

Official journal of the Spanish Society of Chemotherapy

Original

International (CC BY-NC 4.0)(https://creativecommons.org/licenses/by-nc/4.0/).

Rafael Cantón^{1,2} Elena Loza¹ Javier Aznar³ Francisco Javier Castillo⁴ Emilia Cercenado^{5,6} Pablo Arturo Fraile-Ribot⁷ Fernando González-Romo⁸ José Luis López-Hontangas⁹ Jesús Rodríguez-Lozano¹⁰ Ana Isabel Suárez-Barrenechea¹¹ Fe Tubau^{6,12} Jazmín Díaz-Regañón¹³ Diego López-Mendoza¹³ and the SMART-Spain Working Group.

Monitoring the antimicrobial susceptibility of Gramnegative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)

- ¹Servicio de Microbiología, Hospital Universitario Ramón y Cajal and Instituto Ramón y Cajal de Investigación Sanitaria (IRYCIS), Madrid, Spain. ²Red Española de Investigación en Patología Infecciosa (REIPI), Madrid, Spain.
- ³Servicio de Microbiología. Unidad Clínica de Enfermedades Infecciosas, Microbiología y Medicina Preventiva. Departamento de Microbiologia Instituto de Biomedicina de Sevilla, Hospital Universitario Virgen del Rocío, CSIC/Universidad de Sevilla, Spain.
- ⁴Servicio de Microbiología, Hospital Clínico Universitario Lozano Blesa, Zaragoza, Facultad de Medicina, Universidad de Zaragoza. IIS Aragón, Spain.
- ⁵Servicio de Microbiología y Enfermedades Infecciosas, Hospital General Universitario Gregorio Marañón, Madrid, Spain.
 ⁶Centro de Investigación en Red de Enfermedades Respiratorias (CIBERES), Madrid, Spain.
- ⁷Servicio de Microbiología. Hospital Universitario Son Espases e Instituto de Investigación Sanitaria Islas Baleares (IDIS-BA), Mallorca, Spain.
- ⁸Servicio de Microbiología Clínica, Hospital Clínico San Carlos, Madrid, Spain.
- ⁹Servicio de Microbiología, Hospital Universitario y Politécnico La Fe, Valencia, Spain.
- ¹⁰Servicio de Microbiología, Hospital Universitario Marqués de Valdecilla e Instituto de Investigación Valdecilla (IDIVAL), Santander, Spain.
- ¹¹Unidad de Gestión Clínica de Enfermedades Infecciosas, Microbiología y Medicina Preventiva. Hospital Universitario Virgen Macarena. Sevilla, Spain.
- ¹²Servicio de Microbiología Hospital Universitario Bellvitge e Instituto de Investigación Biomédica de Bellvitge (IDIBELL), Hospitalet de Llobregat, Barcelona, Spain.
- ¹³Departamento Médico MSD España, Madrid, Spain.

Article history Received: 9 December 2018; Accepted: 9 January 2019

ABSTRACT

Introduction. Continuous antimicrobial resistance surveillance is recommended by Public Health authorities. We updated data from the SMART (Study for Monitoring Antimicrobial Resistance Trends) surveillance study in Spain.

Material and methods. The antimicrobial susceptibility data and extended-spectrum beta-lactamase (ESBL) production in isolates recovered from intra-abdominal (IAI) (n=1,429) and urinary tract (UTI) (n=937) infections during the 2016-2017 SMART study in 10 Spanish hospitals were analysed.

Results. Escherichia coli was the most frequently microorganism isolated (48.3% and 53.7%) followed by *Klebsiella* spp. (11.5% and 21.9%) in IAIs and UTIs, respectively. Figures for *Pseudomonas aeruginosa* were 9.0% and 6.1%, being more frequently recovered from patients with nosocomial infections. Overall, 9.9% (IAI) and 14.0% (UTI) of *E. coli, Klebsiella* spp. and *Proteus mirabilis* isolates were ESBL-producers, being *Klebsiella* pneumoniae (34.5%) from UTI of nosocomial origin the most frequent. ESBL-producers were higher in patients >60 years in

Correspondence: Rafael Cantón. both IAIs and UTIs. As in previous years, amikacin (96.3%-100% susceptibility), ertapenem (84.2%-100%) and imipenem (70.3%-100%) were the most active antimicrobials tested among Enterobacterales species. The activity of amoxicillin-clavulanic, piperacillin-tazobactam, and ciprofloxacin susceptibility was lower, particularly among ESBL-producers. Ertapenem susceptibility (88.9%-100%) was retained in ESBL-*E. coli* isolates that were resistant to these antimicrobials but decreased (28.6%-100%) in similar isolates of *K. pneumoniae*.

Conclusions. Continuous antimicrobial resistance surveillance from the SMART study reveals overall maintenance of ESBL-producers in Spain, although with higher presence in isolates from UTIs than from IAIs. Moreover, ertapenem activity was high in *E. coli* irrespective of ESBL production but decreased in *K. pneumoniae*, particularly among ESBL-producers.

Key words: antimicrobial resistance surveillance, intra-abdominal infection, urinary tract infection, extended-spectrum-beta-lactamases, carbapenems

Seguimiento de la sensibilidad antimicrobiana de microorganismos gramnegativos procedentes de infecciones intraabdominales y urinarias del estudio SMART (España, 2016 y 2017)

RESUMEN

Introducción. Las autoridades de Salud Pública re-

Servicio de Microbiología. Hospital Universitario Ramón y Cajal. Carretera de Colmenar Km. 9,1. 28034 Madrid. Spain. Phone. +34913368330. E-mail: rafael.canton@salud.madrid.org

R. Cantón, et al.

Monitoring the antimicrobial susceptibility of Gram-negative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)

comiendan la vigilancia continua de la resistencia a los antimicrobianos. Se actualizan los datos del estudio SMART (*Study for Monitoring Antimicrobial Resistance Trends*) en España.

Material y métodos. Se analizaron los datos de sensibilidad antimicrobiana y la producción de betalactamasas de espectro extendido (BLEE) en aislamientos obtenidos en el estudio SMART de infecciones intraabdominales (IIA) (n=1.429) y del tracto urinario (ITU) (n=937) durante 2016-2017 en 10 hospitales españoles.

Resultados. Escherichia coli fue el microorganismo más frecuente (54,5% y 57,5%, respectivamente), seguido de Klebsiella spp. (18,4% y 25,4%) en IIA y en ITU. En Pseudomonas aeruginosa estas cifras fueron 9% y 6%, siendo más frecuente en la infección nosocomial. El 9,9% (IIA) y el 14% (ITU) del total de los aislados de E. coli. Klebsiella spp. v Proteus mirabilis producían BLEE, obteniéndose la tasa más alta en Klebsiella pneumoniae (34.5%) en ITU nosocomial. El mayor porcentaje de aislados con BLEE se observó en pacientes >60 años, tanto en IIA como en ITU. Como en años anteriores, amikacina (sensibilidad 96,3%-100%), ertapenem (84,2%-100%) e imipenem (70,3%-100%) fueron los antimicrobianos más activos en Enterobacterales. La sensibilidad a amoxicilina-ácido clavulánico. piperacilina-tazobactam y ciprofloxacino fue menor, en particular en los productores de BLEE. La sensibilidad a ertapenem (88,9%-100%) se mantuvo en E. coli con BLEE resistente a estos antimicrobianos, pero disminuyó (28,6%-100%) en aislados similares de K. pneumoniae.

