

John von Neumann Institute for Computing



Aggregate Size and Shape Distributions in Amyloid- β Peptide Solutions

L. Nagel-Steger, B. Demeler, D. Willbold

published in

*From Computational Biophysics to Systems Biology (CBSB07),
Proceedings of the NIC Workshop 2007,*
Ulrich H. E. Hansmann, Jan Meinke, Sandipan Mohanty,
Olav Zimmermann (Editors),
John von Neumann Institute for Computing, Jülich,
NIC Series, Vol. 36, ISBN 978-3-9810843-2-0, pp. 235-237, 2007.

© 2007 by John von Neumann Institute for Computing

Permission to make digital or hard copies of portions of this work for personal or classroom use is granted provided that the copies are not made or distributed for profit or commercial advantage and that copies bear this notice and the full citation on the first page. To copy otherwise requires prior specific permission by the publisher mentioned above.

<http://www.fz-juelich.de/nic-series/volume36>

Aggregate Size and Shape Distributions in Amyloid- β Peptide Solutions

Luitgard Nagel-Steger¹, Borries Demeler², and Dieter Willbold¹

¹ Institute for Neurosciences and Biophysics, INB-2, Research Centre Jülich, D-52425 Jülich, and Institut für Physikalische Biologie & BMFZ, Heinrich-Heine-Universität Düsseldorf, D-40225 Düsseldorf

E-mail: nagelst@biophys.uni-duesseldorf.de

² Dept. of Biochemistry, University of Texas Health Science Center
7703 Floyd Curl Drive, US- San Antonio, Tx, USA

E-mail: demeler@biochem.uthscsa.edu

A peptide with 42 amino acid residues (A β (1–42)) plays a key role in the pathogenesis of the Alzheimer's disease. Aggregation of this peptide leads to the formation of assemblies of different sizes and shapes, among which the amyloid fibril is the most prominent member. We used sedimentation velocity centrifugation to analyze A β (1–42) peptide solutions at different time points during aggregation regarding their hydrodynamic properties. New data evaluation software allowed us to determine not only sedimentation coefficients, but also at the same time the related frictional ratios of species present in a multicomponent system. Shortly after sample preparation, a sharp peak dominated the measured *s*-value distribution. Two-dimensional spectrum analysis assigned this species an *s*-value of $28 \cdot 10^{-13}$ s, a molecular weight of $1.23 \cdot 10^6$ and a frictional ratio of 1.44. Under the assumption of a rigid rod model this structural information is in good agreement with a fibril of 50 nm length build out of two neighboring β -sheets with about 110 monomers each, arranged in a parallel in-register fashion. Incubation for 1, 2 and 5 days causes further aggregation, which results in decreasing frictional ratios, implying a growth to more spherical-like particles.

1 Introduction

A peptide with 42 amino acid residues (A β (1–42)) plays a key role in the pathogenesis of the Alzheimer's disease. It is highly prone to self aggregation, leading to the formation of fibrils which are deposited in amyloid plaques in the brain of affected individuals¹. In previous years increasing evidence arose that probably smaller oligomeric assemblies^{2,3} play a more decisive role as neurotoxic agents than the mature fibril. Information about size and shape of A β peptide assemblies formed during aggregation is therefore of high relevance.

Analytical ultracentrifugation is a method for retrieving structural information about macromolecules by direct observation of their hydrodynamic properties in a centrifugal field. Advanced data analysis permits the determination of *s*-value, molecular weight and shape distributions for multicomponent systems.

Our objective is to evaluate the applicability of analytical ultracentrifugation in combination with the newly developed data evaluation software for generating size and shape distributions of solutions of the A β peptide. The method shall be used to monitor the effects of peptide concentration, solvent and added cosolutes upon aggregation.

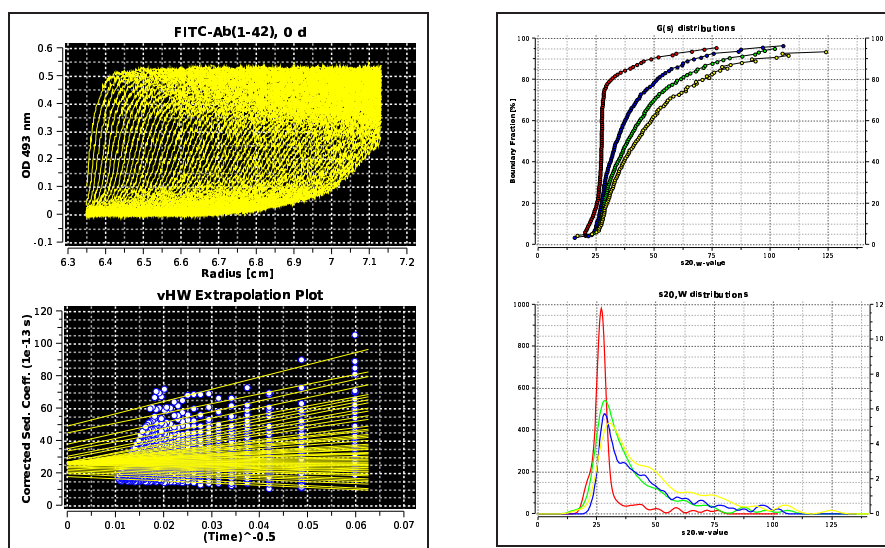


Figure 1. Van-Holde-Weischet Analysis. Left: Raw data (top) and corresponding vHW-analysis plot (bottom) for $A\beta$ aggregation mixture 2 h after sample preparation. Sample volume was 300 μ l. Right: Upper graph: $G(s)$ distribution of sample after 0 d (red), 1 d (blue), 2 d (green), and 5 d (yellow) incubation at 20 $^{\circ}$ C. Lower graph: s -value distribution of sample after 0 d (red), 1 d (green), 2 d (blue), and 5 d (yellow) incubation at 20 $^{\circ}$ C.

