

WestminsterResearch

<http://www.westminster.ac.uk/westminsterresearch>

**Mechanisms underlying glycemic deterioration in type 2 diabetes:
An IMI DIRECT study**

**Bizzotto, R., Jennison, C., Jones, A., Kurbasic, A., Kennedy, G.,
Bell, J.D., Thomas, E.L., Frost, G., Eriksen, R., Koivula, R., Brage,
S., Kaye, J., Hattersley, A., Heggie, A., McEvoy, D., Hart, L.M.,
Beulens, J.W., Elders, P., Musholt, P.B., Ridderstråle, M., Hansen,
T., Allin, K., Hansen, T., Vestergaard, H., Lundgaard, A.T.,
Thomsen, H.S., Masi, F., Tsirigos, K.D., Brunak, S., Viñuela, A.,
Mahajan, A., McDonald, T.J., Kokkola, T., Forgie, I., Giordano,
G.N., Pavo, I., Ruetten, H., Dermitzakis, E., McCarthy, M.,
Pedersen, O., Schwenk, J., Adamski, J., Franks, P., Walker, M.,
Pearson, E. and Mari, A.**

This is an author's accepted manuscript of an article published in Diabetes Care.

The final definitive version is available online at:

<https://doi.org/10.2337/dc20-1567>

The WestminsterResearch online digital archive at the University of Westminster aims to make the research output of the University available to a wider audience. Copyright and Moral Rights remain with the authors and/or copyright owners.

1 **Full title**

2 Processes underlying glycemetic deterioration in type 2 diabetes: An IMI DIRECT study

3

4 **Running title**

5 Glycemetic deterioration in type 2 diabetes

6

7 **Author names**

8 Roberto Bizzotto^{1,*}, Christopher Jennison², Angus G Jones³, Azra Kurbasic⁴, Gwen Kennedy⁵,
9 Jimmy D Bell⁶, Elizabeth L Thomas⁶, Gary Frost⁷, Rebeca Eriksen⁷, Robert W Koivula^{4,8}, Soren
10 Brage⁹, Jane Kaye^{10,11}, Andrew T Hattersley³, Alison Heggie¹², Donna McEvoy¹³, Leen M 't
11 Hart^{14,15,16}, Joline W Beulens¹⁴, Petra Elders¹⁷, Petra B Musholt¹⁸, Martin Ridderstråle¹⁹, Tue H
12 Hansen²⁰, Kristine H Allin²⁰, Torben Hansen²⁰, Henrik Vestergaard^{20,21}, Agnete T Lundgaard²²,
13 Henrik S Thomsen²³, Federico De Masi²⁴, Konstantinos D Tsirigos^{22,25}, Søren Brunak^{22,25}, Ana
14 Viñuela²⁶, Anubha Mahajan^{27,a}, Timothy J McDonald^{3,28}, Tarja Kokkola²⁹, Ian M Forgie³⁰,
15 Giuseppe N Giordano⁴, Imre Pavo³¹, Hartmut Ruetten¹⁸, Emmanouil Dermitzakis²⁶, Mark I
16 McCarthy^{8,27,32,a}, Oluf Pedersen²⁰, Jochen M Schwenk³³, Jerzy Adamski^{34,35,36}, Paul W Franks⁴,
17 Mark Walker³⁷, Ewan R Pearson³⁰, Andrea Mari¹ for the IMI DIRECT consortium³⁸.

18

19 **Affiliations**

20 ¹ CNR Institute of Neuroscience, Padova, Italy;

21 ² Department of Mathematical Sciences, University of Bath, Bath, United Kingdom;

22 ³ Institute of Clinical and Biological Sciences, University of Exeter Medical School, Exeter, United
23 Kingdom;

24 ⁴ Genetic and Molecular Epidemiology Unit, Lund University Diabetes Centre, Department of
25 Clinical Sciences, CRC, Lund University, SUS, Malmö, Sweden;

26 ⁵ Immunoassay Biomarker Core Laboratory, School of Medicine, Ninewells Hospital, Dundee,
27 United Kingdom;

28 ⁶ Research Centre for Optimal Health, Department of Life Sciences, University of Westminster,
29 London, United Kingdom;

30 ⁷ Section for Nutrition Research, Faculty of Medicine, Hammersmith Campus, Imperial College
31 London, London, United Kingdom;

32 ⁸ Oxford Centre for Diabetes, Endocrinology and Metabolism, Radcliffe Department of Medicine,
33 University of Oxford, Oxford, United Kingdom;

34 ⁹ MRC Epidemiology Unit, University of Cambridge, United Kingdom;

35 ¹⁰ Centre for Health, Law and Emerging Technologies (HeLEX), Faculty of Law, University of
36 Oxford, Oxford, United Kingdom;

37 ¹¹ Centre for Health, Law and Emerging Technologies (HeLEX), Melbourne Law School,
38 University of Melbourne, Carlton, Victoria, Australia;

39 ¹² Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, United Kingdom;

40 ¹³ Diabetes Research Network, Royal Victoria Infirmary, Newcastle upon Tyne, United Kingdom;

41 ¹⁴ Department of Epidemiology & Biostatistics, Amsterdam UMC - location VUmc, Amsterdam
42 Public Health Research Institute, Amsterdam, The Netherlands;

