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Evaluating satellite estimates of particulate backscatter in the global open ocean using autonomous profiling floats

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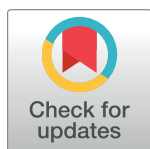
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COMMUNITY PAGE

The *Tara* Pacific expedition—A pan-ecosystemic approach of the “-omics” complexity of coral reef holobionts across the Pacific Ocean

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OPEN ACCESS

Citation: Planes S, Allemand D, Agostini S, Banaigs B, Boissin E, Boss E, et al. (2019) The *Tara* Pacific expedition—A pan-ecosystemic approach of the “-omics” complexity of coral reef holobionts across the Pacific Ocean. *PLoS Biol* 17(9): e3000483. <https://doi.org/10.1371/journal.pbio.3000483>

Published: September 23, 2019

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Funding: We are keen to thank the commitment of the people and the following institutions for their financial and scientific support that made this singular expedition possible: CNRS, PSL, CSM, EPHE, Genoscope/CEA, Inserm, Université Cote d’Azur, ANR, agnès b., UNESCO-IOC, the Veolia Environment Foundation, Région Bretagne, Serge Ferrari, Billerudkorsnas, Amerisource Bergen Company, Lorient Agglomération, Oceans by Disney, the Prince Albert II de Monaco Foundation, L’Oréal, Biotherm, France Collectivités, Kankyo Station, Fonds Français pour l’Environnement Mondial (FFEM), Etienne BOURGOIS, and the Tara Ocean Foundation teams and crew. Tara Pacific would not exist without the continuous support of the participating institutes. This study has been conducted using E.U. Copernicus Marine Service Information and Mercator Ocean products. We

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Abstract

Coral reefs are the most diverse habitats in the marine realm. Their productivity, structural complexity, and biodiversity critically depend on ecosystem services provided by corals that are threatened because of climate change effects—in particular, ocean warming and acidification. The coral holobiont is composed of the coral animal host, endosymbiotic dinoflagellates,

acknowledged funding from the Investissement d'avenir projects France Génomique (ANR-10-INBS-09) and OCEANOMICS (ANR-11-BTBR-0008). RVT was funded by a Dimensions of Biodiversity NSF grant (#1442306) for this work. SS is supported by the ETH Zurich and Helmut Horten Foundation. FL is supported by Sorbonne Université, Institut Universitaire de France, and the Fondation CA-PCA. Finally, we thank the ANR for funding the project CORALGENE, which will support the work the Tara Pacific program. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abbreviations: CBD, Convention on Biological Diversity; FAIR, findable, accessible, interoperable, reusable; GLODAP, Global Ocean Data Analysis Project; MODIS, Moderate Resolution Imaging Spectroradiometer; MS, mass spectrometry; NASA, National Aeronautics and Space Administration; NMR, nuclear magnetic resonance; OTU, operational taxonomic unit; SST, sea surface temperature.

Provenance: Not commissioned; externally peer reviewed

associated viruses, bacteria, and other microeukaryotes. In particular, the mandatory photosymbiosis with microalgae of the family Symbiodiniaceae and its consequences on the evolution, physiology, and stress resilience of the coral holobiont have yet to be fully elucidated. The functioning of the holobiont as a whole is largely unknown, although bacteria and viruses are presumed to play roles in metabolic interactions, immunity, and stress tolerance. In the context of climate change and anthropogenic threats on coral reef ecosystems, the *Tara Pacific* project aims to provide a baseline of the “-omics” complexity of the coral holobiont and its ecosystem across the Pacific Ocean and for various oceanographically distinct defined areas. Inspired by the previous *Tara Oceans* expeditions, the *Tara Pacific* expedition (2016–2018) has applied a pan-ecosystemic approach on coral reefs throughout the Pacific Ocean, drawing an east–west transect from Panama to Papua New Guinea and a south–north transect from Australia to Japan, sampling corals throughout 32 island systems with local replicates. *Tara Pacific* has developed and applied state-of-the-art technologies in very-high-throughput genetic sequencing and molecular analysis to reveal the entire microbial and chemical diversity as well as functional traits associated with coral holobionts, together with various measures on environmental forcing. This ambitious project aims at revealing a massive amount of novel biodiversity, shedding light on the complex links between genomes, transcriptomes, metabolomes, organisms, and ecosystem functions in coral reefs and providing a reference of the biological state of modern coral reefs in the Anthropocene.

Introduction

The 20th century has seen the earth enter into the now widely called Anthropocene [1]. Anthropogenically induced changes are happening on both global and local scales and are altering the physiology of organisms and ecosystems by modifying the entire earth's physical, chemical, and biological processes [2]. Among marine ecosystems, coral reefs have the unfortunate privilege of being highly sensitive to these environmental modifications [3]. In particular, the thermally mediated process of coral bleaching—i.e., the loss of the obligate photosynthetic microalgal endosymbionts—is increasingly decimating corals. Projections estimate that approximately 25% of reefs have already been lost, and up to 99% will be threatened and dramatically transformed by 2050 [4]. Even though they cover only approximately 0.2% of the ocean's surface [5], coral reefs harbor approximately 30% of ocean biodiversity [6], providing ecological services (fisheries, tourism, coastal protection) to nearly 1 billion people [7], and are estimated to be worth USD 30 billion per year [8].

The late 20th century has also seen our world entering into the “-omics” revolution sparked by the development of high-throughput analyses of DNA, RNA, proteins, and metabolites. This revolution has changed our approach to investigate organisms, and it is about to change the descriptions of ecosystems as genes-to-ecosystem modeling improves [9]. It will advance our capabilities to investigate the biodiversity and functioning of our oceans in a holistic way. Holistic approaches, interrogating components across the various levels of organization of an ecosystem, appear accessible today, as exemplified in the *Tara Oceans* project [10], which continues to decipher biodiversity and structural networks across all plankton organisms in the ocean [11].

Tara Pacific is a unique scientific expedition inspired by earlier maritime explorations that uncovered the uncharted territories of marine biodiversity. Coral reef research

started with the contribution of Charles Darwin during “the voyage of the *Beagle*” (1831–1836), when Darwin explored many reefs in the Indian and Pacific Oceans and established his biophysical theory of the formation of coral reefs and atolls [12]. He also highlighted the paradoxical high diversity of reef organisms living in a nutrient-poor system. Later, the Great Barrier Reef Expedition (1928–1929) led by Charles Yonge was a landmark in coral reef research, as it established the scientific basis of coral physiology (nutrition, symbiosis, growth, etc.). The “polypifers,” later called coral polyps, were then demonstrated to host a large population of organisms (protists, viruses, bacteria, archaea, and all sorts of unicellular and multicellular eukaryotes), which are intimately involved in the physiology of the coral animal host in a complex and still mysterious system named the holobiont [13].

The *Tara* Pacific expedition has built on these early tracks, applying the most recent technologies to map the “-omics” complexity of the coral holobiont within its ecosystem and across the Pacific Ocean. Through the exploration of marine biodiversity at scales spanning from organisms to genes to biomolecules, *Tara* Pacific is undertaking the first pan-ecosystemic study of coral reef diversity across an entire ocean basin (Fig 1). Given that a holistic approach integrating all components of the reef biota is unrealistic, the *Tara* Pacific expedition focuses its approach on widely distributed coral and fish holobionts and their contextual biological (plankton) and physicochemical environment, including modifications in the context of global changes (Fig 2).

