

1 **Match and Mismatch between Dietary Switches and Microbial Partners in Plant Sap-**
2 **Feeding Insects**

3

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12 **Abstract**

13 Some animal groups associate with the same vertically-transmitted microbial symbionts over
14 extended periods of evolutionary time, punctuated by occasional symbiont switches to different
15 microbial taxa. Here we test the oft-repeated suggestion that symbiont switches are linked with
16 host diet changes, focusing on hemipteran insects of the suborder Auchenorrhyncha. These insects
17 include the only animals that feed on plant xylem sap through the life cycle, as well as taxa that
18 feed on phloem sap and plant parenchyma cells. Ancestral state reconstruction provides strong
19 statistical support for a xylem-feeding auchenorrhynchan ancestor bearing the dual symbiosis with
20 the primary symbiont *Sulcia* (Bacteroidetes) and companion symbiont “ β -Sym” (β -
21 proteobacteria). We identified 7 dietary transitions from xylem-feeding (six to phloem-feeding,
22 one to parenchyma-feeding), but no reversions to xylem-feeding; five evolutionary losses of
23 *Sulcia*, including replacements by yeast symbionts, exclusively in phloem/parenchyma-feeding
24 lineages; and 14-15 losses of β -Sym, including 9 transitions to a different bacterial companion
25 symbiont. Our analysis indicates that, although companion symbiont switching is not associated
26 with shifts in host diet, *Sulcia* is likely required for xylem-feeding. Furthermore, the ancestral
27 auchenorrhynchan bearing *Sulcia* and β -Sym likely represents the sole evolutionary origin of
28 xylem feeding in the animal kingdom.

29 **1. Introduction**

30 Various ecologically important traits of many animals are mediated by symbiotic
31 microorganisms [1,2]. For example, microbial symbionts may produce toxins that protect their
32 animal host from specific pathogens, or synthesize nutrients that enable their host to utilize
33 otherwise inadequate diets [3,4]. Many of these symbioses display high partner fidelity over long
34 periods of evolutionary time, often with strict co-diversification of host and symbiont lineages
35 [5,6]. However, both symbiont loss and symbiont switching, often to a phylogenetically-distant
36 microbial taxon, has been identified in many host lineages, especially among insects [7–10], as
37 well as in other invertebrate taxa [11]. It has been suggested that symbiont switching may be linked
38 to changes in host traits, e.g. shifts in habitat or diet [12,13], potentially with major ecological and
39 evolutionary consequences for the host [14]. For instance, symbiont switching has been suggested
40 to have been an important driver of host diversification [15,16]. Yet despite their potential
41 importance, hypotheses of links between diet and symbiont switching have rarely been tested
42 formally, limiting our insight into their general applicability across taxa. In recent years, however,
43 our capacity to evaluate such hypotheses has greatly increased by advances in phylogenetic
44 comparative methods, such as ancestral state reconstructions [17].

45 The purpose of this study was to investigate the patterns of symbiont switches by analysis
46 of the temporal relationship between evolutionary shifts in host diet and symbiotic partners.
47 Insects of the order Hemiptera are well suited to this problem for two linked reasons. First,
48 uniquely among animals, some hemipterans can utilize different types of plant sap (i.e. phloem or
49 xylem) through the life cycle; and, second, this trait is correlated with possession of vertically-
50 transmitted microbial symbionts that overproduce essential amino acids, nutrients in short supply
51 in plant sap [1,18]. Our specific focus is the suborder Auchenorrhyncha, which includes the only

52 insects that feed on xylem sap through the life cycle, as well as representatives that feed on phloem
53 sap and the more nutritious diet of whole plant cell contents [19]. The Auchenorrhyncha comprises
54 two infra-orders, the Fulgoromorpha (the planthoppers) and Cicadamorpha (including the
55 leafhoppers, cicadas and froghoppers) [20,21]. Many taxa of both infra-orders bear a dual
56 symbiosis with a bacterium *Candidatus Sulcia muelleri* (Bacteroidetes), also referred to as ‘the
57 primary symbiont’ [1,22], and a β -proteobacterium, which is variously known as: β -Sym, *Ca.*
58 *Nasuia deltocephalinicola* in leafhoppers, *Ca. Vindania fulgoroideae* in planthoppers and *Ca.*
59 *Zinderia insecticola* in spittlebugs, or more generally as a ‘companion symbiont’ to *Sulcia* [23–
60 27]. Some auchenorrhynchans, however, bear *Sulcia* as the sole symbiont, associate with a
61 companion symbiont other than β -Sym [28–31], or have ascomycete fungal symbionts, generically
62 known as yeast-like symbionts [32–35]. This rich diversity of symbionts with functions closely
63 tied to host diet provides a superb basis to test the relationship between switches in diet and
64 symbiont identity.

65 In this study, we generated a detailed phylogeny of the Auchenorrhyncha. We then mapped
66 key symbiont associations (primary and companion symbiont status) and host diet onto this
67 phylogeny, and used a comparative approach to study the association between symbiont and diet.
68 This allowed us to investigate the patterns of association between transitions in symbiont identity
69 - including symbiont losses - and diet across large numbers of clades over long periods of time.