43 ¹⁵ Department of Cell and Chemical Biology, Leiden University Medical Center, Leiden, the
44 Netherlands;
45 ¹⁶ Department of Biomedical Data Sciences, Molecular Epidemiology section, Leiden University
46 Medical Center, Leiden, the Netherlands;
47 ¹⁷ Department of General Practice, Amsterdam UMC - location VUmc, Amsterdam Public Health
48 Research Institute, Amsterdam, The Netherlands;
49 ¹⁸ R&D Global Development, Translational Medicine & Clinical Pharmacology (TMCP), Sanofi-
50 Aventis Deutschland GmbH, Frankfurt, Germany;
51 ¹⁹ Clinical Pharmacology and Translational Medicine, Novo Nordisk A/S, Søborg, Denmark;
52 ²⁰ Novo Nordisk Foundation Center for Basic Metabolic Research, Faculty of Health and Medical
53 Sciences, University of Copenhagen, Denmark;
54 ²¹ Bornholms Hospital, Rønne, Denmark;
55 ²² Novo Nordisk Foundation Center for Protein Research, University of Copenhagen, Copenhagen,
56 Denmark;
57 ²³ Faculty of Medical and Health Sciences, University of Copenhagen, Copenhagen;
58 ²⁴ Department of Health Technology, Technical University of Denmark, Kongens Lyngby,
59 Denmark;
60 ²⁵ Center for Biological Sequence Analysis, Department of Systems Biology, Technical University
61 of Denmark, Kongens Lyngby, Denmark;
62 ²⁶ Department of Genetic Medicine and Development, University of Geneva Medical School,
63 Geneva, Switzerland;
64 ²⁷ Wellcome Centre for Human Genetics, University of Oxford, Oxford, United Kingdom;
65 ²⁸ Blood Sciences, Royal Devon and Exeter NHS Foundation Trust, Exeter, United Kingdom;
66 ²⁹ Department of Medicine, University of Eastern Finland, Kuopio, Finland;
67 ³⁰ Population Health & Genomics, School of Medicine, University of Dundee, Dundee, United
68 Kingdom;
69 ³¹ Eli Lilly Regional Operations GmbH, Vienna, Austria;
70 ³² Oxford NIHR Biomedical Research Centre, Oxford University Hospitals NHS Foundation Trust,
71 John Radcliffe Hospital, Oxford, United Kingdom;
72 ³³ Affinity Proteomics, Science for Life Laboratory, School of Engineering Sciences in Chemistry,
73 Biotechnology and Health, KTH - Royal Institute of Technology, Solna, Sweden;
74 ³⁴ Research Unit of Molecular Endocrinology and Metabolism, Helmholtz Zentrum München,
75 Neuherberg, Germany;
76 ³⁵ Lehrstuhl für Experimentelle Genetik, Technische Universität München, Freising-
77 Weihenstephan, Germany;
78 ³⁶ Department of Biochemistry, Yong Loo Lin School of Medicine, National University of
79 Singapore, Singapore, Singapore;
80 ³⁷ Translational and Clinical Research Institute, Faculty of Medical Sciences, Newcastle University,
81 Newcastle, United Kingdom;
82 ³⁸ Names and affiliations listed in the Supplemental Material.
83 ^a Current address: Genentech, 1 DNA Way, South San Francisco, CA 94080
84

85 ***Corresponding author**

86 Roberto Bizzotto

87 Corso Stati Uniti 4

88 35127 Padova

89 Italy

90 Telephone: 0039 049 8295796

91 Fax: 0039 049 8295763

92 Email address: roberto.bizzotto@cnr.it

93

94

Word count: 3994

95

Tables: 1 – Figures: 3

96 **Abstract**

97 *Objective*

98 We investigated the processes underlying glycemetic deterioration in type 2 diabetes (T2D).

99 *Research Design and Methods*

100 732 recently diagnosed T2D patients from the IMI-DIRECT study were extensively phenotyped
101 over three years, including measures of insulin sensitivity (OGIS), β -cell glucose sensitivity (GS)
102 and insulin clearance (CLIm) from mixed meal tests, liver enzymes, lipid profiles, and baseline
103 regional fat from MRI. The associations between the longitudinal metabolic patterns and HbA_{1c}
104 deterioration, adjusted for changes in BMI and in diabetes medications, were assessed via stepwise
105 multivariable linear and logistic regression.

106 *Results*

107 Faster HbA_{1c} progression was independently associated with faster deterioration of OGIS and GS,
108 and increasing CLIm; visceral or liver fat, HDL-cholesterol and triglycerides had further
109 independent, though weaker, roles ($R^2=0.38$). A subgroup of patients with a markedly higher
110 progression rate (fast progressors) was clearly distinguishable considering these variables only
111 (discrimination capacity from AUROC=0.94). The proportion of fast progressors was reduced from
112 56% to 8-10% in subgroups in which only one trait among OGIS, GS and CLIm was relatively
113 stable (odds ratios 0.07 to 0.09). T2D polygenic risk score and baseline pancreatic fat, GLP-1,
114 glucagon, diet, and physical activity did not show an independent role.

115 *Conclusions*

116 Deteriorating insulin sensitivity and β -cell function, increasing insulin clearance, high visceral or
117 liver fat, and worsening of the lipid profile are the crucial factors mediating glycemetic deterioration

118 of T2D patients in the initial phase of the disease. Stabilization of a single trait among insulin
119 sensitivity, β -cell function, and insulin clearance may be relevant to prevent progression.

120 Maintaining glucose levels within appropriate limits in patients with type 2 diabetes (T2D) is a
121 crucial factor to prevent complications. Effective strategies to slow glyceic progression can be
122 supported by understanding the processes underlying deterioration of glucose control.

123 Few studies have assessed HbA_{1c} trajectories and the possible determinants of glyceic
124 deterioration. An established finding is that β -cell function decline is an important factor (1,2),
125 while contradictory conclusions were drawn for insulin sensitivity (1,3–7). Whether heterogeneous
126 patterns between patients exist in β -cell function and insulin sensitivity decline has not been
127 clarified, an important question for patient stratification and personalized medicine. Other
128 limitations of previous analyses include the incomplete characterization of the metabolic parameters
129 affecting glucose homeostasis (derived using fasting data only (2,4)), the restricted set of traits
130 investigated together, and the lack of potentially relevant measures such as ectopic fat, insulin
131 clearance, or lifestyle. No study has assessed the relationships between the longitudinal trajectories
132 of HbA_{1c} and those of the other metabolic traits.

133 In this analysis, we have used data from the cohort of recently diagnosed and extensively
134 phenotyped T2D patients of the DIRECT study (8,9) to elucidate the processes underlying glyceic
135 deterioration. Specific features of the DIRECT study are the detailed assessment of the glucose
136 homeostasis parameters, and patients all being in the initial phase of the disease. We determined the
137 patterns over a 3-year period of HbA_{1c}, β -cell function, insulin sensitivity and other relevant
138 laboratory, clinical and functional parameters, and assessed their relevance in the deterioration of
139 glucose control.

140 **Research Design and Methods**

141 *Subjects and protocol*

142 The IMI-DIRECT (Innovative Medicines Initiative - Diabetes Research on Patient Stratification)
143 project is a multicenter prospective study on northern European adults (8,9) (ClinicalTrials.gov

144 identifier NCT03814915). The present analysis considers the DIRECT cohort of recently diagnosed
145 T2D patients, who were recruited according to the following criteria: white race, T2D diagnosis
146 according to the American Diabetes Association 2011 criteria (10) not less than 6 months and not
147 more than 24 months before baseline examination, previous treatment via lifestyle measures with or
148 without metformin therapy, age between 35 and 74 years, BMI between 20 and 50 kg/m², estimated
149 glomerular filtration rate >50 ml/min, and HbA_{1c} concentration <7.64 % (60.0 mmol/mol) within
150 the previous 3 months. Participants were studied at baseline (month 0) and at months 9, 18 and 36.
151 Subjects with HbA_{1c} available at least in two visits were included in this analysis (N=750).