Conclusiones. La vigilancia continua de la resistencia a los antimicrobianos en el estudio SMART revela el mantenimiento de la frecuencia de aislados productores de BLEE en España, pero con mayor presencia en las ITUs que en las IIAs. Además, la sensibilidad a ertapenem fue alta en *E. coli* con independencia de la producción de BLEE, pero disminuyó en *K. pneumoniae*, sobre todo en los productores de BLEE.

Palabras clave: vigilancia epidemiológica de la resistencia, infección intraabdominal, infección urinaria, betalactamasas de espectro extendido, carbapenems

INTRODUCTION

The increase in antimicrobial resistance is a worldwide reality that threatens the prevention and effective treatment of an increasing number of infections, challenging clinical microbiologists and infectious disease specialists [1]. Two of the most common infections are urinary tract (UTI) and intraabdominal (IAI) infections caused mainly by Enterobacterales, in particular Escherichia coli and Klebsiella species [2,3]. In the 1980s, extended spectrum beta-lactamase (ESBL)-producing Enterobacterales were considered one of the leading causes of nosocomial infections and later also of those acquired in the community [4]. These enzymes have the ability to hydrolyze beta-lactam antibiotics, including penicillins, cephalosporins and the monobactam aztreonam but not carbapenems [5]. As a consequence, carbapenems were considered the antimicrobials of choice for the treatment of infections caused by ESBL producers, however the prevalence of carbapenemases, enzymes that inactivate them, continue to increase worldwide [6]. In addition, the production of ESBL combined with mutations affecting permeability can also contribute to the carbapenems resistance. This situation warns the need for surveillance of susceptibility to antimicrobials, especially to carbapenems. Global surveillance programs such as SMART (Study for Monitoring Antimicrobial Resistance Trends) that evaluates antimicrobial susceptibility to beta-lactam antibiotics, including carbapenems, and also aminoglycosides and quinolones, against a large number of Gram-negative bacilli species collected from IAI and UTI fulfills this function.

In this study, we analysed the antimicrobial susceptibility data from isolates recovered in 2016 and 2017 in Spain from abdominal samples in patients with diagnosis of IAI and urinary samples from patients with UTI included in the SMART database. The ESBL production of these isolates is also presented.

MATERIAL AND METHODS

Microorganisms and participating sites. All isolates studied were obtained from abdominal samples from patients with diagnosis of IAI and from urinary samples from patients with UTI. Details on sampling and criteria for the inclusion of microorganisms were previously described [7]. During the 2 years of the study (2016 and 2017) a total of 10 Spanish hospitals participated (H. Universitario Gregorio Marañón, Madrid, H. Clínico San Carlos, Madrid, H. Universitario Virgen Macarena, Sevilla, H. Universitario Virgen del Rocío, Sevilla, H. Universitario Marqués de Valdecilla, Santander, H. Universitario Lozano Blesa, Zaragoza, H. Universitario Bellvitge, Hospitalet de Llobregat, Barcelona, H. Universitario y Politécnico La Fe, Valencia, and H. Universitario Ramón y Cajal, Madrid).

A total of 1,429 intra-abdominal isolates were collected; the most frequent were recovered from peritoneal fluid (41%), intra-abdominal abscesses (31%) and gall bladder (18%), and to a lesser extent and in decreasing order, from the liver, appendix, pancreas, colon, rectum, and other sources. Most of the isolates were obtained during surgery procedures and others from paracentesis and percutaneous aspiration of intra-abdominal abscesses. Regarding UTI, a total of 937 isolates were obtained, being virtually all urine samples (98%). Isolates from other locations (i.e. blood, abdominal drainages, superficial wounds or perirectal abscesses) were excluded.

The identification of the isolates was performed at each hospital and sent to a central laboratory (International Health Management Associates, SA. Schaumburg, IL, US) to confirm the identification and to establish the susceptibility to different antimicrobials of choice for the treatment of IAIs or UTIs. All results were included in a centralized database. In addition to the source of the sample, patient's age was considered. Following the standard criteria of the *Centers for Disease Control and Prevention* (CDC) the organisms were also rated as isolates obtained within 48 h after hospitalization Monitoring the antimicrobial susceptibility of Gram-negative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)

(community-acquired infection) and isolates obtained after 48 h of hospital stay (nosocomial infection) [8].

Antimicrobial susceptibility and ESBL Antimicrobial susceptibility production. testing results were obtained at a central laboratory (International Health Management Associates) using the standard ISO broth microdilution method [9]. MIC results were interpreted each year according to the most recent EUCAST guidelines (http://www. eucast.org/clinical breakpoints/). Dried MicroScan (Beckman, West Sacramento, CA, US) microdilution panels were used. The antimicrobials analyzed in this study were: piperacillin-tazobactam, cefotaxime, ceftazidime, cefepime, imipenem, ertapenem, amikacin and ciprofloxacin. In addition, susceptibility to amoxicillinclavulanate was measured with a MIC gradient test (Etest[®], bioMérieux, Lyon, France). The quality controls strains used were Escherichia coli ATCC 25922, E. coli ATCC 35218, Klebsiella pneumoniae ATCC 700603 (positive ESBL control) and P. aeruginosa ATCC 27853. E. coli, Klebsiella spp. and Proteus mirabilis isolates were classified as ESBL following CLSI criteria [10].

Statistical analysis. The frequency comparison (incidence between hospital and community isolates) was performed using the chi-squared test (χ^2) taking *P*<0.05 as statistically significant.

RESULTS

During 2016 and 2017, a total of 1,429 isolates from IAI and 937 isolates from UTI recovered in the 10 Spanish hospitals were included (tables 1 and 2). In IAI, the Enterobacterales (1,265) constituted 85.5% of the total isolates. This figure was 876 isolates (93.4%) in UTI. Overall, *E. coli* was the most frequently isolated microorganism (48.3% and 53.7%), followed by *Klebsiella* spp. (11.5% and 21.8%) in IAIs and UTIs, respectively. Figures for *Pseudomonas aeruginosa* were 9.0% and 6.1%, being more frequently

recovered in patients with nosocomial infections. When the origin of the isolates was considered (tables 1 and 2), 43.2% of IAI isolates were considered to be acquired in the community compared to 56.8% that had their origin in the nosocomial setting. In UTI, there was also a lower number of isolates from community (47.8%) than from nosocomial origin (52.2%). In 1.5% of IAI isolates, their origin was not specified in the data collection sheets.

Table 1

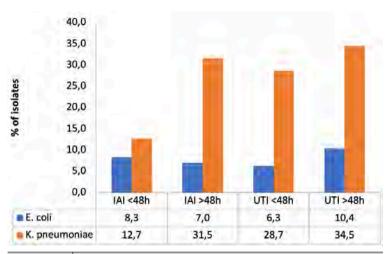
Distribution of the most common Gram-negative organisms collected in intra-abdominal infections in Spain in the SMART Study (2016-2017).

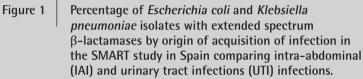
| | - | Communit | y associated | Nosocomia | l associated |
|-----------------------------|--------------|----------|--------------|-----------|--------------|
| Organisms | No. isolates | No. | % | No. | 0/0 |
| Escherichia coli | 690 | 337 | 54.6 | 353 | 43.4 |
| Klebsiella pneumoniae | 165 | 54 | 8.7 | 111 | 13.6 |
| Klebsiella oxytoca | 69 | 39 | 6.3 | 30 | 3.6 |
| Proteus mirabilis | 46 | 17 | 2.7 | 29 | 3.5 |
| Enterobacter cloacae | 75 | 30 | 4.8 | 45 | 5.5 |
| Citrobacter freundii | 31 | 19 | 3.0 | 12 | 1.4 |
| Morganella morganii | 27 | 6 | 0.9 | 21 | 2.5 |
| Serratia marcescens | 25 | 9 | 1.4 | 16 | 1.9 |
| Other Enterobacterales | 137 | 44 | 7.1 | 93 | 11.4 |
| Pseudomonas aeruginosa | 129 | 54 | 8.7 | 75 | 9.2 |
| Other Gram-negative bacilli | 35 | 8 | 1.2 | 27 | 3.3 |
| TOTAL | 1,429 | 617 | 43.2 | 812 | 56.8 |

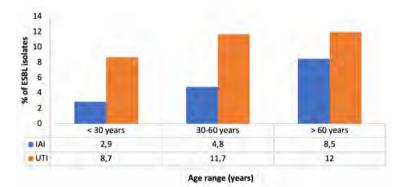
Table 2Distribution of the most common Gram-negative organisms
collected in urinary tract infections in Spain in the SMART
Study (2016-2017).

