

Development and validation of a targeted gene sequencing panel for application to disparate cancers

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Supplementary Figure S1. Sensitivity controls. To test panel sensitivity, controls with known single nucleotide variants (SNVs), multiple nucleotide variants (MNVs), small insertions (INS) and deletions (DEL), substitutions (SUBST), duplications (DUP) and complex variants (CV) were obtained from Kathleen Cunningham Foundation Consortium for Research into Familial Breast Cancer (kConFab) or as a commercially available pool of synthetic oligos against a normal genomic background (AcroMetrix). Presented here are pie charts representing the distribution of the different mutation types across the controls. kConFab samples were provided as germline DNA samples from 35 breast cancer patients (7 other samples had previously been omitted), whilst AcroMetrix represented 34 germline mutations, and 521 somatic mutations at 5-15% variant allele frequency (VAF; n=341), and 15-35% VAF (n=180).



Supplementary Table S1. Panel metrics for targeted regions (coding + non-coding)

	PV1			PV2			Fail criteria
	Mean	Min	Max	Mean	Min	Max	
Number of reads	21,951,481	11,688,460	46,698,912	22,995,280	9,385,708	54,554,608	
Unique reads	9,263,214	2,799,758	31,500,507	17,417,919	5,529,146	46,378,680	
Mean coverage across target	380	100	1425	395	106	1006	<100x
%Bases > 20x across target	0.95	0.91	1.00	0.99	0.97	1.00	<95%
%Bases > 50x across target	0.93	0.88	0.99	0.96	0.86	0.99	
%Bases > 100x across target	0.89	0.54	0.98	0.88	0.48	0.99	
Read enrichment (%)	81	77	86	68	50	80	
Duplication (%)	59	33	78	26	8	68	

Supplementary Figure S2. Potential annotation errors in three of the kConFab samples. Integrative Genomics Viewer (IGV) screenshots of three variants of discrepant annotation provided by kConFab. All three are in *BRCA1* which is on the antisense strand, hence reverse base nomenclature. (A) Expected heterozygous variant occurring at splice site c.135-1G>T. Detected variant is indicated by the black arrow. (B) An insertion was expected in *BRCA1* at c.3194-3195. This position is indicated by the black arrow in the magnified inset. (C) A 2bp insertion was expected at c.1881_1882 in *BRCA1*. This position is indicated by the black arrow in the magnified inset. Individual reads in opposing directions are shown as purple and orange lines. Mutations/insertions of interest are positioned where actually detected, between the dashed black lines. Note insertions (indicated by a purple I) have been grouped together in the insets for ease of visualisation.





Supplementary Figure S3. Improved coverage and missed AcroMetrix variants. Representative Integrative Genomics Viewer (IGV) snapshots for two variants missed in PV1 due to lack of coverage/targeting in (A) kConFab sample *CHEK2* c.441+1G>A and (B) AcroMetrix *PTEN* c.870A>G, p.G293G, that were then readily detectable in the improved PV2 design. Two variants (of 555 targeted), were still missed in the AcroMetrix variant pool including (C) *HNF1A* c.872_873insC, p.G292fs*25 occurring in a highly repetitive region and (D) *PIK3CA* c.1633G>A, p.E545K occurring in a mutational hotspot. Note in C, there are two panels indicating conflicting positioning of the insertion (purple I) either side of the guanine nucleotide. This is the same position as an expected substitution in the background genomic DNA. Individual reads in opposing directions are shown as purple and orange lines. Mutations of interest are positioned between the dashed black lines.



Supplementary Figure S4. Common shared CNVs between PV1 captured DNA and CCLE-reported CNVs. Data presented are an extension upon Fig. 3D-G, and represent all deletions (red) and amplifications (blue) detected by PV1 in the SK-BR-3 and BT-474 breast cancer cell lines in comparison to those reported by the Cancer Cell Line Encyclopedia (CCLE). Deletions were defined as copy numbers of <1.5 and amplifications were defined as copy numbers of >2.5 to allow for errors. Samples which did not fall within these definitions were considered to be diploid and excluded from the plots. Note the log₂ y-axes. Gene names are recorded along the x-axes.