152 All participants provided written informed consent and the study protocol was approved by the
153 regional research ethics review boards. The research conformed to the ethical principles for medical
154 research involving human participants outlined in the declaration of Helsinki.

155 *Collected data*

156 Anthropometric data, HbA_{1c}, blood lipids and liver enzymes were collected at all visits. A 27-month
157 HbA_{1c} sample was collected in 39 patients. A standardized mixed meal test (8) (MMTT) was
158 performed at months 0, 18 and 36 to calculate indices of insulin sensitivity (in fasting conditions,
159 QUICKI (11), and post-MMTT, OGIS (12)), β -cell function (13) (glucose sensitivity, GS, and rate
160 sensitivity), and insulin clearance (in fasting conditions, and post-MMTT, CLIm). From the
161 baseline visit we collected glucagon, proinsulin and glucagon-like peptide 1 (GLP-1), measures of
162 regional fat from MRI (8) (available in 561 participants), of physical activity from accelerometer
163 (8), and of self-reported 24-hour nutrient intake (8), and we computed the fatty liver index (FLI)
164 (14) and a T2D polygenic risk score (PRS) (15). The whole set of traits considered in this study is
165 described in detail in the Supplemental Material (DATA, METHODS, and Table S2).

166 *Assessment of progression rates*

167 We computed the progression rates for HbA_{1c} and several traits available at follow up
168 (Supplemental Table S4). Each trajectory was described with a conditional linear mixed-effect
169 model (16), in which the longitudinal component of the data was described as a proportional
170 function of time, with normally distributed slopes describing individual progression rates. HbA_{1c}
171 progression was adjusted for changes in BMI and diabetes medications, which were recorded at all
172 visits (as dosage and start and end of treatment). The adjustments were assumed to be 1)
173 proportional to BMI; 2) linearly related to the metformin dose, expressed as percentage of a
174 maximal dose of 3 grams; 3) linearly related to the cumulative dose for the other antidiabetic drugs
175 (insulin excluded), expressed as sum of the percentages of the maximum dose of each drug; 4)
176 constant under insulin treatment. A proportional effect of delay in HbA_{1c} assay, i.e. of the difference
177 between the time of measurement and the time of sample collection, was also introduced.
178 Medications were considered to be effective if taken at least 30 days before HbA_{1c} measurement.
179 OGIS and QUICKI trajectories were adjusted for changes in BMI. Further details about the
180 conditional linear mixed-effect models are provided in the Supplemental Material (METHODS).

181 *Statistical analysis*

182 Results are presented for participants ($N=732$) with GAD <11 U/ml and islet antigen-2 antibodies
183 (IA-2) <7.5 U/ml, to exclude other possible forms of diabetes (17). Distributions are described as
184 mean \pm standard deviation. Pairwise associations between continuous variables were assessed using
185 the Spearman correlation coefficient; differences between groups were assessed using the Wilcoxon
186 signed rank test (for two groups) and Kruskal-Wallis test (for three or more groups).

187 We used stepwise multivariable linear regression to determine the set of variables, as baseline
188 values (Table S2) and progression rates (Table S4), independently associated with the HbA_{1c}
189 progression rate, with adjustment for center, sex and age. For baseline variables, both
190 untransformed and transformed values were considered; transformations were logarithmic, or logit
191 when variables were constrained within an interval. The independent variables were included in

192 the regression model when their effects had $p < 0.05$ and produced an increment in the adjusted R^2
193 value. Two stepwise analyses were performed: one on all participants, excluding MRI variables
194 from the analysis, and one on the subset of participants with MRI data, including this data in the
195 analysis. Standardized coefficients were computed per standard deviation of the underlying data
196 distribution.

197 Since the distribution of HbA_{1c} progression rates was skewed to the right with a group of patients
198 with high values, we split the subjects into *average* and *fast* progressors according to a progression
199 rate threshold (see Results). We used multivariable logistic regression to assess the odds ratios of
200 average vs fast progression, using the independent variables identified in the multiple linear
201 regression analysis of HbA_{1c} progression. The logistic analysis provided values for AUROC,
202 sensitivity, specificity and accuracy, to be used as measures of the discrimination capacity of the
203 investigated independent variables over fast vs average progressors. These parameters must not be
204 interpreted as measures of predictive capacity.

205 *Role of the funding source*

206 The funders had no role in study design, in collection, analysis, and interpretation of data, in writing
207 of the report, or in the decision to submit the paper for publication. The corresponding author had
208 full access to all data and had final responsibility for the decision to submit for publication.

209 **Results**

210 *Subjects' baseline characteristics*

211 At baseline, the participants had age of 62 ± 8 years, were moderately obese (30.4 ± 4.9 kg/m² BMI),
212 and had HbA_{1c} of 6.41 ± 0.53 % (46.5 ± 5.8 mmol/mol) and fasting glucose of 7.1 ± 1.4 mmol/l. (Table
213 S2). 34% of the subjects were treated with metformin at baseline, the rest was treatment naïve.

214 *Progression rates of HbA_{1c} and other traits*

215 The individual HbA_{1c} progression rates (Supplemental Figure S1), adjusted for changes in BMI and
216 in diabetes medications, were on average only slightly positive and mostly distributed close to their
217 median (median, first and ninth deciles were 0.041, -0.038 and 0.185 %/year (0.45, -0.41 and 2.02
218 mmol mol⁻¹ year⁻¹), respectively). However, the distribution showed a heavy right tail with values
219 up to 0.897 %/year (9.8 mmol mol⁻¹ year⁻¹). The adjustment of progression rates for BMI changes
220 implied a standardized coefficient for the BMI effect of 0.37.

221 All the other investigated traits had a mean progression rate per year smaller, in absolute value, than
222 5% of the corresponding baseline average (see Table S5 for details). On average, waist
223 circumference, but not BMI, increased very slightly. Insulin sensitivity (as OGIS) and most of the
224 β-cell function parameters decreased. Fasting, but not post-meal, insulin clearance decreased. Total
225 cholesterol did not change, while its fractions showed opposite changes, with HDL increasing and
226 LDL decreasing; TG increased. Creatinine and ALT did not change, while AST and AST/ALT
227 increased.

