Enhanced root carbon allocation through organic farming is restricted to topsoils

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1 Abstract

2 Soils store significant amounts of carbon (C) and thus can play a critical role for mitigating 3 climate change. Crop roots represent the main C source in agricultural soils and are particularly 4 important for long-term C storage in agroecosystems. To evaluate the potential of different farming 5 systems to contribute to soil C sequestration and thus climate change mitigation, it is of great importance 6 to gain a better understanding of the factors influencing root C allocation and distribution. So far, it is 7 still unclear how root C allocation varies among farming systems and whether the choice of management 8 practices can help to enhance root C inputs. In this study, we compared root C allocation in three main 9 arable farming systems, namely organic, no-till, and conventional farming. We assessed root biomass, 10 vertical root distribution to 0.75 m soil depth, and root-shoot ratios in 24 winter wheat fields. We further 11 evaluated the relative importance of the farming system compared to site conditions and quantified the 12 contribution of individual management practices and pedoclimatic drivers. Farming system explained 13 one third of the variation in topsoil root biomass and root-shoot ratios, both being strongly positively 14 related to weed biomass and soil organic C content and negatively to mineral nitrogen fertilization 15 intensity. Root C allocation was significantly higher in organic farming as illustrated by an increase in 16 root biomass (+40%) and root-shoot ratios (+60%) compared to conventional farming. By contrast, the 17 overall impact of no-till was low. The importance of pedoclimatic conditions increased substantially 18 with soil depth and deep root biomass was largely controlled by precipitation and soil texture, while the 19 impact of management was close to zero. Our findings highlight the potential of organic farming in 20 promoting root C inputs to topsoils and thereby contributing to soil organic matter build-up and 21 improved soil quality in agroecosystems.

22 Keywords

root carbon inputs; farming system; agricultural management; on-farm study; root biomass distribution;
subsoil

25 1. Introduction

Soils play a prominent role in the global carbon (C) cycle as they contain substantially more C than the atmosphere and land vegetation combined (Lehmann and Kleber, 2015). Increasing soil organic C therefore holds great promise for mitigating climate change. Agricultural soils could be a key in this effort because 34% of the land surface is currently under agricultural use (Ritchie and Roser, 2020) and management substantially influences soil organic C storage by altering inputs and decomposition rates (Janzen, 2015; Paustian *et al.*, 2016).

32 Root C is one of the most important contributors to soil organic C and constitutes up to 90% of 33 all C inputs to arable soils (Kätterer et al., 2011). Due to its resistant chemical composition (Rasse et 34 al., 2005) and preferential incorporation into more stable fractions (Ghafoor et al., 2017), root C has a 35 longer residence time in soil than C derived from above ground crop residues and manure (Kätterer et 36 al., 2011; Menichetti et al., 2015; Zhang et al., 2015). Particularly, root C inputs to deep soil have been 37 linked to long-term C storage (Russell et al., 2009; Fan et al., 2019) due to the low decomposer 38 abundance and high storage capacity of deep unsaturated layers (Rasse et al., 2005; Rumpel et al., 2012; 39 Sanaullah et al., 2016). Hence, the promotion of more and deeper roots has been proposed as a strategy to mitigate climate change with an estimated potential to remove atmospheric CO_2 of about 1 Pg yr⁻¹ 40 41 (Lynch and Wojciechowski, 2015; Paustian et al., 2016; Pierret et al., 2016). Thus, it is crucial to 42 understand how management can promote root C inputs to agricultural soils in order to sequester C in 43 the long-term, but also to stimulate C dynamics, thereby enhancing the manifold benefits of soil organic 44 matter for agricultural soils (Janzen, 2015; Paustian et al., 2016).

45 Agricultural management affects root biomass allocation in various ways by its impact on crop 46 nutrition and soil properties through e.g. type and amount of fertilization, crop rotation, or soil tillage 47 (Malhi and Lemke, 2007; Chirinda et al., 2012; Qin et al., 2018). For instance, in organic farming, the 48 application of synthetic nutrient inputs is prohibited, which often leads to reduced mineral nitrogen (N) 49 availability (Lorenz and Lal, 2016). It is expected that this increases biomass allocation below ground 50 as crops need to cope with primarily growth-limiting resources (Lynch et al., 2012; Poorter et al., 2012). 51 No-till farming is another alternative to conventional farming and is characterised by reduced or zero 52 soil disturbance through tillage. Hence, it often results in accumulation of organic matter and nutrients

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53 but also increased bulk density in the topsoil (Huggins and Reganold, 2008; Powlson et al., 2014). This 54 may lead to a shift in biomass allocation and increased superficial root proliferation (Qin et al., 2018; 55 Mondal et al., 2020), thereby altering vertical root distribution (Dwyer et al., 1996; Ball-Coelho et al., 1998; Barzegar et al., 2004). So far, the influence of different farming systems on root C allocation has 56 57 still not been clearly established and current knowledge is based on controlled field studies conducted 58 at a small number of sites. In organic farming, both similar (Steingrobe et al., 2001; Lazicki et al., 2016; 59 Hirte et al., 2018a) and higher (Chirinda et al., 2012; Hu et al., 2018) root biomass compared to 60 conventional farming has been reported for cereals. No-till was even found to influence root biomass in 61 any direction for cereals or rapeseed, i.e. tillage effects were negative, absent, or positive (Plaza-Bonilla 62 et al., 2014; Li et al., 2017; Sarker et al., 2017).

63 The unclear picture of how agricultural management influences root C allocation may be linked 64 to the impact of soil and climate characteristics that often overlay management effects. Soil properties 65 such as mechanical impedance or nutrient availability as well as climatic conditions such as precipitation 66 or temperature affect root growth to a large extent and complex interactions of stimuli often obliterate 67 root response to individual drivers (reviewed by Rich and Watt, 2013). Consequently, biomass 68 allocation to roots and shoots can vary by a factor of 10 across environments (Enquist and Niklas, 2002; 69 Poorter et al., 2012). In order to unravel the potential of agricultural management to enhance root C 70 inputs to soil (Paustian et al., 2016; Dignac et al., 2017), management effects need to be assessed over 71 a wide range of pedoclimatic conditions. On-farm measurements over multiple locations can not only 72 provide practice-related, generalizable results but could also allow for quantitative comparisons of the 73 effects of specific management practices on crop parameters beyond classified farming systems 74 (Nkurunziza et al., 2017; Büchi et al., 2019).

