

Supplementary material for the article:

Ilić Đurđić, K.; Ostafe, R.; Prodanović, O.; Đurđević Đelmaš, A.; Popović, N.; Fischer, R.; Schillberg, S.; Prodanović, R. Improved Degradation of Azo Dyes by Lignin Peroxidase Following Mutagenesis at Two Sites near the Catalytic Pocket and the Application of Peroxidase-Coated Yeast Cell Walls. *Front. Environ. Sci. Eng.* **2020**, *15* (2), 19.  
<https://doi.org/10.1007/s11783-020-1311-4>.

## Supporting Information

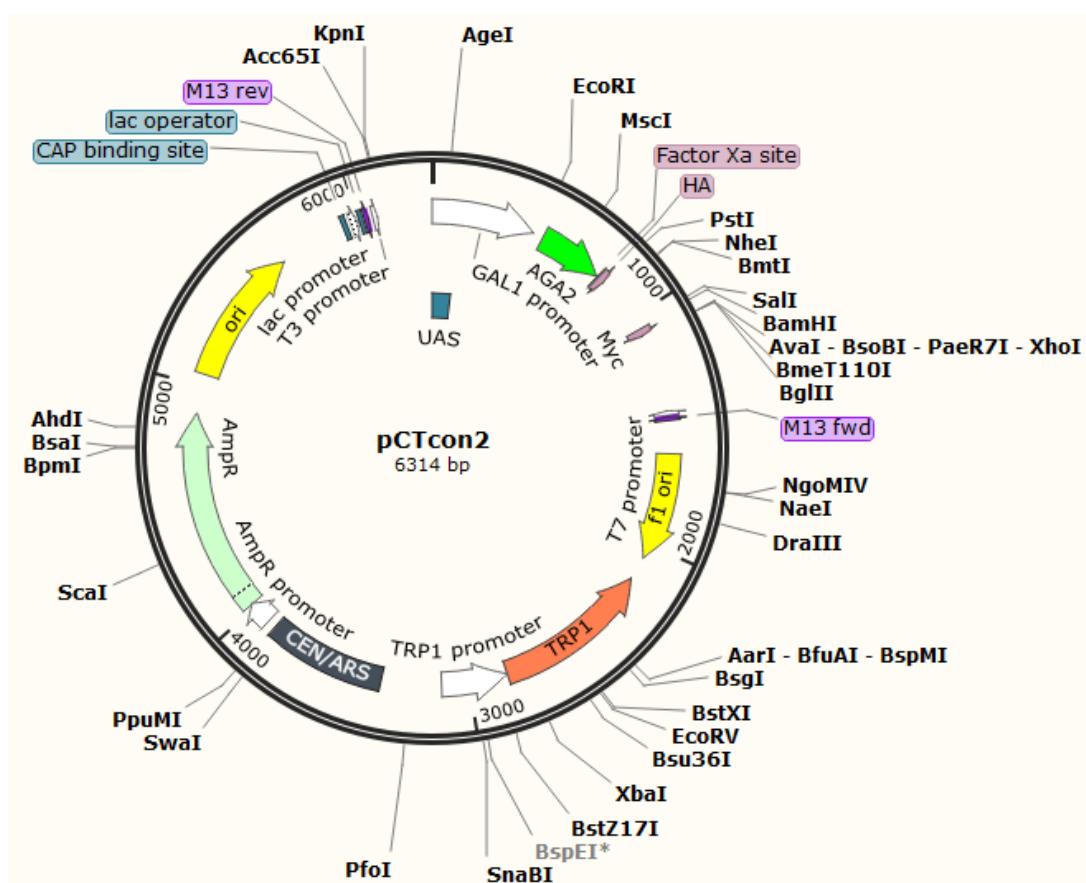
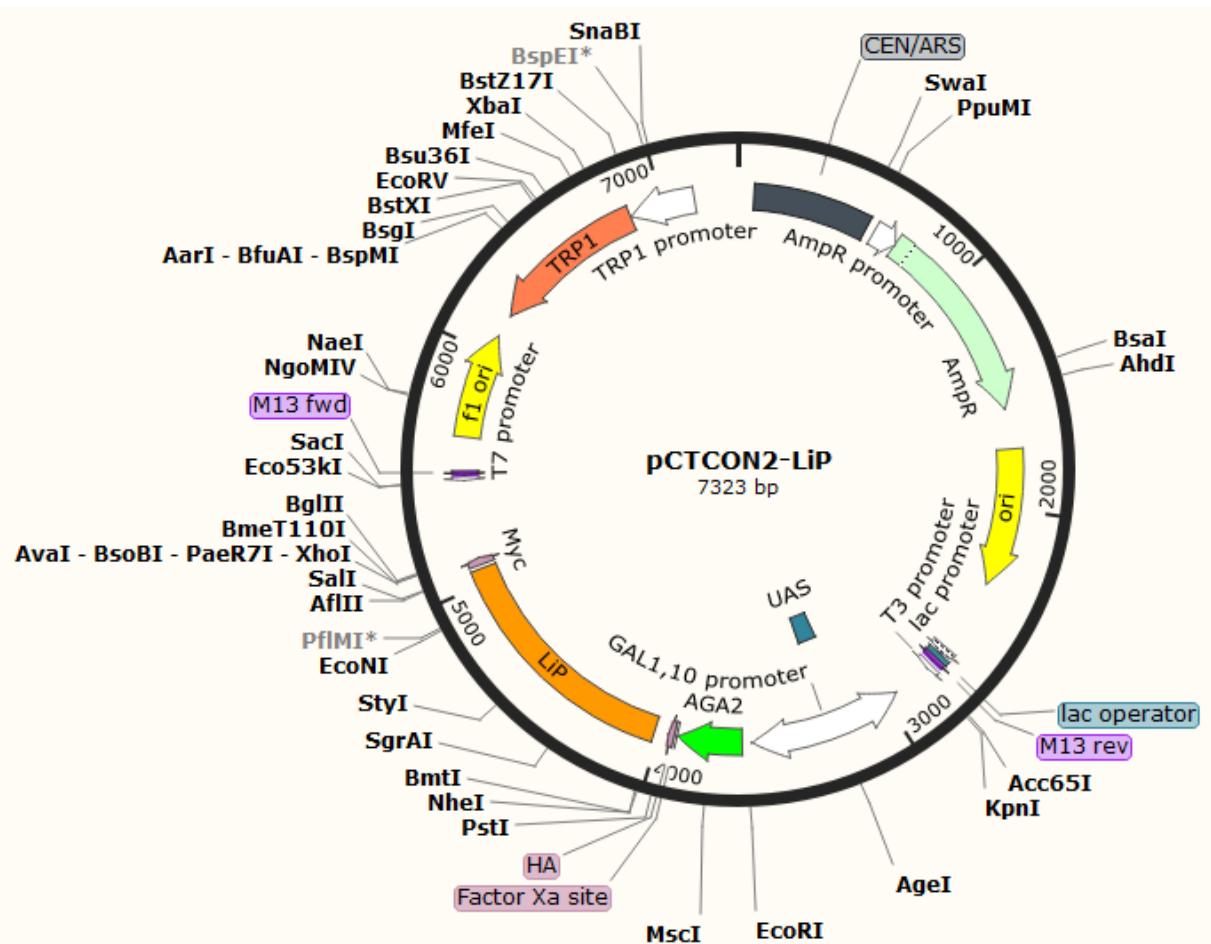