228 Several pairwise associations were observed between HbA_{1c} progression rate and laboratory,
229 clinical, and functional parameters (Supplemental Figure S2). In particular, HbA_{1c} progression rate
230 was clearly associated ($p < 0.01$) with some baseline traits (positively with BMI, waist
231 circumference, triglycerides, glucagon, liver and visceral fat; inversely with age, HDL, insulin
232 sensitivity, and β-cell function) and some progression rates (positively with those of triglycerides
233 and liver enzymes; inversely with those of insulin sensitivity, β-cell function, AST/ALT ratio, and
234 HDL).

235 Several pairwise associations were also observed between the progression rates of the investigated
236 traits (Figure S2, panel B). GS and OGIS progression rates were independent of one another despite
237 HbA_{1c} progression rate being associated with both of them.

238 *Variables associated with HbA_{1c} progression rate: multivariable linear analysis*

239 In multivariable linear analysis of HbA_{1c} progression rate in all patients, the baseline values and the
240 progression rates of several traits provided an independent contribution (adjusted R^2 0.38; Figure 1,
241 panel A). Faster HbA_{1c} progression was independently associated with lower baseline values and
242 faster deterioration of insulin sensitivity (as OGIS) and β -cell function (mostly as glucose
243 sensitivity, GS), with higher baseline values of MMTT insulin clearance, CLIm, and with its
244 increase (all p -values <0.001). Faster HbA_{1c} progression was also independently associated with
245 lower baseline HDL ($p<0.05$) or its slower increase ($p<0.001$), with a quicker increase of TG
246 ($p<0.001$), as well as with higher baseline values of BMI ($p<0.01$) and lower baseline values of
247 HbA_{1c} ($p<0.001$). The variables with strongest effects were the baseline OGIS value and the
248 progression rates of OGIS, GS and CLIm (standardized coefficients, in absolute value, between
249 0.24 and 0.57).

250 In multivariable analysis of the subset of patients with baseline MRI measurements (adjusted R^2
251 0.40; Figure 1, panel B), baseline visceral fat was positively and independently correlated with
252 HbA_{1c} progression rate; moreover, female sex and younger age independently predicted faster
253 HbA_{1c} progression. The role of the other key metabolic parameters, OGIS, GS and CLIm, remained
254 similar. Replacing visceral fat with liver fat produced similar results (standardized coefficient equal
255 to 0.15 for visceral fat, to 0.11 for liver fat); when both visceral and liver fat were included in the
256 model, the latter was not independently associated with HbA_{1c} progression.

257 No independent effects were detected for smoking status, family history, T2D polygenic risk score,
258 baseline values of diet, physical activity, pancreatic fat, GLP-1 (total and intact at fasting, total at 60
259 min), glucagon, and 60-min proinsulin, baseline values and progression rates of AST and ALT.

260 Further details on the multivariable linear analysis are reported in the Supplemental Material
261 (RESULTS).

262 *Variables associated with HbA_{1c} progression rate: multivariable logistic analysis*

263 The threshold selected to separate the heavy right tail of the distribution of HbA_{1c} progression rates
264 was 0.255 %/year (2.79 mmol mol⁻¹ year⁻¹). This threshold split the subjects into average
265 progressors (N=699), with a progression rate of 0.044±0.076 %/year (0.48±0.83 mmol mol⁻¹ year⁻¹),
266 and fast progressors (N=33), with a ~10-fold mean progression rate (0.460±0.185 %/year,
267 5.03±2.02 mmol mol⁻¹ year⁻¹) (Figure 2).

268 We found that the trajectories of most variables independently affecting HbA_{1c} progression as from
269 the linear analysis were clearly different ($p<0.001$) in the two groups (Figure 2): in fast progressors,
270 OGIS and GS strongly declined and TG and CLIm markedly increased. At baseline, fast
271 progressors had lower OGIS ($p<0.05$), CLIm ($p<0.01$) and HDL ($p<0.001$), and higher BMI
272 ($p<0.01$).

273 Logistic analysis substantially confirmed the results of linear regression (Figure 1), with half the
274 investigated variables still contributing ($p<0.05$) to distinguish average and fast progressors (Figure
275 3): fast HbA_{1c} progression independently associated with stronger deterioration and a lower
276 baseline value of OGIS and GS, CLIm increase, and HDL reduction. The discrimination capacity of
277 the logistic model, computed as AUROC, was 0.94 (95% CI between 0.86 and 0.98).

278 Similar outcomes were obtained using lower HbA_{1c} progression rate thresholds, which resulted in
279 larger numbers of patients classified as fast progressors (Supplemental Material - RESULTS,
280 Figures S1 and S3).

281 At baseline, the percentage of patients treated with metformin were not different between fast
282 progressors (39.4% [24.7-56.3%, 95% CI]) and average progressors (33.9% [30.5-37.5%], $p =$
283 0.64). At the last visit, the percentage of patients treated with any diabetes medication was
284 somewhat higher in fast progressors, as expected ($p = 0.048$, details provided in the Supplemental
285 Material - RESULTS). Only 7 average progressors were on insulin at the last visit.

286 *Impact of stable OGIS, GS or CLIm on proportion of fast HbA_{1c} progressors*

287 Because HbA_{1c} progression was associated with worsening of three main factors, OGIS, GS and
288 CLIm, we have evaluated the possible importance of maintaining one of these key traits relatively
289 stable in order to avoid fast progression. For this purpose, we considered each trait as deteriorating
290 if its progression rate fell within its worst tertile (the bottom tertile for OGIS and GS, the top one
291 for CLIm), and as stable if it fell in the other two tertiles. We examined the subgroups of patients in
292 which none or only one of these key traits was relatively stable (Table 1).

293 We found that the proportion of fast progressors was 56% in the patient subgroup where GS, OGIS
294 and CLIm were all deteriorating, and decreased to 8-10% in the subgroups where a single trait,
295 either GS, OGIS or CLIm, was stable. All proportions were different from 0 at 90% confidence
296 level, stressing that fast progression did not imply quick changes for each of the three considered
297 traits. All differences in proportions (one stable trait *vs* none) had $p < 0.001$, and were associated to
298 odds ratio for fast *vs* average progression below 0.1 (Table 1); thus, relatively stable progression
299 rate of one single trait among GS, OGIS and CLIm was strongly associated to reduced glycemc
300 deterioration.