We therefore established a network of 24 farms classified as conventional, no-till, or organic in Switzerland and investigated root biomass in the top and subsoil in winter wheat fields. In addition, we collected detailed information on management practices and soil and climate conditions for each field. Our objectives were (i) to assess the impact of organic, conventional and no-till farming on root biomass and plant biomass allocation and (ii) to evaluate the relative importance of management- and site-related variables for root and shoot biomass, root-shoot ratios, and vertical root distribution.

81 2. Methods

82 2.1. Farming systems and sites

83 The study was conducted in 2016 on 24 commercial farms in the northern part of Switzerland, 84 which were categorized as conventional with tillage (conventional), conventional without tillage (no-85 till), or organic with tillage (organic) according to the farm structure census 2015 (Supplementary table 86 1; FSO, 2017; Büchi et al., 2019). No-till soil management implied that not more than 25% of the soil 87 surface could be disturbed at sowing (Swiss Federal Council, 2013). All farms were managed according 88 to the certification scheme Proof of Ecological Performance PEP (Swiss Federal Council, 2013), the 89 guidelines of the Swiss Farmer Association for Integrated Production IP-Suisse (IP-SUISSE, 2019), or 90 the regulations of the Federation of Swiss Organic Farmers BIO-Suisse (Swiss Federal Council, 1997). 91 The farms were located at eight sites spread over a distance of roughly 100 km arranged in farming 92 system triplets of one conventional, no-till, and organic farm each (Supplementary figure 1). The nearest 93 weather stations operated by the Federal Office of Meteorology and Climatology with recorded long-94 term precipitation data were chosen as reference points for the sites (Supplementary table 2). Annual 95 temperature and precipitation (1981-2010) for Zurich-Affoltern (08°31'04", 47°25'40"), which is 96 centrally located within the study area, are 9.4 °C and 1054 mm, respectively.

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2.2. Growth conditions of winter wheat

On each farm, one field was selected for plant and soil analyses. Winter wheat (*Triticum aestivum*, L.) was sown between 2 and 26 October 2015 and harvested between 18 July and 4 August 2016. Varieties, type of fertilization, weed and pathogen control, and use of growth retardants differed between farms (Supplementary table 1). Organic fertilizers were applied as cattle or pig slurry using an injector or as cattle manure, compost, humus acid suspension, or granulated organic N fertilizer (Büchi et al., 2019).

104 2.3. Root and shoot sampling

Root and shoot biomass of wheat and weeds was sampled at wheat flowering between 14 and
23 June 2016. A circular area with a radius of 10 m and a distance of at least 20 m to the nearest edge
of the field was defined as sampling area and divided into four quarters (Supplementary figure 2). Within

108 each quarter, shoot samples were taken directly above the ground on one randomly selected sampling 109 plot covering four wheat rows of 0.5 m length with electric grass clippers and separated into wheat and 110 weed shoot biomass. On the same sampling plots, root samples were collected by taking two soil cores, 111 one within and one half-way between wheat rows, to a depth of 0.75 m by means of a metal sampling 112 rod (inner diameter: 60 mm; lined with polyethylene film) driven into soil with an electric breaker 113 (EH50, Wacker, Germany) and extracted with a 3-cylinder-lifting unit (ZGM-9E ECO, Nordmeyer 114 Geotool GmbH, Germany). The cores were separated into three layers of 0.25 m length (top: 0–0.25 m, 115 intermediate: 0.25–0.5 m, deep: 0.5–0.75 m) and stored in polyethylene film at 4 °C for a maximum of 116 three weeks until further processing.

117 2.4. Biomass determination

118 Roots were extracted from each soil core separately using an automated root washer 119 (Hydropneumatic Elutriation System GVF 13000, Gillison's Variety Fabrication Inc., USA). The field-120 fresh soil was dispersed for 10 minutes in a high-energy hydrovortex at a water pressure of 121 approximately 350 kPa and roots were separated from the mineral fraction by flotation and recovered 122 on a 0.5 mm mesh (Smucker et al., 1982). The thus retained root samples were transferred to aluminium 123 dishes and extraneous organic matter was visually identified based on shape, structure, colour, and 124 elasticity of particles and removed from the samples using tweezers (Schuurman and Goedewaagen, 125 1971; Hirte et al., 2017). Identifiable weed roots, e.g. tap or rhizomatous roots, were removed from the 126 root samples. However, a certain proportion of weed roots could not be distinguished from wheat roots 127 by eye and remained in the samples. All plant material was dried at 55 °C until constant weight (shoots: 128 72 h; roots: 48 h) and dry weight was recorded.

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2.5. Management and pedoclimatic variables

The following variables and their importance for root biomass and distribution were investigated: mineral N fertilization intensity, sowing density, above ground weed biomass, soil bulk density, soil texture, soil organic C, total N and available P in soil, and precipitation (Supplementary table 3). Mineral N fertilization intensity and sowing density were derived from questionnaires returned by the farmers (Büchi et al., 2019). Mineral N fertilization intensity was calculated from fertilizer-N input (total N in mineral fertilizers and ammonium-N in organic fertilizers as estimated by Büchi *et al.*,
2019) in the wheat season 2015/2016 as the amount of applied N (kg ha⁻¹ season⁻¹) relative to the
recommended amount of available N (kg ha⁻¹ season⁻¹) for wheat according to the Principles of
Agricultural Crop Fertilisation in Switzerland (Richner and Sinaj, 2017). Although wheat variety was
an important aspect of management, this categorical information could not be accounted for due to the
great diversity of 15 different genotypes and, thus, the lack of replications across fields (Supplementary
table 1).