70

71 **2. Materials and Methods**

72 **(a) Phylogeny**

73

74 We generated a phylogeny at the genus-level for the Auchenorrhyncha using a supermatrix
75 approach [36], utilising five marker genes that had previously been used to study hemipteran

76 phylogeny (18S, 28S, COI, Wingless and Histone 3) [20,37,38]. We used phyloGenerator (version
77 2) [39] to obtain sequences for all auchenorrhynchan genera with any of these markers available
78 in NCBI GenBank. We employed the referenceDownload method to select high quality
79 representative sequences for each genus, based on reference sequences from [20]. A genus can
80 thus be represented by sequences from different species within the genus. For each marker gene,
81 the selected sequences were aligned using MAFFT (version 7.310) [40]. We then concatenated the
82 five alignments, and visually verified the quality of the resulting supermatrix of 824 genera and
83 7243 bp using Aliview (version 1.20) [41]. We created a maximum-likelihood (ML) phylogeny in
84 RAxML (version 8.2.8) using a GTRCAT substitution model and rapid bootstrapping (100
85 replicates) followed by a thorough ML search [42]. The branch lengths of the bootstrap replicates
86 were optimised using GTRGAMMA. Based on [20,21], the genera *Ceratocombus* (Heteroptera)
87 and *Xenophyes* (Coleorrhyncha) were used as outgroup to root the tree, and we used the NCBI
88 taxonomy at the subfamily level to constrain our tree search. We used r8s (version 1.8), employing
89 the Langley-Fitch method and a truncated newton (TN) algorithm to convert our ML phylogeny
90 to a timetree [43]. To date the phylogeny, we fixed the root to 299 MYA and constrained a further
91 seventeen nodes based on previous estimates of divergence times (table S1).

92

93 **(b) Trait data set**

94 We conducted a systematic search of the primary literature to compile information on three
95 auchenorrhynchan traits of interest: (1) diet, (2) primary symbiont association status and (3)
96 companion symbiont association status. We obtained data from 34 different studies, covering a
97 total of 162 unique genera (full references table S3). We recorded data and performed all our
98 analyses at the genus level because variation in the traits of interest was not evident at the subgenus

99 level. Primary symbiosis was treated as a binary variable: presence of primary endosymbiont
100 *Candidatus* Sulcia muelleri vs. absence [22]. For the companion symbiosis we recorded the
101 taxonomic identity of the endosymbiont. We then clustered this into a categorical variable with
102 five levels: (i) α -proteobacteria (*Candidatus* Hodgkinia cicadicola), (ii) β -proteobacteria
103 (*Candidatus* Vidania fulgoroideae; *Candidatus* Zinderia insecticola; *Candidatus* Nasuia
104 deltocephalinicola), (iii) γ -proteobacteria (*Candidatus* Purcelliella pentastirinum; a *Sodalis*-like
105 bacterial symbiont; *Candidatus* Baumannia cicadellinicola), (iv) yeast-like symbionts (YLS)
106 (*Ophiocordyceps*, *Entomomyces delphacidicola*) and (v) companion symbionts absent. For the
107 diet variable, each genus was assigned as: phloem-feeder, xylem-feeder, or parenchyma cell-
108 feeder. Insects within the Fulgoromorpha, Membracidae, and Deltocephalinae were inferred as
109 phloem-feeders, and within the Cicadoidea, Cercopoidea, Cicadellinae as xylem-feeders [44–48].
110 One study reported an absence of both primary and companion symbionts in sap-feeding
111 Auchenorrhyncha [35]. Given the nutritionally highly unbalanced composition of plant sap [1],
112 these cases likely represent false negatives and were therefore not included in our dataset.

113

114 (c) Comparative analyses

115 All our comparative analyses were performed in R (version 3.4.4.). The overlap of our full
116 phylogeny containing 824 tips (genera), and our full trait database (162 genera) was equal to 145
117 genera. Of these, we had data on primary symbiont status for 142 genera: this is the dataset we
118 included in our analyses (full analysed genus list available in table S4). The full phylogeny was
119 pruned to comprise the 142 tip genera. We first performed ancestral state reconstructions (ASRs)
120 to elucidate the evolutionary history of our three traits of interest. We then compared correlated

121 and uncorrelated models of evolution to study the potential for evolutionary correlations between
122 traits [49].

123
124 For our ASRs of both diet (three levels) and companion symbiont association status (five levels),
125 we fitted a model of evolution for categorical traits, using the R-package *corHMM* (version 1.22)
126 [50]. We generated ER (equal rates), SYM (symmetrical) and ARD (All-Rates-Different) models
127 of evolution and used AICc-weights to guide our model selection. In our diet ASR, we treated diet
128 state as missing for our outgroup. For our ASR of the primary symbiosis, we analysed hidden rates
129 models, which allow for potential variation in the rate of evolution of a binary trait [50]. We used
130 marginal likelihoods to estimate ancestral states and employed Yang's method to determine the
131 ancestral root state for diet and companion symbiosis [51], while constraining the root to primary
132 symbiont presence. As before, we calculated the number of transitions between evolutionary states
133 assuming parsimony [13,52].

134
135 We used Pagel's method to study potential correlated evolution among binary traits of interest,
136 specifically host diet and symbiotic association status [49]. Given that xylem is under negative
137 pressure and is particularly nutrient poor, using it as a food sources requires a highly specialised
138 set of adaptations including an enlarged muscle-filled head to facilitate active pumping of the
139 xylem sap [53]. We therefore analysed diet as a binary variable of xylem-feeding vs other food
140 sources (*i.e.* phloem or parenchyma-feeding). For the companion symbiosis, we generated three
141 binary variables, representing the presence or absence of the main types of companion symbionts
142 (*i.e.* β -proteobacteria, γ -proteobacteria and YLS; we did not analyse α -proteobacteria this way

143 because they are found in only two genera). In all cases, we used R-package *phytools* (version 0.6-
144 44) [54] to compare correlated and uncorrelated models of evolution using likelihood ratio tests.

