

ORIGINAL ARTICLE

Genome-wide analysis implicates microRNAs and their target genes in the development of bipolar disorder

AJ Forstner^{1,2,50}, A Hofmann^{1,2,50}, A Maaser^{1,2}, S Sumer³, S Khudayberdiev³, TW Mühleisen^{1,2,4}, M Leber⁵, TG Schulze⁶, J Strohmaier⁷, F Degenhardt^{1,2}, J Treutlein⁷, M Mattheisen^{8,9}, J Schumacher^{1,2}, R Breuer⁷, S Meier^{7,10}, S Herms^{1,2,11}, P Hoffmann^{1,2,4,11}, A Lacour¹², SH Witt⁷, A Reif¹³, B Müller-Myhsok^{14,15,16}, S Lucae¹⁴, W Maier¹⁷, M Schwarz¹⁸, H Vedder¹⁸, J Kammerer-Ciernioch¹⁹, A Pfennig²⁰, M Bauer²⁰, M Hautzinger²¹, S Moebus²², L Priebe^{1,2}, S Sivalingam^{1,2}, A Verhaert^{1,2}, H Schulz²³, PM Czerski²⁴, J Hauser²⁴, J Lissowska²⁵, N Szeszenia-Dabrowska²⁶, P Brennan²⁷, JD McKay²⁸, A Wright^{29,30}, PB Mitchell^{29,30}, JM Fullerton^{31,32}, PR Schofield^{31,32}, GW Montgomery³³, SE Medland³³, SD Gordon³³, NG Martin³³, V Krasnov³⁴, A Chuchalin³⁵, G Babadjanova³⁵, G Pantelejeva³⁶, LI Abramova³⁶, AS Tiganov³⁶, A Polonikov³⁷, E Khusnutdinova^{38,39}, M Alda^{40,41}, C Cruceanu^{42,43,44}, GA Rouleau⁴², G Turecki^{43,44,45}, C Laprise⁴⁶, F Rivas⁴⁷, F Mayoral⁴⁷, M Kogevinas⁴⁸, M Grigoriou-Serbanescu⁴⁹, P Propping¹, T Becker^{5,12}, M Rietschel⁷, S Cichon^{1,2,4,11}, G Schratz³ and MM Nöthen^{1,2}

Bipolar disorder (BD) is a severe and highly heritable neuropsychiatric disorder with a lifetime prevalence of 1%. Molecular genetic studies have identified the first BD susceptibility genes. However, the disease pathways remain largely unknown. Accumulating evidence suggests that microRNAs, a class of small noncoding RNAs, contribute to basic mechanisms underlying brain development and plasticity, suggesting their possible involvement in the pathogenesis of several psychiatric disorders, including BD. In the present study, gene-based analyses were performed for all known autosomal microRNAs using the largest genome-wide association data set of BD to date (9747 patients and 14 278 controls). Associated and brain-expressed microRNAs were then investigated in target gene and pathway analyses. Functional analyses of *miR-499* and *miR-708* were performed in rat hippocampal neurons. Ninety-eight of the six hundred nine investigated microRNAs showed nominally significant *P*-values, suggesting that BD-associated microRNAs might be enriched within known microRNA loci. After correction for multiple testing, nine microRNAs showed a significant association with BD. The most promising were *miR-499*, *miR-708* and *miR-1908*. Target gene and pathway analyses revealed 18 significant canonical pathways, including brain development and neuron projection. For *miR-499*, four Bonferroni-corrected significant target genes were identified, including the genome-wide risk gene for psychiatric disorder *CACNB2*. First results of functional analyses in rat hippocampal neurons neither revealed nor excluded a major contribution of *miR-499* or *miR-708* to dendritic spine morphogenesis. The present results suggest that research is warranted to elucidate the precise involvement of microRNAs and their downstream pathways in BD.

Translational Psychiatry (2015) 5, e678; doi:10.1038/tp.2015.159; published online 10 November 2015

¹Institute of Human Genetics, University of Bonn, Bonn, Germany; ²Department of Genomics, Life and Brain Center, University of Bonn, Bonn, Germany; ³Institute of Physiological Chemistry, Philipps-University Marburg, Marburg, Germany; ⁴Institute of Neuroscience and Medicine, Research Center Juelich, Juelich, Germany; ⁵Institute for Medical Biometry, Informatics and Epidemiology, University of Bonn, Bonn, Germany; ⁶Institute of Psychiatric Phenomics and Genomics, Ludwig-Maximilians-University Munich, Munich, Germany; ⁷Department of Genetic Epidemiology in Psychiatry, Central Institute of Mental Health, Medical Faculty Mannheim/University of Heidelberg, Heidelberg, Germany; ⁸Department of Biomedicine, Aarhus University, Aarhus, Denmark; ⁹Institute for Genomics Mathematics, University of Bonn, Bonn, Germany; ¹⁰National Center Register-Based Research, Aarhus University, Aarhus, Denmark; ¹¹Division of Medical Genetics, Department of Biomedicine, University of Basel, Basel, Switzerland; ¹²German Center for Neurodegenerative Diseases, Bonn, Germany; ¹³Department of Psychiatry, Psychosomatic Medicine and Psychotherapy, University Hospital Frankfurt am Main, Frankfurt, Germany; ¹⁴Max Planck Institute of Psychiatry, Munich, Germany; ¹⁵Munich Cluster for Systems Neurology (SyNergy), Munich, Germany; ¹⁶University of Liverpool, Institute of Translational Medicine, Liverpool, UK; ¹⁷Department of Psychiatry, University of Bonn, Bonn, Germany; ¹⁸Psychiatric Center Nordbaden, Wiesloch, Germany; ¹⁹Center of Psychiatry Weinsberg, Weinsberg, Germany; ²⁰Department of Psychiatry and Psychotherapy, University Hospital Carl Gustav Carus, TU Dresden, Dresden, Germany; ²¹Department of Psychology, Clinical Psychology and Psychotherapy, Eberhard Karls University Tübingen, Tübingen, Germany; ²²Institute of Medical Informatics, Biometry and Epidemiology, University Duisburg-Essen, Essen, Germany; ²³Cologne Center for Genomics, University of Cologne, Cologne, Germany; ²⁴Department of Psychiatry, Laboratory of Psychiatric Genetics, Poznan University of Medical Sciences, Poznan, Poland; ²⁵Department of Cancer Epidemiology and Prevention, Maria Skłodowska-Curie Memorial Cancer Centre and Institute of Oncology Warsaw, Warsaw, Poland; ²⁶Department of Epidemiology, Nofer Institute of Occupational Medicine, Lodz, Poland; ²⁷Genetic Epidemiology Group, International Agency for Research on Cancer, Lyon, France; ²⁸Genetic Cancer Susceptibility Group, International Agency for Research on Cancer, Lyon, France; ²⁹School of Psychiatry, University of New South Wales, Randwick, NSW, Australia; ³⁰Black Dog Institute, Prince of Wales Hospital, Randwick, NSW, Australia; ³¹Neuroscience Research Australia, Sydney, NSW, Australia; ³²School of Medical Sciences, Faculty of Medicine, University of New South Wales, Sydney, Australia; ³³Queensland Institute of Medical Research, Brisbane, QLD, Australia; ³⁴Moscow Research Institute of Psychiatry, Moscow, Russian Federation; ³⁵Institute of Pulmonology, Russian State Medical University, Moscow, Russian Federation; ³⁶Russian Academy of Medical Sciences, Mental Health Research Center, Moscow, Russian Federation; ³⁷Department of Biology, Medical Genetics and Ecology, Kursk State Medical University, Kursk, Russian Federation; ³⁸Institute of Biochemistry and Genetics, Ufa Scientific Center of Russian Academy of Sciences, Ufa, Russian Federation; ³⁹Department of Genetics and Fundamental Medicine, Bashkir State University, Ufa, Russian Federation; ⁴⁰Department of Psychiatry, Dalhousie University, Halifax, NS, Canada; ⁴¹National Institute of Mental Health, Klecany, Czech Republic; ⁴²Montreal Neurological Institute, McGill University, Montreal, QC, Canada; ⁴³Department of Human Genetics, McGill University, Montreal, QC, Canada; ⁴⁴McGill Group for Suicide Studies and Douglas Research Institute, Montreal, QC, Canada; ⁴⁵Department of Psychiatry, McGill University, Montreal, QC, Canada; ⁴⁶Département des sciences fondamentales, Université du Québec à Chicoutimi (UQAC), Chicoutimi, QC, Canada; ⁴⁷Department of Psychiatry, Hospital Regional Universitario, Biomedical Institute of Malaga, Malaga, Spain; ⁴⁸Center for Research in Environmental Epidemiology, Barcelona, Spain and ⁴⁹Biometric Psychiatric Genetics Research Unit, Alexandru Obregia Clinical Psychiatric Hospital, Bucharest, Romania. Correspondence: Professor MM Nöthen, Institute of Human Genetics, University of Bonn, Sigmund-Freud-Strasse 25, Bonn 53127, Germany. E-mail: markus.noethen@uni-bonn.de