301 **Conclusions**

302 Leveraging on the detailed participant characterization of the DIRECT study, we have been able to
303 elucidate the processes underlying glycemc deterioration in T2D patients in the initial phase of the
304 disease. We found that HbA_{1c} deterioration was independently associated with 1) a decrease in
305 insulin sensitivity; 2) a decrease in β -cell function (primarily β -cell glucose sensitivity); 3) an
306 increase in insulin clearance; 4) lower values of insulin sensitivity and glucose sensitivity and
307 higher values of insulin clearance at baseline. Further variables independently associated with faster
308 HbA_{1c} progression were declining HDL, increasing TG and high baseline visceral or liver fat.

309 The variables identified by multivariable linear analysis also explained the rapid HbA_{1c}
310 deterioration detected in a subset of patients (identified as fast progressors), the strongest predicting
311 variables of the multivariable linear model being significant also with logistic analysis. Clear

312 differences were evident between fast and average HbA_{1c} progressors (Figure 2), consistent with the
313 associations derived from the multivariable linear analysis. The high discrimination capacity of the
314 logistic analysis suggests that the selected variables capture the most relevant pathophysiological
315 factors underlying glycemic deterioration.

316 The independent associations with HbA_{1c} progression of several variables, in particular the
317 progression rates of insulin sensitivity, β -cell function and insulin clearance, and the existence of
318 fast HbA_{1c} progressors with relatively stable conditions for any of these three traits (Table 1),
319 indicates 1) that the processes of glycemic deterioration are heterogeneous in this population of
320 T2D patients; 2) that fast progression does not imply quick deterioration of a specific trait, e.g.
321 insulin sensitivity or β -cell function.

322 The dichotomous analysis shows that the odds for fast *vs* average progression are substantially
323 reduced when either glucose sensitivity, insulin sensitivity or insulin clearance is relatively stable.
324 Although these findings do not demonstrate causality, they suggest that preventing either high
325 degradation rates of glucose sensitivity or insulin sensitivity, or high increase rates of insulin
326 clearance, may be an effective strategy to slow down glycemic deterioration in the initial phase of
327 the disease. This reemphasizes the importance of lifestyle interventions aiming at controlling insulin
328 resistance, as preventing deterioration of the other traits currently appears more difficult.

329 This study also shows that insulin resistance plays a major role in glycemic deterioration in these
330 T2D patients. In particular, we show associations of glycemic deterioration with baseline insulin
331 sensitivity and its longitudinal change that the Belfast Diet Study (1), UKPDS (4,18) and ADOPT
332 (6) could not identify, possibly due to differences in subject selection or to the use of post-MMTT
333 *vs* fasting insulin sensitivity indices. We also demonstrate that the associations between glycemic
334 deterioration and insulin sensitivity are independent from both the baseline value and the
335 progression rate of the β -cell function, and that insulin resistance progresses independently from β -
336 cell glucose sensitivity. Since in our analysis both HbA_{1c} and insulin sensitivity trajectories were

337 adjusted for BMI changes and BMI did not increase on average, we can conclude that worsening of
338 insulin resistance in T2D and the associated glyceemic deterioration are partly independent from
339 BMI changes. Whether the observed average increases in TG and AST (whose progression rates
340 were inversely correlated with OGIS progression rate) have a role in insulin sensitivity deterioration
341 (19), and whether this is mediated by ectopic fat accumulation (20), deserves further study.

342 UKPDS 25 and 26 (4,18), the Belfast Diet Study (1) and the ADOPT study (6) identified baseline
343 HOMA-%B as a predictor of glyceemic deterioration (insulin requirement within 6 years for
344 UKPDS, time of failure to dietary therapy for the Belfast Diet Study, and monotherapy failure
345 before 4 years for ADOPT). Our study confirms the role of β -cell dysfunction as driver of glyceemic
346 deterioration using a dynamic β -cell function assessment based on a glucose challenge, rather than
347 on fasting data only. We show that both baseline β -cell dysfunction (especially β -cell glucose
348 sensitivity) and its deterioration over time are independently associated with HbA_{1c} worsening.
349 Moreover, we demonstrate that patients with limited or absent deterioration in β -cell function have
350 considerably lower odds of rapid glyceemic deterioration.

351 Another novel finding is the strong and independent association between HbA_{1c} progression and
352 insulin clearance during the MMTT, CLIm. To our knowledge, this is the first study examining
353 insulin clearance trajectories after T2D onset. We found that higher baseline CLIm and faster CLIm
354 increase over time independently associate with faster HbA_{1c} progression. This is consistent with
355 the glucose homeostasis mechanisms, as higher CLIm reduces the average insulin levels. Notably,
356 we found a positive correlation between insulin sensitivity and insulin clearance, considering both
357 the baseline values of the two traits, in agreement with previous findings (21), and their progression
358 rates (Figure S2). However, on average, in spite of a decrease in insulin sensitivity, insulin
359 clearance did not decrease. These findings show that, while in pre-diabetic subjects insulin
360 clearance reduction may be a way to mitigate the effects of insulin resistance (22), in T2D patients
361 this compensation appears present but impaired and contributing to glyceemic deterioration. The

362 reasons underlying these results remain elusive. The lack of decrease in insulin clearance may be
363 explained by the decrease of total MMTT insulin secretion and consequent desaturation of insulin
364 utilization (23) only in fast progressors, as in average progressors total insulin secretion slightly
365 increased (Figure 2). Whether hepatic or extrahepatic mechanisms underlie these findings cannot be
366 determined from this study and deserves further investigation.

367 Our results on TG and HDL effects were partially anticipated by a study of the Genetics of Diabetes
368 Audit and Research (GoDARTS) (24), where the outcome was the risk of progression to insulin
369 treatment. The study identified baseline TG and HDL (besides BMI, sex, and age, year and HbA_{1c}
370 at diagnosis) as independent determinants. A later study on the same data (25), investigating the
371 baseline determinants of HbA_{1c} progression rate over about 9 years, confirmed an independent
372 effect of HDL (together with age, BMI and year at diagnosis) but not of TG. The FIELD study in
373 T2D patients on lifestyle measures only revealed that the HDL effect on initiation of oral
374 hypoglycemic agents survives the adjustment for HOMA-IR (26). Compared to previous studies
375 (24–26) our analysis includes the progression rates of plasma lipid components and baseline MRI
376 assessment of regional fat. We show that baseline HDL and BMI, and the progression rates of TG
377 and HDL are associated with HbA_{1c} progression, even after accounting for the effects of the three
378 main determinants of glucose homeostasis, i.e. insulin sensitivity, β -cell function and insulin
379 clearance. In the subset of participants with MRI data, baseline visceral fat or liver fat was
380 independently correlated with HbA_{1c} progression rate, a further novel observation. These findings
381 suggest that additional lipid-dependent factors contribute to HbA_{1c} deterioration, possible
382 candidates being fat accumulation in the viscera (with excessive supply of fatty acids to the liver
383 (27)), liver fat and consequent hepatic insulin resistance (28), or glucose overproduction (29). The
384 role of visceral/liver fat supports interventions to reduce ectopic fat as a possible way for slowing
385 future glycaemic progression.