145

146 **(d) Sensitivity**

147 All results from phylogenetic comparative analyses are subject to uncertainty in the underlying
148 data [55]. We therefore characterised the sensitivity of our key result of correlated evolution among
149 primary endosymbiosis and diet status to (i) phylogenetic, (ii) trait and (iii) taxon sampling
150 uncertainty. To quantify sensitivity to phylogenetic uncertainty, we regenerated our model across
151 the hundred bootstrap replica that were used to obtain support for our ML phylogeny, and
152 characterised the p-value and Δ -AICc for each rerun (figure S6). To study the effects of trait
153 uncertainty, we simulated trait dataset based on our original assignments, assuming separate false
154 positive and false negative rates for our data of primary symbiont (*Sulcia*) presence (figure S7).
155 This allows us to simulate an unbiased but inaccurate dataset (both high false positive and false
156 negative), as well as biases in the field towards detection of symbionts (high false positive, low
157 false negative) or towards not finding symbionts which are actually there (high false negative rate,
158 low false positive). Third, we tested if our key conclusions are robust to the specific genus set
159 sampled (figure S8). This helps address the potential for influential (groups of) genera driving the
160 results. We used a jack-knifing approach, randomly removing up to 30% of the genera from the
161 analyses [55].

162

163 **3. Results**

164 **(a) Ancestral state reconstructions**

165 In our first analysis of 142 genera of auchenorrhynchan insects that have both phylogenetic
166 information and trait data, we investigated the evolutionary transitions in diet. The evolutionary
167 model with the best statistical support (electronic supplementary material table S2) estimates
168 xylem as the most likely ancestral state of the Auchenorrhyncha (likelihood of xylem-feeder:
169 90.1%; parenchyma-feeder 9.9%). In this model, xylem feeding is subsequently lost at the origin
170 of the Fulgoromorpha (planthoppers), and five times within the the Cicadomorpha (comprising
171 leafhoppers, treehoppers, cicadas and spittlebugs) (figure 1; figures S1 and S2). All diet switches
172 are from xylem to phloem sap, apart from a single switch from xylem feeding to parenchyma
173 feeding near the base of the Typhlocybae (Cicadomorpha: Cicadellidae). All instances of phloem
174 feeding in the Auchenorrhyncha appear to be an evolutionary end-state, but the model predicts one
175 further transition, from parenchyma feeding to phloem feeding in the Ledorinae (Cicadomorpha:
176 Cicadellidae).

177

178 In the best-supported model for the primary symbiont *Sulcia*, the association is initially in a
179 relatively unstable evolutionary state (figures S3 and S4). Specifically, the symbiosis is lost five
180 times (calculated 4.90 losses) across the Auchenorrhyncha (figure 2) but never regained
181 (calculated 0.19 gains). Four of these losses occur predominantly within the Fulgoromorpha, with
182 a single instance in the Cicadomorpha, in the Ledorinae. In the other lineages of the Cicadomorpha,
183 *Sulcia* presence is retained, and transitions towards a stable evolutionary state (figure S4). All
184 losses occur in lineages that have switched from xylem to phloem or parenchyma-feeding.

185

186 For the companion symbionts, the best-supported model is symmetrical, i.e. with equal
187 transition rates between two states in both directions (table S2). The model predicts that the

188 Auchenorrhyncha was ancestrally associated with the β -Sym symbiont (likelihood β -Sym 97.1%;
189 companion absence 2.9%; figure 1 and figure S5), which has been retained in most
190 Fulgoromorpha, as well as many Cercopoidea and Cicadellidae within the Cicadomorpha. In other
191 lineages, however, β -Sym was not stable, with at least nine switches to alternative companion
192 symbionts and a further five or six losses without replacement by a taxonomically-different
193 companion symbiont (figure 1). In contrast to the losses of the primary *Sulcia*-symbiosis, these
194 losses take place across all three diets observed in the Auchenorrhyncha.

195

196 **(b) Correlated evolution**

197 To evaluate the hypothesis that diet changes co-occur with changes in symbiont composition,
198 we compared correlated and uncorrelated models of evolution. This analysis yielded strong
199 evidence of correlated evolution of primary endosymbiont (*Sulcia*) status and diet (Likelihood
200 Ratio Test: $p \ll 0.01$), but no evidence of correlated evolution of diet with any group of companion
201 symbionts (β -Sym, $p=0.49$), γ -proteobacteria, $p=0.06$, yeast-like symbionts $p=0.70$). Specifically,
202 the primary symbiont *Sulcia* is lost exclusively in insects that have switched from feeding on xylem
203 to consuming phloem or parenchyma (figure 1). The associated ancestral state reconstruction
204 confirms that the loss of *Sulcia* is invariably preceded by loss of xylem-feeding, generally by
205 dozens of millions of years (and in one case by up to 185 million years). Thus, loss of xylem-
206 feeding appears to be a required, but probably not sufficient, condition for *Sulcia* loss. In summary,
207 these results indicate an indirect evolutionary link between diet with loss of the *Sulcia* primary
208 symbiont but not with changes of the companion symbiont.

209

210 **(c) Sensitivity analyses**

211 We evaluated the sensitivity of our correlated evolution results to phylogenetic, data and sampling
212 uncertainty using a simulation approach [55]. We found that our key conclusions are robust to
213 phylogenetic uncertainty (figure S6). We also simulated errors in our underlying trait data and
214 found that correlated evolution conclusions were robust even to 25% of our underlying trait data
215 being mistaken, including to biased over- or under- detection of symbioses (figure S7). Lastly, we
216 found that our key results are robust to variation in genus sampling (figure S8).