⁵⁰These authors contributed equally to this work.

Received 20 August 2015; accepted 7 September 2015

INTRODUCTION

Bipolar disorder (BD) is a severe neuropsychiatric disorder with an estimated lifetime prevalence of 1%.¹ BD is characterized by recurrent episodes of mania and depression, and shows a heritability of ~70%.² Molecular genetic candidate studies and—more recently—genome-wide association studies (GWAS) have identified the first BD susceptibility genes.^{3–7} However, the disease pathways and underlying regulatory networks remain largely unknown.⁸

Accumulating evidence suggests that microRNAs (miRNAs) are implicated in the biological pathways that regulate brain development and synaptic plasticity.^{9,10} This in turn suggests their possible involvement in the pathogenesis of several psychiatric disorders,^{11,12} including BD.^{13,14} Studies of the post-mortem brain tissue of BD patients have demonstrated altered miRNA expression profiles in the prefrontal cortex.^{13,14}

The miRNAs are a class of 21–25-nucleotide small noncoding RNAs. In the nucleus they are transcribed by RNA polymerase II to primary miRNA (pri-miRNA) transcripts, which are double-stranded stem loop structures comprising 100–1000 nucleotides.^{15,16} Approximately 50% of all vertebrate miRNAs are processed from the introns of protein-coding genes or from genes encoding other noncoding RNA classes. However, miRNAs can also be encoded in intergenic regions.¹⁷

The pri-miRNAs are then processed by the Drosha-DGCR8 complex to precursor miRNAs.^{18,19} These precursor miRNAs are 60–70 nucleotides in length. The precursor miRNAs are exported to the cytoplasm, where they are cleaved into ~20-base pair (bp) mature miRNAs by the Dicer enzyme.^{16,20} The mature miRNAs are incorporated into the RNA-induced silencing complex, which then targets distinct sets of messenger RNAs (mRNAs).²¹

The miRNAs control the expression of their target genes by binding to target sites within the mRNAs, typically in their 3' untranslated regions.^{22,23} A region of 2–7 or 2–8 consecutive nucleotides from the 5' end of the mature miRNA forms the seed region, which is crucial for the recognition of the target genes.²⁴ In general, each miRNA controls up to several hundred target mRNAs, whereas one mRNA target can be subjected to synergistic regulation by multiple miRNAs.^{25,26} In consequence, miRNAs integrate different intracellular signals and regulate a number of signaling pathways.^{27,28} Interestingly, the miRNA regulatory effect itself has been shown to be a heritable trait in humans.²⁹

The hypothesis that miRNAs are implicated in BD is also supported by the results of the largest GWAS of BD to date.⁶ In this study, a single-nucleotide polymorphism (SNP) in an intergenic region flanking *MIR2113* on chromosome 6q16.1 was the eighth strongest finding. However, no significant enrichment of BD-associated genes within the known or predicted targets of *MIR2113* was observed.⁶

Several studies have investigated the role of single miRNAs in the development of psychiatric disorder,^{30–32} including BD.³³ However, to our knowledge, no systematic, genome-wide analysis of miRNA-coding genes has yet been performed. The aim of the present study was, thus, to determine whether common variants at any of the known miRNA loci contribute to the development of BD.

MATERIALS AND METHODS

Sample description

The gene-based tests were performed using data from our previous GWAS of BD (9747 patients and 14 278 controls).⁶ This GWAS data set combined data from Canada, Australia and four European countries (MooDS) with the GWAS results of the multinational Psychiatric Genomics Consortium (PGC).³ The study was approved by the respective local Ethics Committees. Written informed consent was obtained from all participants.⁶

Genome-wide miRNA association analysis

For the gene-based analyses, a set-based testing approach adapted from the versatile gene-based test for GWAS³⁴ was used. This algorithm is obtainable upon request. The chromosomal positions of all miRNAs ($n = 718$) were obtained from miRBase release 13.0.³⁵ This release contains a high confidence set of miRNAs for which detailed information about miRNA function and predicted target genes is available. Using the summary statistics, gene-wide P -values were calculated for all 636 autosomal miRNAs and their ± 20 kilobase (kb) flanking sequences. Twenty-seven of these miRNA loci contained no common SNP. Therefore, gene-wide P -values were obtained for 609 miRNAs.

The applied statistical algorithm is described in more detail in the article by Liu et al.³⁴ Briefly, SNPs within these boundaries were grouped together, and a set-based test statistic was calculated as the sum of the χ^2 one degree of freedom association P -values within the miRNA. The test statistic was compared with simulated test statistics from the multivariate normal distribution. An empirical miRNA-based P -value was calculated as the proportion of simulated test statistics above the observed test statistic. For the purposes of the present study, the 10% most significant SNPs for each miRNA were summarized. The calculated gene-based P -values were Bonferroni-corrected for multiple testing according to the number of investigated miRNAs ($n = 609$).

As different reference panels were used for the imputation of the MooDS and PGC genotype data (1000 Genomes Project, February 2012 release, and HapMap phase 2 CEU, respectively), we used simulated test statistics on the basis of an intermarker linkage disequilibrium (LD) structure as derived from the HapMap phase 2 population genotypes. However, for miRNAs that showed a significant association with BD after Bonferroni correction, we also calculated gene-based tests based on 1000 Genomes Project phase 3 population genotypes.

Inflation of the observed and expected P -values for different SNP subcategories (SNPs in miRNA loci, SNPs in genes and intergenic SNPs) was defined as the degree of deviation from the expected uniform distribution in the quantile–quantile (Q–Q) plot and tested for significance using Fisher's exact test (one-sided) for different P -value thresholds. Only LD-pruned SNPs ($r^2 < 0.8$) were used for the enrichment analysis.

Follow-up of miRNA association results—regional association plots

A window-based approach that included common variants in miRNAs and flanking sequences was applied. To determine whether the signal was associated with any of the miRNAs of interest, visual inspection of the regional association plots was performed.

