Archival Report

Biological Psychiatry

Pharmacogenomic Study of Clozapine-Induced Agranulocytosis/Granulocytopenia in a Japanese Population

Takeo Saito, Masashi Ikeda, Taisei Mushiroda, Takeshi Ozeki, Kenji Kondo, Ayu Shimasaki, Kohei Kawase, Shuji Hashimoto, Hidenaga Yamamori, Yuka Yasuda, Michiko Fujimoto, Kazutaka Ohi, Masatoshi Takeda, Yoichiro Kamatani, Shusuke Numata, Tetsuro Ohmori, Shu-ichi Ueno, Manabu Makinodan, Yosuke Nishihata, Masaharu Kubota, Takemi Kimura, Nobuhisa Kanahara, Naoki Hashimoto, Kiyoshi Fujita, Kiyotaka Nemoto, Taku Fukao, Taro Suwa, Tetsuro Noda, Yuji Yada, Manabu Takaki, Naoya Kida, Taku Otsuru, Masaru Murakami, Atsushi Takahashi, Michiaki Kubo, Ryota Hashimoto, and Nakao Iwata

ABSTRACT

BACKGROUND: Clozapine-induced agranulocytosis (CIA)/clozapine-induced granulocytopenia (CIG) (CIAG) is a lifethreatening event for schizophrenic subjects treated with clozapine.

METHODS: To examine the genetic factor for CIAG, a genome-wide pharmacogenomic analysis was conducted using 50 subjects with CIAG and 2905 control subjects.

RESULTS: We identified a significant association in the human leukocyte antigen (HLA) region (rs1800625, $p = 3.46 \times 10^{-9}$, odds ratio [OR] = 3.8); therefore, subsequent HLA typing was performed. We detected a significant association of HLA-B*59:01 with CIAG ($p = 3.81 \times 10^{-8}$, OR = 10.7) and confirmed this association by comparing with an independent clozapine-tolerant control group (n = 380, $p = 2.97 \times 10^{-5}$, OR = 6.3). As we observed that the OR of CIA (OR: $9.3 \sim 15.8$) was approximately double that in CIG (OR: $4.4 \sim 7.4$), we hypothesized that the CIG subjects were a mixed population of those who potentially would develop CIA and those who would not develop CIA (non-CIA). This hypothesis allowed the proportion of the CIG who were non-CIA to be calculated, enabling us to estimate the positive predictive value of the nonrisk allele on non-CIA in CIG subjects. Assuming this model, we estimated that 1) ~50% of CIG subjects would be non-CIA; and 2) ~60% of the CIG subjects without the risk allele would be non-CIA and therefore not expected to develop CIA.

CONCLUSIONS: Our results suggest that HLA-B*59:01 is a risk factor for CIAG in the Japanese population. Furthermore, if our model is true, the results suggest that rechallenging certain CIG subjects with clozapine may not be always contraindicated.

Keywords: Genome-wide association study, Human leukocyte antigen, Pharmacogenomics, Schizophrenia, Side effect, Single nucleotide polymorphism

http://dx.doi.org/10.1016/j.biopsych.2015.12.006

Schizophrenia is a chronic, serious, and disabling mental disorder with a lifetime prevalence of approximately 1% of the world population (1). Antipsychotics are the most useful therapeutic option for schizophrenia; however, approximately one third of patients do not respond adequately to first-line antipsychotics, resulting in treatment-resistant schizophrenia (TRS) (2,3).

Clozapine (CLZ) is a gold standard drug for managing TRS, with a lower incidence of movement abnormality and efficacy superior to that of other antipsychotics in the management of TRS (2-5). Despite its efficacy with TRS, the use of CLZ is significantly restricted by severe side effects such as

CLZ-induced agranulocytosis (CIA)/CLZ-induced granulocytopenia (CIG) (CIAG), which is rare but potentially life threatening. CIAG events are observed in approximately 1% (for CIA) and 3% (for CIG) of CLZ-treatment patients, although the prevalence of CIA differs between populations (Caucasian: $.3\% \sim .9\%$; Asian: greater than 1%; Japanese: 1.1%) (6-11).

In the clinical setting, patients treated with CLZ require frequent blood monitoring for reduced neutrophil count, an indication of CIG and CIA. One of the main reasons for this is that, in general, CIG and CIA are considered to be a continuous phenotype; psychiatrists expect to control the development of CIA by detecting patients at the less serious CIG stage (12).

636 © 2016 Society of Biological Psychiatry. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
Biological Psychiatry October 15, 2016; 80:636–642 www.sobp.org/journal

However, as psychiatrists recognize based on experience, and as Hummer *et al.* (13) reported, some patients with CIG may not go on to experience a serious outcome such as immediate development of CIA (i.e., benign or transient neutropenia) (12,13). For example, a routine blood examination showing an absolute neutrophil count (ANC) of 1400 cells/mm³ may result in discontinuation of CLZ (as this is below the Japanese cutoff level of 1500 cells/mm³ at which discontinuing CLZ treatment is mandatory), but the ANC may then recover quickly to an acceptable value. This may be because an ANC of around 1500 cells/mm³ can fluctuate; for example, it may increase with exercise (14,15) or decrease with viral infection (16).

In such clinical situations, a perfect genetic test to screen for the risk of CIAG before treatment by CLZ is not always mandatory. As described, frequent blood monitoring can be used to detect CIA before CIG develops. Also, CLZ is one of the last treatment options for TRS; the psychiatrist may have little choice other than CLZ treatment. In this regard, a screening test may be useful for reducing the frequency of blood monitoring, thus relieving the burden on quality of life and reducing cost. More importantly, however, psychiatrists need more information on whether or not rechallenging is possible or need a screening test to select CIAG patients who can be treated again with CLZ as a rechallenge.