217

218 **3. Discussion**

219 It has been argued in much of the symbiosis literature that evolutionary acquisition, loss or
220 switching of microbial symbionts have been drivers of major evolutionary changes in host traits,
221 especially diets of animals [1,2,14,56–58]. Despite the seemingly close link between host diet and
222 symbiont function in the Auchenorrhyncha, the correlations identified in this study provide limited
223 support for this proposition. Although our finding of a perfect association between xylem-feeding
224 and retention of the primary symbiont *Sulcia* is consistent with the proposed relationship between
225 symbiont and diet, two other findings of this study are inconsistent. First, we obtain no association
226 between change or loss of companion symbionts and diet switching; both occur across all
227 auchenorrhynchan diets. Second, the loss of *Sulcia* in some phloem-feeding lineages occurs tens
228 of millions of years after the dietary switch from xylem feeding (figure 1), suggesting that the
229 change in diet is permissive for – and not a consequence of - symbiosis change.

230 We have also obtained strong statistical support for a xylem-feeding auchenorrhynchan
231 ancestor. This conclusion contrasts with the hypothesis that xylem-feeding evolved in the ancestor
232 of the Cicadomorpha, proposed in a recent study [59] that was not based on a formal ancestral
233 trait reconstruction of insect diet and contained only few Auchenorrhyncha species. Our analyses

234 indicate multiple switches from xylem-feeding to phloem- and parenchyma-feeding, but no
235 reversions back to a diet of xylem sap. Given that no other animal group is known to feed through
236 the life cycle on xylem [60], we conclude that the ancestral auchenorrhynchan represents the sole
237 evolutionary origin of the xylem-feeding habit in the animal kingdom.

238 This study has provided a quantitative phylogenetic framework for the many instances of
239 symbiont gain, loss and switching reported for the Auchenorrhyncha in the literature [10,27]. In
240 particular, our quantitative data validate the long-held belief that the xylem-feeding habit in the
241 Auchenorrhyncha is perfectly correlated with the possession of bacterial symbionts [1] and
242 confirm the inference from previous studies that the ancestral symbiosis comprised the primary
243 symbiont *Sulcia* and companion symbiont β -Sym [22,27,28]. The retention of *Sulcia* in all xylem-
244 feeding lineages, despite multiple switches of the companion symbiont, suggests that *Sulcia* has
245 unique traits that permit xylem-feeding. A possible explanation comes from recent metabolic
246 modeling, which has revealed that *Sulcia* converts host-derived nitrogen sources into essential
247 amino acids with very high efficiency [61]. This quantitative metabolic trait may be especially
248 valuable for host utilization of xylem sap, which has a markedly lower total nitrogen content than
249 phloem sap [60]. In other words, the fitness consequences of a symbiont switch from *Sulcia* to a
250 less efficient symbiont may be less severe for a phloem-feeding insect, thereby explaining how
251 *Sulcia* has been replaced by other taxa in some phloem-feeding auchenorrhynchan lineages, but in
252 no xylem-feeding lineages. Intriguingly, most replacements of *Sulcia* are evolutionarily rather
253 ancient, yielding a pattern of increased stability of the *Sulcia* symbiosis over time (figure S4).
254 Further research is required to investigate whether other ancient symbionts display a similar pattern
255 of increasing stabilization over time, and to identify the contribution of metabolic and other co-
256 evolved host-symbiont interactions to this trait.

257 What are the factors driving the many instances of symbiont loss and replacement that are
258 not correlated with dietary switches in the Auchenorrhyncha? These losses may be mediated by
259 ecological factors other than diet. For example, many insect symbionts are relatively intolerant of
260 thermal stress [62], and the replacement of thermally-intolerant symbionts by tolerant strains has
261 been demonstrated in insects exposed to elevated temperatures in the laboratory [63].
262 Evolutionary or co-evolutionary processes internal to the symbiosis may also be involved. In
263 particular, these symbionts are subject to genomic decay because obligate vertical transmission of
264 small numbers of symbiont cells results in inefficient selection against deleterious mutations and
265 reduced symbiont functionality [64–66], favoring replacement of a deteriorating symbiont by an
266 alternative taxon [10,14,28]. The incidence of symbiont replacements may, however, be
267 constrained by the high metabolic cost of maintaining a symbiont of large genome size [61] or by
268 traits of the incoming symbiont that may be deleterious to the host [33].

269 Is the poor correspondence between evolutionary transitions in symbiont identity and host
270 diet in the Auchenorrhyncha generalizable to other symbioses? A key feature of this system is that
271 the symbiont services to the host relate to insect nutrition, involving the provisioning of essential
272 amino acids and vitamins, and are broadly equivalent for insects feeding on xylem and phloem sap
273 [67]. We predict that, as for the Auchenorrhyncha, the many instances of symbiont switches in
274 other hemipteran groups with nutritional symbioses [10,14] are largely uncoupled to diet-related
275 factors. Symbiont switches in aphids of the subfamily Lachninae (Hemiptera: suborder
276 Sternorrhyncha) are fully consistent with this prediction [68]. Future studies on the patterns of
277 symbiosis switches across larger phylogenetic scales in hemipterans other than the
278 Auchenorrhyncha, as well as in other animal groups with nutritional symbioses, would provide
279 valuable insights into this topic. We hypothesize that the evolutionary co-incidence of symbiont

280 switches and shifts in ecologically-important traits of the host may be more evident for protective
281 functions, e.g. symbionts that either produce toxins which confer resistance to natural enemies or
282 detoxify dietary allelochemicals [3,69]. Here, acquisition of a symbiont capable of synthesizing or
283 degrading novel toxins could facilitate the exploitation of novel habitats or diets, potentially with
284 consequences for evolutionary patterns of speciation and adaptive diversification in the host
285 lineage. Other future research could check for symbiont switching between closely-related taxa
286 using phylogenomic analysis to test for incongruencies between phylogenies of hosts and both
287 primary and companion endosymbionts [22]. These hypotheses are increasingly becoming testable
288 due to (i) the routine availability of molecular tools for phylogenetics and symbiont identification,
289 and (ii) development of sophisticated comparative methods to test for large scale patterns in the
290 relationships between symbiont switches and host divergence and trait evolution.