Regional association results from our BD GWAS⁶ were plotted for all associated miRNAs and their ± 500 -kb flanking regions using LocusZoom.³⁶ A signal was considered miRNA-associated if the top SNP of the region was located at, or was in high or moderate LD ($r^2 > 0.6$) with, the miRNA locus.

Follow-up of miRNA association results—miRNA brain expression

To investigate expression of the associated miRNAs in the human brain, data from a recent study of miRNA expression patterns in the developing human brain were re-analyzed.³⁷ A miRNA was defined as showing brain expression if it had a total read count of > 120 across all investigated samples.³⁷

In addition, miRNA expression was measured in rat cortical neurons and forebrain. All procedures involving animals followed the guidelines of the German Animal Protection Legislation and the experiments were approved by the Local Committee for Animal Health (RP Gießen). Total RNA was isolated from the postnatal day 15 rat forebrain or synaptosomes, as described elsewhere.³⁸ Briefly, the total RNA from the forebrain of postnatal day-15 Sprague–Dawley rat pups was extracted using peqGOLD TriFast reagent (Peqlab, Erlangen, Germany) in accordance with the manufacturer's instructions. Small RNA libraries were constructed and sequenced at the EMBL genomic core facility (Heidelberg, Germany) using the HiSeq platform (Illumina, San Diego, CA, USA). The web-based software MiRanalyzer was used to determine miRNA expression levels (<http://bioinfo2.ugr.es/miRanalyzer/miRanalyzer.php>).³⁹

miRNA target gene analysis

Targets of the associated miRNAs 499, 708 and 1908 were obtained from TargetScan (Release 6.2).⁴⁰ The Allen human brain atlas (<http://www.brain-map.org/>)⁴¹ was consulted to determine whether predicted target genes are expressed in the human brain. Target genes were considered brain-

expressed if they had shown expression in the hippocampal formation in at least four of the six donor brains. Gene-based *P*-values for all brain-expressed miRNA targets were calculated using versatile gene-based test for GWAS,³⁴ and our BD GWAS data set.⁶ To capture regulatory regions, the default settings in versatile gene-based test for GWAS were used. Enrichment of associated targets was calculated as follows: the number of associated target genes for each miRNA was compared with the number of associated genes from 100 000 random target sets of brain-expressed genes. Each target gene set comprised the same number of genes as the miRNA target genes itself.

Pathway analysis of target genes

The subsequent analyses were restricted to brain-expressed target genes of *miR-499*, *miR-708* and *miR-1908*, with a gene-based association *P*-value of < 0.05. If the chromosomal distance between two target genes was below 100 kb or if the top SNPs of two target genes were in strong or moderate LD ($D' > 0.4$), only the target gene with the lowest gene-based

P-value was retained in the pathway analysis to ensure the independency of association signals. In total, 107 target genes were included in the pathway analyses (Supplementary Box 1). Gene ontology (GO) and Kyoto Encyclopaedia of Genes and Genomes pathway testing was performed using the WebGestalt (Web-based Gene Set Analysis Toolkit) for the brain-expressed, BD-associated target genes of the three associated miRNAs. Bonferroni correction was used to adjust for multiple testings. Significant pathways were filtered to achieve a minimum of three genes per set.

Functional analyses of miR-499 and miR-708 in rat hippocampal neurons

To test the possible involvement of *miR-499* or *miR-708* in the regulation of synaptic function, experiments were performed to investigate the effect of *miR-499* and *miR-708* overexpression on dendritic spine morphogenesis in primary rat hippocampal neurons. We initially focused on overexpression, as this can be easily achieved by the transfection of expression plasmids containing pri-miRNA cassettes. miRNA overexpression constructs were generated by inserting the respective pri-miRNA sequences into the 3'-untranslated repeat of the luciferase reporter gene within pmirGLO (Promega, Madison, WI, USA). Thereby, luciferase reporter assays could be used to monitor the efficiency of pri-miRNA processing. To investigate the potential involvement of *miR-499-5p* and *miR-708-5p* in dendritic spine morphogenesis, hippocampal neurons of embryonic day-18 Sprague-Dawley rats (Charles River Laboratories, Sulzfeld, Germany) were transfected with miRNA-overexpressing constructs for 6 days before fixation. Images with a resolution of 1024 × 1024 pixels were obtained using a LSM5 Zeiss Pascal confocal microscope (Jena, Germany) and in a magnification of × 63 / 1.4. A maximum projection was reconstructed with the Zeiss LSM 510 Meta software from a z-stack consisting of seven optical slices at 0.45-µm interval. The average intensity of an area of 2180 nm² containing 250–300 spines per cell was measured using the ImageJ 1.48v software (National Institutes of Health, Bethesda, MD, USA), as described elsewhere.³⁸ During imaging and analysis, the investigator was blind to the transfection condition.

RESULTS

Overall, the nominal *P*-values of SNPs at miRNA loci were enriched with lower values than would be expected with a uniform *P*-value distribution (Figure 1). This deviation from the expected normal Q–Q plot distribution indicates a general enrichment for miRNAs among BD-associated SNPs. Category testing for different *P*-value thresholds revealed a significant enrichment for BD-associated SNPs in miRNA loci for *P*-values < 1×10^{-4} (Supplementary Table 1). This deviation was also observed among SNPs in genes but not for intergenic SNPs.

Gene-based analysis in our BD GWAS data⁶ generated nominally significant *P*-values for 98 of the 609 miRNAs. These included *miR-2113*, which was located at the genome-wide significant locus on chromosome 6q16.1 in the original BD GWAS analyses.⁶ After correction for multiple testing, nine miRNAs

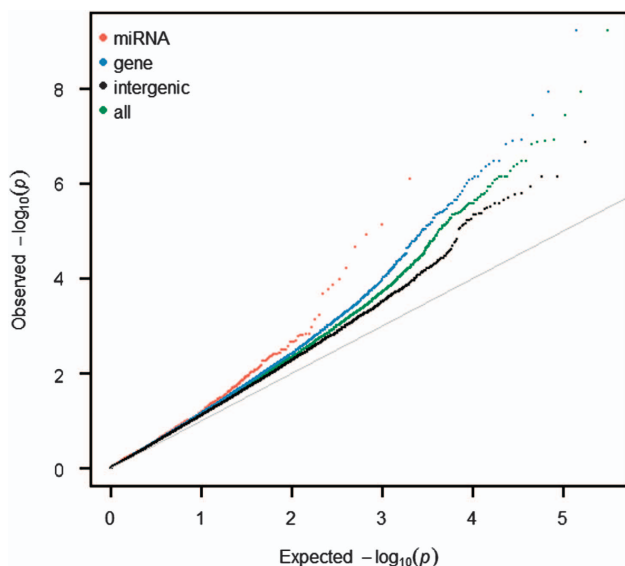


Figure 1. Quantile–quantile (Q–Q) plot of single-nucleotide polymorphism (SNP) *P*-values. The $-\log_{10}$ of the observed genome-wide association studies (GWAS) *P*-values for linkage disequilibrium (LD)-pruned SNPs (on the y axis) are plotted versus the $-\log_{10}$ of the expected *P*-values (under null, on the x axis). The solid line represents expected uniform distribution. Red dots represent the data distribution of *P*-values of SNPs at microRNA loci; blue dots represent SNPs in genes; black dots represent *P*-values of intergenic SNPs; and green dots represent the data distribution of all SNPs.