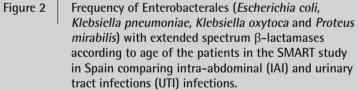
| | | Community | associataed | Nosocomial associated | | | | |
|-----------------------------|--------------|-----------|-------------|-----------------------|------|--|--|--|
| Organisms | No. isolates | No. | % | No. | 0/0 | | | |
| Escherichia coli | 504 | 284 | 63.3 | 220 | 44.9 | | | |
| Klebsiella pneumoniae | 205 | 66 | 14.7 | 139 | 28.4 | | | |
| Klebsiella oxytoca | 18 | 9 | 2.0 | 9 | 1.8 | | | |
| Proteus mirabilis | 61 | 31 | 6.9 | 30 | 6.1 | | | |
| Enterobacter cloacae | 16 | 5 | 1.1 | 11 | 2.2 | | | |
| Citrobacter freundii | 11 | 6 | 1.3 | 5 | 1.0 | | | |
| Morganella morganii | 21 | 8 | 1.7 | 13 | 2.6 | | | |
| Serratia marcescens | 7 | 3 | 0.6 | 4 | 0.8 | | | |
| Other Enterobacterales | 33 | 13 | 2.9 | 20 | 4.0 | | | |
| Pseudomonas aeruginosa | 57 | 22 | 4.9 | 35 | 7.1 | | | |
| Other Gram-negative bacilli | 4 | 1 | 0.2 | 3 | 0.6 | | | |
| TOTAL | 937 | 448 | 47.8 | 489 | 52.2 | | | |

Tables 1 and 2 also show the distribution of the most frequent microorganisms according with their origin. The percentage of *E. coli* of isolates in IAI (table 1) acquired in the community (54.6%) was higher than in those of nosocomial origin (43.4%) (P<0.01). On the contrary, the percentage in *P. aeruginosa* was higher in infections acquired in the hospital (9.2% vs. 8.7%) but without statistical significance (P=0.751). The same situation occurs, even to a greater extent, in the UTIs Monitoring the antimicrobial susceptibility of Gram-negative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)









(Table 2). In *E. coli*, the corresponding numbers are 63.3% in the community and 44.9% in nosocomial infection (p<0.01). In *P. aeruginosa* these percentages were 4.9 and 7.1, respectively (P=0.150).

Overall, the Enterobacterales with AmpC-type inducible chromosomal β -lactamases, such as *Enterobacter cloacae*, *Morganella morganii* and *Serratia marcescens*, were mainly recovered in infections of hospital origin, both in IAI and in UTI (tables 1 and 2).

The presence of ESBL in Enterobacterales such as *E. coli*, *Klebsiella* spp. and *Proteus mirabilis* was specifically studied in IAI and in UTI. In IAI a total of 96 (9.9%) were ESBL producers.

The highest frequency was found in *K. pneumoniae* (25.4%), followed by *E. coli* (7.6%) and *K. oxytoca* (1.4%). In *P. mirabilis* none was found. In UTI the same pattern was followed with higher percentages: *K. pneumoniae* had a higher percentage of ESBL (32.6%) followed by *E. coli* (8.1%), *K. oxytoca* (5.5%) and *P. mirabilis* (1.6%). In all microorganisms with ESBL, the frequency of these enzymes was higher in nosocomially acquired than in community infections (figure 1), with the exception of *E. coli* and *P. mirabilis* in IAI. Likewise, an increase of the ESBL isolates was observed in parallel with the increase of the age of the patients, reaching a frequency higher than 8% in those over 60 years in both types of infection (figure 2).

The susceptibility profile for the antibiotics studied of the most common microorganisms is detailed in table 3. In IAI, the most active antibiotics in Enterobacterales were amikacin (susceptibility rates range: 96.3%-100%), ertapenem (84.2%-100%) and imipenem (70.3%-100%). Ciprofloxacin demonstrated less activity with a percentage of resistance in E. coli greater than 25% and close to 40% in K. pneumoniae. Regarding the associations of penicillins with beta-lactamase inhibitors, piperacillin-tazobactam susceptibility ranged from 66.6% to 100% and amoxicillin-clavulanic acid from 58.3% to 81.5% (table 3). In P. aeruginosa, amikacin, imipenem and ceftazidime, were the most active compounds (96.9%, 76.7% and 72.8% susceptible, respectively).

In UTI the most active antibiotics against Enterobacterales were the same as in IAI, with similar figures for amikacin (97%-100% susceptibility) and higher ones for ertapenem (94.7%-100%) and imipenem (90.4%-100%). Regarding ciprofloxacin, the loss of activity against isolates from urine is noteworthy: only 63% of *E. coli*, 57% of *K. pneumoniae* and 54.1% of *P. mirabilis* were susceptible to this fluoroquinolone.

On the other hand, considering the most frequent microorganisms recovered from IAI (n=1,429), 43.2% were of community origin compared to 56.8% of hospital origin. Of those responsible for the UTIs (n=937), 47.8% were community acquired and 52.2% were of hospital origin. Tables 4 and 5 comparatively analyze the activity of the different antibiotics against community and hospital isolates. Systematically, in the isolates with higher numbers (E. coli and K. pneumoniae), the activity of all antimicrobials was higher in those originated in the community. However, in the remaining species, there were some exceptions. In those from IAI (table 4), the opposite occurs in C. freundii with piperacillin-tazobactam and the third-generation cephalosporins and in *M. morganii* with ciprofloxacin. In UTI (table 5), exceptions occurred with amoxicillin-clavulanate and K. pneumoniae, with the third-generation cephalosporins and P. mirabilis, C.

Table 3

Activity of different antimicrobial used in intra-abdominal (IAI) and urinary tract infections (UTI) against the most common microorganisms collected in Spain in the SMART study (2016-2017).