To date, several pharmacogenetic/pharmacogenomic (PGx) studies have been carried out, mainly in Caucasian populations; however, there is no report to clarify the difference between CIA and CIG. For example, classical candidate gene-based pharmacogenetic studies suggested that a specific sequence variant in human leukocyte antigen (HLA) (HLA-DQB1, 6672G > C) was associated with CIA, but the significance was modest ($p \sim .001$) (17,18). A recent PGx study using a genome-wide association approach and Exome sequencing analysis reported a large impact on CIAG genetics: it showed that CIAG was associated with two independent amino acid changes in HLA-B (158T: $p = 6.4 \times 10^{-10}$ and HLA-DQB1 (126Q: $p = 4.7 \times 10^{-14}$) (19). It is of note that the p values surpassed the genome-wide significance threshold (5 \times 10⁻⁸), although these associations were detected by imputation of nonsynonymous single nucleotide polymorphisms (SNPs) of HLA genes and not by the genotyping of classical HLA alleles.

In this study, we aimed to explore the genetic risk for CIAG in the Japanese population by a genome-wide SNP survey. Although the sample size of CIAG subjects was small, we detected significant SNPs in the HLA region compared with healthy control subjects drawn from the general population. To detect the responsible risk allele, we conducted classical HLA typing for the CIAG subjects and for CLZ-tolerant control subjects, which showed that a specific allele in HLA-B was significantly associated with CIAG. We investigated the clinical utility of this risk allele and whether it could increase posterior probability in the screening test for CIAG or could provide information regarding the possibility of CLZ rechallenging for CIG subjects.

METHODS AND MATERIALS

Ethical Statement

After providing a complete description of the study to the subjects, written informed consent was obtained. The ethics

committees of each university, institute, and hospital participating in this project approved this study.

Participants

CIAG Subjects. Fifty-two patients with CIAG were included (28 male and 24 female patients, age 44.0 \pm 15.6 years old). Of these, 23 were diagnosed with CIA, defined as a decrease in ANC to less than 500 cells/mm³, and 29 were diagnosed with CIG, defined as an ANC between 500 cells/mm³ and 1500 cells/mm³ or for three of these subjects as leukopenia (white blood cell count less than 3000 cells/mm³). These criteria were followed by the Clozaril Patient Monitoring Service in Japan and many other countries, which can reflect a real-world clinical setting. All subjects experienced CIAG within 180 days of first being prescribed CLZ, in accordance with the definition of CIAG in a previous paper (20). All of the subjects were diagnosed as TRS, and all identified themselves as Japanese.

Healthy Comparison Subjects. A total of 2948 subjects (1120 male and 1828 female subjects; age 37.0 ± 15.2 years old) were genotyped as healthy control subjects; they had no personal history of mental disorders and were Japanese descent by self-report. Among them, 1108 subjects were nurses at Fujita Health University Hospital (21), while the others were recruited from the general population.

CLZ-Tolerant Control Subjects. For the classical HLA association analysis, we recruited 380 CLZ-tolerant individuals (210 male and 170 female individuals, age 42.7 \pm 11.8 years old) who had been treated with CLZ for more than 180 days without suffering from CIAG. Again, all of the subjects were diagnosed as having TRS and self-reported as Japanese.

Genotyping and Quality Control

In the PGx analysis, we genotyped the CIAG cases and the healthy comparison subjects using the Illumina HumanOmniExpressExome v1.0 (for 825 healthy comparison subjects) or v1.2 (for the rest of the subjects) (Illumina, San Diego, California).

The following quality control (QC) procedure was applied: 1) we extracted overlapping SNPs between the v1.0 and v1.2 chips; 2) we ensured gender consistency by investigating the SNPs on chromosome X (one CIAG subject); 3) we removed the subjects with a low call rate < .99 (null subjects); and 4) we removed subjects with two degrees or less of relatedness using an identity-by-state analysis (22 healthy comparison subjects). After this first QC filtering, 51 CIAG and 2926 healthy comparison subjects were still eligible, with 643,234 SNPs with a minor allele frequency of more than 1%. To investigate the population structures as following QC, we carried out principal component analysis (22). Using HapMap datasets for four populations (Japanese in Tokyo; Han Chinese in Beijing; Yoruba in Ibadan, Nigeria; and Utah residents with ancestry from northern and western Europe), we confirmed that samples in the East Asian population were grouped in one cluster (Supplemental Figure S1). Then, using only the East Asian HapMap populations (Japanese in Tokyo and Han Chinese in Beijing), we classified the population clusters, including Chinese, Japanese-Mainland, and Japanese-Ryukyu clusters (Supplemental Figure S1) (23). We confirmed that the 11 CIAG subjects recruited from Ryukyu Hospital were classified in the Japanese-Ryukyu clusters. The subjects in these two Japanese clusters were selected by visual inspection, with 22 subjects (one CIAG and 21 healthy comparison subjects) being excluded as outliers. To calculate more precise eigenvectors for association analysis, we carried out further principal component analysis for the selected CIAG and healthy comparison subjects. The SNPs with a call rate <.99 and with large deviations from the Hardy Weinberg equilibrium (p value of the exact test $<1 \times 10^{-5}$) in the control group were also removed. Finally, we filtered SNPs by minor allele frequency of more than 1% in control subjects, leaving 522,694 SNPs, 50 CIAG subjects (22 CIA and 28 CIG), and 2905 healthy comparison subjects.