142 Soil was sampled on each farm between 20 April and 27 May 2016 for determination of soil 143 texture, organic C, total N, available P, and bulk density. Except for bulk density, 15-20 samples were 144 taken in five soil layers (0-0.05 m, 0.05-0.2 m, 0.2-0.25 m, 0.25-0.5 m, 0.5-0.75 m) on transect lines 145 that ran in 45° angles to the seedling rows and divided the quarters for root and shoot biomass sampling. 146 Composite samples per layer were dried and soil texture (sedimentation), organic C (oxidation with 147 potassium dichromate), and available P (CO2-saturated water extraction and colorimetry) were 148 determined on 2-mm sieved fine soil according to the Swiss reference methods (Agroscope, 1996). Total 149 soil N was measured after dry oxidation according to the Dumas method (Bremner, 1965). For soil bulk 150 density measurements, undisturbed samples of 100 ml volume and 50 mm height were taken in the 151 middle of each layer except the 0.5–0.75 m layer and oven-dried at 105°C for at least 72 h (Colombi et 152 al., 2019). Bulk density values of the 0.25–0.5 m layer were used for the 0.5–0.75 m layer. The weighted 153 averages of variables measured on samples from the upper three layers (0–0.05 m, 0.05–0.2 m, 0.2–0.25 154 m) served as composite values for the 0–0.25 m layer for further analyses.

155 Precipitation during the wheat growing season (October 2015 to June 2016) was retrieved from 156 the nearest local weather station to each farm operated by either MeteoSwiss, the Federal Roads Office, 157 the Cantons of Lucerne, Thurgovia, or Zurich, or MeteoGroup Switzerland. Due to clustering of farms 158 within sites and limited spatial distribution of local weather stations, 12 data sets for the total of 24 farms 159 were available. We tested the effect of cumulative precipitation during several time periods on the 160 investigated response variables and found the strongest effect for precipitation between March and mid-161 June, i.e. between tillering and flowering, corresponding to the main part of the vegetative growth phase. 162 From here on, we refer to this time period when we report values and the effect of precipitation.

163 2.6. Calculations and statistics

To extrapolate to field scale, root biomass sampled within and between rows was weighted with
respect to row width for each layer individually (adapted from Frasier et al., 2016):

$$RB_{within} = \frac{M_{within}}{\pi * (\frac{D}{2})^2} * \frac{D}{s}$$
(1)

$$RB_{between} = \frac{M_{between}}{\pi * (\frac{D}{2})^2} * \frac{(s-D)}{s}$$
(2)

166 where RB_{within} and $RB_{between}$ are root biomass (g m⁻²) within and between rows, respectively, 167 M_{within} and $M_{between}$ are the dry weights of roots (g) extracted from the soil cores taken within and between 168 rows, respectively, D is the inner diameter of the sampling rod (m), and s is the distance between rows 169 (m). Root biomass was obtained by summing RB_{within} and $RB_{between}$. Root-shoot ratios were calculated 170 for each subplot from averaged total root (0–0.75 m) and shoot biomass and were ln-transformed prior 171 to statistical analysis (Poorter and Sack, 2012). Unless otherwise stated, root-shoot ratios relate to wheat 172 shoot biomass (excluding weed) but were also analysed for wheat plus weed shoot biomass.

A few data points (12 out of 576) needed to be eliminated when problems with sampling or sample processing occurred (e.g. sieve clogging and root loss in the root washer). Consequently, root biomass could not be estimated for those instances and only 3 out of 4 field replications were used. Root and shoot biomass and root-shoot ratios of individual subplots on each farm were treated as lower-level replicates for statistical analysis and were averaged per farm for data presentation. Mean data for farming systems and sites are presented as averages of farming system/site and farm and standard errors of farming system/site.

We analysed the data in a three-step procedure and thereby investigated the following response variables: root biomass and the proportion of root biomass in the individual layers (0–0.25 m, 0.25–0.5 m, 0.5–0.75 m) and total root biomass (0–0.75 m) of wheat and weeds, wheat shoot biomass, and rootshoot ratio. (i) To test for differences in response variables between farming systems and sites, we fitted 184 the data to mixed effects models (fixed factors: farming system and site; random factor: farm) and 185 determined differences between group means by ANOVA and subsequent simultaneous multiple 186 comparison of estimated marginal means of group pairs with Tukey-adjustment of p-values. (ii) To 187 further evaluate the effects of the management and pedoclimatic variables on the response variables, we 188 used mixed effects models (fixed factor: management or pedoclimatic variable; random factor: farm) in 189 univariate analyses and ANOVA. (iii) To determine the relative importance of (a) farming system and 190 site and (b) management and pedoclimatic variables for the response variables, we conducted 191 multivariate linear regressions without prior variable selection and calculated variance decomposition 192 metrics: (a) LMG metrics for uncorrelated categorical regressors (Lindeman Merenda Gold; Lindeman, 193 1980) and (b) CAR scores for correlated numerical regressors (Correlation-Adjusted coRrelation; Zuber 194 and Strimmer, 2011). While LMG metrics are unweighted averages over orderings of sequential 195 contributions of explanatory variables to models of different sizes (Grömping, 2015), CAR scores are 196 based on simultaneous orthogonalization of correlated explanatory variables and subsequent estimation 197 of marginal correlations between response and decorrelated explanatory variables (Zuber and Strimmer, 198 2011). Shoot biomass and root-shoot ratios were related to soil variables in the top layer. We considered 199 a significance level of p < 0.05.

We used the software R version 3.4.2 (R Core Team, 2019) and the R packages lme4 (Bates et al., 2015), lmerTest (Kuznetsova et al., 2017), pbkrtest (Halekoh and Højsgaard, 2014), emmeans (Lenth, 2018), and relaimpo (Grömping and Lehrkamp, 2018) for statistical analyses and the R packages ggplot2 (Wickham, 2016), GGally (Schloerke et al., 2018), gridExtra (Auguie, 2017), and lemon (Edwards, 2019) for data visualization.