| | Percentage of susceptible isolates ^a | | | | | | | | | | | | | | | | | |
|------------------------|---|-----------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|------|------|
| Organism | A | /C ^a | P | /T | C. | ГΧ | C/ | AZ | FI | EP | IP | M | E | ГP | A | K | С | IP |
| Type of infection | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI | IAI | UTI |
| Escherichia coli | 81.5 | 77.7 | 90.0 | 90.9 | 90.5 | 90.1 | 89.8 | 89.1 | 92.0 | 90.9 | 99.7 | 99.8 | 99.4 | 99.4 | 97.9 | 99.0 | 72.4 | 63.0 |
| Klebsiella pneumoniae | 58.3 | 94.1 | 66.6 | 69.7 | 72.7 | 64.3 | 67.8 | 64.8 | 72.7 | 65.3 | 95.1 | 97.0 | 84.2 | 86.8 | 98.7 | 97.0 | 62.4 | 57.0 |
| Klebsiella oxytoca | 76.3 | 100.0 | 85.5 | 84.2 | 97.1 | 94.7 | 97.1 | 94.7 | 100.0 | 94.7 | 100.0 | 100.0 | 100.0 | 94.7 | 100.0 | 100.0 | 97.1 | 89.4 |
| Proteus mirabilis | 74.1 | 100.0 | 100.0 | 100.0 | 100,0 | 96.7 | 100.0 | 93.4 | 100.0 | 100.0 | 91.3 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 60.8 | 54.1 |
| Enterobacter cloacae | _b | _b | 78.6 | 58.8 | 73.3 | 52.9 | 72.0 | 58.8 | 84.0 | 82.3 | 96.0 | 94.1 | 85.3 | 94.1 | 97.3 | 100.0 | 90.6 | 70.5 |
| Citrobacter freundii | _b | _b | 70.9 | 90.9 | 70.9 | 72.7 | 54.8 | 63.6 | 87.1 | 90.9 | 93.5 | 90.9 | 96.7 | 90.9 | 100.0 | 100.0 | 93.5 | 81.8 |
| Morganella morganii | _b | _b | 100,0 | 95.2 | 51.8 | 71.4 | 74.0 | 66.6 | 96.3 | 95.2 | 70.3 | 90.4 | 100.0 | 100.0 | 96.3 | 100.0 | 70.3 | 66.6 |
| Serratia marcescens | _b | _b | 88,0 | 100,0 | 72.0 | 100.0 | 96.0 | 100.0 | 92.0 | 100.0 | 92.0 | 100.0 | 92.0 | 100.0 | 100.0 | 100.0 | 96.0 | 85.7 |
| Other Enterobacterales | 36.3 | 60.0 | 79.8 | 74.1 | 82.4 | 84.8 | 72.8 | 78.7 | 98.2 | 93.9 | 99.1 | 100.0 | 96.4 | 100.0 | 98.2 | 100.0 | 91.2 | 87.8 |
| Pseudomonas aeruginosa | _b | _b | 66.6 | 81.8 | _b | _b | 72.8 | 77.5 | 72.0 | 74.1 | 76.7 | 81.0 | _b | _b | 96.9 | 91.3 | 70.5 | 67.2 |

^aEUCAST criteria except A/C in which CLSI criteria were considered. A/C: amoxicillin-clavulanic acid, P/T: piperacillin/tazobactam; CTX: cefotaxime; CAZ: ceftazidime; FEP: cefepime; IPM: imipenem; ETP: ertapenem; AK: amikacin; CIP: ciprofloxacin

^bThis antimicrobial is not considered adequate against the microorganism tested.

freundii and M. morganii, with ciprofloxacin in P. mirabilis and M. morganii and with imipenem in S. marcescens. Moreover, in P. aeruginosa recovered from IAI, all the antibiotics tested were more active when this pathogen was originated in the community, but in the UTI this premise was not observed with piperacillin-tazobactam, ceftazidime and cefepime.

When ESBL producers were considered and compared with non-ESBL producers in IAI (figure 3), the activity of imipenem (99.6% non-ESBL, 100% ESBL) and ertapenem (99.3% non-ESBL, 100% ESBL) remained about at the same level in E. coli whereas amikacin was slightly affected (98.9% non ESBL, 86.7% ESBL). On the contrary, the associations of penicillins with the beta-lactamase inhibitors, as well as third generation cephalosporins and ciprofloxacin importantly decreased their activity. In K. pneumoniae, amikacin susceptibility (100%) non-ESBL, 95.2% ESBL) was little affected compared with that of imipenem (97.5% non-ESBL, 88.1% ESBL) and especially with ertapenem (97.5% non ESBL, 45.2% ESBL) and decreases drastically in the rest of antibiotics as described in E. coli. In UTI. E. coli isolates showed similar results than those described for IAI. In K. pneumoniae, the activity of ertapenem was affected (96.3% non ESBL, 67.1% ESBL), although to a lesser extent than in the IAI isolates.

Finally, when analyzing the activity of carbapenems both in ESBL and in non-ESBL producing *E. coli* and *K. pneumoniae* that were resistant to amoxicillin-clavulate, piperacillin-tazobactam or ciprofloxacin from IAI and UTI (table 6), it was observed that in *E. coli* both the activity of imipenem (data not shown) and that of ertapenem was scarcely modified with susceptibility values higher than 88%. However, in *K. pneumo*- *niae*, ertapenem activity was retained to a lesser extent. In IAI, 28.6% of ESBL producers that were also resistant to amoxicillin-clavulanate were susceptible to ertapenem and in UTI 38.9% of ESBL producers that were resistant to piperacillin-tazobactam were susceptible to ertapenem.

DISCUSSION

Antimicrobial resistance is a global increased problem and poses challenges for the effective treatment of many types of infections, including IAI and UTI. This situation, mainly due to its wide dispersion, is especially alarming in relation to microorganisms that produce ESBL. As a consequence, carbapenems are generally considered the treatment of choice for these infections [11,12], although a decrease in the susceptibility to these compounds have been observed due to the production of carbapenemases or alterations in the porins combined with the production of ESBL or AmpC cephalosporinases [13,14]. Epidemiological surveillance studies analyze trends in resistance but also allow data to progressively adapt treatment guidelines over time, providing valuable information for the selection of initial antibiotic treatment, often empirical. The SMART study (Study for Antimicrobial Resistance Trends), initiated in 2002, is a worldwide program designed to longitudinally monitor the involvement of aerobic and facultative Gram-negative bacilli in IAI, both from community and nosocomial acquisition, as well as their patterns of resistance [15-18]. As of 2009, microorganisms isolated from UTI were also included. The program has been developed in Spain uninterruptedly since 2002 and has had the participation of a significant number of Microbiology Departments of Spanish University Hospitals. Previous Table 4

Table 5

Susceptibility of community-associated (CA) and hospital-associated (HA) microorganisms collected of IAI in Spain in the SMART study (2016-2017).

| Organism | | | | | | | Р | ercenta | ge of sus | sceptible | e isolate | s ^a | | | | | | |
|------------------------|------|---------|-------|-------|---------|-------|-------|---------|-----------|-----------|-----------|----------------|---------|-------|-------|-------|-------|------|
| Organism | A/ | Ca | P | /T | C | TX | C/ | ٩Z | FE | P | IP | M | E | ΓP | A | K | С | IP |
| Type of infection | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA |
| Escherichia coli | 88.7 | 75.9 | 93.4 | 86.6 | 91.3 | 89.8 | 91.0 | 88.6 | 91.6 | 92.3 | 100,0 | 99.3 | 99.4 | 99.4 | 98.5 | 97.4 | 75.3 | 69.9 |
| Klebsiella pneumoniae | 83.8 | 48.7 | 85.4 | 57.6 | 87.2 | 65.7 | 85.4 | 59.4 | 87.2 | 65.7 | 100.0 | 92.7 | 96.3 | 78.3 | 100.0 | 98.2 | 74.5 | 55.8 |
| Klebsiella oxytoca | 84.2 | 68.4 | 92.3 | 76.6 | 97.4 | 96.6 | 97.4 | 96.6 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 97.4 | 96.6 |
| Proteus mirabilis | 62.5 | 78.2 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 94.1 | 89.6 | 100.0 | 100.0 | 100.0 | 100.0 | 64.7 | 58.6 |
| Enterobacter cloacae | _b | _b | 93.3 | 68.8 | 83.3 | 66.6 | 80.0 | 66.6 | 90.0 | 80.0 | 100.0 | 93.3 | 93.3 | 80.0 | 100.0 | 95.5 | 96.6 | 86.6 |
| Citrobacter freundii | _b | _b | 68.4 | 75.0 | 68.4 | 75.0 | 57.8 | 50.0 | 89.4 | 83.3 | 94.7 | 91.6 | 100.0 | 91.6 | 100.0 | 100.0 | 94.7 | 91.6 |
| Morganella morganii | _b | _b | 100.0 | 100.0 | 66.6 | 47.6 | 66.6 | 76.1 | 100.0 | 95.2 | 83.3 | 66.6 | 100.0 | 100.0 | 100.0 | 95.2 | 50.0 | 76.1 |
| Serratia marcescens | _b | _b _ | 88.8 | 87.5 | 66.6 | 75.0 | 100.0 | 93.7 | 100.0 | 87.5 | 100.0 | 87.5 | 100.0 | 87.5 | 100.0 | 100.0 | 100.0 | 93.7 |
| Pseudomonas aeruginosa | _b | _b _ | 79.6 | 57.3 | _b _ | _b | 85.1 | 64.0 | 88.8 | 60.0 | 88.8 | 68.0 | _b _ | _b | 98.1 | 96.0 | 79.6 | 64.0 |