291

292 **Competing Interests.** We declare no competing interests.

293 **Data accessibility.** Our full analysed insect trait database, phylogeny and alignment files,
294 sequence accession table and all R-code to repeat our analyses and generate figures are available
295 on GitHub (https://github.com/gijsbertwerner/aucho_endosymbionts) and on Dryad (link upon
296 publication).

297 **Authors' Contributions** L.B-R A.E.D. and G.D.A.W. designed the study, L.B-R and G.D.A.W.
298 compiled the database, generated the phylogeny and performed the comparative analyses.

299 G.D.A.W. and A.E.D. wrote the manuscript, and all authors commented on the manuscript.

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305

306 **References**

- 307 1. Buchner P. 1965 *Endosymbiosis of animals with plant microorganisms*. New York: John
308 Wiley & Sons.
- 309 2. Douglas AE. 2010 *The Symbiotic Habit*. Princeton University Press.
- 310 3. Flórez L V., Biedermann PHW, Engl T, Kaltenpoth M. 2015 Defensive symbioses of
311 animals with prokaryotic and eukaryotic microorganisms. *Nat. Prod. Rep.* **32**, 904–936.
312 (doi:10.1039/c5np00010f)
- 313 4. Douglas AE. 2009 The microbial dimension in insect nutritional ecology. *Funct. Ecol.* **23**,
314 38–47. (doi:10.1111/j.1365-2435.2008.01442.x)
- 315 5. Bright M, Bulgheresi S. 2010 A complex journey: transmission of microbial symbionts.
316 *Nat. Rev. Microbiol.* **8**, 218–230. (doi:10.1038/nrmicro2262)
- 317 6. Moran NA, Munson MA, Baumann P, Ishikawa H. 1993 A molecular clock in
318 endosymbiotic bacteria is calibrated using the insect hosts. *Proc. R. Soc. London. Ser. B*
319 *Biol. Sci.* **253**, 167–171. (doi:10.1098/rspb.1993.0098)
- 320 7. Cafaro MJ *et al.* 2011 Specificity in the symbiotic association between fungus-growing
321 ants and protective *Pseudonocardia* bacteria. *Proc. R. Soc. B Biol. Sci.* **278**, 1814–1822.
322 (doi:10.1098/rspb.2010.2118)
- 323 8. Manzano-Marín A, Szabó G, Simon J-C, Horn M, Latorre A. 2017 Happens in the best of
324 subfamilies: establishment and repeated replacements of co-obligate secondary
325 endosymbionts within Lachninae aphids. *Environ. Microbiol.* **19**, 393–408.

- 326 (doi:10.1111/1462-2920.13633)
- 327 9. Toju H, Tanabe AS, Notsu Y, Sota T, Fukatsu T. 2013 Diversification of endosymbiosis:
328 replacements, co-speciation and promiscuity of bacteriocyte symbionts in weevils. *ISME*
329 *J.* **7**, 1378–1390. (doi:10.1038/ismej.2013.27)
- 330 10. Douglas AE. 2016 How multi-partner endosymbioses function. *Nat. Rev. Microbiol.* **14**,
331 731–743. (doi:10.1038/nrmicro.2016.151)
- 332 11. Ozawa G, Shimamura S, Takaki Y, Takishita K, Ikuta T, Barry JP, Maruyama T, Fujikura
333 K, Yoshida T. 2017 Ancient occasional host switching of maternally transmitted bacterial
334 symbionts of chemosynthetic vesicomid clams. *Genome Biol. Evol.* **9**, 2226–2236.
335 (doi:10.1093/gbe/evx166)
- 336 12. Lefèvre C, Charles H, Vallier A, Delobel B, Farrell B, Heddi A. 2004 Endosymbiont
337 phylogenesis in the dryophthoridae weevils: evidence for bacterial replacement. *Mol. Biol.*
338 *Evol.* **21**, 965–973. (doi:10.1093/molbev/msh063)
- 339 13. Werner GDA, Cornelissen JHC, Cornwell WK, Soudzilovskaia NA, Kattge J, West SA,
340 Kiers ET. 2018 Symbiont switching and alternative resource acquisition strategies drive
341 mutualism breakdown. *Proc. Natl. Acad. Sci.* **115**, 5229–5234.
342 (doi:10.1073/pnas.1721629115)
- 343 14. Sudakaran S, Kost C, Kaltenpoth M. 2017 Symbiont acquisition and replacement as a
344 source of ecological innovation. *Trends Microbiol.* **25**, 375–390.
345 (doi:10.1016/j.tim.2017.02.014)
- 346 15. Sudakaran S, Retz F, Kikuchi Y, Kost C, Kaltenpoth M. 2015 Evolutionary transition in
347 symbiotic syndromes enabled diversification of phytophagous insects on an imbalanced
348 diet. *ISME J.* **9**, 1–18. (doi:10.1038/ismej.2015.75)