205 **3. Results**

We analysed total root biomass, vertical root distribution, wheat shoot biomass, and root shoot ratios from 24 farms arranged in farming system triplets (conventional, no-till, organic) that were located at eight sites in Switzerland. The sites spread over a distance of just 100 km, yet pedoclimatic characteristics varied considerably among farms (Supplementary table 3). Total root biomass in the 0– 0.75 m soil profile ranged among individual farms from 87–274 g m⁻². Root biomass varied between 211 55–178 g m⁻² in the top layer, 12–53 g m⁻² in the intermediate layer, and 7–43 g m⁻² in the deep layer, 212 corresponding to 55–78%, 10–28%, and 8–22% in the respective layers of total root biomass. Wheat 213 shoot biomass ranged among farms from 909–1692 g m⁻² and root-shoot ratios from 0.07–0.22.

214 3.1. Differences in root parameters between farming systems

Total root biomass was 132 g m⁻² in conventional, 156 g m⁻² in no-till, and 182 g m⁻² in organic 215 216 farming and was significantly higher in organic than conventional (p = 0.018) and intermediate in no-217 till farming (Figure 1). Differences between farming systems were limited to the top layer, where root 218 biomass was 87, 101, and 132 g m⁻² in conventional, no-till, and organic farming, respectively, and 219 significantly higher in organic compared to both conventional (p = 0.003) and no-till farming (p = 0.032; 220 Figure 1). The proportion of topsoil root biomass was highest in organic (73%), lowest in no-till (64%; 221 p = 0.017), and intermediate in conventional farming (66%; Supplementary figure 2). In the intermediate 222 and deep layer, respectively, root biomass and its proportion were similar among farming systems, averaging 27 g m⁻² (18%) and 23 g m⁻² (14%; Figure 1; Supplementary figure 2). 223

224 Wheat shoot biomass at flowering was similar among farming systems and averaged 1311 g m⁻ 225 ² (Figure 1). Consequently, root-shoot ratios were significantly higher in organic farming than in both 226 conventional and no-till farming, irrespective of whether shoot biomass referred to wheat shoot biomass 227 only (organic 0.15; conventional 0.09, p < 0.001; no-till 0.11, p = 0.002) or wheat plus weed shoot 228 biomass (organic 0.14; conventional 0.09, p < 0.001; no-till 0.10, p = 0.005; Figure 1).

229 3.2. Variation in root parameters among sites

230 Total root biomass ranged from 105-221 g m⁻² among the eight farming system triplets and 231 differed significantly between sites (p = 0.011). In addition to the large variation in topsoil root biomass 232 $(75-151 \text{ g m}^{-2}; p = 0.015)$, significant differences between sites also occurred in deep root biomass (11– 35 g m⁻²; p = 0.014), while root biomass was similar in the intermediate layer (27 g m⁻²). Vertical root 233 234 distribution was not significantly affected by site conditions as the proportion of root biomass was similar among sites in all layers (top: 68%, intermediate: 18%, deep: 15%). Similar to the farming 235 system comparison, wheat shoot biomass at flowering was similar among sites (1311 g m⁻²) but root-236 237 shoot ratios differed significantly (0.07-0.18; p < 0.001; Supplementary table 4).

238 3.3. Differences in management and pedoclimatic variables between farming systems and sites

239 Compared to conventional and no-till farming, organic farming involved lower N fertilization 240 intensity (p = 0.003 and 0.025, respectively) but higher weed biomass (p = 0.011 and 0.009, respectively; 241 Supplementary table 3). Topsoil bulk density was higher in no-till than in conventional and organic 242 farming (p < 0.001 each). All other variables were similar among farming systems except for organic C 243 and total N in the intermediate layer, which were higher in organic than in no-till (p = 0.009 and 0.017, 244 respectively) and intermediate in conventional farming (Supplementary table 3). The sites differed in 245 mineral N fertilization intensity, topsoil bulk density, precipitation, soil organic C, total soil N, sand, 246 silt, and clay content in the top and intermediate layer (see Supplementary table 3 for p-values). In the 247 deep layer, all soil variables were similar among both farming systems and sites (data not shown).

248 3.4. Explained variation in root and shoot biomass and root-shoot ratio

249 *Farming system and site*

Farming system and site as explanatory variables accounted for 19 and 54%, respectively, of the variation in total root biomass. In the top, intermediate, and deep layer, respectively, the variation in root biomass was by 32, 11, and <1% explained by farming system and by 44, 39, and 66% by site (Figure 2a). The variation in the proportion of root biomass was by 37, 26, and 20% explained by farming system and 22, 12, and 46% by site in the three soil layers (Figure 2b). Farming system and site, respectively, accounted for 15 and 40% of the variation in shoot biomass (Figure 2c) and 28 and 57% of the variation in root-shoot ratios (Figure 2d).

257 Management and pedoclimatic variables

The outcomes of the two evaluation methods (univariate and multivariate analyses) were largely in agreement, i.e. explanatory variables with high relative importance were also significantly related to the respective response variable, with few exceptions. Relative importance metrics and relations of all variables are shown in Figures 3 and 4 and corresponding p-values in Supplementary table 5. Here, we focus on concordant results for both evaluation methods.

In the top, intermediate, and deep layer, respectively, the investigated management and pedoclimatic variables explained together 78, 74, and 72% of the variation in root biomass and 68, 51,

265 and 70% of the variation in the proportion of root biomass (Figure 3). In the top layer, root biomass and 266 the proportion of root biomass were strongest related to weed biomass (positive) and mineral N 267 fertilization intensity (negative; Figure 3). High importance for root biomass was also assigned to soil 268 organic C (positive) and for the proportion of root biomass to soil bulk density (negative; Figure 3). In 269 the intermediate layer, sowing density explained the largest part of the variation in root biomass and its 270 proportion (positive), while root biomass was additionally strongly related to silt content (negative) and 271 the proportion of root biomass to mineral N fertilization intensity (positive; Figure 3). In the deep layer, 272 precipitation had the highest importance for root biomass and a strong positive effect, while the 273 proportion of root biomass was not significantly related to any variable (Figure 3).