386 Previous studies have reported an inverse correlation between baseline age and HbA_{1c} progression
387 (1,4,6,24,25,30). In our analysis, baseline age does not have a clear independent role in the
388 multivariable model, most likely because the age range is relatively narrow relative to other studies,
389 or because the stronger predictors of HbA_{1c} progression are correlated with age. The latter
390 explanation would suggest that the age univariate effect on glycemic deterioration is indirect. We
391 do not find a clear sex effect in glycemic deterioration, in agreement with most previous studies
392 (1,4,6,24,25).

393 In the multivariable model, baseline HbA_{1c} was independently and inversely correlated with HbA_{1c}
394 progression rate, in contrast with previous findings (1,4,6,24,30). However, baseline HbA_{1c} was not
395 significant in the logistic model. The most likely explanation of this finding is regression to the
396 mean: indeed, a random decrease in baseline HbA_{1c} can produce a higher estimate of HbA_{1c}
397 progression rate, particularly when the follow-up period is not long, as in our study. Tight glycemic
398 control, an inclusion criterion, may have enhanced this effect.

399 This study does not find a relevant role of other variables often associated with glucose control. In
400 particular, we did not find an effect of smoking status (reported in GPRD (30)), T2D polygenic risk
401 score (in agreement with GoDARTS (24)), baseline values of diet, physical activity, pancreatic fat,
402 GLP-1, and glucagon. Several of these variables were not associated with HbA_{1c} progression rate
403 even in simple correlation analysis (Figure S2). The lack of association for pancreatic fat is
404 particularly relevant, and contributes to the ongoing discussion on the role of pancreas fat in T2D
405 management (31).

406 In spite of the unique extensive phenotyping of our study and the consistent results, a significant
407 limitation is the relatively short follow-up period (3 years). The accuracy of the estimated HbA_{1c}
408 progression rate over this time frame may be limited, and in a longer time period the factors
409 contributing to progression may differ. In this study, we could not assess the changes over time of
410 relevant variables such as regional fat by MRI, diet and physical activity. MRI measurements were

411 available only for a subset of subjects. Insulin sensitivity was not derived from the gold standard
412 euglycemic clamp. As the cohort included only patients of white race, our findings are not
413 generalizable to other racial/ethnic groups. Causal relationships could not be inferred from our
414 regression analyses. The study of the mechanisms underlying the deterioration of the factors
415 affecting HbA_{1c} progression, an important aspect to envisage optimal treatment strategies, also
416 requires further investigation.

417 In summary, based on the extensively phenotyped cohort of white European diabetic patients of the
418 DIRECT study, we identified decreasing insulin sensitivity, deteriorating β -cell function, increasing
419 insulin clearance, high liver or visceral fat, and worsening of the lipid profile as the most important
420 factors independently associated with HbA_{1c} deterioration in the early phase of the disease. We also
421 showed that patients with a relatively stable value over time of at least one of insulin sensitivity, β -
422 cell glucose sensitivity, or insulin clearance have considerably reduced odds of fast HbA_{1c} increase.
423 This study contributes to the understanding of the factors underlying diabetes progression,
424 elucidating the processes that might be targeted for personalized treatments.

425 **Acknowledgments.**

426 The authors thank the participants across all IMI DIRECT study centers for their contributions to
427 the study. We also thank the staff involved in the design, implementation and conduction of the
428 study.

429 **Funding.**

430 The work leading to this publication has received support from the Innovative Medicines Initiative
431 Joint Undertaking under grant agreement n°115317 (DIRECT), resources of which are composed of
432 financial contribution from the European Union's Seventh Framework Programme (FP7/2007-2013)
433 and EFPIA companies' in kind contribution. Information on the project can be found at
434 <http://www.direct-diabetes.org/>.

435 **Duality of Interest.**

436 Dr. Jennison reports grants from EU Innovative Medicines Initiative, during the conduct of the
437 study; personal fees from NovoNordisk, Sanofi, AstraZeneca, and Boehringer-Ingelheim, outside
438 the submitted work. Dr. 't Hart reports grants from IMI-JU, during the conduct of the study. Dr.
439 Beulens reports grants from IMI-EU, during the conduct of the study. Dr. Musholt reports to be an
440 employee of Sanofi-Aventis Deutschland GmbH. Dr. Martin Ridderstråle reports to be an employee
441 of Novo Nordisk A/S. Dr. Brunak reports personal fees from Intomics A/S and Proscion A/S,
442 outside the submitted work. As of January 2020, Dr. Mahajan is an employee of Genentech, and a
443 holder of Roche stock. Dr. Ruetten is an employee of Sanofi, and shareholder. Dr. McCarthy reports
444 employment and stock from Genentech, grants and personal fees from Merck, Novo Nordisk, Eli
445 Lilly, and Pfizer, grants from Roche, Servier, Sanofi Aventis, Abbvie, AstraZeneca, Boehringer
446 Ingelheim, Janssen, and Takeda, outside the submitted work. Dr. Schwenk reports grants from
447 KTH, during the conduct of the study. Dr. Franks reports research funding from Boehringer
448 Ingelheim, Eli Lilly, Janssen, Novo Nordisk A/S, Sanofi Aventis and Servier, consulting fees from

449 Eli Lilly, Novo Nordisk and Zoe Global Ltd; he also reports stock options in Zoe Global Ltd. All
450 other co-authors have nothing to disclose.

451 **Author Contributions.**

452 R.B. and A.M. designed the analysis, analyzed the data, and wrote the manuscript. R.B., C.J.,
453 A.G.J., M.W., E.R.P. and A.M. interpreted the results. E.R.P. and A.M. supervised the analysis.
454 C.J., A.G.J., A.K., M.W. and E.R.P. reviewed the manuscript. All authors were involved in the
455 DIRECT study at different levels, and were essential for the production, release and management of
456 the data analyzed here. R.B. is the guarantor of this work and, as such, takes full responsibility for
457 the work as a whole, including the study design, access to data, and the decision to submit and
458 publish the manuscript.