The geographic and sequencing extent of *Tara* Pacific will bring an exceptionally comprehensive description of coral holobionts, together with the surrounding fish and plankton biota. Such unprecedented sequencing coverage and depth will place coral reefs among the first ecosystems with a comprehensive description of their hologenomic diversity (i.e., the totality of genomes making up the coral holobiont) across their natural environments. We will go beyond the mere description of species composition to investigate in detail the gene content and gene expression of these communities, as well as the interconnectedness with their hosts and habitats. Broad emerging patterns will then be accessible through large-scale data mining and network analyses.

The coral holobiont

The coral host

Scleractinian corals evolved over 450 million years ago [14] and belong to the phylum Cnidaria, animals located close to the root of the metazoans. Because of their place in the early Eumetazoa and as a sister group of bilaterians, the cnidarians allow interesting evolutionary analyses regarding the origins of animal complexity and the coevolution of early animals with microbes. Shinzato and colleagues [15] were the first to show that the coral genome is as complex as the one of vertebrates and that it has retained many ancestral genes lost in other lineages. Subsequent studies confirmed the deep genomic differences between coral taxa [16]. Moreover, the exceptional longevity of corals, reaching several hundred to thousands of years, makes them attractive emerging models for aging studies.

The coral endosymbiont microalgae

Reef corals host symbiotic unicellular algae of the family Symbiodiniaceae inside their endodermal cells [17], raising intriguing questions on animal–plant interactions, photosymbiosis, coevolution, and speciation [18]. The divergence and genetic variation within and between populations of coral species is also investigated, allowing us to firmly ground the exact taxonomic status of each coral host [19].

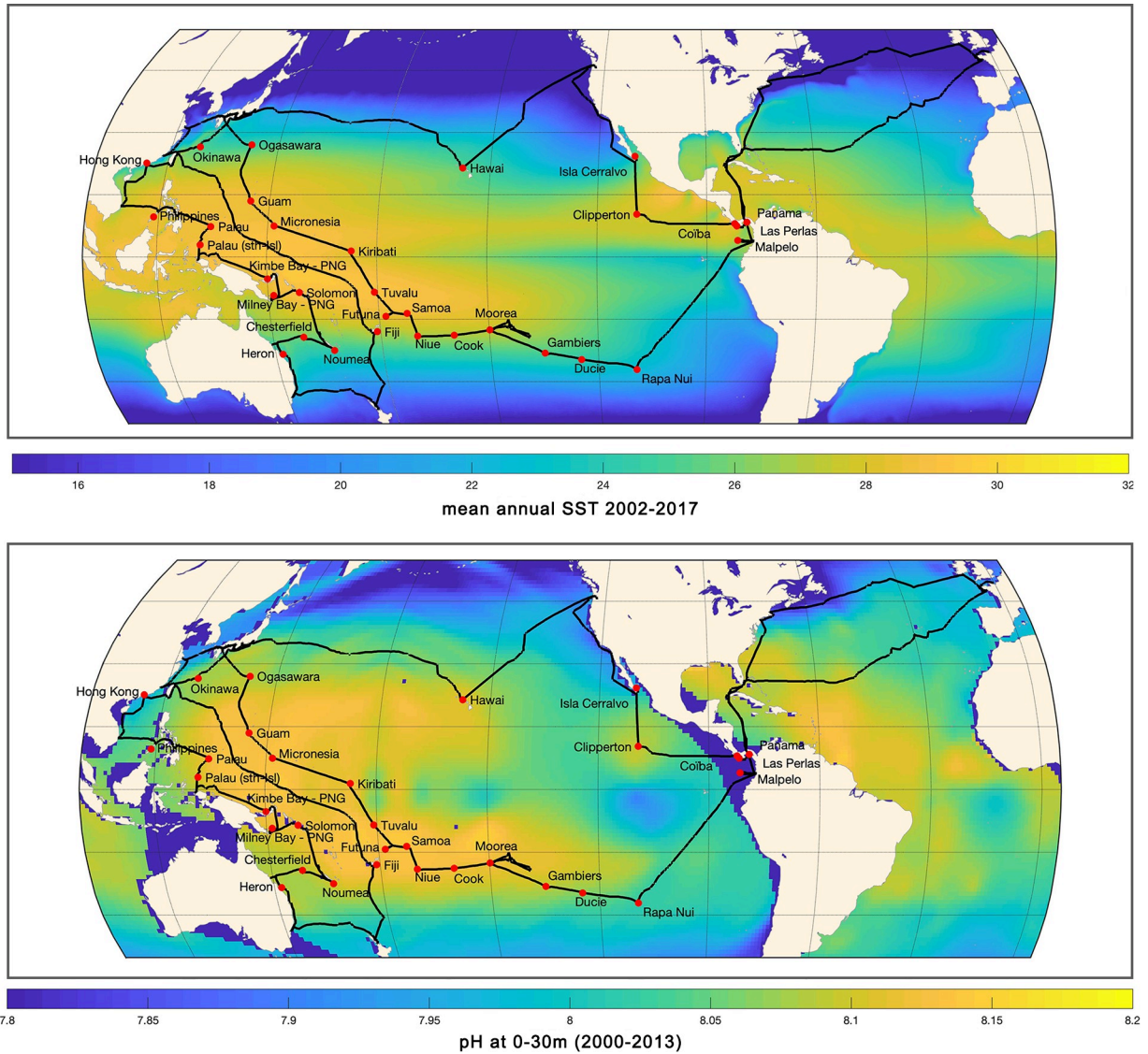


Fig 1. Map showing the route of the *Tara Pacific* expedition and the sampling sites (red spots) throughout the Pacific Ocean (July 2016 to October 2018) as well as the mean annual SST (top) and pH (bottom). Global annual SSTs were extracted from the MODIS-Aqua satellite and correspond to global mapped climatologies of the period from 2002 to 2018 (NASA Goddard Space Flight Center), whereas pH values originate from the GLODAPv2 database [59, 60], with mean data displayed corresponding to the 0–30 m depths between 2000 and 2013. GLODAP, Global Ocean Data Analysis Project; MODIS, Moderate Resolution Imaging Spectroradiometer; NASA, National Aeronautics and Space Administration; PNG, Papua New Guinea; SST, sea surface temperature.