Table 1. Results of the gene-based tests for the nine microRNAs that withstood Bonferroni correction

| miRNA | Chr | nSNPs | Top SNP | <i>p</i> Top SNP | <i>p</i> Corr Gene | miRNA-assoc. signal | Expr. hum. brain |
|-------------------|-----|-------|-----------|-----------------------|--------------------|---------------------|------------------|
| <i>miR-499</i> | 20 | 27 | rs3818253 | 6.58×10^{-7} | 0.0012 | Yes | Yes |
| <i>miR-640</i> | 19 | 21 | rs2965184 | 7.23×10^{-7} | 0.0012 | Yes | No |
| <i>miR-708</i> | 11 | 72 | rs7108878 | 3.45×10^{-7} | 0.0012 | Yes | Yes |
| <i>miR-581</i> | 5 | 36 | rs697112 | 3.61×10^{-6} | 0.0073 | Yes | No |
| <i>miR-644</i> | 20 | 12 | rs7269526 | 1.22×10^{-5} | 0.0104 | No | No |
| <i>miR-135a-1</i> | 3 | 20 | rs9311474 | 2.16×10^{-5} | 0.0122 | No | Yes |
| <i>let-7 g</i> | 3 | 9 | rs6445358 | 2.23×10^{-5} | 0.0305 | No | Yes |
| <i>miR-1908</i> | 11 | 16 | rs174575 | 2.85×10^{-5} | 0.0353 | Yes | Yes |
| <i>miR-611</i> | 11 | 23 | rs174535 | 5.03×10^{-5} | 0.0457 | No | No |

Abbreviations: Chr, chromosome; expr. hum. brain, expression in the human brain according to Ziats and Rennert;³⁷ miRNA, microRNA; miRNA-assoc. signal, specificity of the associated finding in the regional association plot; *p* Corr Gene, Bonferroni-corrected gene-based *P*-value; *p* Top SNP, *P*-value of the Top SNP within gene; nSNPs, number of investigated SNPs; SNP, single-nucleotide polymorphism.

showed a significant association with BD (Table 1). The additional calculation of gene-based tests for these nine miRNAs on the basis of 1000 Genomes LD structure generated nominal P -values of $\leq 7.20 \times 10^{-5}$ (Supplementary Table 2).

Visual inspection of the regional association plots revealed a miRNA-associated signal for five of the nine miRNAs (Figure 2, Supplementary Figures 1–4).

The re-analysis of the expression data from Ziats and Rennert³⁷ revealed that five of the nine miRNAs were expressed in the human brain (Table 1).

Three of these (*miR-499*, *miR-708* and *miR-135a-1*) were also found to be expressed in the rat forebrain. This method could not be used to investigate the expression of the other miRNAs, as they are not expressed in rats.³⁵

The regional association plots and the miRNA expression data in human brain tissue suggest that the three brain-expressed miRNAs, that is, *miR-499*, *miR-708* and *miR-1908*, are the most promising candidates for further analyses. The three miRNAs had 296, 181 and 67 target genes, respectively. Of these 286, 174 and 56 showed brain expression (Table 2).

The target gene enrichment analysis showed no significant enrichment of BD-associated genes within the targets of *miR-499*, *miR-708* or *miR-1908* (Table 2). After Bonferroni correction, *miR-1908* had one (*KLC2*) and *miR-708* had two significant target genes (*NRAS* and *CREB1*), whereas *miR-499* had four significant target genes (*GPC6*, *C16orf72*, *WDR82* and *CACNB2*).

Pathway testing revealed 18 significant canonical pathways that are driven by brain-expressed target genes of the three miRNAs (Table 2). For each miRNA, the results of the GO analysis are presented as directed acyclic graphs (Supplementary Figure 5). The target genes that drive a particular pathway are listed in Supplementary Table 3.

Luciferase assays revealed efficient processing of *pri-miR-499*, but not *pri-miR-708*, upon transfection of the respective constructs in neurons (Supplementary Figure 6). Overexpression of *miR-499* led to a small and statistically nonsignificant increase in spine volume (Figure 3), but no effect on spine density was observed. As expected, transfection of the non-effective *miR-708* expression construct had no significant effect on spine morphological parameters. Taken together, these results suggest that increasing levels of the BD-associated *miR-499* have no—or only minimal—modulatory function during dendritic spine morphogenesis.

DISCUSSION

The present genetic association results for miRNA-coding genes suggest that miRNAs and their target genes may be implicated in the development of BD. The nominal P -values of SNPs at miRNA loci showed early deviation from the expected null line in the Q–Q plot, and this leftward shift reflects an enrichment of BD-associated SNPs at miRNA loci.

For the nine miRNAs that withstood Bonferroni correction, we additionally calculated the gene-based tests on the basis of the 1000 Genomes LD structure. This analysis revealed nominally gene-based P -values $\leq 7.20 \times 10^{-5}$ for all nine miRNAs, indicating that the results of gene-based tests on the basis of either HapMap phase 2 or 1000 Genomes Project data are highly comparable using our BD GWAS data.

Eight of the nine associated miRNAs were located in a host gene, including the three brain-expressed miRNAs *miR-499*, *miR-708* and *miR-1908*. Recent studies have reported a high correlation between the expression of a host gene and the resident miRNA.^{15,42} Previous authors have hypothesized that this finding may be because of the fact that miRNAs residing in introns are likely to share their regulatory elements and primary transcript with their host gene.²⁴ Some authors point out that host genes and their resident miRNAs may even have synergistic effects, which would have important implications for the fine-tuning of

gene expression patterns in the genome.^{43,44} On the basis of the present genetic association results, it is impossible to determine whether the association was attributable to the host gene, the miRNA or both. Further analyses are therefore warranted to clarify this, which was beyond the scope of the present analysis. However, the general enrichment of BD-associated SNPs at miRNA loci (Figure 1) and the results of our target gene analyses support the hypothesis that the majority of the associated miRNAs are implicated in BD etiology.

Regional association plots and expression data suggest that the miRNAs *miR-499*, *miR-708* and *miR-1908* are the most promising candidates in terms of the development of BD.

The miRNA *miR-499* is located in a region on chromosome 20q11 that showed genome-wide significant association in a previous GWAS of BD.⁴⁵ As *miR-499* is located in a region of high LD, which includes the genes *GSS*, *MYH7B* and *TRPC4AP* (Figure 2), further analyses of this chromosomal region are required to refine the association signal.⁴⁵ However, *miR-499* represents a very promising candidate in this region.

MiR-499 regulates apoptotic pathways involving the calcium-dependent protein phosphatase calcineurin.⁴⁶ A recent study demonstrated an upregulation of *miR-499* in the prefrontal cortex of patients with depression.⁴⁷ In a study of exosomal miRNA expression, *miR-499* showed differential expression in the post-mortem brains of BD patients compared with controls.⁴⁸ When considering a possible pathomechanism, it is important to note that a common SNP (rs3746444) is located in the seed region of the mature *miR-499-3p*.⁴⁹ This seed region is crucial for both the recognition of the target sites and the binding of the target genes. The SNP rs3746444 was not among the 2 267 487 SNPs analyzed in our large BD meta-analysis.⁶ However, rs3746444 achieved a nominally significant P -value of 0.0023 (risk allele: rs3746444-G) in a combined analysis of the seven MoodDS samples (2266 patients and 5028 controls),⁶ which excluded the PGC data set.³ Furthermore, the allele rs3746444-G has been associated with hallucinations and lack of motivation in schizophrenia patients.⁵⁰ This suggests that this SNP may confer susceptibility to BD by influencing depressive and psychotic endophenotypes. However, it may only partly explain the association signal at this locus.