The investigated management and pedoclimatic variables explained 53 and 88% of the variation in shoot biomass and root-shoot ratios, respectively (Figure 4). Available soil P was the only variable with a significant relation (positive) to shoot biomass with high importance, while large parts of the variation in root-shoot ratios were explained by mineral N fertilization intensity (negative) and weed biomass (positive; Figure 4).

279 4. Discussion

280 4.1. Management effects on root biomass allocation to agricultural soils

In this comprehensive on-farm study, we found 40% higher total root biomass under organic 281 282 compared to conventional farming. This is to our knowledge the first study highlighting this substantial 283 farming system effect on root biomass allocation in an on-farm setting characterized by a wide range of 284 management and pedoclimatic conditions across fields. The results thus allow particularly robust 285 conclusions on farming system effects on root biomass allocation. Moreover, conventional agriculture 286 in Switzerland relies to a high degree on cultivation practices that are also typical of organic farming 287 such as long and diverse crop rotations, inclusion of cover crops, and frequent organic fertilization 288 (Nitsch and Osterburg, 2005). A comparison of more divergent systems (e.g. mono-cropping with sole 289 mineral fertilization vs. long crop rotations with sole organic fertilization) might reveal even more 290 pronounced farming system effects. Hence, the here presented results constitute rather conservative 291 estimates for enhanced root C allocation through organic farming in agroecosystems.

292 This study therefore provides supportive evidence for higher root C inputs into organic 293 compared to conventional soils, which has also been found by Chirinda et al. (2012) and Hu et al. (2018) 294 at several long-term field sites in Denmark. Those and our findings suggest an effect size of plus 20-295 40% root biomass in organic compared to conventional systems and thereby oppose the currently 296 prevailing view that organic farming reduces root C inputs along with yields (Lorenz and Lal, 2016). In 297 our study, shoot biomass at flowering showed only a small, non-significant difference among organic 298 and conventional farming and grain yield at harvest was even about 30% lower on the organic than 299 conventional fields (Büchi et al., 2019). Consequently, biomass allocation below and above ground 300 follows different patterns in organic and conventional systems.

301 The farming system effect on total root biomass was mainly a composite of effects of three 302 management-related factors on root biomass in the topsoil. Among the most important drivers was weed 303 biomass, which was an order of magnitude higher in organic (56 g m⁻²) than conventional farming (5 g 304 m^{-2}). Weed roots can trigger over-proliferation of crop roots (Depuydt, 2014) when crops and weeds 305 compete for the same below ground resources (Kiær et al., 2013). However, information on root biomass 306 of weeds would be inevitable to clearly disentangle physiological and methodological causes. As fibrous 307 roots of weeds and crops are often not distinguishable by eye, precise classification requires elaborate 308 methods (Watt et al., 2008; Hirte et al., 2017). As we could remove only clearly identifiable weed roots 309 from the root samples, we assume that weed roots have partly altered sample weight. As a conservative 310 estimate from our weed shoot biomass data and published root-shoot ratios of weeds that correspond to 311 total weed root biomass (Blackshaw et al., 2003; Moreau et al., 2017; Hu et al., 2018), we consider 312 weed root biomass in the organically managed soils to be at most 25 g m^{-2} , thus potentially accounting 313 for up to 50% of the surplus root biomass in organic compared to conventional farming. The presence 314 of weeds, however, is an important aspect of management and contributes in real terms to root biomass 315 and thus organic C inputs to soil.

Similarly important for topsoil root biomass was mineral N fertilization intensity, which was
40% lower on the organic than conventional farms. Low mineral N availability in soil has previously
been found as the main reason for higher root biomass in organic compared to conventional farming
(Chirinda et al., 2012; Hu et al., 2018). In mineral N limited systems, crops invest a larger proportion of

assimilates in below ground organs in order to increase plant interception of soil-borne resources (Lynch *et al.*, 2012). By contrast, total soil N was not related to root biomass in our study, indicating that this
variable, unlike mineral N fertilization intensity, did not represent available soil N fractions adequately.
The importance of available soil P for root biomass was similarly low despite its strong positive effect
on shoot biomass. Phosphorus supply influences rooting characteristics predominantly by altering
topsoil root proliferation, whereas root biomass is only affected under severe P shortage (Hermans *et al.*, 2006). This highlights the outstanding role of N nutrition in the studied farming systems.

327 Soil organic C was the third factor that was prominently related to topsoil root biomass. 328 Although it differed more strongly among sites than farming systems, it was elevated in the organic 329 compared to the conventional soils. This difference proved to be significant in the extended farm 330 network which also included the farms from this study (Colombi et al., 2019). Higher soil organic C can 331 be a consequence of higher root biomass or vice versa as the underlying processes can be bi-directional. 332 On the one hand, continuously increased root biomass enhances soil organic C in the long-term (Lajtha 333 et al., 2014) due to its strong influence on soil organic matter formation (Rasse et al., 2005; Kätterer et 334 al., 2011; Menichetti et al., 2015). On the other hand, higher soil organic C can improve soil aeration 335 and thus stimulate root growth (Colombi et al., 2019). Methodological aspects of sample processing can 336 also entail spurious relationships between soil organic C and root biomass when root samples contain 337 large amounts of extraneous organic matter due to e.g. frequent organic fertilization (Hirte et al., 2017). 338 However, as C inputs to soil by crop residues and organic fertilizers were not substantially increased on 339 the organic compared to the conventional farms (Colombi *et al.*, 2019), we assume a causal relationship 340 between higher root C inputs and increased organic C content in the organically managed soils.