<https://doi.org/10.1371/journal.pbio.3000483.g001>

The coral microbiome

In addition to the Symbiodiniaceae, corals host diverse groups of microorganisms, bacteria, archaea, fungi, and protists, together forming the so-called coral holobiont [13]. The importance of the microbiome in the normal steady-state functioning of corals or during environmental stress is not well understood, but recent research suggests that microbiome structure aligns with and contributes to stress resilience [20, 21, 22] (in particular, bleaching [23, 24]), as seen in humans [25]. Recent work provides evidence that corals have coevolved with certain groups of microbes [26–28], but the exact nature of these host–microbe symbioses and their involvement in coral resilience remains unclear. Therefore, *Tara Pacific* will draw, using

Coral Health and Resilience

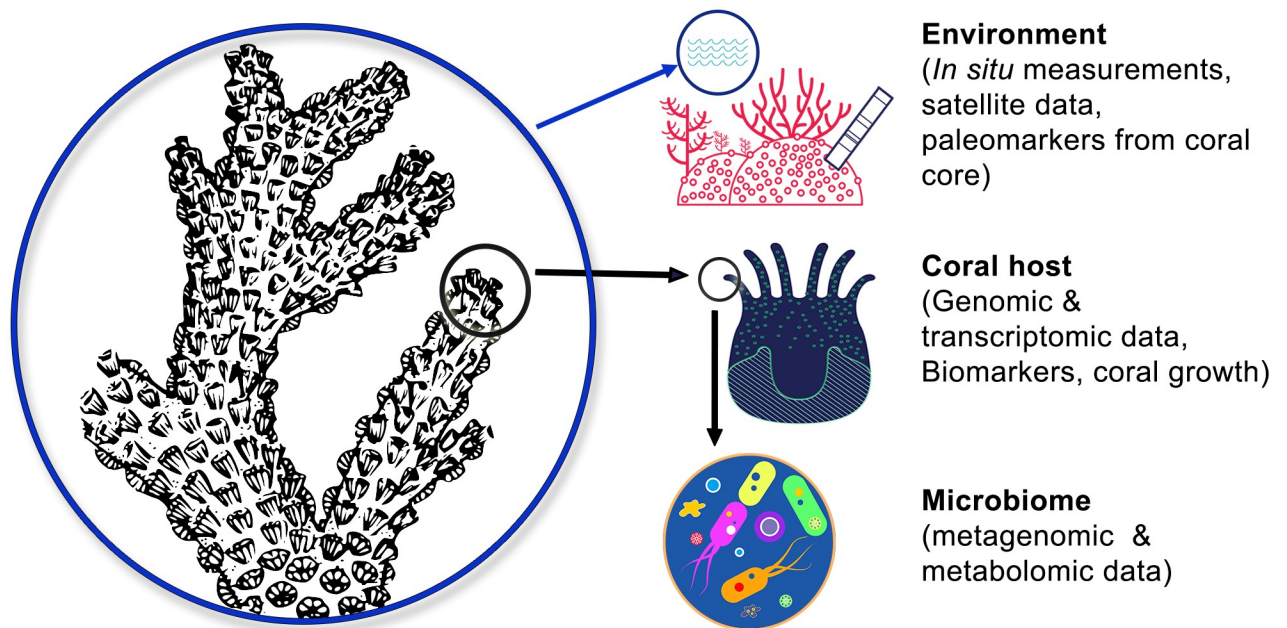


Fig 2. Schematic representation of the frame and major goals of Tara Pacific, which is investigating jointly the coral, microbiome, and environment.

<https://doi.org/10.1371/journal.pbio.3000483.g002>

marker gene and metagenomic/metatranscriptomic approaches, a nearly exhaustive census of the microbial diversity associated with targeted coral taxa, allowing the understanding of the major drivers of the microbiome diversity and its role in coral health, resilience, and evolution.

The coral virome

Viruses infect all known cellular organisms and are the most abundant biological entities on our planet [29]. Yet even the basic diversity of viruses associated with marine species and habitats is shockingly unknown. This is particularly true in tropical reef systems, where research on viral abundance, diversity, and dynamics is still in its infancy (for review, see [30]). *Tara Pacific* is using metagenomics and microscopy to interrogate the diversity of viruses across Pacific waters and within the foundation species of coral reefs.

Research objectives of *Tara Pacific*

Our goal is to unveil the entire organismal diversity of eukaryotes, prokaryotes, and viruses associated with the targeted coral and fish holobionts and their surrounding waters and to assess variation across the explored ecological and geographical gradients. The main objectives of *Tara Pacific* are as follows:

- Draw a near-exhaustive census of the biodiversity composing and surrounding coral holobionts. This includes the coral hosts, their endosymbiotic microalgae (zooxanthellae of the family Symbiodiniaceae), and the associated microeukaryotes, bacteria, archaea, and viruses found either as endo- or exosymbionts, or in the plankton in surrounding water, using a metabarcoding approach.

- Determine the spatial patterns of the holobionts and the environmental diversity within coral reefs throughout the Pacific Ocean and derive large-scale biogeographic patterns to be compared with macrofauna patterns of diversity (i.e., the Pacific biodiversity gradient).
- Compare metatranscriptomes and metagenomes of coral reefs throughout the Pacific Ocean and elucidate the contribution of microbial diversity to local versus basin-scale adaptation in response to climate change. Importantly, the entire plankton community surrounding the corals and the islands has been sampled and integrated with physicochemical contextual data, providing critical context to advance our understanding of the environmental contribution to coral reef resilience.
- Investigate divergence and standing genetic variation within and between populations of the targeted coral species and their Symbiodiniaceae symbionts across the Pacific Ocean basin via marker gene barcoding as well as genome and transcriptome sequencing (population genomics using SNPs). Extensive metadata resources collected during *Tara* Pacific will allow the coral holobiont composition to be modeled, taking environmental and coral trait data into account (e.g., temperature, pH, bleaching, symbiont acquisition, reproduction cycle, etc.) to identify drivers of adaptation/selection or acclimatization, providing insights into coral stress resilience.
- Investigate the health status of corals by measuring growth parameters of the recent coral skeleton and stress biomarkers (antioxidant capacity, apoptosis, stress response pathways, transcriptomic signatures) as well as by determining the telomere status of each sample (telomeric DNA length and damage as potential proxies for stress). These data will be studied in relation to holobiont biodiversity, transcriptomic and metabolomics data, modern and/or historical environmental parameters, and stresses determined from a geochemical analysis of the coral skeleton.
- Generate a holistic coral metabolome for coral species as a foundation to the identification of the links between holobiont metabolism and prevailing environmental conditions.
- Identify how the environment (physical, chemical, biogeochemical, and biological) is influencing coral holobiont diversity, physiology, and evolution using an extensive compilation of environmental conditions.

A unique, pan-ecosystemic sampling strategy

The Pacific Ocean covers approximately one-third of the earth's surface, with nearly 25,000 islands, most of which harbor coral reefs. *Tara* Pacific equipped the schooner *Tara* to explore 32 islands across the entire Pacific over a period of 2.5 years (Fig 1). The route of the *Tara* schooner was chosen to maximize the number of visited remote islands and atolls and to perform the widest possible comparative survey, from the equator to the temperate and more acidic regions, encompassing most of the environmental range where scleractinian coral species can live. It also covers a biodiversity gradient, from the low diversity present in the eastern Pacific reefs to the highly diverse western Pacific "warm pool" [14]. This sampling strategy also encompasses a wide variety of environments, from high temperature/low seasonality to low temperature/high seasonality, and also a full range of trophic status and physicochemical environmental parameters that affect coral reef ecosystems (sea surface temperature [SST], pH, nutrients, lights, pollutants, etc.).

Across its entire route, the *Tara* Pacific expedition targeted two species of scleractinian corals (Cnidaria, Anthozoa: *Pocillopora meandrina* and *Porites lobata*), one species of hydrocoral

(Cnidaria, Hydrozoa: *Millepora platyphylla*), and two species of reef fish (*Acanthurus triostegus* and *Zanclus cornutus*) (Fig 3). The chosen taxa are among the few species that occur across most of the Pacific Ocean and are usually abundant on reefs. Around each of the 32 islands, three sites were sampled, collecting 10 colonies of each coral species and 5–10 individuals of each fish species, as well as water samples (see below) (Fig 3). Coral fragments and fish samples were preserved immediately on board, using specific buffers (e.g., DNA/RNA shield for genomics, glutaraldehyde for microscopy, etc.), and/or flash frozen for subsequent laboratory analyses (e.g., metabolomics, biomarkers, etc.).