Our target gene analysis revealed that *miR-499* had four significant target genes, including the previously reported genome-wide significant risk gene for psychiatric disorders *CACNB2*.⁵¹

Brain-expressed target genes of *miR-499-5p* exhibited an enrichment in biological processes related to cerebral development, which might however, at least partly, reflect the fact that our pathway analysis was restricted to brain-expressed genes. In addition, our pathway analysis indicates a potential role of *miR-499* in the regulation of the actin cytoskeleton. Interestingly, this pathway has been identified in a previous investigation of differentially and concordantly expressed genes enriched in association signals for schizophrenia and BD.⁵² Substantial research evidence suggests that the rearrangement of the cytoskeleton is crucial for neuronal cell migration and maturation, neurite outgrowth and maintenance of synaptic density and plasticity.^{53–56} These combined data suggest that *miR-499* is an interesting candidate for BD pathogenesis.

The miRNA *miR-708* is located in the first intron of *ODZ4* (odd Oz/ten-m homolog 4, *TENM4*), which has been reported as a genome-wide significant susceptibility gene for BD.³

A recent study of postpartum psychosis—a disorder that often heralds the incipient onset of BD⁵⁷—suggested differential expression of *miR-708* in the monocytes of affected patients compared with controls.⁵⁸ In another study, Xu *et al.*⁵⁹ demonstrated an altered expression profile for *miR-708* in mouse hippocampal neurons and showed that this was mediated by oxidative stress. Another recent study found that *miR-708* regulated the expression of neuronatin, which is a membrane

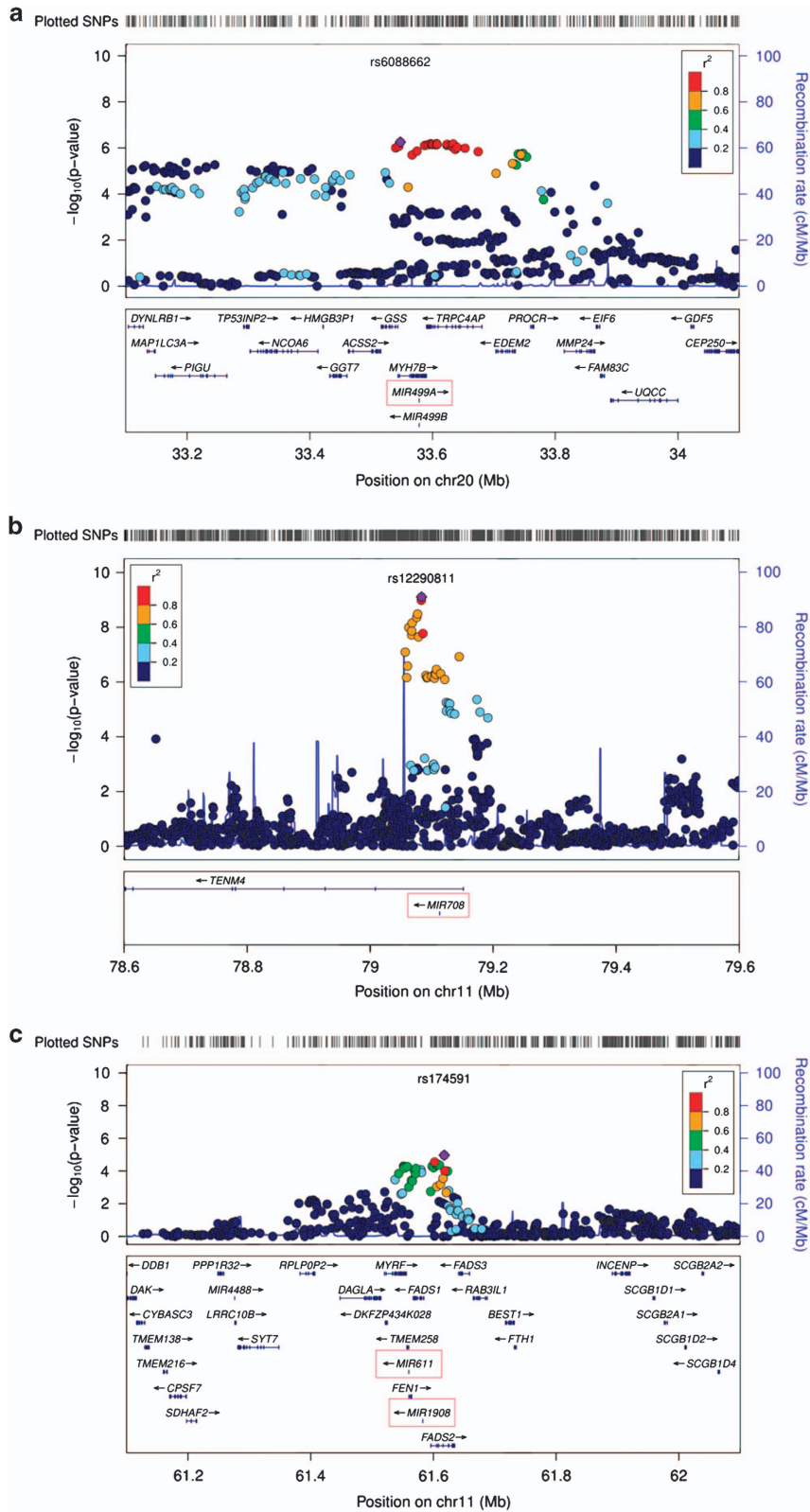


Figure 2. Regional association plots of *miR-499*, *miR-708* and *miR-1908*. Regional association results for the three most promising associated microRNAs *miR-499* (a), *miR-708* (b) and *miR-1908* (c), and their ± 500 -kb flanking regions were plotted using LocusZoom (Pruim et al.³⁶). The plot of *miR-1908* (c) includes *miR-611*, which is also localized at the depicted chromosomal locus.

Table 2. Target gene and pathway analysis for *miR-499*, *miR-708* and *miR-1908*

| MicroRNA | No. of brain-expressed target genes | No. of brain-expressed target genes, $P < 0.05$ | P enrichment | No. of significant targets (corr) | No. of significant pathways |
|--------------------|-------------------------------------|---|----------------|-----------------------------------|-----------------------------|
| <i>miR-499-5p</i> | 286 | 59 | 0.7172 | 4 | 12 |
| <i>miR-708-5p</i> | 174 | 37 | 0.9265 | 2 | 1 |
| <i>miR-1908-5p</i> | 56 | 17 | 0.1422 | 1 | 5 |

Abbreviations: No. of significant pathways, number of significant pathways at $P \leq 0.05$; No. of significant targets (corr), number of significant target genes after Bonferroni correction for multiple testing; P enrichment, P -value of the enrichment analysis (χ^2 -test). Results of the target gene analysis for the three brain-expressed microRNAs *miR-499*, *miR-708* and *miR-1908* that were associated with bipolar disorder after correction for multiple testing.