In the HLA analysis, genotyping of the HLA alleles of HLA-A, HLA-C, HLA-B, and HLA-DRB1 genes was performed using WAKFlow HLA typing kit (Wakunaga, Hiroshima, Japan) and a Luminex Multi-Analyte Profiling system (xMAP; Luminex, Austin, Texas).

Statistical Analysis in the PGx

The association of the direct genotyped SNPs was assessed by a logistic regression analysis with adjustment of sex and the first two eigenvectors as covariates using PLINK ver1.9 (https://www.cog-genomics.org/plink2/) (24). Regional plots were generated using LocusZoom (25). The association of HLA alleles (dominant model) between CIAG and healthy comparison/CLZ-tolerant control subjects was evaluated using a 2 × 2 Fisher's exact test in R (www.r-project.org). The statistical level of significance was set at $p < 5 \times 10^{-8}$ for the genome-wide SNP analysis and the HLA analysis.

Clinical Performance of HLA-B*59:01 as a Genetic Test

To assess the clinical utility of the detected risk in HLA-B (HLA-B*59:01), we calculated sensitivity and specificity, a standard statistical measure of the performance of the test. We also estimated positive predictive value (PPV) and negative predictive value (NPV) assuming 1%, 3%, and 4% prevalence for CIA, CIG, and CIAG, respectively, as prior probabilities.

Based on the PGx results, described later in Results, we hypothesized that CIG subjects were a mixed population of those with the potential to go on to develop CIA (CIA_{estimated}) and those whose neutrophil count had simply decreased below the recommended cutoff line and who would not develop CIA (non-CIA_{estimated}) (12,13). We therefore focused on the CIG subjects to estimate the proportion who were non-CIA_{estimated}.

Assume that ϕ is the proportion of the CIG who are non-CIA_{estimated} (i.e., ϕ = non-CIA_{estimated}/CIG) and that P(CIG_{observed}), P(CIA_{observed}), and P(Control_{observed}) are the proportions of patients observed to have the risk allele in the CIG, CIA, and control (i.e., healthy comparison subjects and CLZ-tolerant control subjects) groups, respectively. In line with this assumption, we hypothesized that the proportion between the probability of CIAestimated with the risk allele in the CIG subjects and that without the risk allele in the CIG subjects corresponds with the proportion between the CIA subjects observed to have the risk allele and those without the risk allele. This would also be the case with the non-CIA_{estimated} subjects, which is compatible with the healthy comparison subjects and CLZ-tolerant control subjects (Supplemental Figure S2). To calculate ϕ , we used the following formulas:

$$\begin{aligned} \mathsf{P}(\mathsf{CIG}_{\mathsf{observed}}) &= \phi \times \mathsf{P}(\mathsf{non-CIA}_{\mathsf{estimated}}) + (1 - \phi) \\ \times \mathsf{P}(\mathsf{CIA}_{\mathsf{estimated}}) &\approx \phi \times \mathsf{P}(\mathsf{Control}_{\mathsf{observed}}) + (1 - \phi) \\ &\times \mathsf{P}(\mathsf{CIA}_{\mathsf{observed}}) \end{aligned}$$

After solving for φ , we could estimate the number of subjects with the risk allele (under the dominant model) for non-CIA_{estimated} [=number of CIG_{observed} $\times \varphi \times P(non-CIA_{observed})$] and CIA_{estimated} [=number of CIG_{observed} $\times (1-\varphi) \times P(CIA_{observed})$] in the CIG group. Then, we examined the statistical measure of the performance of the test to assess the utility of the nonrisk allele on the non-CIA_{estimated}, who are the safest subjects for rechallenging: sensitivity, specificity are assumed in the theoretical model (we could not divide subjects into non-CIA and CIA in the CIG group in the real world), but PPV (the proportion of non-CIA in the CIG without the risk allele) is the most informative value because we can speculate the validity of CLZ rechallenging to the CIG.

RESULTS

Genome-wide PGx Analysis

A quantile-quantile plot was generated using the logistic regression test (Supplemental Figure S3) and the genomic inflation factor (λ_{GC}) based on median chi-square statistics was 1, which indicates that population stratification was controlled by the eigenvectors.

Figure 1 shows a Manhattan plot of the data. The top SNPs of the association are listed in Table 1. SNPs in the major histocompatibility complex (MHC) were redundant due to linkage disequilibrium, so we listed only SNPs with genome-wide significance. For regions other than the MHC, SNPs with $p < 1 \times 10^{-5}$ were listed; all of the top SNPs with $p < 1 \times 10^{-5}$ are listed in Supplemental Table S1.

The SNPs in the MHC region showed genome-wide significance (i.e., $p < 5 \times 10^{-8}$), and the top hit in this region was rs1800625 located around 32 megabase in chromosome

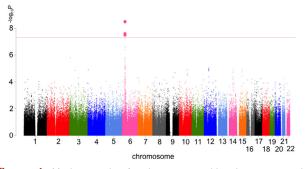


Figure 1. Manhattan plot for the genome-wide pharmacogenomic results. Red line indicates genome-wide significance (5 \times 10⁻⁸). The y axis is $-\log_{10}(p \text{ values})$ of the single nucleotide polymorphisms and the x axis is chromosomal position (hg19).