In addition, seawater biogeochemistry and plankton microbiomes (viruses to zooplankton) were assessed in oceanic waters upstream and downstream of each island, as well as from water above each site (i.e., “surface water”) and surrounding two colonies of sampled *P. meandrina* corals at each site (i.e., “coral-surrounding water”). Underwater propeller-driven plankton nets (for surface water) and water pumping systems (for coral-surrounding water) were developed to collect plankton on reef sites, whereas the preparation of plankton samples into different size classes (<0.2, 0.2–3, 3–20, 20–2000 μm) and their preservation followed the *Tara* Oceans protocols [31] in order to maximize comparability and integration of data. Samples will be analyzed using high-throughput imaging and genomic techniques.

Finally, key physical, chemical, and biological parameters were measured continuously from surface water and from aerosols using specific instruments installed on board (e.g., thermosalinograph, mass- and spectrophotometers), as well as samples for imaging and genetic analysis of aerosol particles. Environmental context will also be obtained from satellite images and operational oceanographic products from the European Copernicus Marine Service and Mercator Ocean.

Tara Pacific samples and data resources for the immediate and long-term future

Tara Pacific endorses the findable, accessible, interoperable, reusable (FAIR) principles for scientific data management [32] as well as ethical and responsible use of data. With a few exceptions, all physical samples of coral, plankton, fish, and sediments were preserved on board *Tara* and sent back to partner laboratories for subsequent analyses. Controlled vocabularies describing sampling devices and sample preparation protocols were used throughout the expedition to capture provenance metadata on customized log sheets, and samples were assigned unique identifiers to facilitate their traceability. Legal documents regulating the collection, export, and import of samples, as well as links to the Convention on Biological Diversity’s (CBD) access and benefit-sharing clearing house (<https://absch.cbd.int/>) are in the process of being provided for each sample as a first step toward ethical and responsible use of the *Tara* Pacific data. The detailed registry of all samples, including their provenance and environmental and legal context, is curated manually using simple semantics that enable machine- and human-readable data discovery services. Sequencing data will be deposited at the free, open-access European Nucleotides Archive (<https://www.ebi.ac.uk/ena>); environmental data are deposited at the free, open-access PANGAEA database (<https://www.pangaea.de/>); and both archives will be interlinked *via* the sample registry available online at BioSamples (<https://www.ebi.ac.uk/biosamples/>). Metabolomic data (mass spectrometry [MS] and nuclear magnetic resonance [NMR]) and their annotations will also be accessible through the Metabolights portal (<https://www.ebi.ac.uk/metabolights/>). The submission of data in a relational, open-access, updated, and cured database is critical and necessary for our aim to establish a reference of the biological state of coral reefs in the Anthropocene for the broader research community.

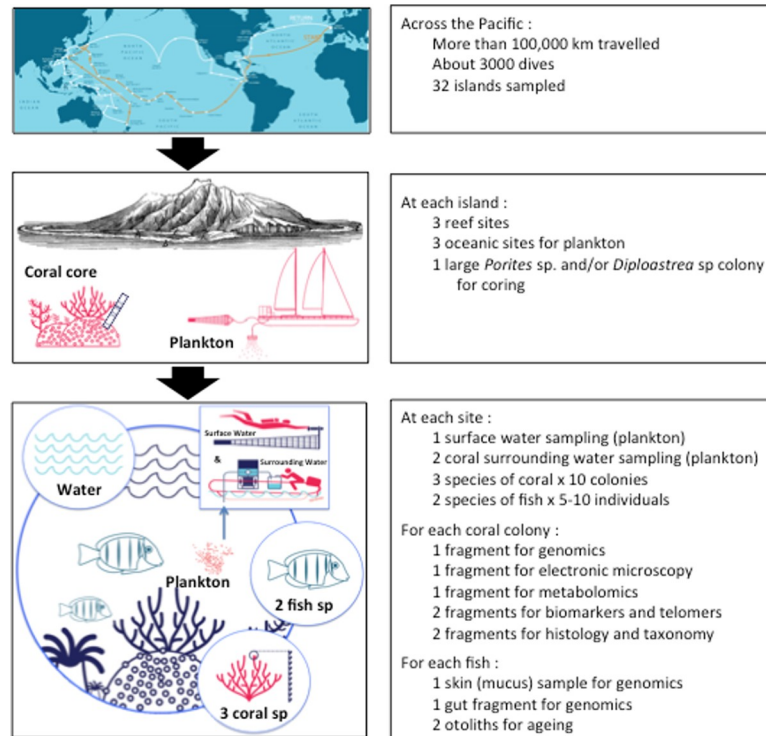


Fig 3. Schematic representation of the sampling design allowing the comparison of various components of coral reef ecosystems. Using the developed protocol, we collected 2,500 oceanic samples (32 islands \times 3 oceanic sites \times 5 size fractions \times 2–5 protocols), 7,500 coral-surrounding and surface water samples (32 islands \times 3 coral sites \times 3 environments \times 5 size fractions \times 2–5 protocols), 40 coral core samples (32 islands \times 1 *Porites* sp. and/or *Diploastrea* sp \times 1–2 core samples), 20,160 coral fragments (32 islands \times 3 coral sites \times 3 species of corals \times 10 colonies \times 7 protocols), and 9,600 fish tissue samples (32 islands \times 3 coral sites \times 2 species of fish \times 5–10 individuals \times 5 protocols). Taken together, the dataset comprises a total of approximately 40,000 samples. *Bottom figure copyright to Agence DATCHA/ taraexpeditions.org.*

<https://doi.org/10.1371/journal.pbio.3000483.g003>

Metagenomics and metatranscriptomics to explore the diversity and physiology of coral reef holobionts

Building upon expertise gained in the previous *Tara* Oceans project [33–36], we are applying Illumina HiSeq technology to sequence a series of barcodes designed recently to explore the diversity of bacteria and archaea (V4–V5 region of the nuclear 16S rRNA gene [37]), eukaryotes (V9 region of nuclear 18S rRNA gene [33]), Symbiodiniaceae (ITS2 region of the nuclear ribosomal DNA [38–40]), and metazoan species (mitochondrial COX1 gene [41]), as well as metagenome assembly–defined viral populations [42, 43]. Barcode-specific PCRs are performed on DNA from coral tissues, coral-surrounding water, fish tissues, and surface water above the reef in order to assess interkingdom diversity associated with the holobiont at different degrees of proximity from the coral animal host, from endosymbiotic to drifting in coral-surrounding and surface waters. Biogeographic gradients will also be investigated between the studied island systems. The sequence reads are assembled into operational taxonomic units (OTUs) for bacteria, archaea, and eukaryotes and into ITS2-type profiles (using the SymPortal framework at symportal.org [40]) for Symbiodiniaceae and taxonomically annotated by comparison to reference databases. Classical numerical ecology methods are applied to employ OTU richness and abundance data to (1) assess and compare the total diversity of bacteria, archaea, and eukaryotes associated with various compartments of the different coral

holobionts; (2) compare biodiversity patterns across coral colonies, hosts, reefs, islands (geographic and Lagrangian oceanic circulation distances), and all contextual environmental parameters measured; (3) integrate the *Tara* Pacific data into the *Tara* Oceans global plankton dataset. In addition, co-occurrence graph techniques inspired from systems biology [44, 9] are used to reconstruct OTU interaction (sub)networks, and we will aim to disentangle the intrinsic (symbiotic auto-organization) from the extrinsic (environmental, historical contingencies) forces on holobiont biodiversity. Overall, this study is the first attempt to characterize coral reef and holobiont diversity across comprehensive taxonomic, ecological, and geographical scales over an entire oceanic basin. Comparison with *Tara* Oceans plankton data will also reveal which part of the coral holobiont biodiversity is found in the open ocean and thus bring fundamental information about reef connectivity, as recently shown for Symbiodiniaceae, which are found all over the plankton [34].