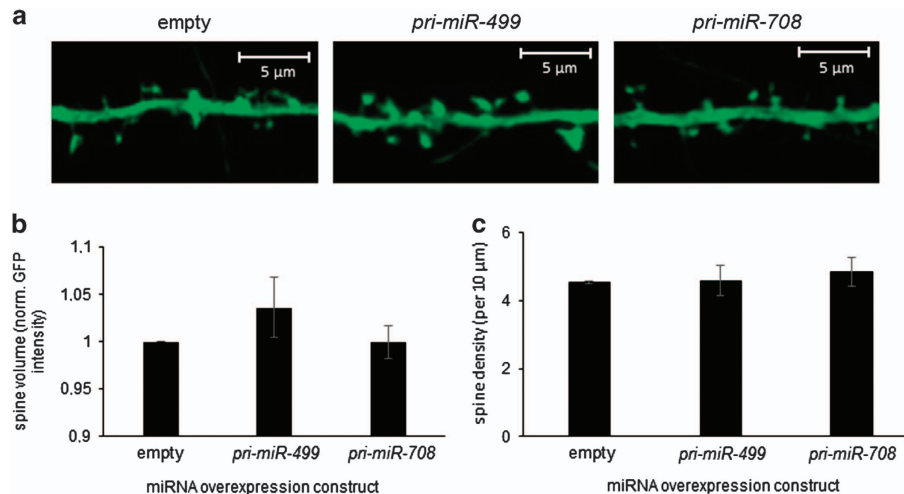


Figure 3. Effect of the overexpression of *miR-499* and *miR-708* on dendritic spine size and density in primary rat hippocampal neurons. DIV14 primary hippocampal neurons were transfected with: (i) empty pmirGLO (250 ng) or (ii) pmirGLO (250 ng) containing *pri-miR-499* or *pri-miR-708* in the 3'-untranslated repeat of the Firefly luciferase gene and green fluorescent protein (GFP). The transfected neurons were then cultured until DIV19 and fixed for fluorescence microscopy. (a) Representative images for cells transfected with the indicated pmirGLO constructs or GFP only. A three-dimensional reconstruction was made from seven 45- μm stacks; scale bars, 5 μm . (b) Spine volume quantification of hippocampal neurons transfected with the indicated pmirGLO constructs. Values are represented as means \pm s.d. ($n = 3$; 24 neurons per condition with a 200–250 spine count per cell). (c) Spine density of hippocampal neurons transfected with the indicated pmirGLO constructs. Values are represented as means \pm s.d. per 10 μm dendritic length ($n = 3$; 24 neurons per condition). Data are presented as the mean of three independent transfections normalized to the empty pmirGLO condition \pm s.d.

protein in the endoplasmic reticulum. Interestingly, the neuronatin-mediated regulation of intracellular Ca^{2+} levels has been implicated in cell migration and neural induction within embryonic stem cells.⁶⁰

Our target gene analysis revealed that *miR-708* had two significant target genes. These include *CREB1* that has previously been identified as a susceptibility gene for major depressive disorder.^{61–63} In addition, *CREB1* was found to be associated with BD in a recent study of large-scale BD samples⁶⁴ that included 8403 patients and 11 588 controls of our BD GWAS.⁶ However, the present pathway analysis provided no strong evidence for an enrichment of biological processes of relevance to psychiatric disorder.

miR-1908 is located in the first intron of the fatty acid desaturase 1 (*FADS1*) gene on chromosome 11. To date, few published studies have investigated the function of *miR-1908*. One recent study implicated *miR-1908* as a cancer biomarker.⁶⁵ A further study found that *miR-1908* belonged to a miRNA cluster that down-regulates the MARK1 signaling pathway, thus altering cell proliferation and differentiation.⁶⁶

Pathway analysis results for *miR-1908* indicate a potential role of the miRNA-regulated target gene network in key neuronal processes (GO subcategories: neuron projection and nervous system development). As these pathways showed the strongest

enrichment, further research into *miR-1908* and its regulated network appears to be warranted.

Although initial efforts have been made to elucidate the regulation of miRNA expression,⁶⁷ the manner in which miRNA expression and processing are regulated remains largely unknown. Given that pri-miRNAs have a length of 100–1000-bp,¹⁶ the present study investigated common variants at the miRNA loci and ± 20 kb flanking sequences in order to capture possible regulatory regions. However, further analyses of the regulation of miRNA expression by common variants are required to determine whether, and how, the presently described association signals influence the expression levels and function of the implicated miRNAs. The present approach did not allow investigation of SNPs with *trans*-expression quantitative trait loci (eQTL) effects on miRNAs. As recent studies suggest that $\sim 50\%$ of the identified miRNA eQTLs are *trans*-eQTLs,⁶⁸ investigations into the association between miRNA *trans*-eQTLs and BD are indicated.

The results of the functional analyses of *miR-499* and *miR-708* in rat hippocampal neurons revealed no major contribution of these miRNAs to the morphogenesis of dendritic spines, which represent the major sites of synaptic contact. However, only the results for *miR-499* can be considered robust, as the *miR-708* expression construct did not increase *miR-708* in primary neurons effectively. Alternative strategies for *miR-708* expression, together

with *miR-499/708* loss-of-function approaches, must be tested before definite conclusions regarding the role of these miRNAs in dendritic spine morphogenesis can be drawn. Moreover, to obtain more comprehensive insights into the potential effects of these miRNAs on synaptic function, future experiments should be complemented by immunocytochemistry analyses of synaptic marker proteins and electrophysiological recordings. Beyond a potential involvement in dendritic spine morphogenesis, these miRNAs could also regulate other aspects of neuronal morphology, such as dendrite arborization or axon growth, which could be tested in future studies.

CONCLUSION

The results of the present miRNA and target gene analyses suggest that the brain-expressed miRNAs *miR-499*, *miR-708* and *miR-1908* may contribute to the development of BD. Further research is warranted to elucidate the involvement of these miRNAs and their downstream pathways in BD.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

We are grateful to all of the patients and control subjects who contributed to this study. The study was supported by the German Federal Ministry of Education and Research (BMBF) through the Integrated Network IntegraMent (Integrated Understanding of Causes and Mechanisms in Mental Disorders), under the auspices of the e:Med Programme (grant 01ZX1314A to MMN and SC, grant 01ZX1314G to MR, grant 01ZX1314J to BMM), and through e:AtheroSysMed (Systems medicine of myocardial infarction and stroke, grant 01ZX1313B to BMM). MMN is a member of the DFG-funded Excellence-Cluster ImmunoSensation. MMN also received support from the Alfried Krupp von Bohlen und Halbach-Stiftung. The study was supported by the German Research Foundation (DFG; grant FOR2107; RI908/11-1 to MR; SCHR1136/3-1 to GS; NO246/10-1 to MMN). MG-S received the grant no. 89/2012 from UEFISCDI, Romania. Canadian patients were genotyped within the ConLiGen project (www.ConLiGen.org), with the support of a grant from the Deutsche Forschungsgemeinschaft to MR, MB and TGS (RI 908/7-1). Controls for Germany II were drawn from the Heinz Nixdorf Recall Study (HNR) cohort, which was established with the support of the Heinz Nixdorf Foundation. Recruitment of the Australian sample was supported by an Australian NHMRC program grant (number 1037196). The recruitment of the Canadian patients was supported by a grant from the Canadian Institutes of Health Research #64410 to MA. The study also used data generated by the GABRIEL consortium (controls for the sample Russia). Funding for the generation of these data was provided by the European Commission as part of GABRIEL contract number 018996 under the Integrated Program LSH-2004-1.2.5-1. Post-genomic approaches to understand the molecular basis of asthma aiming at a preventive or therapeutic control and the Wellcome Trust under award 084703. Canadian controls were drawn from the French Canadian study (SLSJ), which was supported in part by the Canada Research Chair Environment and genetics of respiratory diseases and allergy, the Canadian Institutes of Health Research (Operating grant No. MOP-13506) and the Quebec Respiratory Network of the Fonds de recherche en Santé du Québec (FRQS). Polish controls were recruited by the International Agency for Research on Cancer (IARC)/Centre National de Genotypage (CNG) GWAS Initiative. We thank the Bipolar Disorder Working Group of the PGC (PGC-BD) for providing access to the relevant data.