Table 1. Results for the Association between CIAG and Control Subjects										
CHR	SNP	BP	A1	A2	F_A	F_U	OR (95% Cls)	p	Closest Gene	
6	rs1800625	32152442	G	А	.30	.084	3.78 (2.43–5.87)	3.46E-09	PBX2 (MHC)	
6	rs2269418	32180120	G	Т	.30	.094	3.60 (2.29–5.66)	2.69E-08	NOTCH4 (MHC)	
6	rs3213468	32183158	А	G	.30	.094	3.58 (2.28–5.62)	2.89E-08	NOTCH4 (MHC)	
6	rs2071282	32188943	Т	С	.30	.095	3.56 (2.27–5.60)	3.45E-08	NOTCH4 (MHC)	
3	rs3749448	7188116	А	G	.37	.20	2.96 (1.90–4.61)	1.60E-06	GRM7	
12	rs646782	50482211	G	А	.14	.051	3.94 (2.15–7.23)	9.59E-06	SMARCD1	

Table 1. Results for the Association Between CIAG and Control Subjects

Due to tight LD in MHC, only SNPs with *p* less than 5E-8 for MHC region were listed. For other region, SNPs with *p* less than 1E-5 were listed. A1, nonreference allele; A2, reference allele; BP, base position based on hg19; CHR, chromosome; CI, confidence interval; CIAG, clozapine-induced agranulocytosis/granulocytopenia; F_A, frequency of A1 in cases; F_U, frequency of A1 in healthy comparison subjects; LD, linkage disequilibrium; MHC, major histocompatibility complex; OR, odds ratio for A1; SNP, single nucleotide polymorphism.

6 ($p = 3.5 \times 10^{-9}$, odds ratio [OR] = 3.8; Supplemental Figure S4). The other regions of top association were seen at rs3749448 in the glutamate receptor, metabotropic 7 (*GRM7*, $p = 1.6 \times 10^{-6}$, OR = 3.0; Supplemental Figure S5), and rs646782 in SWI/SNF related, matrix associated, action dependent regulator of chromatin, subfamily D, member 1 (*SMARCD1*) ($p = 9.1 \times 10^{-6}$, OR = 3.9; Supplemental Figure S5).

To detect the responsible HLA allele, a classical HLA typing (HLA-A, HLA-C, HLA-B, and HLA-DRB1) was conducted to assess the association of HLA alleles between CIAG (n = 50) and a subset of healthy comparison subjects (n = 1891) of which most were from the general population (n = 1739) and the rest were nurses (n = 152). We detected that a specific allele of HLA-B*59:01 showed significant association with CIAG in the Japanese population (dominant model: p = 3.8 \times 10⁻⁸, OR = 10.7; Table 2; all of the association results are listed in Supplemental Table S2). Interestingly, the narrow phenotype subjects with CIA (n = 22) showed a greater effect size (OR = 15.8), indicating that the core phenotype might extract a purer risk for CIAG despite the lower statistical power due to the small sample size ($p = 2.8 \times 10^{-6}$; Table 2). We also found a trend for significant association of HLA-DRB1*04:05 ($p = 5.2 \times 10^{-5}$, OR = 3.4; Supplemental Table S2); however, this allele is in linkage disequilibrium with HLA-B*59:01 (D' \sim .95 and $r^2 \sim$.1). The conditional analysis with adjustment of HLA-B*59:01 revealed only a nominal significant association between HLA-DRB1*04:05 and CIAG (p = .034), and so this association was correlated with that of HLA-B*59:01.

To validate the significant association of HLA-B*59:01 between the CIAG and healthy comparison subjects and to find novel risk alleles, we genotyped a total of 380 CLZ-tolerant control subjects as an independent comparison dataset (HLA-A, HLA-C, HLA-B, and HLA-DRB1). Again, a significant association was detected between the CIAG group and the CLZ-tolerant control subjects in HLA-B*59:01 ($p = 3.0 \times 10^{-5}$, OR = 6.3; Table 2; all of the association results are listed in Supplemental Table S3). The narrow phenotype, CIA, again showed a greater effect size (OR = 9.3) than that for CIG (OR = 4.4).

To assess the clinical utility for the risk allele of HLA-B*59:01, standard statistical measures of test performance were calculated (Table 3). We obtained low sensitivity and PPV but high specificity and NPV; however, the acceptable line for clinical use was not reached.

Simulated Performance of HLA-B*59:01 for CLZ Rechallenging

The PGx results showed a large difference in effect size between CIA and CIG (CIA > CIG). We therefore assumed a simple model where CIG consists of a mixture of subjects on the way to developing CIA (CIA_{estimated}) and subjects whose ANC had simply decreased to less than 1500 cell/mm³ by chance or due to an unknown risk factor and who would not develop CIA (non-CIA_{estimated}) (12,13). Using this model, we examined as an explorative analysis its clinical utility, i.e., whether this information can be a reasonable predictor for identifying among CIG subjects those that are non-CIA_{estimated} and those that are CIA_{estimated}. The values for φ (non-CIA_{estimated}/CIG) were

Table 2. Association of HLA-B*59:01 With CIAG

	Allele Count for Case ^a		Allele Count for Control ^a				
Comparison	2 Allele + 1 Allele	0 Allele	2 Allele + 1 Allele	0 Allele	р	OR	95% Cls
CIAG ($n = 50$) vs. Healthy Comparison Subjects ($n = 1891$)	12	38	54	1837	3.81E-08	10.7	4.8-22.4
CIA ($n = 22$) vs. Healthy Comparison Subjects ($n = 1891$)	7	15			2.78E-06	15.8	5.2-43.2
CIG ($n = 28$) vs. Healthy Comparison Subjects ($n = 1891$)	5	23			1.32E-03	7.4	2.1–20.9
CIAG ($n = 50$) vs. Tolerant Control Subjects ($n = 380$)	12	38	18	362	2.97E-05	6.3	2.6–15.1
CIA ($n = 22$) vs. Tolerant Control Subjects ($n = 380$)	7	15			1.39E-04	9.3	2.8–28.1
CIG ($n = 28$) vs. Tolerant Control Subjects ($n = 380$)	5	23			.015	4.4	1.2–13.7

CI, confidence interval; CIA, clozapine-induced agranulocytosis; CIAG, clozapine-induced agranulocytosis/granulocytopenia; CIG, clozapine-induced granulocytopenia; OR, odds ratio.