459 **References**

- 460 1. Levy J, Atkinson AB, Bell PM, McCance DR, Hadden DR. Beta-cell deterioration determines
461 the onset and rate of progression of secondary dietary failure in Type 2 diabetes mellitus: the
462 10-year follow-up of the Belfast Diet Study. *Diabet Med.* 1998 Apr;15(4):290–6.
- 463 2. U.K. Prospective Diabetes Study Group. U.K. Prospective Diabetes Study 16: Overview of 6
464 Years' Therapy of Type II Diabetes: A Progressive Disease. *Diabetes.* 1995 Nov
465 1;44(11):1249–58.
- 466 3. Best JD, Drury PL, Davis TME, Taskinen M-R, Kesäniemi YA, Scott R, et al. Glycemic
467 Control Over 5 Years in 4,900 People With Type 2 Diabetes. *Diabetes Care.* 2012
468 May;35(5):1165–70.
- 469 4. Matthews DR, Cull CA, Stratton IM, Holman RR, Turner RC. UKPDS 26: sulphonylurea
470 failure in non-insulin-dependent diabetic patients over six years. *Diabet Med.* 1998;15(4):297–
471 303.
- 472 5. Kahn SE, Haffner SM, Heise MA, Herman WH, Holman RR, Jones NP, et al. Glycemic
473 Durability of Rosiglitazone, Metformin, or Glyburide Monotherapy. *N Engl J Med.* 2006 Dec
474 7;355(23):2427–43.
- 475 6. Kahn SE, Lachin JM, Zinman B, Haffner SM, Aftring RP, Paul G, et al. Effects of
476 Rosiglitazone, Glyburide, and Metformin on β -Cell Function and Insulin Sensitivity in
477 ADOPT. *Diabetes.* 2011 May;60(5):1552–60.
- 478 7. Festa A, Williams K, D'Agostino R, Wagenknecht LE, Haffner SM. The Natural Course of
479 beta-Cell Function in Nondiabetic and Diabetic Individuals: The Insulin Resistance
480 Atherosclerosis Study. *Diabetes.* 2006 Apr 1;55(4):1114–20.
- 481 8. Koivula RW, Heggie A, Barnett A, Cederberg H, Hansen TH, Koopman AD, et al. Discovery
482 of biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes:
483 rationale and design of the epidemiological studies within the IMI DIRECT Consortium.
484 *Diabetologia.* 2014 Jun;57(6):1132–42.
- 485 9. Koivula RW, Forgie IM, Kurbasic A, Viñuela A, Heggie A, Giordano GN, et al. Discovery of
486 biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes:
487 descriptive characteristics of the epidemiological studies within the IMI DIRECT Consortium.
488 *Diabetologia.* 2019 Sep;62(9):1601–15.
- 489 10. American Diabetes Association. Standards of Medical Care in Diabetes. *Diabetes Care.*
490 2011;34(Suppl 1):S11–61.
- 491 11. Katz A, Nambi SS, Mather K, Baron AD, Follmann DA, Sullivan G, et al. Quantitative insulin
492 sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans.
493 *J Clin Endocrinol Metab.* 2000 Jul;85(7):2402–10.
- 494 12. Mari A, Pacini G, Murphy E, Ludvik B, Nolan JJ. A Model-Based Method for Assessing
495 Insulin Sensitivity From the Oral Glucose Tolerance Test. *Diabetes Care.* 2001 Mar
496 1;24(3):539–48.
- 497 13. Mari A, Tura A, Gastaldelli A, Ferrannini E. Assessing Insulin Secretion by Modeling in
498 Multiple-Meal Tests: Role of Potentiation. *Diabetes.* 2002 Feb 1;51(Supplement 1):S221–6.

- 499 14. Bedogni G, Bellentani S, Miglioli L, Masutti F, Passalacqua M, Castiglione A, et al. The Fatty
500 Liver Index: a simple and accurate predictor of hepatic steatosis in the general population.
501 BMC Gastroenterol. 2006 Dec;6(1):33.
- 502 15. Mahajan A, Taliun D, Thurner M, Robertson NR, Torres JM, Rayner NW, et al. Fine-mapping
503 type 2 diabetes loci to single-variant resolution using high-density imputation and islet-
504 specific epigenome maps. Nat Genet. 2018 Nov;50(11):1505–13.
- 505 16. Verbeke G. Conditional Linear Mixed Models. Am Stat. 2001;55(1):25–34.
- 506 17. Hitman GA. The message for MODY. Diabet Med J Br Diabet Assoc. 2011 Sep;28(9):1009.
- 507 18. Turner R, Stratton I, Horton V, Manley S, Zimmet P, Mackay IR, et al. UKPDS 25:
508 autoantibodies to islet-cell cytoplasm and glutamic acid decarboxylase for prediction of insulin
509 requirement in type 2 diabetes. The Lancet. 1997 Nov 1;350(9087):1288–93.
- 510 19. Ginsberg HN, Zhang Y-L, Hernandez-Ono A. Regulation of Plasma Triglycerides in Insulin
511 Resistance and Diabetes. Arch Med Res. 2005 May 1;36(3):232–40.
- 512 20. Kotronen A, Juurinen L, Tiikkainen M, Vehkavaara S, Yki-Järvinen H. Increased Liver Fat,
513 Impaired Insulin Clearance, and Hepatic and Adipose Tissue Insulin Resistance in Type 2
514 Diabetes. Gastroenterology. 2008 Jul 1;135(1):122–30.
- 515 21. Lorenzo C, Hanley AJG, Wagenknecht LE, Rewers MJ, Stefanovski D, Goodarzi MO, et al.
516 Relationship of Insulin Sensitivity, Insulin Secretion, and Adiposity With Insulin Clearance in
517 a Multiethnic Population. Diabetes Care. 2013 Jan;36(1):101–3.
- 518 22. Jung S-H, Jung C-H, Reaven GM, Kim SH. Adapting to insulin resistance in obesity: role of
519 insulin secretion and clearance. Diabetologia. 2018 Mar;61(3):681–7.
- 520 23. Ferrannini E, Cobelli C. The kinetics of insulin in man. II. Role of the liver. Diabetes Metab
521 Rev. 1987 Apr 1;3(2):365–97.
- 522 24. Zhou K, Donnelly LA, Morris AD, Franks PW, Jennison C, Palmer CNA, et al. Clinical and
523 Genetic Determinants of Progression of Type 2 Diabetes: A DIRECT Study. Diabetes Care.
524 2014 Mar;37(3):718–24.
- 525 25. Donnelly LA, Zhou K, Doney ASF, Jennison C, Franks PW, Pearson ER. Rates of glycaemic
526 deterioration in a real-world population with type 2 diabetes. Diabetologia. 2018;61(3):607–
527 15.
- 528 26. Waldman B, Jenkins AJ, Davis TME, Taskinen M-R, Scott R, O’Connell RL, et al. HDL-C
529 and HDL-C/ApoA-I Predict Long-Term Progression of Glycemia in Established Type 2
530 Diabetes. Diabetes Care. 2014 Aug 1;37(8):2351–8.
- 531 27. Sattar N, McConnachie A, Ford I, Gaw A, Cleland SJ, Forouhi NG, et al. Serial metabolic
532 measurements and conversion to type 2 diabetes in the west of Scotland coronary prevention
533 study: specific elevations in alanine aminotransferase and triglycerides suggest hepatic fat
534 accumulation as a potential contributing factor. Diabetes. 2007 Apr;56(4):984–91.
- 535 28. Birkenfeld AL, Shulman GI. Non Alcoholic Fatty Liver Disease, Hepatic Insulin Resistance
536 and Type 2 Diabetes. Hepatol Baltim Md. 2014 Feb;59(2):713–23.