Root biomass in no-till soils was intermediate and not significantly different from that in conventionally and organically managed soils. Interestingly, it was markedly elevated by data from one farm (274 g m⁻²) that used a seed mix of two wheat varieties. Knowledge on root traits in mixed wheat stands is scarce but findings for other crops suggest that competition between genotypes in mixed stands increases biomass allocation below ground compared to single stands (Ninkovic, 2003; Lin et al., 2014). As revealed by the medians, root biomass in no-till farming (138 g m⁻²) was actually much closer to that in conventional (118 g m⁻²) than that in organic farming (178 g m⁻²). This lack of tillage effects on root biomass and, consequently, root-shoot ratios supports previous findings (Anderson, 1988; Williams *et al.*, 2013; Plaza-Bonilla *et al.*, 2014). However, several studies have reported a shift in vertical root distribution due to no-till (Dwyer *et al.*, 1996; Ball-Coelho *et al.*, 1998; Barzegar *et al.*, 2004), which we did not observe. Despite a clear relation to soil bulk density in the top layer, the proportion of topsoil root biomass differed by only 2% between no-till and conventional farming in our study. Instead, weed biomass and mineral N fertilization intensity were the main drivers of vertical root distribution and accounted for the increased proportion of topsoil root biomass by 8% in the organically managed soils.

355 4.2. Pedoclimatic drivers of root biomass

356 Management effects on total root biomass resulted solely from the large differences in root 357 biomass between organic and conventional fields in the topsoil, where farming system explained 32% 358 of the variation. This decreased to basically zero in the subsoil, reflecting the lack of differences in root 359 biomass between farming systems below 0.25 m depth. In contrast to farming system, site governed root 360 biomass not only in the top layer but most prominently in the deep layer, where it accounted for 66 and 361 46% of the variation in root biomass and the proportion of root biomass, respectively. Although the sites 362 spread over a distance of just 100 km, their edaphic characteristics varied strongly, representing the 363 diversity of European soils (Ballabio et al., 2016; Ballabio et al., 2019).

364 Below 0.25 m soil depth, spring precipitation became increasingly important for root biomass 365 and explained even 40% of its variation in the deep layer. We infer that water was not limiting at any of 366 the studied fields as rainfall was 150 mm (50%) higher than mean annual precipitation (30-year climate 367 norm) from April to June 2016. The particularly moist spring conditions even caused below-average 368 yields (Büchi et al., 2019), which was possibly linked to fewer sunshine hours, higher pest and disease 369 pressure, and fewer opportunities for farmers to perform mechanical soil cultivation for e.g. weeding. 370 Instead, since rainfall is one of the most important driving forces of nitrate leaching in agroecosystems 371 (Goulding et al., 2000; Jabloun et al., 2015), the strong positive relation between precipitation and deep 372 root biomass could be an indication of root response to relocation of N.

373 Subsoil root biomass was also prominently linked to soil texture, in particular silt content in the374 intermediate layer and sand content in the deep layer, which ranged between sites from 29 to 40% and

375 31 to 54%, respectively. Their respective negative and positive effects on subsoil root biomass support 376 findings of greater rooting depth in coarse- than medium-textured soils in temperate climate (Schenk 377 and Jackson, 2005). The unfavourable capacity of sandy soils to hold plant-available water and nutrients 378 forces plants to root deeper in order to meet their demand for those resources. In our study, higher 379 nutrient availability in silty soils was likely to result in lower investment of wheat in root growth below 380 the topsoil, which has also been reported from two Swiss long-term field trials (Hirte *et al.*, 2018a).

381 Sowing density, which was the only driver of root biomass entirely independent of farming 382 system and site, had a strong positive impact in the intermediate soil layer. While it has previously been 383 shown that root biomass in the topsoil increases with sowing density, no effects have so far been found 384 in the subsoil (Marcinkevičienė et al., 2013; Hecht et al., 2016). We assume that fertilization and weed 385 control were the main drivers of root response in the topsoil and overlaid the potential influence of 386 sowing density on topsoil root biomass in our study. Our results indicate that effects of sowing density 387 are not confined to topsoils but might easily be masked by concurring drivers, which will need to be 388 addressed in detail in future research.

389 This on-farm study drew on a clustered design with a range of varying cultivation measures to 390 reflect standard agricultural practice. Hence, unexplored management practices constitute an additional 391 source of variation in root biomass, both between and beyond farming systems. For instance, our data 392 were obtained from 15 wheat genotypes, which differed distinctly among and within farming systems. 393 Most genotypes cultivated in organic farming, such as the variety "Wiwa", are long-stalked and thus 394 superior in weed suppression (Dierauer and Klaiss, 2020), but their rooting patterns have yet to be 395 investigated in detail. Wheat genotypes can vary by a factor of five in root biomass (Mathew et al., 396 2019), suggesting that the genotype-environment-management triad that profoundly governs above 397 ground crop parameters (Hillel and Rosenzweig, 2013; Hatfield and Walthall, 2015), also plays a 398 significant role in below ground biomass allocation. We therefore argue that a major part of the 30% 399 variation in root biomass, which remained unexplained in our study, may be assigned to genetic drivers. 400 Thus, future research employing multidimensional networks with completely crossed designs of 401 genotype x environment x management can allow to disentangle the complex interactions of farming 402 system and variety in biomass allocation.

403 4.3. Implications for soil C dynamics, soil C modelling, and climate change mitigation

404 Higher root biomass in organic than conventional topsoils implies considerably larger total below ground C inputs via root biomass and rhizodeposition. The surplus of roughly 25 g m⁻² wheat root 405 406 biomass (excluding weeds) in organic farming can be extrapolated to 25 g m⁻² total below ground C 407 inputs that are additionally allocated to soil by organic compared to conventional wheat in Swiss 408 agricultural practice (C concentration in wheat roots: 44%; rhizodeposition-root ratio: 1.3; Hirte et al., 409 2018a; Hirte et al., 2018b). On top of that, weeds provide an extra source of substantial C inputs to 410 organically managed soils. This stimulates soil organic matter dynamics profoundly, thereby releasing 411 plant nutrients, providing energy for soil microbes, and contributing to soil organic matter build-up 412 (Janzen, 2015; Lorenz and Lal, 2016). Hence, by increased topsoil root C inputs, organic farming fosters 413 soil chemical, biological, and physical processes that enhance soil quality and sustainability of this 414 agroecosystem.