^a2 allele, samples with 2 alleles of HLA-B*59:01; 1 allele, samples with 1 allele of HLA-B*59:01; 0 allele, samples with null allele of HLA-B*59:01.

Comparison	Sensitivity	Specificity	PPV ^a	NPV ^a	PAR ^a	PARP ^a (%)			
CIAG ($n = 50$) vs. Healthy Comparison Subjects ($n = 1891$)	.240	.971	.259	.968	.00843	21.1			
CIA ($n = 22$) vs. Healthy Comparison Subjects ($n = 1891$)	.318	.971	.101	.993	.00296	29.6			
CIG ($n = 28$) vs. Healthy Comparison Subjects ($n = 1891$)	.179	.971	.162	.975	.00451	15.0			
CIAG ($n = 50$) vs. Tolerant Control Subjects ($n = 380$)	.240	.953	.174	.968	.00783	19.6			
CIA ($n = 22$) vs. Tolerant Control Subjects ($n = 380$)	.318	.953	.064	.993	.00282	28.2			
CIG ($n = 28$) vs. Tolerant Control Subjects ($n = 380$)	.179	.953	.104	.974	.00402	13.4			

Table 3. Diagnostic Performance of HLA-B*59:01 on CIAG

CIA, clozapine-induced agranulocytosis; CIAG, clozapine-induced agranulocytosis/granulocytopenia; CIG, clozapine-induced granulocytopenia; NPV, negative predictive value; PAR, population attributable risk; PARP, population attributable risk percent; PPV, positive predictive value. ^aAssuming 1%, 3%, and 4% prevalence for CIA, CIG, and CIAG, respectively.

around ~50% (48.2% based on the calculation for healthy comparison subjects and 51.6% for the CLZ-tolerant control subjects), indicating that half of the subjects in the CIG group had minimal risk of developing CIA. The estimated allelic distributions of HLA-B*59:01 are listed in Table 4, calculated based on P(CIA_{observed}) and P(non-CIA_{observed}). In this 2 × 2 table, it is of note that 1) 82% of the CIG subjects did not have the risk allele; and 2) the PPV for detecting non-CIA_{estimated} subjects in the CIG group without the risk allele was moderate (~60%).

DISCUSSION

Through the genome-wide SNP and classical HLA analyses, we identified a risk allele for CIAG at HLA-B in people of Japanese ancestry. It is of note that HLA-B was reported as a risk factor for CIAG in Caucasian subjects in a recent PGx study (19); our results therefore support this association. However, SNP 158T in HLA-B, the risk allele detected by Goldstein *et al.* (19), is not compatible with HLA-B*59:01 detected in this study.

HLA-B*59:01, which has not been reported as a risk for side effects induced by CLZ or other drugs, is not common in the Japanese population (the allele frequency is $\sim 1.5\%$ in our control subjects and 2.0% in the Japanese population based on the HLA laboratory dataset URL: http://www.hla.or.jp/haplo/haplonavi.php?type=haplo&lang=en). Interestingly, based on the publicly available database (http://www.allelefre quencies.net/), the allele is virtually nonexistent in other populations such as Caucasian and African, the exception being East Asian populations (e.g., in the Chinese and Korean populations the allele frequency is .1 $\sim 2\%$). This worldwide allelic distribution might be the cause of the different

prevalences of CIAG (for example, the East Asian population demonstrates a higher prevalence than the Caucasian), if this allele poses a genuine risk not only for those of Japanese ancestry but also for other populations.

Although risk allele HLA-B*59:01 showed a significant association with CIAG, its clinical utility as a screening test is limited with regard to predicting future CIAG in patients being treated by CLZ for the first time. Sensitivity and PPV were low, indicating this allele is not suitable for a screening test to target CIAG, especially for CIG. Specificity and NPV were high, indicating that the test in general could be useful for making a definitive diagnosis. However, the information provided by this risk allele is not necessary for the diagnosis of CIAG, as definitive diagnosis is made by blood count. Taken together, the predictive power estimated by our results indicates this would not be useful for the clinical diagnosis of CIAG (and/or CIA) when first treating with CLZ.

However, our results based on explorative estimates may provide evidence that CLZ rechallenging is a possible option for patients when they respond only to CLZ. Using the estimated proportion of non-CIA_{estimated}/CIA_{estimated}, we observed the high sensitivity (to assess the non-CIA_{estimated} patients with the nonrisk allele) that can provide a key to personalized medicine for CIG patients. However, in the real world, we do not, of course, have a predictor to divide CIG patients into non-CIA and CIA but only have allele information of the risk with HLA-B*59:01. Therefore, the PPV is the most informative value for CIG patients: if a patient has no risk allele, indicating a lower risk for developing subsequent CIA (as with approximately 80% of the CIG subjects [n = 23/28], they would be a potent candidate for rechallenging with CLZ treatment. Among these, it is of note that 60% of the CIG subjects without HLA-B*59:01 have the potential to avoid