- 349 16. Joy JB. 2013 Symbiosis catalyses niche expansion and diversification. *Proc. R. Soc. B*
350 *Biol. Sci.* **280**, 20122820–20122820. (doi:10.1098/rspb.2012.2820)
- 351 17. Garamszegi LZ, editor. 2014 *Modern Phylogenetic Comparative Methods and Their*
352 *Application in Evolutionary Biology*. Berlin, Heidelberg: Springer Berlin Heidelberg.
353 (doi:10.1007/978-3-662-43550-2)
- 354 18. Douglas AE. 1998 Nutritional interactions in insect-microbial symbioses: aphids and their
355 symbiotic bacteria *Buchnera*. *Annu. Rev. Entomol.* **43**, 17–37.
356 (doi:10.1146/annurev.ento.43.1.17)
- 357 19. Dolling WR. 1991 *The Hemiptera*. London, UK: The Natural History Museum
358 Publications.
- 359 20. Cryan JR, Urban JM. 2012 Higher-level phylogeny of the insect order Hemiptera: is
360 Auchenorrhyncha really paraphyletic? *Syst. Entomol.* **37**, 7–21. (doi:10.1111/j.1365-
361 3113.2011.00611.x)
- 362 21. Li H, Leavengood JM, Chapman EG, Burkhardt D, Song F, Jiang P, Liu J, Zhou X, Cai
363 W. 2017 Mitochondrial phylogenomics of Hemiptera reveals adaptive innovations driving
364 the diversification of true bugs. *Proc. R. Soc. B Biol. Sci.* **284**, 20171223.
365 (doi:10.1098/rspb.2017.1223)
- 366 22. Moran NA, Tran P, Gerardo NM. 2005 Symbiosis and insect diversification: an ancient
367 symbiont of sap-feeding insects from the bacterial phylum Bacteroidetes. *Appl. Environ.*
368 *Microbiol.* **71**, 8802–8810. (doi:10.1128/AEM.71.12.8802-8810.2005)
- 369 23. Bennett GM, Moran N a. 2013 Small, smaller, smallest: the origins and evolution of
370 ancient dual symbioses in a phloem-feeding insect. *Genome Biol. Evol.* **5**, 1675–1688.
371 (doi:10.1093/gbe/evt118)

- 372 24. Gonella E *et al.* 2011 Bacterial endosymbiont localization in *Hyalesthes obsoletus*, the
373 insect vector of Bois Noir in *Vitis vinifera*. *Appl. Environ. Microbiol.* **77**, 1423–1435.
374 (doi:10.1128/AEM.02121-10)
- 375 25. McCutcheon JP, Moran NA. 2010 Functional convergence in reduced genomes of
376 bacterial symbionts spanning 200 My of evolution. *Genome Biol. Evol.* **2**, 708–718.
377 (doi:10.1093/gbe/evq055)
- 378 26. Noda H, Watanabe K, Kawai S, Yukuhiro F, Miyoshi T, Tomizawa M, Koizumi Y, Nikoh
379 N, Fukatsu T. 2012 Bacteriome-associated endosymbionts of the green rice leafhopper
380 *Nephotettix cincticeps* (Hemiptera: Cicadellidae). *Appl. Entomol. Zool.* **47**, 217–225.
381 (doi:10.1007/s13355-012-0110-1)
- 382 27. Muller HJ. 1962 Neuere vorstellungen uber verbreitung und phylogenie der
383 endosymbiosen der zikaden. *Zeitschrift fur Morphol. und oekologie der Tiere* **51**, 190–
384 210. (doi:10.1007/BF00409635)
- 385 28. Koga R, Bennett GM, Cryan JR, Moran NA. 2013 Evolutionary replacement of obligate
386 symbionts in an ancient and diverse insect lineage. *Environ. Microbiol.* **15**, 2073–2081.
387 (doi:10.1111/1462-2920.12121)
- 388 29. Mao M, Yang X, Poff K, Bennett G. 2017 Comparative genomics of the dual-obligate
389 symbionts from the treehopper, *Entylia carinata* (Hemiptera: Membracidae), provide
390 insight into the origins and evolution of an ancient symbiosis. *Genome Biol. Evol.* **9**,
391 1803–1815. (doi:10.1093/gbe/evx134)
- 392 30. McCutcheon JP, McDonald BR, Moran NA. 2009 Origin of an alternative genetic Code in
393 the extremely small and GC-rich genome of a bacterial symbiont. *PLoS Genet.* **5**,
394 e1000565. (doi:10.1371/journal.pgen.1000565)