Figure S1. Map of pCTCON2 vector



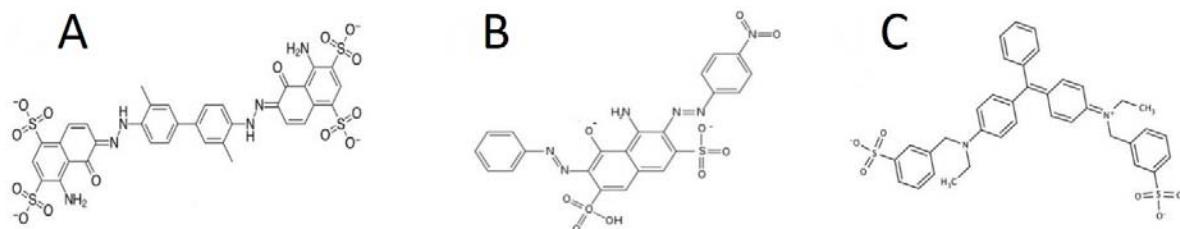
**Figure S2.** Schematic representation of LiP-pCTCON2 construct.

**Table S1.** Primers sequences

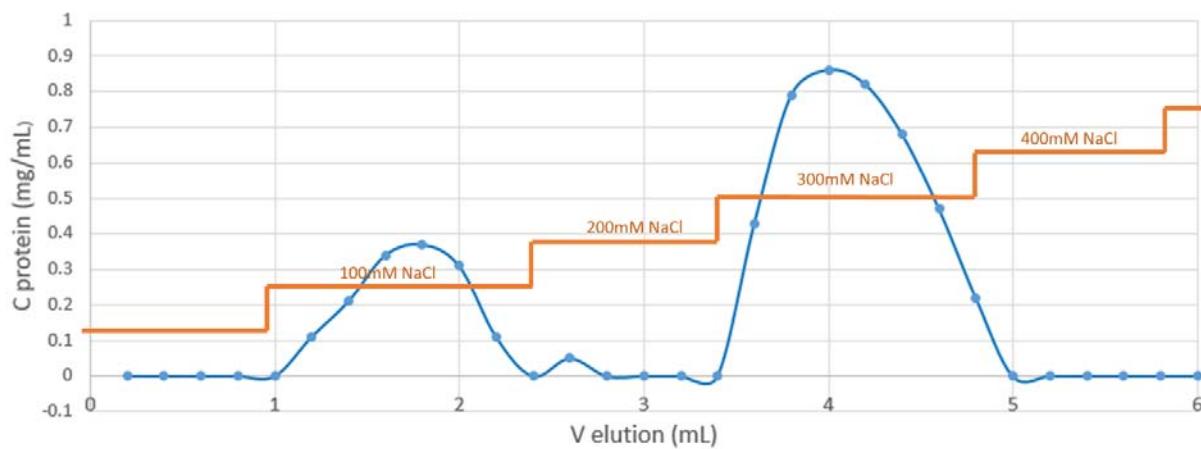
| Application                            | Sequence (5' → 3')            |
|--|-------------------------------|
| Forward primer for cloning of LiP gene | AATGCTAGCGAACCTGTGCTAATGGTAAG |
| Reverse primer for cloning of LiP gene | AATGTCGACTTAAGCCTGTGGAGGTATT  |
| Forward primer for sequencing          | CCCATACGACGTTCCAGACTACGC      |
| Reverse primer for sequencing          | GATCTCGAGCTATTACAAGTCCTCTCAG  |
| Primer for mutation of D165N           | GCTGGAGAATTNNNNGAATTAGAATTGG  |
| Primer for mutation of D264N           | CAAAGTTGGTAGATNNNTTCAATTCAATT |

GCAACCTGTGCTAATGGTAAGACAGTAGGTGATGCTTCTTGCCTGGTCATGCTTAGATGAC  
 ATTCAAGCAAATATGTTCACGGTGGTCAGTGTGGGCCGAAGCTCACGAATCTATTAGATTGGCTTT  
 CACGATTCTATAGCAATATCTCCTGCTATGGAGGCTAAAGGAAAGTTGGTGGAGGTGGAGCCGATGG  
 TTCAATCATGATATTGATAACCAGAACCGCTTCCACCCAACACATAGGATTGGATGAAGTAGTTGC  
 TATGCAAAAACCATTGTCCAAAACACGGAGTAACACCAAGGTGACTTATCGCTTCGCCGGTCCGT  
 CGCCTTATCTAATTGCCCTGGTGCCTCAGATGAACCTCTTACCGGAAGGAAGGCCAGCTACACAACC  
 AGCACCTGACGGTTAGTACCAAGAACCTTTCATACTGTAGACCAAATTATAGCTAGAGTCAACGACGC  
 TGGAGAATTGATGAATTAGAATTGGTATGGATGTTGCTGCTCACTCTGTTGCAGCAGTCATGACGT  
 TGATCCAACCGTCCAAGGATTACCTTTGATTCAACTCCAGGAATTTCGATTCTCAGTTTCGTTGAA  
 ACTCAATTCAAGAGGAACCTTGTTCAGGTTCAGGAGGAATCAAGGTGAAGTTGAGTCAGGTATGGC  
 TGGTGAATCAGGATCCAGACAGATCATACATTGGCTAGAGATTCTAGGACTGCCTGTGAATGGCAAT  
 CTTCGTCGTAATCAATCAAAGTTGGTAGATTTCAATTCAATTGTTGGCTTAACCCAGTTGGG  
 TCAAGATCCAATGCAATGACAGATTGTTCTGATGTAATCCCATTATCAAACCTATACCAAGGTAATGG  
 TCCATTTCATTCTCCCTGGTAAATCACATTCTGATATTGAAACAGGCTTGCTGAGACTCCTTT  
 CCATTTAGTTACTTGCAGGTCCAGCCACTTCAGTCGCAAGAATACCTCACACAAGGCTTAAGTC  
 GA