2 Methods

The aggregation mixture contained 70 μ M unlabeled and 14 μ M FITC-labeled $A\beta$ (1–42) in 10 mM sodium phosphate buffer, pH 7.4 with 6 % final DMSO concentration. Sedimentation velocity experiments were performed with an X-LA analytical ultracentrifuge (Beckman-Coulter), equipped with absorption optics. Samples were measured in standard double-sector aluminum cells at 20,000 rpm, 20 $^{\circ}$ C. Radial step size was set to 0.001 cm.

Raw data from the analytical ultracentrifuge were processed and evaluated using the UltraScan software package⁴ running on a 44 node AMD Opteron cluster under Linux. After determination of suitable start parameters by a van-Holde-Weischet analysis⁵ the 2-dimensional spectrum analysis⁶ (2DSA), implemented in the software package, was started. 2DSA solves the inverse problem of fitting sedimentation velocity data to a linear combination of finite element solutions of the Lamm equation. Each term of the linear combination reflects a solute in the 2-dimensional space over s and f/f_0 . Finally Monte-Carlo simulations were used to identify statistically significant solutes.

3 Results

Data analysis of sedimentation velocity runs performed at 20,000 rpm, 20 $^{\circ}$ C at 4 time points during aggregation of $A\beta$ (1–42) according to van-Holde-Weischet as depicted in Fig. 1 showed an increase of the mean s -value over the incubation time of 57 %. Aggregation of the amyloid- β peptide starts with a narrow distribution with an s_{avg} -value of 30 S. The distribution is dominated by a species of 28 S, a molecular weight of $1.22 \cdot 10^6$

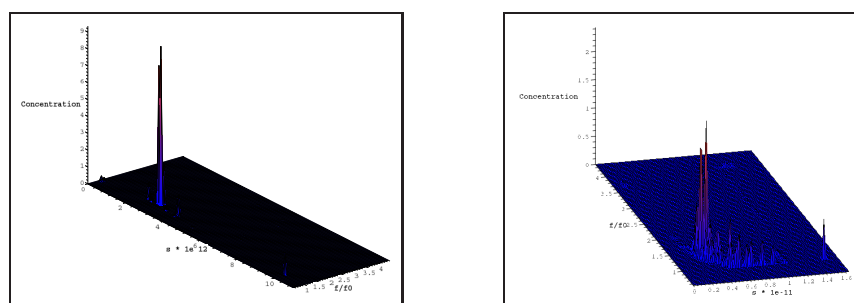


Figure 2. 3D-Plots of s -value vs f/f_0 distributions. Left: $A\beta(1-42)$ after 2 h incubation at 20 °C. Right: $A\beta(1-42)$ after 5 d incubation at 20 °C.

and a frictional ratio of 1.44, which comprises about 40 % of the sedimentation boundary (Fig. 2, left). This aggregate species is reproducibly detected in repeated experiments. It can be modeled by a rod-like particle with an axial ratio of 8.6. Under the assumption of a rigid rod model and molecule dimensions for β -strands this structural information is in very good agreement with a fibril of 50 nm length build out of two neighboring β -sheets with about 110 monomers each, arranged in a parallel in-register fashion, as proposed by⁷.

Upon incubation, the measured distribution is asymmetrically broadening to higher s -values with a still distinguishable maximum at about 28 S. During the 5 d incubation of the sample, the f/f_0 -value of detected solutes decreases from 1.6 to around 1 with s -values increasing from 28 S to 150 S. Particles are becoming more spherical during the incubation time within the observed growth range (Fig. 2, right). The method is a tool to study the action of drugs on the aggregation of $A\beta(1-42)$ peptide.

References

1. J.A. Hardy and G.A. Higgins, *Alzheimer's disease: the amyloid cascade hypothesis*, Science, **256**, 184–185, 1992.
2. W.L. Klein, *Fibrils, proteofibrils and $A\beta$ -derived diffusible ligands: How $A\beta$ causes neuron dysfunction and death in Alzheimer's disease*, Humana Press, Totowa, NJ, pp. 1–49, 2001.
3. D.M. Walsh, J.V. Klyubin, I. Fadeeva, W.K. Cullen, R. Anwyl, M.J. Wolfe, M.S. Rowan, and D.J. Selkoe, *Natural secreted oligomers of amyloid β protein potently inhibit hippocampal longterm potentiation in vivo*, Nature, **416**, 535–539, 2002.
4. B. Demeler, *UltraScan*, <http://www.ultrascan.uthscsa.edu>, version 9.0.
5. B. Demeler, and K.E. van Holde, *Sedimentation velocity analysis of highly heterogeneous systems*, Anal. Biochem., **335**, 279–288, 2004.
6. E.H. Brookes, R.V. Boppana, and B. Demeler, *Computing Large Sparse Multivariate Optimization Problems with an Application in Biophysics*, in: SC '06: Proceedings of the 2006 ACM/IEEE Conference on Supercomputing, vol. 6, p. 81, 2006.
7. A.T. Petkova, W.-M. Yau, and R. Tycko, *Experimental Constraints on Quaternary Structure in Alzheimer's β -Amyloid Fibrils*, Biochemistry, **45**, 498–512, 2006.