Table 4. Diagnostic Performance of Nonrisk Allele (Alleles Except for HLA-B*59:01) on non-CIA Among CIG Subjects

Calculated Based on	Samples	non-CIA _{estimated} ^a (%)	CIA _{estimated} ^a (%)	Total (%)	Sensitivity	Specificity	PPV ^b	NPV ^b
CIA vs. Healthy Comparison Subjects	Risk allele (-)	46.8	35.3	82.1	.971	.318	.570	.923
(φ = 48.2%)	Risk allele (+)	1.4	16.5	17.9				
	Total	48.2	51.8	100				
CIA vs. Tolerant Control Subjects	Risk allele (-)	49.1	33.0	82.1	.953	.318	.598	.863
(φ = 51.6%)	Risk allele (+)	2.4	15.4	17.8				
	Total	51.6	48.4	100				

CIA, clozapine-induced agranulocytosis; CIG, clozapine-induced granulocytopenia; NPV, negative predictive value; PPV, positive predictive value.

^aProportion of the non-CIA_{estimated} and CIA_{estimated} samples were calculated by ϕ .

^bAssuming 48.2% and 51.6% incidence based on healthy comparison and tolerant controls subjects, respectively, as prior probability.

future CIA. Conversely, this means that 40% of the CIG subjects without the risk allele may develop CIA. It is a matter of clinical judgment whether or not this has a high value, but given that there is no treatment strategy other than CLZ to control the patient's symptoms (30% to 40 % of patients with CLZ treatment showed a very good response) (26,27), and that mortality from CIG is not high under the current monitoring system (2% to 4% even among CIA subjects) (6), rechallenging will be one of the options to control the severe symptoms for TRS. We stress, of course, that frequent monitoring of the white blood cells and ANC is essential regardless.

Although we replicated the association in the HLA-B gene detected in the report by Goldstein et al. (19), several inclusion criteria were not identical. For example, Goldstein et al. (19) used a more stringent definition of CIG, the upper limit of the ANC being less than 1000 cells/mm³. Whereas the definition of the ANC we used was less than 1500 cells/mm³, which is in accord with the Clozaril Patient Monitoring Service, it thus could reflect the real-world clinical setting. To validate the findings by Goldstein et al. (19), we conducted an explorative analysis using the stringent ANC cutoff level (less than 1000 cells/mm³); we could replicate the significant association between HLA-B*59:01 and the CIAG subjects (CIA + stringent CIG) with the ANC less than 1000 cells/mm³ (Supplemental Table S4). Also, the effect size showed similar magnitude between CIA (OR: 15.8/9.3 for comparison with healthy control subjects/tolerant control subjects, respectively) and the stringent CIG (OR: 13.5/7.9 for comparison with healthy control subjects/tolerant control subjects, respectively; further analysis by stratifying the subjects based on ANC ranges is described in the Supplement text and Supplemental Table S4), supporting HLA-B*59:01 as a genuine risk factor for CIA.

Our study has several limitations. First, the definition of the tolerant control was not perfect: epidemiological surveys suggested that CIAG was observed at 80% in the first 18 weeks and 90% by 1 year (8,10). Therefore, we would have to monitor our subjects again to validate them as genuine tolerant subjects; theoretically, however, approximately three subjects out of 380 subjects ($380 \times .04 \times .2 \approx 3$) would be the misclassified subjects under the assumptions of the prevalence of CIAG as 4% and of the 20% (in a worst-case scenario) of the subjects undetected at the earlier stage, expecting low possibility to have a large impact on our results. Lastly, our sample size was limited; therefore, further replication with greater power is essential for clinical application.

In summary, we identified the HLA risk for CIAG in the Japanese population. The effect size is relatively large, but due to the modest predictive power and population frequency of the allele, it is difficult to use this result as a predictive factor for patients exposed to CLZ for the first time. However, our results indicate that rechallenging is a possible option for CIG patients who have no risk allele when they respond only to CLZ. Future identification of risk factors other than HLA-B*59:01 could provide a more accurate indicator to predict CIAG, including environment factors, such as lower ANC in the stable condition, drug interaction, or gene-environment interplay.

This study was supported by grants of part of the BioBank Japan Project from the Ministry of Education, Culture, Sports, and Technology (MEXT) of Japan; the Health and Labour Sciences Research Grants for Comprehensive Research on Persons with Disabilities from Japan Agency for Medical Research and Development (AMD); Integrated Research Program for Brain Sciences from the MEXT of Japan and Japan Agency for Medical Research and Development (AMED); Grant-in-Aid for Scientific Research on Innovative Areas (Comprehensive Brain Science Network, Glia assembly) from the MEXT of Japan; SENSHIN Medical Research (B) from the MEXT of Japan.