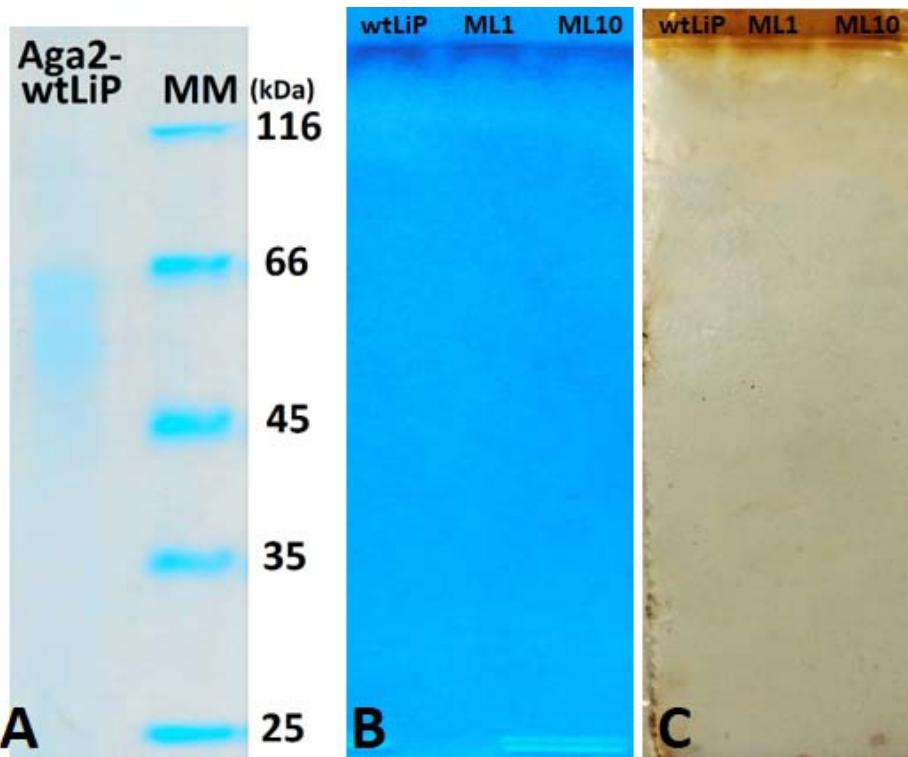
**Figure S3.** Sequence of lignin peroxidase H8 gene (5' → 3').



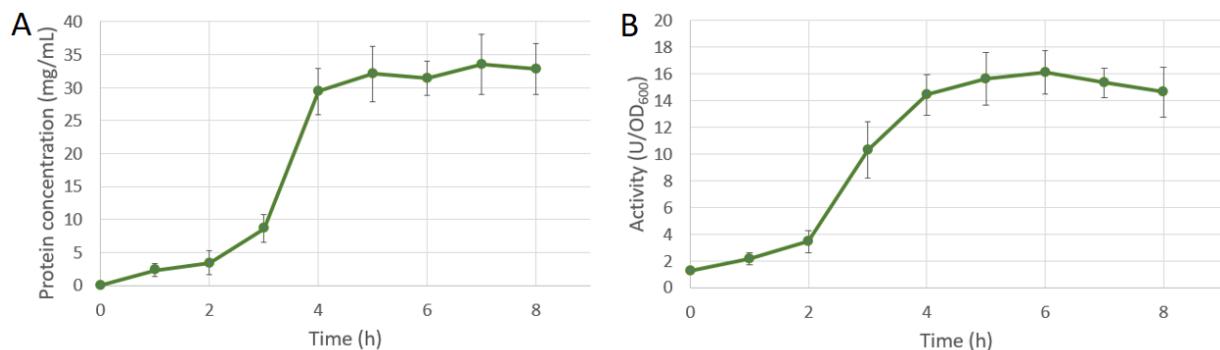
**Figure S4:** Structures of used azo dyes (A) Evans blue (B) Amido black 10B (C) Guinea green



**Figure S5.** Chromatogram showing the elution of Aga2-wtLiP chimera using Vivapure mini spin columns with optimized NaCl step elution.



**Figure S6.** Polyacrylamide gel electrophoresis **A.** SDS-PAGE of purified aga2-wtLiP compared with molecular weight markers (MM) **B.** Native 12% polyacrylamide gel electrophoresis with protein bands after CBB R250 staining for Aga2-wtLiP and two selected mutants. **C.** Native 12% polyacrylamide gel electrophoresis with activity bands in the gel after incubation with 0.5 mM H<sub>2</sub>O<sub>2</sub> and 9 mM guaiacol for Aga2-wtLiP and two selected mutants



**Figure S7.** Concentration of intercellular proteins released from the cells during cell lysis and activity of wtLiP during cell lysis. (A) In order to optimize lysis of toluene-induced cell lysis we followed concentration of released proteins during 8 h using Bradford reagent. (B) Activity of wtLiP was followed during cell lysis with 2,4-DCP assay (0.2 mM DCP, 80mM 4-AAP and 1 mM H<sub>2</sub>O<sub>2</sub>). Data are means of triplicate experiments with error bars indicating standard deviations. Error bars are not visible when smaller than the symbol size.