415 As a consequence of higher root-shoot ratios in organic farming, the well-established approach 416 in soil C modelling of deriving root biomass from shoot biomass at harvest and plant C allocation 417 coefficients usually inferred at flowering (Bolinder et al., 1997) may therefore not be suitable for 418 different farming systems. This is supported by recent studies reporting only poor agreement between 419 estimated and actually measured root biomass in organic farming (Taghizadeh-Toosi et al., 2016; Hirte 420 et al., 2018b; Hu et al., 2018). While it has previously been suggested that the major source of this 421 mismatch is the higher shoot biomass in conventional than organic systems at harvest (Hirte *et al.*, 422 2018b; Hu et al., 2018), our findings provide evidence that it is further amplified by management-423 induced differences in root biomass at flowering. The current use of plant C allocation coefficients in 424 soil C modelling therefore needs to be revisited, both with regard to farming systems and plant ontogeny. 425 Among the proposed strategies to mitigate climate change through increased C inputs to 426 agricultural soils (Smith et al., 2014; Paustian et al., 2016), an increase in deep root C is least susceptible 427 to rapid reversal and therefore of particular importance for long-term C sequestration (Kell, 2012). This 428 study provides the first robust data on the potential of agricultural management practices to alter deep 429 root C inputs in the most prevalent arable farming systems in Europe. We give evidence that 430 pedoclimatic drivers substantially govern root biomass below 0.5 m depth, where the impact of farming 431 system is close to zero. Yet, more than one-third of the variation in subsoil root biomass remains 432 unexplained, leaving room for prospects to control crop root C inputs to deep layers. We expect that 433 insights into genetic diversity will contribute to fill this gap and that multidimensional genotype– 434 environment–management networks should become a central part of future research on soil C 435 management.

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445 **Conflict of interest**

446 The authors declare that they have no known competing financial interests or personal relationships that

447 could have appeared to influence the work reported in this paper.

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Figure 1: Root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–0.75 m) soil
layers, wheat shoot biomass, and root-shoot ratios in conventional (CON), no-till (NT), and organic
(ORG) winter wheat fields at flowering in Switzerland (n = 8 sites; average of 4 field replications each).
Error bars refer to standard errors of total root (0–0.75 m) and shoot biomass of 8 sites. Different letters
denote significant differences between estimated marginal means of root biomass in the individual soil
layers (lower case letters) and total root biomass and root-shoot ratios (upper case letters) at p <0.05
(Tukey HSD).



457

458 Figure 2: Explained variation (R² * 100) by farming system and site in (a) root biomass and (b) the
459 proportion of root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–0.75 m) soil
460 layer, respectively, (c) wheat shoot biomass, and (d) root-shoot ratios in 24 winter wheat fields in
461 Switzerland. R² decomposition method: LMG metrics.





463 *Figure 3: Explained variation* ($R^2 * 100$) *by management and pedoclimatic variables in (a) root biomass* **464** *and (b) the proportion of root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–*

465 0.75 m) soil layer, respectively, in 24 winter wheat fields in Switzerland. R^2 decomposition method: CAR

466 scores. Negative / positive relations refer to univariate relations between each management and
467 pedoclimatic variable and root biomass (see Supplementary table 5 for p-values).



469 Figure 4: Explained variation (R² * 100) by management and pedoclimatic variables in (a) shoot
470 biomass and (b) root-shoot ratios in 24 winter wheat fields in Switzerland (soil variables: top layer). R²
471 decomposition method: CAR scores. Negative / positive relations refer to univariate relations between
472 each management and pedoclimatic variable and shoot biomass or root-shoot ratio (see Supplementary
473 table 5 for p-values).