The sequencing of the holotranscriptomes of coral holobionts sampled along the transect will provide important genetic information to uncover ecological and evolutionary questions at both intraspecific and community levels. Two protocols are successively applied on total RNA from three specimens per species and per site. In a first step, a polyA⁺ enrichment protocol is applied to provide eukaryotic mRNA in order to study the coral and Symbiodiniaceae dual transcriptome, and in a second step, rRNA is removed from the remaining polyA⁻ fraction to sequence microbial mRNA. We are applying different metatranscriptomics protocols to capture the gene expression of the Cnidaria, Symbiodiniaceae, bacteria and archaea, other microbes, and even viruses from the same initial tissue samples [45, 46]. In addition, we are using similar analysis procedures after filtration of coral-surrounding water samples. This may potentially reveal how microbes and viruses interact with the coral host and how the coral host adapts or acclimatizes to their presence. On top of providing deep insight on gene expression networks at the holobiont level, mRNA samples will also be used to generate transcriptome-wide SNPs for the coral host, allowing in-depth characterization of the patterns of diversity, connectivity, demographic history, and local adaptation of sampled coral species. Hence, the resulting patterns will provide a solid foundation on which to base the study of partner relationships within the coral holobiont, and possibly even allow for the identification of the selective forces acting on key genes for these symbiotic associations, plus an account on standing genetic diversity within and between reefs to highlight regions of high genetic endemism and diversity important for conservation efforts.

A metabolomic approach

We will also assess the metabolome (i.e., the metabolic diversity) of the three targeted coral species (two stony and one fire coral, see above). Small molecules (i.e., specialized metabolites) are the end products of unique metabolic pathways. Most of these presumably possess an ecological role, and as such, they represent key phenotypic traits for the specimens. The broad geographical cover of the collected coral species will enable us to provide some insight into the relative contribution of environmental factors and the genetic information expressed in the metabolome by comparison with genomic data. Recent advances in analytical techniques and bioinformatics led to the development of global metabolomics approaches capable of providing an overview of the thousands of metabolites present in a minute amount of sample [47, 48]. A nontargeted metabolomics approach is applied to the three species of corals collected across the Pacific Ocean using both MS and NMR. The subsequent identification of the chemomarkers unveiled by these comparative approaches are performed using not only databases of experimental spectra but also comparison with *in silico* databases of metabolites. The results will finally contribute to the assessment of the reef health status when combined with other “-omics” data.

Identifying the interaction among environmental stressors, the holobiont, and coral resilience

A key question to be addressed by *Tara Pacific* is how reef corals change geographically and how corals locally adapt or acclimate to environmental changes and increasing stress. This question has been rarely studied at the scale of a complete ocean and across multiple populations under consideration of phenotypic traits [49]. The physiological status of corals will be assessed by measuring the parameters of coral growth such as density, linear extension, and calcification rates of *P. lobata* (with added *Diploastrea heliophora* in few places) colonies by analyzing coral cores recovering the last 50–150 years by sclerochronology. The annual density bands and growth parameters of the massive coral sampled along the transect will be determined using X-ray radiography and 3D-computed tomography of growth rings on sliced coral cores [50–52], allowing the calculation of recent reef growth over the last decades and century. In parallel, ambient sea temperature and pH changes over the same period of time will be documented by analyzing, at an annual or monthly timescale, the most advanced geochemical tracers along the cores, such as boron isotopes or trace elements, trapped along the cores during the coral life [53–56]. Recent stress events will be identified by a series of conventional markers of physiological stress or damage (see above) [57]. In addition, telomere length measurements of coral and microalgal symbiont genomes will be conducted. Notably, although measurements of telomere length as a method of assessing stress accumulation is currently being employed in numerous human cohort studies, the importance of telomere length for stress resilience and biodiversity in ecological studies is in its infancy and remains to be determined [58]. Thus, it will be highly informative to compare telomere length variations with coral growth measurements assessed by sclerochronology.

More broadly, the integration of environmental and physicochemical with biological data will allow us to pinpoint the specific adaptations that enable corals to live and survive across environmental gradients. Comparison of current to historical data throughout the Pacific will then show the biological cost of this adaptation and the associated time span. This in turn will make for a much better understanding of the capacity of coral holobionts to adapt to adverse environmental conditions and the required time scales.

In summary, *Tara Pacific* is poised to build the most comprehensive morphomolecular inventory of the phenotypic and genotypic biodiversity of coral reef ecosystems, including a wide spectrum of life from viruses to bacteria and from unicellular eukaryotes to metazoans and covering nested spatial scales from coral holobiont colonies, their surrounding water (i.e., coral-surrounding water), reef surface water, and neighboring oceanic surface waters (upstream and downstream of the sampled islands). The project focuses on selected key species that are ubiquitous in coral reef ecosystems throughout the Pacific and can be traced from an environmental, physicochemical, biological perspective down to the gene. This ambitious project has the potential to reveal substantial uncharted biodiversity; to shed light on the complex links between genomes, transcriptomes, metabolomes, organisms, and ecosystem functions in coral reef systems; and to provide a basis for the biodiversity and biological state of modern coral reefs for the research community at large. In addition, this project will contribute significantly to other fields of research such as stress and aging biology. We expect that the unprecedented scale of this project will help to decipher the complex interactions that together ensure a healthy state of coral reefs, a quest for which advances are urgently needed.

Acknowledgments

We are keen to thank the *Tara Ocean Foundation* teams and crew, without which *Tara Pacific* would not exist.