- 395 31. McCutcheon JP, Moran NA. 2007 Parallel genomic evolution and metabolic
396 interdependence in an ancient symbiosis. *Proc. Natl. Acad. Sci.* **104**, 19392–19397.
397 (doi:10.1073/pnas.0708855104)
- 398 32. Kobińska M, Michalik A, Walczak M, Szklarzewicz T. 2018 Dual “bacterial-fungal”
399 symbiosis in Deltocephalinae leafhoppers (Insecta, Hemiptera, Cicadomorpha:
400 Cicadellidae). *Microb. Ecol.* **75**, 771–782. (doi:10.1007/s00248-017-1075-y)
- 401 33. Matsuura Y, Moriyama M, Łukasik P, Vanderpool D, Tanahashi M, Meng X-Y,
402 McCutcheon JP, Fukatsu T. 2018 Recurrent symbiont recruitment from fungal parasites in
403 cicadas. *Proc. Natl. Acad. Sci.* **115**, E5970–E5979. (doi:10.1073/pnas.1803245115)
- 404 34. Noda H, Nakashima N, Koizumi M. 1995 Phylogenetic position of yeast-like symbiotes of
405 rice planthoppers based on partial 18S rDNA Sequences. *Insect Biochem. Mol. Biol.* **25**,
406 639–646. (doi:10.1016/0965-1748(94)00107-S)
- 407 35. Urban JM, Cryan JR. 2012 Two ancient bacterial endosymbionts have coevolved with the
408 planthoppers (Insecta: Hemiptera: Fulgoroidea). *BMC Evol. Biol.* **12**, 87.
409 (doi:10.1186/1471-2148-12-87)
- 410 36. de Queiroz A, Gatesy J. 2007 The supermatrix approach to systematics. *Trends Ecol.*
411 *Evol.* **22**, 34–41. (doi:10.1016/j.tree.2006.10.002)
- 412 37. Cryan JR. 2005 Molecular phylogeny of Cicadomorpha (Insecta: Hemiptera: Cicadoidea,
413 Cercopoidea and Membracoidea): adding evidence to the controversy. *Syst. Entomol.* **30**,
414 563–574. (doi:10.1111/j.1365-3113.2004.00285.x)
- 415 38. Urban JM, Cryan JR. 2009 Entomologically famous, evolutionarily unexplored: The first
416 phylogeny of the lanternfly family Fulgoridae (Insecta: Hemiptera: Fulgoroidea). *Mol.*
417 *Phylogenet. Evol.* **50**, 471–484. (doi:10.1016/j.ympev.2008.12.004)

- 418 39. Pearse WD, Purvis A. 2013 phyloGenerator: an automated phylogeny generation tool for
419 ecologists. *Methods Ecol. Evol.* **4**, 692–698. (doi:10.1111/2041-210X.12055)
- 420 40. Katoh K, Standley DM. 2013 MAFFT Multiple Sequence Alignment Software Version 7:
421 Improvements in Performance and Usability. *Mol. Biol. Evol.* **30**, 772–780.
422 (doi:10.1093/molbev/mst010)
- 423 41. Larsson A. 2014 AliView: A fast and lightweight alignment viewer and editor for large
424 datasets. *Bioinformatics* **30**, 3276–3278. (doi:10.1093/bioinformatics/btu531)
- 425 42. Stamatakis A. 2014 RAxML version 8: A tool for phylogenetic analysis and post-analysis
426 of large phylogenies. *Bioinformatics* **30**, 1312–1313. (doi:10.1093/bioinformatics/btu033)
- 427 43. Sanderson MJ. 2003 r8s: inferring absolute rates of molecular evolution and divergence
428 times in the absence of a molecular clock. *Bioinformatics* **19**, 301–302.
429 (doi:10.1093/bioinformatics/19.2.301)
- 430 44. Zahniser JN, Dietrich CH. 2010 Phylogeny of the leafhopper subfamily Deltocephalinae
431 (Hemiptera: Cicadellidae) based on molecular and morphological data with a revised
432 family-group classification. *Syst. Entomol.* **35**, 489–511. (doi:10.1111/j.1365-
433 3113.2010.00522.x)
- 434 45. Redak RA, Purcell AH, Lopes JRS, Blua MJ, Mizell III RF, Andersen PC. 2004 The
435 biology of xylem fluid-feeding insect vectors of *Xylella fastidiosa* and their relation to
436 disease epidemiology. *Annu. Rev. Entomol.* **49**, 243–270.
437 (doi:10.1146/annurev.ento.49.061802.123403)
- 438 46. Dietrich C. 2003 Auchenorrhyncha (Cicadas, spittlebugs, leafhoppers, treehoppers and
439 planthoppers). In *Encyclopedia of Insects*, pp. 66–74. New York: Academic Press.
- 440 47. Young DA. 1968 Taxonomic Study of the Cicadellinae (Homoptera: Cicadellidae) pt. 1:

- 441 Proconiini. *Bull. United States Natl. Museum* , 1–287. (doi:10.5479/si.03629236.261.1)
- 442 48. Cryan JR, Svenson GJ. 2010 Family-level relationships of the spittlebugs and froghoppers
443 (Hemiptera: Cicadomorpha: Cercopoidea). *Syst. Entomol.* **35**, 393–415.
444 (doi:10.1111/j.1365-3113.2009.00520.x)
- 445 49. Pagel M. 1994 Detecting correlated evolution on phylogenies: a general method for the
446 comparative analysis of discrete characters. *Proc. R. Soc. B Biol. Sci.* **255**, 37–45.
447 (doi:10.1098/rspb.1994.0006)
- 448 50. Beaulieu JM, O’Meara BC, Donoghue MJ. 2013 Identifying hidden rate changes in the
449 evolution of a binary morphological character: the evolution of plant habit in campanulid
450 angiosperms. *Syst. Biol.* **62**, 725–737. (doi:10.1093/sysbio/syt034)
- 451 51. Yang Z. 2006 *Computational Molecular Evolution*. Oxford, UK: Oxford University Press.
- 452 52. Werner GDA, Cornwell WK, Spreti JI, Kattge J, Kiers ET. 2014 A single evolutionary
453 innovation drives the deep evolution of symbiotic N₂-fixation in angiosperms. *Nat.*
454 *Commun.* **5**, 4087. (doi:10.1038/ncomms5087)
- 455 53. Novotny V, Wilson MR. 1997 Why are there no small species among xylem-sucking
456 insects? *Evol. Ecol.* **11**, 419–437. (doi:10.1023/A:1018432807165)
- 457 54. Revell LJ. 2012 phytools: An R package for phylogenetic comparative biology (and other
458 things). *Methods Ecol. Evol.* **3**, 217–223. (doi:10.1111/j.2041-210X.2011.00169.x)
- 459 55. Paterno GB, Penone C, Werner GDA. 2018 sensiPhy: An r-package for sensitivity
460 analysis in phylogenetic comparative methods. *Methods Ecol. Evol.* **9**, 1461–1467.
461 (doi:10.1111/2041-210X.12990)
- 462 56. Hansen AK, Moran N a. 2014 The impact of microbial symbionts on host plant utilization
463 by herbivorous insects. *Mol. Ecol.* **23**, 1473–96. (doi:10.1111/mec.12421)