We thank the following doctors for their participation in this study: Drs. Kentaro Mizuno, M.D., and Toshiaki Shiratsuchi, M.D. (Wakakusa Hospital); Dr. Keishi Ueda, M.D. (Kumamotoseimei Hospital); Drs. Yasuhide Fukuji, M. D., and Minori Nakai, M.D. (National Hospital Organization Ryukyu Hospital); attending psychiatrists of Tosa Hospital; attending psychiatrists of Baba Hospital; attending psychiatrists of Matsuyama Kinen Hospital; attending psychiatrists of Kochi Medical School Hospital; Drs. Yutaka Sawa, M.D., and Haruo Watanabe, M.D. (Sawa Hospital); Drs. Sadakazu Tanaka, M.D., and Yoshiki Kishi, M.D. (Okayama Psychiatric Medical Center); attending psychiatrists of Manyo Clinic; Dr. Toshiya Murai, M.D., Ph.D. (Department of Psychiatry, Graduate School of Medicine, Kyoto University); Drs. Akiko Mamoto, M.D., Ph.D., and Norio Taniguchi, M.D., Ph.D. (Asakayama General Hospital); Dr. Kyuryoku Hirakawa, M.D. (Matsubara Hospital); Dr. Hideaki Tabuse, M.D. (Holy Cross Hospital); Dr. Yuuhei Amano, M.D. (Kakamigahara Hospital); Dr. Katsushi Kon, M.D. (Gakuji-kai Kimura Hospital); Drs. Yasuo Fujii, Ph.D., M.D., and Ryoji Miyata, M.D. (Yamanashi Prefectural Kita Hospital); attending psychiatrists of Okinawa Prefectural Seiwa Hospital; Dr. Nobutoshi Kawai, M.D., Ph.D. (Department of Psychiatry, Mitsukaido Kosei Hospital); Ms. Masami Miyata (Center for Research Promotion and Support, Fujita Health University); Dr. Ichiro Kusumi, M.D., Ph.D. (Department of Psychiatry, Hokkaido University Graduate School of Medicine); Dr. Hitoshi Kiyoi, M.D., Ph.D. (Department of Hematology and Oncology, Nagoya University Graduate School of Medicine); and Dr. Norio Ozaki, M.D., Ph.D. (Department of Psychiatry, Nagoya University Graduate School of Medicine). No compensation was provided.

Dr. N. Iwata has received research support or speakers' honoraria from or has served as a consultant to Janssen Pharmaceutical K.K., GlaxoSmithKline, Eli Lilly, Otsuka, Shionogi, Dainippon Sumitomo, Tanabe Mitsubishi, and Daiichi-Sankyo. Dr. Hashimoto has received research support or speakers' fees from Pfizer Inc; Yoshitomiyakuhin Corporation; Dainippon Sumitomo Pharma Co., Ltd.; Novartis Pharma K.K.; Glaxo-SmithKline K.K.; Hisamitsu Pharmaceutical Co., Inc.; Janssen Pharmaceutical K.K.; Nippon Zoki Pharma Inc.; Eli Lilly Japan K.K.; and Abbot Japan Co., Ltd. Dr. Ikeda has received research support from Janssen Pharmaceutical K.K. The remaining authors report no biomedical financial interests or potential conflicts of interest.

ARTICLE INFORMATION

From the Department of Psychiatry (TSa, MI, KKo, AS, KKa, NI), Fujita Health University School of Medicine, Toyoake, Aichi; Research Group for Pharmacogenomics (TM, TOz), RIKEN Center for Integrative Medical Sciences, Yokohama; Department of Hygiene (SH), Fujita Health University School of Medicine, Toyoake, Aichi; Department of Psychiatry (HY, YYas, MF, KO, MTake, RH), Osaka University Graduate School of Medicine, Suita, Osaka: Molecular Besearch Center for Children's Mental Development (MTake, RH), United Graduate School of Child Development, Osaka University, Suita, Osaka; Laboratory for Statistical Analysis (YK, AT), RIKEN Center for Integrative Medical Sciences, Yokohama; Department of Psychiatry (SN, TOh), Institute of Biomedical Sciences, Tokushima University Graduate School, Tokushima; Department of Neuropsychiatry (SU), Ehime University Graduate School of Medicine, Shitsukawa; Department of Psychiatry (MMa, YN), Faculty of Medicine, Nara Medical University, Kashihara, Nara; Kusakabe Memorial Hospital (MKubot), Yamanasi, Yamanasi; Division of Clinical Research (TK), National Hospital Organization Kikuchi Hospital, Koshi, Kumamoto; Chiba University Center for Forensic

Mental Health (NKa), Chiba, Chiba; Department of Psychiatry (NH), Hokkaido University Graduate School of Medicine, Sapporo, Hokkaido; Department of Psychiatry (KF), Okehazama Hospital, Toyoake, Aichi; Department of Psychiatry (KN), Division of Clinical Medicine, Faculty of Medicine, University of Tsukuba, Tsukuba, Ibaraki; Department of Psychiatry (TF), Gifu University Graduate School of Medicine, Gifu; Department of Psychiatry (TSu), Graduate School of Medicine, Kyoto University, Sakyo-ku, Kyoto; Osaka Psychiatric Medical Center (TN), Hirakata, Osaka; Okayama Psychiatric Medical Center (TN), Hirakata, Osaka; Okayama Psychiatric Medical Center (YYad), Kita-ku, Okayama; Department of Neuropsychiatry (MTaka), Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama City, Okayama; National Hospital Organization Ryukyu Hospital (NKi, TOt, MMu), Kunigamigun, Okinawa; Laboratory for Omics Informatics (AT), Omics Research Center, National Cerebral and Cardiovascular Center, Osaka; and RIKEN Center for Integrative Medical Sciences (MKubo), Yokohama, Japan.

TSa and MI contributed equally to this article.

Address correspondence to Ryota Hashimoto, M.D., Ph.D., Molecular Research Center for Children's Mental Development, United Graduate School of Child Development, Osaka University and Department of Psychiatry, Osaka University Graduate School of Medicine, D3, 2-2, Yamadaoka, Suita, Osaka 565-0871, Japan; E-mail: hashimor@psy.med.osaka-u.ac.jp.

Received Jul 22, 2015; revised Oct 31, 2015; accepted Dec 3, 2015.

Supplementary material cited in this article is available online at http://dx.doi.org/10.1016/j.biopsych.2015.12.006.

