus heteroclitus (L.) at various salinities. Trans. Amer. Fish. Soc. 104(1):129-134.
128. Voyer R.A. 1982.
Viability of embryos of the winter flounder Pseudopleuronectes americanus exposed to mixtures of cadmium and silver in combination with selected fixed salinities. Aquat. Toxicol. 2:223-233.
129. Voyer R.A., J.F. Heltsche, and R.A. Kraus. 1979.
Hatching success and larval mortality in an estuarine teleost, Menidia menidia, exposed to Cd in constant and fluctuating salinity regimes. Bull. Environ. Contam. Toxicol. 23(4-5):475-481.
130. Voyer R.A., C.E. Wentworth, Jr., E.P. Barry, and R.J. Hennekey. 1977.
Viability of embryos of the winter flounder (Pseudopleuronectes americanus) exposed to combinations of cadmium and salinity at selected temperatures. Mar. Biol. 44(2):117-124.
131. Walsh G.E. 1980.
Toxicity of textile mill effluents to fresh water and estuarine algae, crustaceans and fishes. Environ. Pollut. Ser. Amer. 21(3):169-179.
132. Ward G.S., G.C. Cramm, P.R. Parrish, and S.R. Petrocelli. 1982.
Effect of ammonium jarosite on early life stages of a saltwater fish (Cyprinodon variegatus). Mar. Pollut. Bull. 13(6):191-195.
133. Wang R.T. and J.A.C. Nicol. 1977.
Effect of fuel oil on sea catfish: feeding activity and cardiac responses. Bull. Environ. Contam. Toxicol. 18(2):170-176.
134. Weis J.S. and P. Weis. 1977.
Effects of heavy metals on development of the killifish, Fundulus heteroclitus. J. Fish. Biol. 11:49-54.
135. Weis J.S., P. Weis, M. Heber, and S. Vaidya. 1981.
Methylmercury tolerance of killifish (Fundulus heteroclitus) embryos from a polluted environment. Mar. Biol. 65(3):283-287.
136. Weis P. and J.S. Weis. 1976.
Effects of heavy metals on fin regeneration in the killifish Fundulus. Bull. Environ. Contam. Toxicol. 16:197-202.
137. Wildish D.J. 1972.
Acute toxicity of polyoxyethelene esters and polyoxyethelene esthers to S. salar and G. oceanicus. Water. Res. 6(7):759-762.

138. Wildish D.J., H. Akagi, and N.J. Poole. 1977.
Avoidance by herring of dissolved components in pulp mill effluents. *Bull. Environ. Contam. Toxicol.* 18(5):521-525.
139. Wilson K.W. 1976.
Effects of oil dispersants on the developing embryos of marine fish. *Mar. Biol.* 36:256-268.
140. Wilson K.W. 1977.
Acute toxicity of oil to dispersants to marine fish larvae. *Mar. Biol.* 40(1):65-74.
141. Zitko V., D.W. McLeese, W.G. Carson, and H.E. Welch. 1976.
Toxicity of alkyl-dinitrophenols to some aquatic organisms. *Bull. Environ. Contam. Toxicol.* 16(5):508-518.

GENERAL LITERATURE

Note : The authors used these references to compile the list of international conventions and Tables I, II, and XVI.

Anonymous.

Standard methods for the evaluation of water and wastewater. (15th ed.). Amer. Publ. Health Ass., Washington DC.

Hart J.L. 1973.

Pacific fishes of Canada. *Fish. Res. Bd Can., Bull.* 180. 740 p.

IRPTC. 1978.

Data profiles for chemicals for the evaluation of their hazards to the environment of the Mediterranean Sea. Vol. 1. Data Profile Ser. 1, Int. Register Potentially Toxic Chemicals (IRPTC), UN Environ. Program, Geneva. 928 p.

Leim A.H. and W.B. Scott. 1966.

Fishes of the Atlantic coast of Canada. *Fish. Res. Bd Can., Bull.* 155. 485 p.

Sterba G. 1966.

Freshwater fishes of the world. Studio Vista Ltd, London. 877 p.

Wheeler A. 1969.

The fishes of the British Isles and North West Europe. Macmillan and Co. Ltd, London. 613 p.