The members of the Tara Pacific Consortium are as follows: Planes S. (1), Allemand D. (2), Agostini S. (15), Armstrong E. (20), Audrain S. (12), Aury J.-M. (20), Banaig B. (1), Barbe V. (20), Belser C. (20), Beraud E. (2), Boissin E. (1), Bonnival E. (22), Boss E. (13), Bourdin G. (13), Bourgois E. (12), Bowler C. (25), Carradec Q. (20), Cassar N. (27, 28), Cohen N. R. (30), Conan P. (10), Cronin D. R. (19), da Silva O. (17), de Vargas C. (22), Djerbi N. (3), Dolan J. R. (17), Dominguez Herta G. (19), Douville E. (14), Du J. (19), Filée J. (32), Flores J. M. (8), Forcioli D. (3), Friedrich R. (26), Furla P. (3), Galand P. E. (11), Ghigliione J.-F. (10), Gilson E. (3), Gorsky G. (17), Guinther M. (15), Haëntjens N. (13), Henry N. (22), Hertau M. (12), Hochart C. (11), Hume B. C. C. (4), Iwankow G. (1), John S. G. (29), Karp-Boss L. (13), Kelly R. L. (29), Kitano Y. (16), Klings G. (21), Koren I. (8), Labadie K. (20), Lancelot J. (12), Lang-Yona N. (9), Lê-Hoang J. (20), Lemee R. (17), Lin Y. (27), Lombard F. (17), Marie D. (22), McMind R. (3), Miguel-Gordo M. (24), Trainic M. (8), Monmarche D. (12), Moulin C. (12), Mucherie Y. (12), Noel B. (20), Ottaviani A. (3), Paoli L. (7), Pedrotti M.-L. (17), Pesant S. (23), Pogoreutz C. (4), Poulain J. (20), Pujo-Pay M. (10), Reverdin G. (31), Reynaud S. (2), Romac S. (22), Röthig T. (5), Rottinger E. (3), Rouan A. (3), Ruscheweyh H.-J. (7), Salazar G. (7), Sullivan M. B. (18), Sunagawa S. (7), Thomas O. P. (24), Troublé R. (12), Vardi A. (9), Vega-Thunder R. (21), Woolstra C. R. (4), Wincker P. (20), Zahed A. (19), Zamoum T. (3), Ziegler M. (6), and Zoccola D. (2) on behalf of the consortium Tara Pacific. With (1) PSL Research University: EPHE-UPVD-CNRS, USR 3278 CRIOBE, Université de Perpignan, France; (2) Centre Scientifique de Monaco, Principality of Monaco; (3) Université Côte d'Azur, CNRS, Inserm-IRCAN; (4) Department of Biology, University of Konstanz, Konstanz, Germany; (5) Aquatic Research Facility, Environmental Sustainability Research Centre, University of Derby, Derby, United Kingdom; (6) Department of Animal Ecology & Systematics, Justus Liebig University, Giessen, Germany; (7) Department of Biology, Institute of Microbiology and Swiss Institute of Bioinformatics, ETH Zürich, Zürich, Switzerland; (8) Weizmann Institute of Science, Dept. Earth and Planetary Science, Israel; (9) Weizmann Institute of Science, Dept. Plant and Environmental Science, Israel; (10) Sorbonne Université, CNRS, LOMIC, Observatoire Océanologique de Banyuls; (11) Sorbonne Université, CNRS, LECOB, Observatoire Océanologique de Banyuls; (12) Tara Ocean Foundation, Paris, France; (13) School of Marine Sciences, University of Maine, United States of America; (14) Laboratoire des Sciences du Climat et de l'Environnement, LSCE/IPSL, CEA-CNRS-UVSQ, Université Paris-Saclay, France; (15) Shimoda Marine Research Center, University of Tsukuba, Shizuoka, Japan; (16) National Institute of Environmental Science, Japan; (17) Sorbonne Université, Institut de la Mer de Villefranche sur mer, Laboratoire d'Océanographie de Villefranche, France; (19) The Ohio State University, Departments of Microbiology and Civil, Environmental and Geodetic Engineering, Columbus, Ohio, United States of America and The Ohio State University, Departments, Columbus, Ohio, United States of America; (20) Génomique Métabolique, Genoscope, Institut François Jacob, CEA, CNRS, Univ Evry, Université Paris-Saclay, Evry, France; (21) Oregon State University, Department of Microbiology, Oregon, United States of America; (22) Sorbonne Université, CNRS, Station Biologique de Roscoff, AD2M, UMR 7144, ECOMAP, Roscoff, France; (23) PANGAEA, Data Publisher for Earth and Environment Science, Bremen, Germany & MARUM—Center for Marine Environmental Sciences, Universität Bremen, Bremen, Germany; (24) Marine Biodiscovery Laboratory, School of Chemistry and Ryan Institute, National University of Ireland, Galway, Ireland; (25) Institut de Biologie de l'Ecole Normale Supérieure (IBENS), Ecole normale supérieure, CNRS, INSERM, Université PSL, Paris, France; (26) World Courier, an AmerisourceBergen Company, Russelsheim, Germany; (27) Division of Earth and Ocean Sciences, Duke University, Durham, North Carolina, United States of America; (28) Laboratoire des Sciences de l'Environnement Marin (LEMAR), UMR 6539 UBO/CNRS/IRD/IFREMER, Institut Universitaire Européen de la Mer (IUEM), Brest, France; (29)

Department of Earth Sciences, University of Southern California, Los Angeles, California, United States of America; (30) Marine Chemistry and Geochemistry Department, Woods Hole Oceanographic Institution; (31) Institut Pierre Simon Laplace, CNRS/IRD/MNHN (LOCEAN) Sorbonne-Université Paris Cedex 05, France; (32) Laboratoire Evolution, Génomes, Comportement et Ecologie, CNRS/Université Paris-Saclay, Avenue de la Terrasse, Gif sur Yvette, France. S. Planes and D. Allemand are joint scientific directors of *Tara Pacific*.