REFERENCES

- 1. Picchioni MM, Murray RM (2007): Schizophrenia. BMJ 335:91-95.
- Dold M, Leucht S (2014): Pharmacotherapy of treatment-resistant schizophrenia: A clinical perspective. Evid Based Ment Health 17:33–37.
- Hasan A, Falkai P, Wobrock T, Lieberman J, Glenthoj B, Gattaz WF, et al. (2012): World Federation of Societies of Biological Psychiatry (WFSBP) Guidelines for Biological Treatment of Schizophrenia, part 1: Update 2012 on the acute treatment of schizophrenia and the management of treatment resistance. World J Biol Psychiatry 13:318–378.
- Essali A, Al-Haj Haasan N, Li C, Rathbone J (2009): Clozapine versus typical neuroleptic medication for schizophrenia. Cochrane Database Syst Rev; CD000059.
- Lewis SW, Barnes TR, Davies L, Murray RM, Dunn G, Hayhurst KP, et al. (2006): Randomized controlled trial of effect of prescription of clozapine versus other second-generation antipsychotic drugs in resistant schizophrenia. Schizophr Bull 32:715–723.
- Cohen D, Bogers JP, van Dijk D, Bakker B, Schulte PF (2012): Beyond white blood cell monitoring: Screening in the initial phase of clozapine therapy. J Clin Psychiatry 73:1307–1312.
- Copolov DL, Bell WR, Benson WJ, Keks NA, Strazzeri DC, Johnson GF (1998): Clozapine treatment in Australia: A review of haematological monitoring. Med J Aust 168:495–497.
- Munro J, O'Sullivan D, Andrews C, Arana A, Mortimer A, Kerwin R (1999): Active monitoring of 12,760 clozapine recipients in the UK and Ireland. Beyond pharmacovigilance. Br J Psychiatry 175:576–580.
- Alvir JM, Lieberman JA, Safferman AZ, Schwimmer JL, Schaaf JA (1993): Clozapine-induced agranulocytosis. Incidence and risk factors in the United States. N Engl J Med 329:162–167.

- Atkin K, Kendall F, Gould D, Freeman H, Liberman J, O'Sullivan D (1996): Neutropenia and agranulocytosis in patients receiving clozapine in the UK and Ireland. Br J Psychiatry 169:483–488.
- 11. Inada K, Ishigooka J (2013): [Clozapine]. Nihon Rinsho71:678-683.
- Whiskey E, Taylor D (2007): Restarting clozapine after neutropenia: Evaluating the possibilities and practicalities. CNS Drugs 21: 25–35.
- Hummer M, Kurz M, Barnas C, Saria A, Fleischhacker WW (1994): Clozapine-induced transient white blood count disorders. J Clin Psychiatry 55:429–432.
- Phillips D, Rezvani K, Bain BJ (2000): Exercise induced mobilisation of the marginated granulocyte pool in the investigation of ethnic neutropenia. J Clin Pathol 53:481–483.
- Nooijen PM, Carvalho F, Flanagan RJ (2011): Haematological toxicity of clozapine and some other drugs used in psychiatry. Hum Psychopharmacol 26:112–119.
- Dunk LR, Annan LJ, Andrews CD (2006): Rechallenge with clozapine following leucopenia or neutropenia during previous therapy. Br J Psychiatry 188:255–263.
- Zhang JP, Malhotra AK (2013): Pharmacogenetics of antipsychotics: Recent progress and methodological issues. Expert Opin Drug Metab Toxicol 9:183–191.
- Athanasiou MC, Dettling M, Cascorbi I, Mosyagin I, Salisbury BA, Pierz KA, et al. (2011): Candidate gene analysis identifies a polymorphism in HLA-DQB1 associated with clozapine-induced agranulocytosis. J Clin Psychiatry 72:458–463.
- Goldstein JI, Jarskog LF, Hilliard C, Alfirevic A, Duncan L, Fourches D, et al. (2014): Clozapine-induced agranulocytosis is associated with rare HLA-DQB1 and HLA-B alleles. Nat Commun 5:4757.
- Pirmohamed M, Park K (1997): Mechanism of clozapine-induced agranulocytosis: Current status of research and implications for drug development. CNS Drugs 7:139–158.
- 21. Ikeda M, Shimasaki A, Takahashi A, Kondo K, Saito T, Kawase K, *et al.* (2015): Genome-wide environment interaction between depressive state and stressful life events. J Clin Psychiatry.
- Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D (2006): Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 38:904–909.
- Yamaguchi-Kabata Y, Nakazono K, Takahashi A, Saito S, Hosono N, Kubo M, et al. (2008): Japanese population structure, based on SNP genotypes from 7003 individuals compared to other ethnic groups: Effects on population-based association studies. Am J Hum Genet 83: 445–456.
- Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ (2015): Second-generation PLINK: Rising to the challenge of larger and richer datasets. Gigascience 4:7.
- Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, et al. (2010): LocusZoom: Regional visualization of genome-wide association scan results. Bioinformatics 26:2336–2337.
- Butcher NJ, Fung WL, Fitzpatrick L, Guna A, Andrade DM, Lang AE, et al. (2015): Response to clozapine in a clinically identifiable subtype of schizophrenia. Br J Psychiatry 206:484–491.
- Naber D, Riedel M, Klimke A, Vorbach EU, Lambert M, Kuhn KU, *et al.* (2005): Randomized double blind comparison of olanzapine vs. clozapine on subjective well-being and clinical outcome in patients with schizophrenia. Acta Psychiatr Scand 111:106–115.