- 464 57. Rolshausen G, Dal Grande F, Sadowska-Deś AD, Otte J, Schmitt I. 2018 Quantifying the
465 climatic niche of symbiont partners in a lichen symbiosis indicates mutualist-mediated
466 niche expansions. *Ecography (Cop.)*. **41**, 1380–1392. (doi:10.1111/ecog.03457)
- 467 58. McFall-Ngai M *et al.* 2013 Animals in a bacterial world, a new imperative for the life
468 sciences. *Proc. Natl. Acad. Sci.* **110**, 3229–3236. (doi:10.1073/pnas.1218525110)
- 469 59. Johnson KP *et al.* 2018 Phylogenomics and the evolution of hemipteroid insects. *Proc.*
470 *Natl. Acad. Sci.* **2018**, 201815820. (doi:10.1073/pnas.1815820115)
- 471 60. Raven JA. 1983 Phytophages of xylem and phloem: a comparison of animal and plant
472 sap-feeders. *Adv. Ecol. Res.* **13**, 135–234. (doi:10.1016/S0065-2504(08)60109-9)
- 473 61. Ankrah NYD, Chouaia B, Douglas AE. 2018 The cost of metabolic interactions in
474 symbioses between insects and bacteria with reduced genomes. *MBio* **9**, 1–15.
475 (doi:10.1128/mBio.01433-18)
- 476 62. Wernegreen JJ. 2012 Mutualism meltdown in insects: bacteria constrain thermal
477 adaptation. *Curr. Opin. Microbiol.* **15**, 255–62. (doi:10.1016/j.mib.2012.02.001)
- 478 63. Moran N a, Yun Y. 2015 Experimental replacement of an obligate insect symbiont. *Proc.*
479 *Natl. Acad. Sci.* **112**, 2093–2096. (doi:10.1073/pnas.1420037112)
- 480 64. Bennett GM, Moran NA. 2015 Heritable symbiosis: the advantages and perils of an
481 evolutionary rabbit hole. *Proc. Natl. Acad. Sci.* **112**, 10169–10176.
482 (doi:10.1073/pnas.1421388112)
- 483 65. Campbell MA, Łukasik P, Simon C, McCutcheon JP. 2017 Idiosyncratic genome
484 degradation in a bacterial endosymbiont of periodical cicadas. *Curr. Biol.* **27**, 3568–
485 3575.e3. (doi:10.1016/j.cub.2017.10.008)
- 486 66. Moran NA. 1996 Accelerated evolution and Muller’s ratchet in endosymbiotic bacteria.

- 487 *Proc. Natl. Acad. Sci.* **93**, 2873–2878. (doi:10.1073/pnas.93.7.2873)
- 488 67. Douglas AE. 2015 Multiorganismal insects: diversity and function of resident
489 microorganisms. *Annu. Rev. Entomol.* **60**, 17–34. (doi:10.1146/annurev-ento-010814-
490 020822)
- 491 68. Meseguer AS, Manzano-Marín A, Coeur d'Acier A, Clamens A-L, Godefroid M,
492 Jousselein E. 2017 Buchnera has changed flatmate but the repeated replacement of co-
493 obligate symbionts is not associated with the ecological expansions of their aphid hosts.
494 *Mol. Ecol.* **26**, 2363–2378. (doi:10.1111/mec.13910)
- 495 69. van den Bosch TJM, Welte CU. 2017 Detoxifying symbionts in agriculturally important
496 pest insects. *Microb. Biotechnol.* **10**, 531–540. (doi:10.1111/1751-7915.12483)
- 497
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499 **Figure Legend**

500

501 **Figure 1: Analysed phylogeny, trait data and key ancestral state reconstructions of the Auchenorrhyncha.** Vertical bands at
502 the tips of the 142-genus phylogeny indicate – from left to right – primary endosymbiosis status, host diet and companion symbiosis
503 status. In all bands, missing data (i.e. traits could not be estimated reliably) are represented in white. Branch colours indicate inferred
504 companion symbiont status (full model shown in figure S5). Stars indicate the branch along which transitions from the ancestral
505 xylem-feeding state towards either phloem feeding (filled stars), or parenchyma feeding (open star) are inferred to have occurred
506 (full model in figures S1 and S2). The inset figure shows the model of correlated evolution among host diet (xylem vs non-xylem
507 feeding) and primary endosymbiosis status (presence and absence), indicating transition rates between the four potential
508 combinations of these two binary traits. Transitions rates are multiplied by hundred, and can thus be interpreted as number of
509 transitions per lineage per 100 million years. Transitions that are possible under the model, but were inferred to be non-existent,
510 are indicated with dashed arrows. The corresponding ancestral state reconstruction is represented on the phylogeny by pie charts.
511 From the ancestral state of feeding on xylem and associating with *Sulcia* (orange), primary symbiosis loss (transition towards
512 brown) occurred five times and was always preceded by a diet shift away from xylem feeding (purple), sometimes tens of millions
513 of years earlier.