References

1. Crutzen PJ, Stoermer EF (2000) The “Anthropocene”. *Global Change* 41: 17–18.
2. Steffen W, Grinevald J, Crutzen P, McNeill J (2011) The Anthropocene: conceptual and historical perspectives. *Phil Trans R Soc A* 369: 842–867. <https://doi.org/10.1098/rsta.2010.0327> PMID: 21282150
3. Gattuso JP, Magnan A, Billé R, Cheung WWL, Howes EL, Joos F, et al. (2015) Contrasting futures for ocean and society from different anthropogenic CO₂ emissions scenarios. *Science* 349: aac4722. <https://doi.org/10.1126/science.aac4722> PMID: 26138982
4. IPCC (2018) Global Warming of 1.5°C: An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty. Masson-Delmotte V, Zhai P, Pörtner H-O, Roberts D, Skea J, Shukla PR, Pirani A, Moufouma-Okia W, Péan C, Pidcock R, Connors S, Matthews JBR, Chen Y, Zhou X, Gomis MI, Lonnoy E, Maycock T, Tignor M, Waterfield T, editors. Geneva, Switzerland: IPCC.
5. Smith L (1978) Coral reef area and the contributions of reefs to processes and resources of the world's oceans. *Nature* 273: 225–226.
6. Porter JW, Tougas JI (2001) Reef ecosystems: threats to their biodiversity. In: *Encyclopedia of Biodiversity*, Volume 5, Levin S.A., editor. San Diego: Academic Press. p. 73–95.
7. Wilkinson C, editor (2004) Status of Coral Reefs of the World: 2004. Volume 1. Townsville, Queensland, Australia: Australian Institute of Marine Science. p. 301.
8. Chen P-Y, Chen C-C, Chu LF, McCarl B (2015) Evaluating the economic damage of climate change on global coral reefs. *Global Environ Change* 30: 12–20.
9. Guidi L, Chaffron S, Bittner L, Eveillard D, Larhlimi A, Roux S, et al. (2016) Plankton networks driving carbon export in the oligotrophic ocean. *Nature* 532: 465–470. <https://doi.org/10.1038/nature16942> PMID: 26863193
10. Karsenti E, Acinas SG, Bork P, Bowler C, De Vargas C, Raes J, et al. (2011) A holistic approach to marine eco-systems biology. *PLoS Biol* 9(10): e1001177. <https://doi.org/10.1371/journal.pbio.1001177> PMID: 22028628
11. Bork P, Bowler C, de Vargas C, Gorsky G, Karsenti E, Wincker P (2015) Tara Oceans studies plankton at planetary scale. *Science* 348: 873–873. <https://doi.org/10.1126/science.aac5605> PMID: 25999501
12. Darwin C (1842) The Structure and Distribution of Coral Reefs. Being the First Part of the Geology of the Voyage of the ‘Beagle’, under the command of capt. Fitzroy R. N. London: Smith, Elder & Co.
13. Rohwer F, Seguritan V, Azam F, Knowlton N (2002) Diversity and Distribution of Coral-Associated Bacteria. *Mar Ecol Progress Ser* 243: 1–10.
14. Huang D, Roy K (2015) The future of evolutionary diversity in reef corals. *Phil Trans R Soc B* 370: 20140010
15. Shinzato C, Shoguchi E, Kawashima T, Hamada M, Hisata K, Tanaka M, et al. (2011) Using the *Acropora digitifera* genome to understand coral responses to environmental change. *Nature* 476: 320–323. <https://doi.org/10.1038/nature10249> PMID: 21785439
16. Voolstra CR, Li Y, Liew YJ, Baumgarten S, Zoccola D, Flot J-F, et al. (2017) Comparative analysis of the genomes of *Stylophora pistillata* and *Acropora digitifera* provides evidence for extensive differences between species of corals. *Scientific Reports* 7: 17583. <https://doi.org/10.1038/s41598-017-17484-x> PMID: 29242500
17. LaJeunesse TC, Parkinson JE, Gabrielson PW, Jeong HJ, Reimer JD, Voolstra CR, Santos SR (2018) Systematic Revision of Symbiodiniaceae Highlights the Antiquity and Diversity of Coral Endosymbionts. *Current Biol* 28: 873–875.
18. Thornhill DJ, Howells EJ, Wham DC, Steury TD, Santos SR (2017) Population genetics of reef coral endosymbionts (Symbiodinium, Dinophyceae). *Mol Ecol* 26: 2640–2659. <https://doi.org/10.1111/mec.14055> PMID: 28188662

19. Pante E, Puillandre E, Viricel A, Arnaud-Haond S, Aurelle D, Castelin M, et al. (2015) Species are hypotheses: avoid connectivity assessments based on pillars of sand. *Mol Ecol* 24: 525–544. <https://doi.org/10.1111/mec.13048> PMID: 25529046
20. Jessen C, Villa Lizcano JF, Bayer T, Roder C, Aranda M, Wild C, Voolstra CR (2013) *In situ* Effects of Eutrophication and Overfishing on Physiology and Bacterial Diversity of the Red Sea Coral *Acropora hemprichii*. *PLoS ONE* 8(4): e62091. <https://doi.org/10.1371/journal.pone.0062091> PMID: 23630625
21. Zaneveld JR, Burkepille DE, Shantz AA, Pritchard CE, McMinds R, Payet JP, et al. (2016) Overfishing and nutrient pollution interact with temperature to disrupt coral reefs down to microbial scales. *Nature Communications* 7: 11833.
22. Pogoreutz C, Rådecker N, Cárdenas A, Gärdes A, Wild C, Voolstra CR (2018) Dominance of *Endozoicomonas* bacteria throughout coral bleaching and mortality suggests structural inflexibility of the *Pocillopora verrucosa* microbiome. *Ecology and Evolution* 8: 2240–2252. <https://doi.org/10.1002/ece3.3830> PMID: 29468040
23. Ziegler M, Seneca FO, Yum LK, Palumbi SR, Voolstra CR (2017) Bacterial community dynamics are linked to patterns of coral heat tolerance. *Nature Communications* 8: 14213. <https://doi.org/10.1038/ncomms14213> PMID: 28186132
24. Pogoreutz C, Rådecker N, Cárdenas A, Gärdes A, Voolstra CR, Wild C (2017) Sugar enrichment provides evidence for a role of nitrogen fixation in coral bleaching. *Global Change Biology* 23: 3838–3848. <https://doi.org/10.1111/gcb.13695> PMID: 28429531
25. McDervitt-Irwin J, Garren M, Baums J, Vega Thurber R (2017) Responses of coral-associated bacterial community responses to local and global stressors. *Frontiers of Marine Science* 4(262): e00143.
26. Pollock JF, McMinds R, Smith S, Bourne DG, Willis BL, Medina M, et al. (2018) Coral-associated bacteria demonstrate phyllosymbiosis and cophylogeny. *Nature Com* 9: 4921.
27. Neave MJ, Michell CT, Apprill A, Voolstra CR (2017) *Endozoicomonas* genomes reveal functional adaptation and plasticity in bacterial strains symbiotically associated with diverse marine hosts. *Scientific Reports* 7: 40579. <https://doi.org/10.1038/srep40579> PMID: 28094347
28. Neave MJ, Rachmawati R, Xun L, Michell CT, Bourne DG, Apprill A, Voolstra CR (2017) Differential specificity between closely related corals and abundant *Endozoicomonas* endosymbionts across global scales. *ISME J* 11: 186–200. <https://doi.org/10.1038/ismej.2016.95> PMID: 27392086
29. Güemes AGB, Youle M, Cantu VA, Felts B, Nulton J, Rohwer F (2016) Viruses as the Winners in the Game of Life. *Annual Reviews of Virology* 3: 197–214.
30. Vega Thurber R, Payet JP, Thurber AR, Correa AMS (2017) Virus–host interactions and their roles in coral reef health and disease. *Nature Reviews Microbiology* 15: 205–216. <https://doi.org/10.1038/nrmicro.2016.176> PMID: 28090075
31. Pesant S, Not F, Picheral M, Kandels-Lewis S, Le Bescot N, Gorsky G, et al. (2015) Open science resources for the discovery and analysis of Tara Oceans data. *Sci Data* 2: 150023. <https://doi.org/10.1038/sdata.2015.23> PMID: 26029378
32. Wilkinson MD, Dumontier M, Aalbersberg IJ, Appleton G, Axton A, Blomberg N, et al. (2016) The fair guiding principles for scientific data management and stewardship. *Science Data* 3: 160018.
33. de Vargas C, Audic S, Henry N, Decelle J, Mahe F, Logares R, et al. (2015) Ocean plankton. Eukaryotic plankton diversity in the sunlit ocean. *Science* 348: 1261605. <https://doi.org/10.1126/science.1261605> PMID: 25999516
34. Decelle J, Carradec Q, Pochon X, Henry N, Romac S, Mahé S, et al. (2018) Worldwide Occurrence and Activity of the Reef-Building Coral Symbiont Symbiodinium in the Open Ocean. *Current Biol* 28: 3625–3633.
35. Sunagawa S, Karsenti E, Bowler C, Bork P (2015) Computational eco-systems biology in Tara Oceans: translating data into knowledge. *Mol Syst Biol* 11: 809. <https://doi.org/10.15252/msb.20156272> PMID: 25999085
36. Brum JR, Ignacio-Espinoza JC, Roux S, Doucier G, Acinas SG, Alberti A, et al. (2015) Ocean plankton. Patterns and ecological drivers of ocean viral communities. *Science* 348: 1261498. <https://doi.org/10.1126/science.1261498> PMID: 25999515
37. Walters W, Hyde ER, Berg-Lyons D, Ackermann G, Humphrey G, Parada A, et al. (2015) Improved bacterial 16S rRNA gene (V4 and V4-5) and fungal internal transcribed spacer marker gene primers for microbial community surveys. *mSystems* 1(1):e00009–15. <https://doi.org/10.1128/mSystems.00009-15> PMID: 27822518
38. Arif C, Daniels C, Bayer T, Banguera-Hinestroza E, Barbrook A, Howe CJ, et al. (2014) Assessing *Symbiodinium* diversity in scleractinian corals via next-generation sequencing-based genotyping of the ITS2 rDNA region. *Molecular Ecology* 23(17): 4418–33. <https://doi.org/10.1111/mec.12869> PMID: 25052021