REFERENCES

- Craddock N, Sklar P. Genetics of bipolar disorder. *Lancet* 2013; **381**: 1654–1662.
- Nothen MM, Nieratschker V, Cichon S, Rietschel M. New findings in the genetics of major psychoses. *Dialogues Clin Neurosci* 2010; **12**: 85–93.
- Psychiatric GWAS Consortium Bipolar Disorder Working Group. Large-scale genome-wide association analysis of bipolar disorder identifies a new susceptibility locus near ODZ4. *Nat Genet* 2011; **43**: 977–983.
- Baum AE, Akula N, Cabanero M, Cardona I, Corona W, Klemens B et al. A genome-wide association study implicates diacylglycerol kinase eta (DGKH) and several other genes in the etiology of bipolar disorder. *Mol Psychiatry* 2008; **13**: 197–207.

- Ferreira MA, O'Donovan MC, Meng YA, Jones IR, Ruderfer DM, Jones L et al. Collaborative genome-wide association analysis supports a role for ANK3 and CACNA1C in bipolar disorder. *Nat Genet* 2008; **40**: 1056–1058.
- Muhleisen TW, Leber M, Schulze TG, Strohmaier J, Degenhardt F, Treutlein J et al. Genome-wide association study reveals two new risk loci for bipolar disorder. *Nat Commun* 2014; **5**: 3339.
- Sullivan PF, Daly MJ, O'Donovan M. Genetic architectures of psychiatric disorders: the emerging picture and its implications. *Nat Rev Genet* 2012; **13**: 537–551.
- Nurnberger JI Jr., Koller DL, Jung J, Edenberg HJ, Foroud T, Guella I et al. Identification of pathways for bipolar disorder: a meta-analysis. *JAMA Psychiatry* 2014; **71**: 657–664.
- Fineberg SK, Kosik KS, Davidson BL. MicroRNAs potentiate neural development. *Neuron* 2009; **64**: 303–309.
- Schratt G. MicroRNAs at the synapse. *Nat Rev Neurosci* 2009; **10**: 842–849.
- Forstner AJ, Degenhardt F, Schratz G, Nöthen MM. MicroRNAs as the cause of schizophrenia in 22q11.2 deletion carriers, and possible implications for idiopathic disease: a mini-review. *Front Mol Neurosci* 2013; **6**: 47.
- Xu B, Karayiorgou M, Gogos JA. MicroRNAs in psychiatric and neurodevelopmental disorders. *Brain Res* 2010; **1338**: 78–88.
- Kim AH, Reimers M, Maher B, Williamson V, McMichael O, McClay JL et al. MicroRNA expression profiling in the prefrontal cortex of individuals affected with schizophrenia and bipolar disorders. *Schizophr Res* 2010; **124**: 183–191.
- Moreau MP, Bruse SE, David-Rus R, Buyske S, Brzostowicz LM. Altered microRNA expression profiles in postmortem brain samples from individuals with schizophrenia and bipolar disorder. *Biol Psychiatry* 2011; **69**: 188–193.
- Godnic I, Zorc M, Jevsinek Skok D, Calin GA, Horvat S, Dovc P et al. Genome-wide and species-wide in silico screening for intragenic MicroRNAs in human, mouse and chicken. *PLoS One* 2013; **8**: e65165.
- Maffioletti E, Tardito D, Gennarelli M, Bocchio-Chiavetto L. Micro spies from the brain to the periphery: new clues from studies on microRNAs in neuropsychiatric disorders. *Front Cell Neurosci* 2014; **8**: 75.
- Rodriguez A, Griffiths-Jones S, Ashurst JL, Bradley A. Identification of mammalian microRNA host genes and transcription units. *Genome Res* 2004; **14**: 1902–1910.
- Han J, Lee Y, Yeom KH, Kim YK, Jin H, Kim VN. The Drosha-DGCR8 complex in primary microRNA processing. *Genes Dev* 2004; **18**: 3016–3027.
- Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003; **425**: 415–419.
- Krol J, Loedige I, Filipowicz W. The widespread regulation of microRNA biogenesis, function and decay. *Nat Rev Genet* 2010; **11**: 597–610.
- Gregory RI, Chendrimada TP, Cooch N, Shiekhattar R. Human RISC couples microRNA biogenesis and posttranscriptional gene silencing. *Cell* 2005; **123**: 631–640.
- He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004; **5**: 522–531.
- Meola N, Gennarino VA, Banfi S. microRNAs and genetic diseases. *Pathogenetics* 2009; **2**: 7.
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; **116**: 281–297.
- Didiano D, Hobert O. Molecular architecture of a miRNA-regulated 3' UTR. *RNA* 2008; **14**: 1297–1317.
- Sathyan P, Golden HB, Miranda RC. Competing interactions between micro-RNAs determine neural progenitor survival and proliferation after ethanol exposure: evidence from an *ex vivo* model of the fetal cerebral cortical neuroepithelium. *J Neurosci* 2007; **27**: 8546–8557.
- Choi WY, Giraldez AJ, Schier AF. Target protectors reveal dampening and balancing of Nodal agonist and antagonist by miR-430. *Science* 2007; **318**: 271–274.
- Johnston RJ, Hobert O. A microRNA controlling left/right neuronal asymmetry in *Caenorhabditis elegans*. *Nature* 2003; **426**: 845–849.
- Geeleher P, Huang SR, Gamazon ER, Golden A, Seoighe C. The regulatory effect of miRNAs is a heritable genetic trait in humans. *BMC Genomics* 2012; **13**: 383.
- Forstner AJ, Basmanav FB, Mattheisen M, Bohmer AC, Hollegaard MV, Janson E et al. Investigation of the involvement of MIR185 and its target genes in the development of schizophrenia. *J Psychiatry Neurosci* 2014; **39**: 386–396.
- Lopez JP, Lim R, Cruceanu C, Crapper L, Fasano C, Labonte B et al. miR-1202 is a primate-specific and brain-enriched microRNA involved in major depression and antidepressant treatment. *Nat Med* 2014; **20**: 764–768.
- Strazisar M, Cammaerts S, van der Ven K, Forero DA, Lenaerts AS, Nordin A et al. MIR137 variants identified in psychiatric patients affect synaptogenesis and neuronal transmission gene sets. *Mol Psychiatry* 2015; **20**: 472–481.
- Wang Z, Zhang C, Huang J, Yuan C, Hong W, Chen J et al. MiRNA-206 and BDNF genes interacted in bipolar I disorder. *J Affect Disord* 2014; **162**: 116–119.
- Liu JZ, McRae AF, Nyholt DR, Medland SE, Wray NR, Brown KM et al. A versatile gene-based test for genome-wide association studies. *Am J Hum Genet* 2010; **87**: 139–145.