^aEUCAST criteria except A/C in which CLSI criteria were considered. A/C: amoxicillin-clavulanic acid, P/T: piperacillin/tazobactam; CTX: cefotaxime; CAZ: ceftazidime; FEP: cefepime; IPM: imipenem; ETP: ertapenem; AK: amikacin; CIP: ciprofloxacin

Susceptibility of community-associated (CA) and hospital-associated (HA) microorganisms

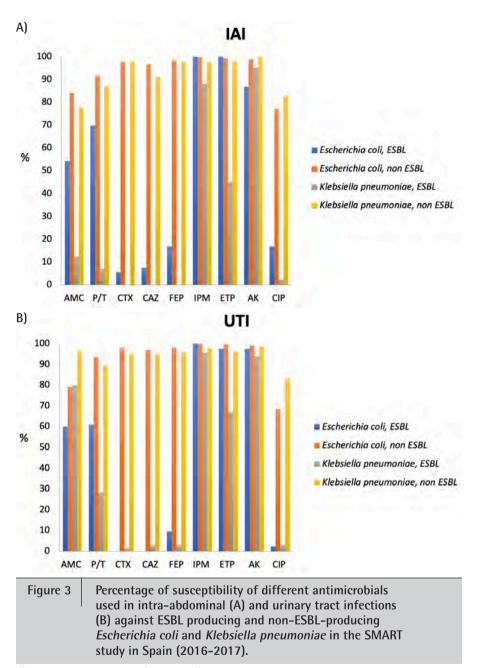
^bThis antimicrobial is not considered adequate against the microorganism tested.

| | ted of | f UTI i | in Spa | in in | the S | SMAR | T stud | dy (20 | 016-2 | 2017). | | | | | | | | |
|------------------------|--------|-----------------|--------|-------|-------|--------|--------|---------|----------|-----------|-----------|----------------|---------|---------|-------|-------|-------|------|
| Organism | | | | | | | Р | ercenta | ge of su | sceptible | e isolate | S ^a | | | | | | |
| | A | 'C ^a | Р | /T | C | TX | C/ | AZ | FI | EP | IP | M | E | ſP | A | K | С | IP |
| Type of infection | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA | CA | HA |
| Escherichia coli | 77.6 | 78.2 | 91.5 | 90.0 | 92.6 | 86.8 | 91.5 | 85.9 | 92.6 | 88.6 | 100.0 | 99.5 | 99.6 | 99.0 | 99.3 | 98.6 | 64.0 | 61.3 |
| Klebsiella pneumoniae | 90.0 | 100.0 | 71.2 | 69.0 | 66.6 | 63.3 | 69.7 | 62.5 | 69.7 | 63.3 | 100.0 | 95.6 | 92.4 | 84.1 | 98.4 | 96.4 | 59.0 | 56.1 |
| Klebsiella oxytoca | 100.0 | 0.0 | 88.8 | 77.7 | 100.0 | 88.8 | 100.0 | 88.8 | 100.0 | 88.8 | 100.0 | 100.0 | 100.0 | 88.8 | 100.0 | 100.0 | 100.0 | 77.7 |
| Proteus mirabilis | 100.0 | 100.0 | 100.0 | 100.0 | 93.5 | 100.0 | 90.3 | 96.6 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 51.6 | 56.6 |
| Enterobacter cloacae | _b | _b | 100.0 | 36.3 | 80.0 | 36.3 | 100.0 | 36.3 | 100.0 | 72.7 | 100.0 | 90.9 | 100.0 | 90.9 | 100.0 | 100.0 | 100.0 | 54.5 |
| Citrobacter freundii | _b | _b _ | 100.0 | 80.0 | 66.6 | 80.0 | 50.0 | 80.0 | 100.0 | 80.0 | 100.0 | 80.0 | 100.0 | 80.0 | 100.0 | 100.0 | 83.3 | 80.0 |
| Morganella morganii | _b | _b | 100.0 | 92.3 | 50.0 | 84.6 | 37.5 | 84.6 | 100.0 | 92.3 | 87.5 | 92.3 | 100.0 | 100.0 | 100.0 | 100.0 | 62.5 | 69.2 |
| Serratia marcescens | _b | b | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 66.6 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 75.0 |
| Pseudomonas aeruginosa | _b | _b | 72.7 | 77.1 | _b | ь — | 77.2 | 80.0 | 72.7 | 77.1 | 81.8 | 80.0 | _b _ | _b _ | 95.4 | 88.5 | 68.1 | 68.5 |

^aEUCAST criteria except A/C in which CLSI criteria were considered. A/C: amoxicillin-clavulanic acid, P/T: piperacillin/tazobactam; CTX: cefotaxime; CAZ: ceftazidime; FEP: cefepime; IPM: imipenem; ETP: ertapenem; AK: amikacin; CIP: ciprofloxacin

^bThis antimicrobial is not considered adequate against the microorganism tested.

articles represent the general picture of antimicrobial susceptibility in our country; the last one (7) updates up to 2015 the evolution of ESBL producing isolates in IAIs in Spain. In the present study, the following two years (2016 and 2017) were analyzed but also including information from UTI pathogens. In general, the results are in line with those obtained in the 2011-2015 period and with others from different regions of the world [13,19-21]. We confirm the relevance of *E. coli* in IAI and UTI and in both cases it is isolated in greater proportion in community-acquired infections than in nosocomial infections, in line with other recent publications [20-22]. *K. pneumoniae* is the second microorganism in order of frequency in both types of infections and unlike the previous period (2011-2015) a greater proportion of isolates was found in nosocomial compared to community infections, both in IAI and in UTI.



A/C: amoxicillin-clavulanic acid, P/T: piperacillin/tazobactam; CTX: cefotaxime; CAZ: ceftazidime; FEP: cefepime; IPM: imipenem; ETP: ertapenem; AK: amikacin; CIP: ciprofloxacin

Given its epidemiological importance, knowledge of the antimicrobial susceptibility of *E. coli* is crucial regarding empirical therapy, as well as for attempts to control the spread of ESBL and, more recently, of carbapenemases. As in other studies [3,13,19,21], imipenem, ertapenem and amikacin were the most active antimicrobials tested against *E. coli* in both IAIs (>97%), and UTIs (>99%) (21) and there is no evidence of loss of activity in 2016 and 2017 compared to 2011-2015 [7]. On the contrary, in *K. pneumoniae* a decrease in the activity of ertapenem in IAI is verified by comparing the two time periods (95.5% in 2011-2015 *versus* 84.2% in 2016-2017) [7]. In UTI, the percentage of susceptibility is 86.8%, slightly lower to that published in studies from other countries [3,21].