- 537 29. Samuel VT, Liu Z-X, Qu X, Elder BD, Bilz S, Befroy D, et al. Mechanism of hepatic insulin
538 resistance in non-alcoholic fatty liver disease. *J Biol Chem*. 2004 Jul 30;279(31):32345–53.
- 539 30. Cook MN, Girman CJ, Stein PP, Alexander CM, Holman RR. Glycemic Control Continues to
540 Deteriorate After Sulfonylureas Are Added to Metformin Among Patients With Type 2
541 Diabetes. *Diabetes Care*. 2005 May 1;28(5):995–1000.
- 542 31. Lim EL, Hollingsworth KG, Aribisala BS, Chen MJ, Mathers JC, Taylor R. Reversal of type 2
543 diabetes: normalisation of beta cell function in association with decreased pancreas and liver
544 triacylglycerol. *Diabetologia*. 2011 Oct 1;54(10):2506–14.

546 **Table 1.** Proportion of fast HbA1c progressors with different combinations of stable/deteriorating conditions for GS, OGIS and CLIm progression
 547 rates.

Condition*			Average progressors (N)	Fast progressors (N)	Fast progressors (%) [95% CI]	Odds ratio [95% CI]	p-value [†]
GS	OGIS	CLIm					
Deteriorating	Deteriorating	Stable	47	5	9.6 [4.2,20.6]	0.09 [0.02,0.32]	2E-4
Deteriorating	Stable	Deteriorating	56	6	9.7 [4.5,19.5]	0.09 [0.02,0.30]	8E-5
Stable	Deteriorating	Deteriorating	34	3	8.1 [2.8,21.3]	0.07 [0.02,0.32]	4E-4
Deteriorating	Deteriorating	Deteriorating	8	10	55.6 [33.7,75.4]	-	-

548 * The progression rate thresholds dividing stable and deteriorating traits for OGIS, GS and CLIm are $-16.68 \text{ ml min}^{-1} \text{ m}^{-2} \text{ year}^{-1}$, $-4.07 \text{ pmol min}^{-1} \text{ m}^{-2} \text{ mmol}^{-1} \text{ l}$
 549 year^{-1} and $0.0184 \text{ l min}^{-1} \text{ m}^{-2} \text{ year}^{-1}$, respectively.

550 [†] Two-sided Chi-square test ($\alpha=0.05$), with Yates continuity correction, on the proportion of fast progressors in the row compared to the same proportion in the
 551 last row.

552 GS: β -cell glucose sensitivity; OGIS: oral insulin sensitivity; CLIm: mixed meal test insulin clearance.

553 **Figure legends**

554 Figure 1. Variables independently associated with HbA_{1c} progression rate from multivariable linear
555 analysis. Panel A: all subjects are included in the analysis (625 with all variables), and MRI
556 measurements are not considered; panel B: only subjects with MRI are included in the analysis (374
557 with all variables), and MRI measurements are taken into consideration. For each variable, the
558 figure shows the standardized coefficients \pm 95% CI of the effect. Age and HDL were log-
559 transformed. OGIS: oral insulin sensitivity; CLIm: mixed meal test insulin clearance; GS: β -cell
560 glucose sensitivity; TG: fasting triacylglycerol; HDL: fasting HDL-cholesterol; RS: β -cell rate
561 sensitivity; progr: progression rate; bas: baseline value; *: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$.

562 Figure 2. Temporal trajectories or baseline values (bar graphs) of HbA_{1c} and other key traits in fast
563 (red lines) and average (blue lines) progressors. Data are mean \pm standard error. Simple
564 comparisons between fast and average progressors (Wilcoxon rank sum test) are shown for baseline
565 values (asterisks at month 0) and progression rates (asterisks at month 18). These comparisons may
566 differ from the results of the multivariable analyses (Figures 2 and 4). Sex is not included in the
567 figure: males were 42% and 36% in average and fast progressors, respectively (non-significant,
568 Chi-squared test). HbA_{1c} values at 27 months are not displayed as they were collected in a subgroup
569 of individuals. In average progressors, HbA_{1c} increases from 46.4 ± 0.2 mmol/mol to 46.7 ± 0.3
570 mmol/mol; in fast progressors, from 48.9 ± 1.21 mmol/mol to 75.7 ± 2.5 mmol/mol. OGIS: insulin
571 sensitivity; CLIm: mixed meal test insulin clearance; GS: β -cell glucose sensitivity; RS: β -cell rate
572 sensitivity; TG: fasting triacylglycerol; HDL: fasting HDL-cholesterol; ISRtot: total mixed meal
573 test insulin secretion; bas: baseline value; *: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$.

574 Figure 3. Odds ratios \pm 95% CI from the multivariable logistic analysis of fast vs average HbA_{1c}
575 progressors. The independent variables are those identified by multivariable linear analysis of
576 HbA_{1c} progression, excluding MRI variables ($N=625$, with 32 fast progressors and 593 average
577 progressors). Age and HDL were log-transformed. Values for sensitivity, specificity and accuracy

578 were derived via maximization of balanced accuracy. OGIS: insulin sensitivity; CLIm: mixed meal
579 test insulin clearance; GS: β -cell glucose sensitivity; TG: fasting triacylglycerol; HDL: fasting
580 HDL-cholesterol; RS: β -cell rate sensitivity; progr: progression rate; bas: baseline value; AUROC:
581 area under the receiver operating characteristics; *: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$.