- 35 Kozomara A, Griffiths-Jones S. miRBase: annotating high confidence microRNAs using deep sequencing data. *Nucleic Acids Res* 2014; **42**: D68–D73.
- 36 Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP et al. LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* 2010; **26**: 2336–2337.
- 37 Ziats MN, Rennett OM. Identification of differentially expressed microRNAs across the developing human brain. *Mol Psychiatry* 2014; **19**: 848–852.
- 38 Siegel G, Obernosterer G, Fiore R, Oehmen M, Bicker S, Christensen M et al. A functional screen implicates microRNA-138-dependent regulation of the depalmitoylation enzyme APT1 in dendritic spine morphogenesis. *Nat Cell Biol* 2009; **11**: 705–716.
- 39 Hackenberg M, Rodriguez-Ezpeleta N, Aransay AM. miRanalyzer: an update on the detection and analysis of microRNAs in high-throughput sequencing experiments. *Nucleic Acids Res* 2011; **39**: W132–W138.
- 40 Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005; **120**: 15–20.
- 41 Hawrylycz MJ, Lein ES, Guillozet-Bongaarts AL, Shen EH, Ng L, Miller JA et al. An anatomically comprehensive atlas of the adult human brain transcriptome. *Nature* 2012; **489**: 391–399.
- 42 Baskerville S, Bartel DP. Microarray profiling of microRNAs reveals frequent coexpression with neighboring miRNAs and host genes. *RNA* 2005; **11**: 241–247.
- 43 Lutter D, Marr C, Krumsiek J, Lang EW, Theis FJ. Intronic microRNAs support their host genes by mediating synergistic and antagonistic regulatory effects. *BMC Genomics* 2010; **11**: 224.
- 44 Rearick D, Prakash A, McSweeney A, Shepard SS, Fedorova L, Fedorov A. Critical association of ncRNA with introns. *Nucleic Acids Res* 2011; **39**: 2357–2366.
- 45 Green EK, Hamshere M, Forty L, Gordon-Smith K, Fraser C, Russell E et al. Replication of bipolar disorder susceptibility alleles and identification of two novel genome-wide significant associations in a new bipolar disorder case-control sample. *Mol Psychiatry* 2013; **18**: 1302–1307.
- 46 Wang JX, Jiao JQ, Li Q, Long B, Wang K, Liu JP et al. miR-499 regulates mitochondrial dynamics by targeting calcineurin and dynamin-related protein-1. *Nat Med* 2011; **17**: 71–78.
- 47 Smalheiser NR, Lugli G, Zhang H, Rizavi H, Cook EH, Dwivedi Y. Expression of microRNAs and other small RNAs in prefrontal cortex in schizophrenia, bipolar disorder and depressed subjects. *PLoS One* 2014; **9**: e86469.
- 48 Banigan MG, Kao PF, Kozubek JA, Winslow AR, Medina J, Costa J et al. Differential expression of exosomal microRNAs in prefrontal cortices of schizophrenia and bipolar disorder patients. *PLoS One* 2013; **8**: e48814.
- 49 Gong J, Tong Y, Zhang HM, Wang K, Hu T, Shan G et al. Genome-wide identification of SNPs in microRNA genes and the SNP effects on microRNA target binding and biogenesis. *Hum Mutat* 2012; **33**: 254–263.
- 50 Zou M, Li D, Lv R, Zhou Y, Wang T, Liu J et al. Association between two single nucleotide polymorphisms at corresponding microRNA and schizophrenia in a Chinese population. *Mol Biol Rep* 2012; **39**: 3385–3391.
- 51 Cross-Disorder Group of the Psychiatric Genomics Consortium. Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *Lancet* 2013; **381**: 1371–1379.
- 52 Zhao Z, Xu J, Chen J, Kim S, Reimers M, Bacanu SA et al. Transcriptome sequencing and genome-wide association analyses reveal lysosomal function and actin cytoskeleton remodeling in schizophrenia and bipolar disorder. *Mol Psychiatry* 2015; **20**: 563–572.
- 53 Auer M, Hausott B, Klimaschewski L. Rho GTPases as regulators of morphological neuroplasticity. *Ann Anat* 2011; **193**: 259–266.
- 54 Belenchi GC, Gurniak CB, Perlas E, Middei S, Ammassari-Teule M, Witke W. N-cofilin is associated with neuronal migration disorders and cell cycle control in the cerebral cortex. *Genes Dev* 2007; **21**: 2347–2357.
- 55 de Curtis I. Functions of Rac GTPases during neuronal development. *Dev Neurosci* 2008; **30**: 47–58.
- 56 Kreis P, Barnier JV. PAK signalling in neuronal physiology. *Cell Signal* 2009; **21**: 384–393.
- 57 Jones I, Craddock N. Familiality of the puerperal trigger in bipolar disorder: results of a family study. *Am J Psychiatry* 2001; **158**: 913–917.
- 58 Weigelt K, Bergink V, Burgerhout KM, Pescatori M, Wijkhuijs A, Drexhage HA. Down-regulation of inflammation-protective microRNAs 146a and 212 in monocytes of patients with postpartum psychosis. *Brain Behav Immun* 2013; **29**: 147–155.
- 59 Xu S, Zhang R, Niu J, Cui D, Xie B, Zhang B et al. Oxidative stress mediated-alterations of the microRNA expression profile in mouse hippocampal neurons. *Int J Mol Sci* 2012; **13**: 16945–16960.
- 60 Ryu S, McDonnell K, Choi H, Gao D, Hahn M, Joshi N et al. Suppression of miRNA-708 by polycomb group promotes metastases by calcium-induced cell migration. *Cancer Cell* 2013; **23**: 63–76.
- 61 Carlezon WA Jr., Duman RS, Nestler EJ. The many faces of CREB. *Trends Neurosci* 2005; **28**: 436–445.
- 62 Maher BS, Hughes HB 3rd, Zubenko WN, Zubenko GS. Genetic linkage of region containing the CREB1 gene to depressive disorders in families with recurrent, early-onset, major depression: a re-analysis and confirmation of sex-specific effect. *Am J Med Genet B Neuropsychiatr Genet* 2010; **153B**: 10–16.
- 63 Wallace TL, Stellitano KE, Neve RL, Duman RS. Effects of cyclic adenosine monophosphate response element binding protein overexpression in the basolateral amygdala on behavioral models of depression and anxiety. *Biol Psychiatry* 2004; **56**: 151–160.
- 64 Li M, Luo XJ, Rietschel M, Lewis CM, Mattheisen M, Muller-Myhsok B et al. Allelic differences between Europeans and Chinese for CREB1 SNPs and their implications in gene expression regulation, hippocampal structure and function, and bipolar disorder susceptibility. *Mol Psychiatry* 2014; **19**: 452–461.
- 65 Rawlings-Goss RA, Campbell MC, Tishkoff SA. Global population-specific variation in miRNA associated with cancer risk and clinical biomarkers. *BMC Med Genomics* 2014; **7**: 53.
- 66 Ye SB, Li ZL, Luo DH, Huang BJ, Chen YS, Zhang XS et al. Tumor-derived exosomes promote tumor progression and T-cell dysfunction through the regulation of enriched exosomal microRNAs in human nasopharyngeal carcinoma. *Oncotarget* 2014; **5**: 5439–5452.
- 67 Gamazon ER, Ziliak D, Im HK, LaCroix B, Park DS, Cox NJ et al. Genetic architecture of microRNA expression: implications for the transcriptome and complex traits. *Am J Hum Genet* 2012; **90**: 1046–1063.
- 68 Borel C, Deutsch S, Letourneau A, Migliavacca E, Montgomery SB, Dimas AS et al. Identification of cis- and trans-regulatory variation modulating microRNA expression levels in human fibroblasts. *Genome Res* 2011; **21**: 68–73.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

Supplementary Information accompanies the paper on the Translational Psychiatry website (<http://www.nature.com/tp>)