In a recent publication, small decreases, although statistically significant, of ertapenem susceptibility in Enterobacterales isolated from IAI and UTI were observed in most regions of the world. Nevertheless, the susceptibility remains above 90% in all regions, except in Asia [22]. In community infections, the activity was >92% in all regions against Enterobacterales [22] despite the existence of communications that alert of the increase in resistance [6]. Another recent study, unrelated to SMART,

Table 6

Activity of ertapenem in ESBL producing *Escherichia coli* and *Klebsiella pneumoniae* isolates resistant to amoxicillin-clavulanate, piperacillin-tazobactam and ciprofloxacin in intra-abdominal (IAI) and urinary tract infections (UTI) of the SMART study (2016–2017) in Spain.

| | | | | | IAI | | | | UTI | | |
|--|----------|---------------|---------------------------|-------------|--------------|-----------|---------------------------|-------------|--------------|-----------|--|
| Mioroorgonisms | ESBL | Antimicrobial | No. | | Ertapenem | | No. | Ertapenem | | | |
| wicroorganisms | ESDL | Antimicrooiai | (% of resistant isolates) | Susceptible | Intermediate | Resistant | (% of resistant isolates) | Susceptible | Intermediate | Resistant | |
| | | | | No. (%) | No. (%) | No. (%) | | No. (%) | No. (%) | No. (%) | |
| Escherichia coli | Negative | | 65 (15.8) | 64 (98.4) | | 1 (1.6) | 26 (20.8) | 26 (100) | | | |
| | | A/C | | | | | | | | | |
| scherichia coli Ne <u>Pe</u> Ne <u>Pi</u> lebsiella pneumoniae Ne <u>Pr</u> | Positive | | 16 (45.7) | 16 (100) | | | 4 (40) | 4 (100) | | | |
| | Negative | | 46 (7.2) | 43 (93.4) | 1 (2.2) | 2 (4.4) | 18 (3.8) | 17 (94.4) | 1 (5.6) | | |
| | | P/T | | | | | | | | | |
| | Positive | | 10 (18.8) | 10 (100) | | | 9 (21.9) | 8 (88.9) | 1 (11.1) | | |
| | Negative | | 126 (19.7) | 123 (97.7) | | 3 (2.3) | 137 (29.4) | 136 (99.3) | 1 (0.7) | | |
| | | CIP | | | | | | | | | |
| | Positive | | 42 (79.2) | 42 (100) | | | 39 (95.1) | 38 (97.4) | 1 (2.6) | | |
| Klebsiella pneumoniae | Negative | | 17 (22) | 14 (82.4) | | 3 (17.6) | 1 (3.4) | 1 (100) | | | |
| | | A/C | | | | | | | | | |
| (lebsiella pneumoniae | Positive | | 28 (87.5) | 8 (28.6) | | 20 (71.4) | 1 (20) | 1 (100) | | | |
| | Negative | | 14 (33.3) | 11 (78.6) | | 3 (21.4) | 11 (7.9) | 6 (54.5) | | 5 (45.5) | |
| | | P/T | | | | | | | | | |
| | Positive | | 38 (30.6) | 16 (42.1) | | 22 (57.9) | 36 (53.7) | 14 (38.9) | 2 (5.5) | 20 (55.6) | |
| | Negative | | 15 (35.7) | 13 (86.7) | | 2 (13.3) | 16 (11.5) | 12 (75) | | 4 (25) | |
| | | CIP | | | | | | | | | |
| | Positive | | 40 (32.2) | 17 (42.5) | 1 (2.5) | 22 (55) | 61 (91) | 40 (65.6) | 2 (3.3) | 19 (31.1) | |

A/C: amoxicillin-clavulanate; P/T: piperacillin/tazobactam; CIP: ciprofloxacin

reported a percentage of susceptibility to ertapenem in the Enterobacterales group of 94.5% (98.7% in *E. coli* and 87.4% in *K. pneumoniae*) [23]. In the study of Lob et al. [22], susceptibility to ertapenem significantly decreased in *K. pneumoniae* between 2012 and 2016 in Africa (6%), Europe (8%) and US/Canada (2.5%). Despite this fact, in 2016 the susceptibility of *K. pneumoniae* to ertapenem remains above 90% in the US/Canada and in the South Pacific area, being greater than 80% in the rest of the world.

In recent years, there is a continuous increase in the rates of Enterobacterales with ESBL around the world, especially in Asia [24]. In a recent review of the global epidemiology, the prevalence of CTX-M ESBLs increased over time in all geographic regions, especially in community isolates [25]. In our study, in IAI the percentage of ESBL in *E. coli* is overall 7.6% (8.3% in community and 7% in nosocomial infection), keeping the total figures in line with the period 2011-2015 [7]. It is noteworthy that the rate is somewhat higher in community-acquired infections, a fact not communicated in most of the published surveillance studies [13,21], although the reports on the spread of ESBL in the community are worrisome [26,27]. In *K. pneumoniae*, the ESBL rate increased

with respect to previous years, from 18.6% in 2015 to 25.4% in 2016-2017, especially at the expense of infections of nosocomial origin (12.7% community and 31.5% nosocomial). In UTI, the figures in ESBL producing E. coli are slightly higher (overall 8.1%; 6.3% community and 10.4% nosocomial) and much higher in K. pneumoniae (overall 32.6%; 28.7% community and 34.5% nosocomial). Our rates of ESBL in K. pneumoniae are difficult to compare with those published in other regions where there are large variations, although it can be summarized that they are lower than those of most countries in Asia, especially China and Thailand [3], and higher than those of the US/Canada [28]. Our study also shows that the highest percentage of ESBL isolates occurs in IAI of hospital origin and in patients of advanced ages. Both circumstances have already been indicated as risk factors for the acquisition of infections due to ESBL producers [29]. In this line, in a recent study in UTI in the US when data are stratified by sex, age and time of hospital stay, there is a higher percentage of ESBL isolations in men, patients \geq 65 years and in nosocomial infections [28].

In IAI, the activity of imipenem, ertapenem and amikacin in ESBL-producing *E. coli* isolates remains practically at the

R. Cantón, et al.

Monitoring the antimicrobial susceptibility of Gram-negative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)

same level in relation to those that do not produce ESBLs. This fact is also confirmed in other publications [13,21,22]. However, one of these articles [13] found some evidence of increased resistance among isolates from the community, in addition to the known decreasing trends in susceptibility to guinolones and third-generation cephalosporins. In ES-BL-producing K. pneumoniae, the activity of imipenem decreased by almost 10% and that of ertapenem by more than 50%. This decrease is not reflected so strongly in any other study and follows the trend already mentioned in the study of the years 2010-2016 in Spain [7]. Ertapenem susceptibility figures below 90% (83.6% in Africa and 85.5% in Europe) have already been published, although data came from a joined analysis including E. coli, K. pneumoniae, K. oxytoca and P. mirabilis ESBL producers from IAI and UTI and not from an individualized analysis [22].

In UTI, the behavior of imipenem, ertapenem and amikacin in *E coli*. and *K. pneumoniae* is similar to that commented for IAI. However, the activity of ertapenem decreased to a lesser extent (somewhat less than 30%) in *K. pneumoniae* being higher than in other publications [3,21]. Regarding the origin of the isolates, *E. coli* slightly decreased their susceptibility to the most active compounds (imipenem, ertapenem and amikacin) when having a hospital origin both in IAI and in UTI, in line with what it is reflected in other studies [3,19,21]. In *K. pneumoniae*, in IAI, the susceptibility decreased to a greater extent, data not sufficiently confirmed in other studies to date [3,19,21].