39. Hume CC, Ziegler M, Poulain J, Pochon X, Romac S, Boissin E., et al. (2018) An improved primer set and amplification protocol with increased specificity and sensitivity targeting the Symbiodinium ITS2 region. *PeerJ* 6: e4816. <https://doi.org/10.7717/peerj.4816> PMID: 29844969
40. Hume BCC, Smith EG, Ziegler M, Warrington HJM, Burt JA, LaJeunesse TC, et al. (2019) SymPortal: A novel analytical framework and platform for coral algal symbiont next-generation sequencing ITS2 profiling. *Molecular Ecology Resources* 19(4): 1063–80. <https://doi.org/10.1111/1755-0998.13004> PMID: 30740899
41. Leray M, Knowlton N (2015) DNA barcoding and metabarcoding of standardized samples reveal patterns of marine benthic diversity. *Proc Natl Acad Sci U S A* 112(7): 2076–81. <https://doi.org/10.1073/pnas.1424997112> PMID: 25646458
42. Gregory A, Zayed A, Jang HB, Conceicao-Neto N, Temperton B, Bolduc B, et al. (2019) Marine DNA viral macro- and micro-diversity from pole to pole. *Cell* 177: 1109–1123. <https://doi.org/10.1016/j.cell.2019.03.040> PMID: 31031001
43. Roux S, Brum JR, Dutilh BE, Sunagawa S, Duhaime MB, Loy A, et al. (2016) Ecogenomics and potential biogeochemical impacts of globally abundant ocean viruses. *Nature* 537: 689–693. <https://doi.org/10.1038/nature19366> PMID: 27654921
44. Lima-Mendez G, Faust K, Henry N, Decelle J, Colin S, Carcillo F, et al. (2015) Determinants of community structure in the global plankton interactome. *Science* 348: 1262073. <https://doi.org/10.1126/science.1262073> PMID: 25999517
45. Alberti A, Poulain J, Engelen S, Labadie K, Romac S, Ferrera I, et al. (2017) Viral to metazoan marine plankton nucleotide sequences from the *Tara* Oceans expedition. *Scientific Data* 4: 170093. <https://doi.org/10.1038/sdata.2017.93> PMID: 28763055
46. Carradec Q, Pelletier E, Da Silva C, Alberti A, Seeleuthner Y, Blanc-Mathieu R, et al. (2018) A global ocean atlas of eukaryotic genes. *Nat Com* 9: 373.
47. Wegley KL, Haas AF, Nelson CE (2018) Ecosystem microbiology of coral reefs: linking genomic, metabolomic, and biogeochemical dynamics from animal symbioses to reefscape processes. *mSystems* 3: e00162–17. <https://doi.org/10.1128/mSystems.00162-17> PMID: 29556542
48. Quinn RA, Vermeij MJA, Hartmann AC, Galtier d'Auriac I, Benler S, Haas A, et al. (2016) Metabolomic of reef benthic interactions reveals a bioactive lipid involved in coral defence. *Proc R Soc B* 283: 20160469. <https://doi.org/10.1098/rspb.2016.0469> PMID: 27122568
49. Houle D, Govindaraju DR, Omholt S (2010) Phenomics: the next challenge. *Nature Reviews Genetics* 11: 855–866. <https://doi.org/10.1038/nrg2897> PMID: 21085204
50. Buddemeier RW, Kinzie RA (1976) Coral growth. *Oceanogr Mar Biol Ann Rev* 14: 183–225.
51. Bessat F, Buigues D (2001) Two centuries of variation in coral growth in a massive *Porites* colony from Moorea (French Polynesia): a response of ocean-atmosphere variability from south central Pacific. *Palaeogeography, Palaeoclimatology, Palaeoecology* 175: 381–392.
52. Lough JM, Cantin NE (2014) Perspectives on Massive Coral Growth Rates in a Changing Ocean. *Biol Bull* 226: 187–202. <https://doi.org/10.1086/BBLv226n3p187> PMID: 25070864
53. Montagna P, McCulloch M, Douville E, López Correa M, Trotter J, Rodolfo-Metalpa R, et al. (2014) Li/Mg systematics in scleractinian corals: Calibration of the thermometer. *Geochim Cosmochim Acta* 132: 288–310.
54. D'Olivo JP, Sinclair DJ, Rankenburg K, McCulloch MT (2018) A universal multi-trace element calibration for reconstructing sea surface temperatures from long-lived *Porites* corals: Removing 'vital-effects'. *Geochim Cosmochim Acta* 239: 109–135.
55. Wu HC, Dissard D, Douville E, Blamart D, Bordier L, Tribollet A, et al. (2018) Surface ocean pH variations since 1689 CE and recent ocean acidification in the tropical south Pacific. *Nature Communications* 9: 2543. <https://doi.org/10.1038/s41467-018-04922-1> PMID: 29959313
56. D'Olivo JP, Georgiou L, Falter J, DeCarlo TM, Irigoien X, Voolstra CR, et al. (2019) Long-Term Impacts of the 1997–1998 Bleaching Event on the Growth and Resilience of Massive *Porites* Corals From the Central Red Sea. *Geochemistry, Geophysics, Geosystems* 2019. <https://doi.org/10.1029/2019GC008312>
57. Hook SE, Gallagher EP, Batley GE (2014) The role of biomarkers in the assessment of aquatic ecosystem health. *Integr Environ Assess Manag* 10: 327–341. <https://doi.org/10.1002/ieam.1530> PMID: 24574147
58. Ye J, Renault V, Jamet K, Gilson É (2014) Transcriptional outcome of telomere signalling. *Nature Rev Genetics* 15: 491–503.
59. Key RM, Olsen A, van Heuven S, Lauvset SK, Velo A, Lin X, et al. (2015) Global Ocean Data Analysis Project, Version 2 (GLODAPv2), ORNL/CDIAC-162, NDP-P093. Oak Ridge, Tennessee: Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, US Department of Energy. https://doi.org/10.3334/CDIAC/OTG.NDP093_GLODAPv2
60. Lauvset SK, Key RM, Olsen A, van Heuven S, Velo A, Lin X, Schirnack C, et al. (2016) A new global interior ocean mapped climatology: the 1°x1° GLODAP version 2. *Earth System Science Data* 8, 325–340.