As in the 2011-2015 study the co-resistance analysis, which is relevant to designing antimicrobial treatment protocols [30], showed that both imipenem (data not shown) and ertapenem have a good activity against ESBL-producing E. coli recovered from IAI and UTI that were also resistant to amoxicillin-clavulanic acid, piperacillin-tazobactam or fluoroquinolones. Nevertheless, the same did not occur in the case of ESBL-producing K. pneumoniae, although ertapenem retained its activity in 28.6%, 42.1% and 42.5% of amoxicillin-clavulanic acid, piperacillin-tazobactam or ciprofloxacin resistant isolates, respectively. These figures were more favorable in UTI, particularly for ciprofloxacin resistant isolates (65.6% of ertapenem susceptibility). The reason for the increased loss of susceptibility to ertapenem in K. pneumoniae was analyzed in a recent study and concluded that it was not only due to production of carbapenemases but to permeability defects [31]. The genes encoding the OmpK35 and OmpK36 porins of the outer membrane were studied and most of the isolates (83.0%) had one or both genes affected. In isolates with higher ertapenem MICs (>4 mg/L), 60.5% of the total isolates, a mutation was found in both porin genes.

Despite the above observations, carbapenems are still considered as empirical therapy of choice in infections suspected to be caused by ESBL producers or AmpC hyperproducers both in IAI and UTI [12,32,33]. Regardless of the spread of ESBL worldwide, a very recent study showed that ertapenem was active against more than 90% of Enterobacterales isolates recovered from IAI and UTI with the ESBL phenotype in Latin America, Middle East, South Pacific, US and Canada. Our study also shows that ertapenem continue to exhibit good activity, despite the emergence of carbapenemases in Spain [34,35], when compared to broad spectrum cephalosporins and associations of penicillins with beta-lactamase inhibitors. This activity is higher in isolates from community origin and may be a viable option to reduce the length of hospitalization of stable patients together with its easy once-a-day dosing, safety and tolerability [36,37]. Continuous surveillance efforts should be performed at local and global levels, since knowledge of the patterns and resistance trends are essential for making decisions about empirical treatment and support infection control efforts.

ACKNOWLEGDEMENTS

The SMART Spain working group is represented by the following investigators who have participated in the study: J. Rodriguez-Lozano and J. Calvo (Hospital Universitario Margués de Valdecilla, Santander); F. Tubau and M.A. Domínguez (Hospital Universitari Bellvitge-IDIBELL. Hospitalet de Llobregat. Barcelona); J.L. Pérez Sáenz, P.A. Fraile-Ribot and A. Oliver (Hospital Universitario Son Espases, Mallorca); F.J. Castillo and C. Seral (Hospital Clínico Universitario Lozano Blesa, Zaragoza); J.L. López-Hontangas (Hospital Universitario y Politécnico La Fe, Valencia); R. Cantón, M. García-Castillo, E. Loza, (Hospital Universitario Ramón y Cajal-IRYCIS, Madrid); E. Cercenado (Hospital Universitario Gregorio Marañón, Madrid); F. González Romo, José Prieto (Hospital Clínico San Carlos, Madrid); J. Aznar and A. Rodríguez-Rey (Hospital Universitario Virgen del Rocío, Sevilla); A. Pascual and A.I. Suárez-Barrenechea (Hospital Universitario Virgen Macarena, Sevilla).

FUNDING

SMART surveillance program is sponsored by MSD. Writing of this manuscript has been performed with an unrestricted grant from MSD-Spain. We thank MSD and IHMA (International Health Management Associates, S.A., Schaumburg, Illinois, U.S.) for providing access to the database of the SMART epidemiological surveillance study.

CONFLICTS OF INTEREST

Rafael Cantón has collaborated in educational meetings sponsored by MSD and Pfizer. He has also had research grants from MSD. F. Javier Castillo has collaborated in educational meetings sponsored by MSD. All other authors declare that they have no conflicts of interest regarding this publication.

REFERENCES

- Prestinaci F, Pezzotti P, Pantosti A. Antimicrobial resistance: a global multifaceted phenomenon. Pathog Glob Health. 2015; 109:309-18. PMID: 26343252.
- 2. Sartelli M, Chichom-Mefire A, Labricciosa FM, Hardcastle T, Abu-

Monitoring the antimicrobial susceptibility of Gram-negative organisms involved in intraabdominal and urinary tract infections recovered during the SMART study (Spain, 2016 and 2017)

Zidan FM, Adesunkanmi AK et al. The management of intra-abdominal infections from a global perspective: 2017 WSES guidelines for management of intra-abdominal infections. World J Emerg Surg. 2017; 12:29. PMID: 28702076.

- Jean SS, Coombs G, Ling T, Balaji V, Rodrigues C, Mikamo H, et al. Epidemiology and antimicrobial susceptibility profiles of pathogens causing urinary tract infections in the Asia-Pacific region: Results from the Study for Monitoring Antimicrobial Resistance Trends (SMART), 2010-2013. Int J Antimicrob Agents. 2016; 47:328-34. PMID: 27005459.
- 4. Pitout JD, Laupland KB. Extended-spectrum beta-lactamase-producing Enterobacteriaceae: an emerging public-health concern. Lancet Infect Dis. 2008; 8:159-66. PMID: 18291338.
- 5. Rawat D, Nair D. Extended-spectrum β-lactamases in Gram negative bacteria. J Glob Infect Dis. 2010; 2:263-74. PMID: 20927289.
- Logan LK, Weinstein RA. The epidemiology of carbapenem-resistant Enterobacteriaceae: the impact and evolution of a global menace. J Infect Dis. 2017. 215 (Suppl 1):S28–36. PMID: 28375512.
- 7. Cantón R, Loza E, Aznar J, Barrón-Adúriz R, Calvo J, Castillo FJ, et al. Antimicrobial susceptibility trends and evolution of isolates with extended spectrum β -lactamases among Gram-negative organisms recovered during the SMART study in Spain (2011-2015). Rev Esp Quimioter. 2018; 31:136-45. PMID: 29532655.
- Horan TC, Andrus M, Dudeck MA. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. Am J Infect Control. 2008; 36:309-32. PMID: 18538699.
- 9. International Organization for Standardization (ISO). 2006 Clinical laboratory testing and *in vitro* diagnostic test systems Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices Part 1: Reference method for testing the *in vitro* activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases. International Standard 20776-1, ISO, Geneva.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Document M100-S27. Wayne, PA: CLSI, 2017.
- Sartelli M, Catena F, Abu-Zidan FM, Ansaloni L, Biffl WL, Boermeester MA, et al. Management of intra-abdominal infections: recommendations by the WSES 2016 consensus conference. World J Emerg Surg. 2017; 12:22. PMID: 28484510.
- Rodríguez-Baño J, Gutiérrez-Gutiérrez B, Machuca I, Pascual A. Treatment of infections caused by extended-spectrum-beta-lactamase-, AmpC-, and carbapenemase-producing Enterobacteriaceae. Clin Microbiol Rev. 2018; 31(2). pii: e00079-17. PMID:29444952.
- Lob SH, Kazmierczak KM, Badal RE, Hackel MA, Bouchillon SK, Biedenbach DJ, Sahm DF. Trends in susceptibility of *Escherichia coli* from intra-abdominal infections to ertapenem and comparators in the United States according to data from the SMART program, 2009 to 2013. Antimicrob Agents Chemother. 2015; 59:3606-10. PMID: 25801558.
- 14. Biedenbach D, Bouchillon S, Hackel M, Hoban D, Kazmierczak K, Hawser S, et al. Dissemination of NDM metallo- β -lactamase genes

among clinical isolates of Enterobacteriaceae collected during the SMART Global Surveillance Study from 2008 to 2012. Antimicrob Agents Chemother 2015; 59:826-30. PMID:27216054

- Morrissey I, Hackel M, Badal R, Bouchillon S, Hawser S, Biedenbach D. A Review of Ten Years of the Study for Monitoring Antimicrobial Resistance Trends (SMART) from 2002 to 2011. Pharmaceuticals (Basel). 2013; 6:1335-46. PMID: 24287460.
- Guembe M, Cercenado E, Alcalá L, Marín M, Insa R, Bouza E. Evolution of antimicrobial susceptibility patterns of aerobic and facultative gram-negative bacilli causing intra-abdominal infections: results from the SMART studies 2003-2007. Rev Esp Quimioter. 2008; 21:166-73. PMID: 18792817.
- 17. Cantón R, Loza E, Aznar J, Calvo J, Cercenado E, Cisterna R, et al. Antimicrobial susceptibility of Gram-negative organisms from intraabdominal infections and evolution of isolates with extended spectrum β -lactamases in the SMART study in Spain (2002-2010). Rev Esp Quimioter. 2011; 24:223-32. PMID: 22173194.
- Babinchak T, Badal R, Hoban D, Hackel M, Hawser S, Lob S et al. Trends in susceptibility of selected gram-negative bacilli isolated from intra-abdominal infections in North America: SMART 2005-2010. Diagn Microbiol Infect Dis. 2013; 76:379-81. PMID: 23541118.
- Hawser S, Hoban DJ, Badal RE, Bouchillon SK, Biedenbach D, Hackel M, et al. Epidemiology and antimicrobial susceptibility of Gram-negative aerobic bacteria causing intra-abdominal infections during 2010-2011. J Chemother. 2015; 27:67-73. PMID: 24548089.
- 20. Bouchillon SK, Badal RE, Hoban DJ, Hawser SP. Antimicrobial susceptibility of inpatient urinary tract isolates of Gram-negative bacilli in the United States: results from the study for monitoring antimicrobial resistance trends (SMART) program: 2009-2011. Clin Ther. 2013; 35:872-7. PMID: 23623624.
- Ponce-de-Leon A, Rodríguez-Noriega E, Morfín-Otero R, Cornejo-Juárez DP, Tinoco JC, Martínez-Gamboa A, et al. Antimicrobial susceptibility of gram-negative bacilli isolated from intra-abdominal and urinary-tract infections in Mexico from 2009 to 2015: Results from the Study for Monitoring Antimicrobial Resistance Trends (SMART). PLoS One. 2018; 13(6):e0198621. PMID: 29927958.
- 22. Lob SH, Hackel MA, Hoban DJ, Young K, Motyl MR, Sahm DF. Activity of ertapenem against Enterobacteriaceae in seven global regions-SMART 2012-2016. Eur J Clin Microbiol Infect Dis. 2018; 37:1481-9. PMID: 29754209.
- Karlowsky JA, Biedenbach DJ, Kazmierczak KM, Stone GG, Sahm DF (2016) Activity of ceftazidime-avibactam against extended-Spectrum- and AmpC beta-lactamase-producing Enterobacteriaceae collected in the INFORM global surveillance study from 2012 to 2014. Antimicrob Agents Chemother 60:2849-57. PMID:26926635.
- Hawser SP, Bouchillon SK, Hoban DJ, Badal RE, Hsueh PR, Paterson DL. Emergence of high levels of extended-spectrum-beta-lactamase-producing gram-negative bacilli in the Asia-Pacific region: data from the Study for Monitoring Antimicrobial Resistance Trends (SMART) program, 2007. Antimicrob Agents Chemother. 2009; 53:3280-4. PMID: 19506060.
- 25. Bevan ER, Jones AM, Hawkey PM. Global epidemiology of CTX-M β -lactamases: temporal and geographical shifts in genotype. J An-

R. Cantón, et al.

timicrob Chemother. 2017; 72:2145-55. PMID: 28541467.

- Pitout JD, Nordmann P, Laupland KB, Poirel L. Emergence of Enterobacteriaceae producing extended-spectrum beta-lactamases (ESBLs) in the community. J Antimicrob Chemother. 2005; 56:52-9. PMID: 15917288.
- 27. Pitout JD. Enterobacteriaceae that produce extended-spectrum β -lactamases and AmpC β -lactamases in the community: the tip of the iceberg? Curr Pharm Des. 2013; 19:257-63. PMID: 22934977.
- Lob SH, Nicolle LE, Hoban DJ, Kazmierczak KM, Badal RE, Sahm DF. Susceptibility patterns and ESBL rates of *Escherichia coli* from urinary tract infections in Canada and the United States, SMART 2010-2014. Diagn Microbiol Infect Dis. 2016; 85:459-65. PMID: 27306116.
- Ofner-Agostini M, Simor A, Mulvey M, McGeer A, Hirji Z, McCracken M, Gravel D, Boyd D, Bryce E. Risk factors for and outcomes associated with clinical isolates of *Escherichia coli* and *Klebsiella* species resistant to extended-spectrum cephalosporins among patients admitted to Canadian hospitals. Can J Infect Dis Med Microbiol. 2009; 20(3):e43-8. PMID: 20808455
- WHO. Global Antimicrobal Resistance Surveillance System (GLASS) (http://www.who.int/glass/en), last access October 12th, 2018.
- Wise MG, Horvath E, Young K, Sahm DF, Kazmierczak KM. Global survey of *Klebsiella pneumoniae* major porins from ertapenem non-susceptible isolates lacking carbapenemases. J Med Microbiol. 2018; 67:289-95. PMID: 29458684.
- Mazuski JE, Tessier JM, May AK, Sawyer RG, Nadler EP, Rosengart MR, Chang PK, O'Neill PJ, Mollen KP, Huston JM, Diaz JJ Jr, Prince JM. The Surgical Infection Society revised guidelines on the management of intra-abdominal infection. Surg Infect (Larchmt). 2017; 18:1-76. PMID: 28085573.
- Bader MS, Loeb M, Brooks AA. An update on the management of urinary tract infections in the era of antimicrobial resistance. Postgrad Med. 2017; 129:242–58. PMID: 27712137.
- Pérez-Vázquez M, Oteo J, García-Cobos S, Aracil B, Harris SR, Ortega A, et al. Phylogeny, resistome and mobile genetic elements of emergent OXA-48 and OXA-245 *Klebsiella pneumoniae* clones circulating in Spain. J Antimicrob Chemother. 2016; 71:887-96. PMID: 26769896.
- Oteo J, Pérez-Vázquez M, Bautista V, Ortega A, Zamarrón P, Saez D, et al. The spread of KPC-producing Enterobacteriaceae in Spain: WGS analysis of the emerging high-risk clones of *Klebsiella pneumoniae* ST11/KPC-2, ST101/KPC-2 and ST512/KPC-3. J Antimicrob Chemother. 2016; 71:3392-9. PMID: 27530752.
- Rattanaumpawan P, Werarak P, Jitmuang A, Kiratisin P, Thamlikitkul V. Efficacy and safety of de-escalation therapy to ertapenem for treatment of infections caused by extended-spectrum-β-lactamase-producing Enterobacteriaceae: an open-label randomized controlled trial. BMC Infect Dis. 2017; 17:183. PMID: 28249572.
- Seo YB, Lee J, Kim YK, Lee SS, Lee JA, Kim HY, et al. Randomized controlled trial of piperacillin-tazobactam, cefepime and ertapenem for the treatment of urinary tract infection caused by extended-spectrum beta-lactamase-producing *Escherichia coli*. BMC Infect Dis. 2017; 17:404. PMID: 28592240.