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474 References

- 475 Agroscope, 1996. Schweizerische Referenzmethoden der Forschungsanstalten Agroscope,
- 476 Band 1: Bodenuntersuchungen zur Düngeberatung.
- 477 Anderson, E., 1988. Tillage and N fertilization effects on maize root growth and root: shoot
 478 ratio. Plant and Soil 108, 245-251.
- 479 Auguie, B., 2017. ggridExtra: Miscellaneous Functions for "Grid" Graphics. R package480 version 2.3.
- 481 Ball-Coelho, B.R., Roy, R.C., Swanton, C.J., 1998. Tillage alters corn root distribution in 482 coarse-textured soil. Soil and Tillage Research 45, 237-249.
- 483 Ballabio, C., Lugato, E., Fernández-Ugalde, O., Orgiazzi, A., Jones, A., Borrelli, P.,
- 484 Montanarella, L., Panagos, P., 2019. Mapping LUCAS topsoil chemical properties at
- 485 European scale using Gaussian process regression. Geoderma 355, 113912.
- Ballabio, C., Panagos, P., Monatanarella, L., 2016. Mapping topsoil physical properties at
 European scale using the LUCAS database. Geoderma 261, 110-123.
- Barzegar, A.R., Mossavi, M.H., Asoodar, M.A., Herbert, S.J., 2004. Root mass distribution of
 winter wheat as influenced by different tillage systems in semi arid region. J. Agron 3, 223228.
- Bates, D., Maechler, M., Bolker, B., Walker, S., 2015. Fitting Linear Mixed-Effects Models
 Using Ime4. Journal of Statistical Software 67, 48.
- 493 Blackshaw, R.E., Brandt, R.N., Janzen, H.H., Entz, T., Grant, C.A., Derksen, D.A., 2003.
- 494 Differential response of weed species to added nitrogen. Weed Science 51, 532-539.
- Bolinder, M., Angers, D., Dubuc, J., 1997. Estimating shoot to root ratios and annual carbon
- 496 inputs in soils for cereal crops. Agriculture, ecosystems & environment 63, 61-66.
- 497 Bremner, J., 1965. Total nitrogen. Methods of Soil Analysis: Part 2 Chemical and
- 498 Microbiological Properties 9, 1149-1178.
- Büchi, L., Georges, F., Walder, F., Banerjee, S., Keller, T., Six, J., van der Heijden, M.,
- 500 Charles, R., 2019. Potential of indicators to unveil the hidden side of cropping system
- 501 classification: Differences and similarities in cropping practices between conventional, no-till 502 and organic systems. European Journal of Agronomy 109, 125920.
- 503 Chirinda, N., Olesen, J.E., Porter, J.R., 2012. Root carbon input in organic and inorganic 504 fertilizer-based systems. Plant and soil 359, 321-333.
- 505 Colombi, T., Walder, F., Büchi, L., Sommer, M., Liu, K., Six, J., Van der Heijden, M.G.,
- 506 Charles, R., Keller, T., 2019. On-farm study reveals positive relationship between gas
- 507 transport capacity and organic carbon content in arable soil. Soil 5, 91-105.
- 508 Depuydt, S., 2014. Arguments for and against self and non-self root recognition in plants. 509 Frontiers in plant science 5, 614.
- 509 Frontiers in plant science 5, 614.
- 510 Dierauer, H., Klaiss, M., 2020. Biogetreide. Sortenliste für die Ernte 2020. Ausgabe Schweiz.
- 511 Forschungsinstitut für biologischen Landbau FibL und Bio Suisse, Frick und Basel, pp. 1-6.
- 512 Dignac, M.-F., Derrien, D., Barré, P., Barot, S., Cécillon, L., Chenu, C., Chevallier, T.,
- 513 Freschet, G.T., Garnier, P., Guenet, B., 2017. Increasing soil carbon storage: mechanisms,
- 514 effects of agricultural practices and proxies. A review. Agronomy for sustainable 515 development 37, 14.
- 516 Dwyer, L.M., Ma, B.L., Stewart, D.W., Hayhoe, H.N., Balchin, D., Culley, J.L.B., McGovern,
- 517 M., 1996. Root mass distribution under conventional and conservation tillage. Canadian
- 518 Journal of Soil Science 76, 23-28.
- 519 Edwards, S., 2019. lemon: Freshing up your 'ggplot2'plots.'.
- 520 Enquist, B.J., Niklas, K.J., 2002. Global allocation rules for patterns of biomass partitioning in 521 seed plants. Science 295, 1517-1520.
- 522 Fan, J., McConkey, B.G., Liang, B.C., Angers, D.A., Janzen, H.H., Kröbel, R., Cerkowniak,
- 523 D.D., Smith, W.N., 2019. Increasing crop yields and root input make Canadian farmland a
- 524 large carbon sink. Geoderma 336, 49-58.
- 525 Frasier, I., Noellemeyer, E., Fernández, R., Quiroga, A., 2016. Direct field method for root
- 526 biomass quantification in agroecosystems. MethodsX 3, 513-519.

- 527 FSO, 2017. Agriculture and Forestry: Farm Structure Survey (STRU) (STAT-TAB -
- 528 interactive tables (FSO)). Federal Statistical Office, Neuchatel.
- 529 Ghafoor, A., Poeplau, C., Kätterer, T., 2017. Fate of straw-and root-derived carbon in a
- 530 Swedish agricultural soil. Biology and Fertility of Soils 53, 257-267.
- 531 Goulding, K.W.T., Poulton, P.R., Webster, C.P., Howe, M.T., 2000. Nitrate leaching from the
- 532 Broadbalk Wheat Experiment, Rothamsted, UK, as influenced by fertilizer and manure inputs
- and the weather. Soil use and management 16, 244-250.
- 534 Grömping, U., 2015. Variable importance in regression models. Wiley Interdisciplinary 535 Reviews: Computational Statistics 7, 137-152.
- 536 Grömping, U., Lehrkamp, M., 2018. Package "relaimpo". Relative importance of regressors in 537 linear models. R package version.
- 538 Halekoh, U., Højsgaard, S., 2014. A Kenward-Roger Approximation and Parametric
- 539 Bootstrap Methods for Tests in Linear Mixed Models The R Package pbkrtest. Journal of 540 Statistical Software 59, 30.
- Hatfield, J., Walthall, C., 2015. Meeting Global Food Needs: Realizing the Potential via
 Genetics × Environment × Management Interactions. Agronomy Journal 0.
- 543 Hecht, V.L., Temperton, V.M., Nagel, K.A., Rascher, U., Postma, J.A., 2016. Sowing density:
- a neglected factor fundamentally affecting root distribution and biomass allocation of field
- 545 grown spring barley (Hordeum vulgare L.). Frontiers in plant science 7, 944.
- Hermans, C., Hammond, J.P., White, P.J., Verbruggen, N., 2006. How do plants respond tonutrient shortage by biomass allocation? Trends in Plant Science 11, 610-617.
- 548 Hillel, D., Rosenzweig, C., 2013. Handbook of climate change and agroecosystems: global 549 and regional aspects and implications. World Scientific.
- 550 Hirte, J., Leifeld, J., Abiven, S., Mayer, J., 2018a. Maize and wheat root biomass, vertical
- distribution, and size class as affected by fertilization intensity in two long-term field trials.
 Field Crops Research 216, 197-208.
- Hirte, J., Leifeld, J., Abiven, S., Oberholzer, H.-R., Mayer, J., 2018b. Below ground carbon
 inputs to soil via root biomass and rhizodeposition of field-grown maize and wheat at harvest
 are independent of net primary productivity. Agriculture, Ecosystems & Environment 265,
- 556 556-566.
- 557 Hirte, J., Leifeld, J., Abiven, S., Oberholzer, H.R., Hammelehle, A., Mayer, J., 2017.
- 558 Overestimation of Crop Root Biomass in Field Experiments Due to Extraneous Organic 559 Matter. Front Plant Sci 8, 284.
- Hu, T., Sørensen, P., Wahlström, E.M., Chirinda, N., Sharif, B., Li, X., Olesen, J.E., 2018.
 Root biomass in cereals, catch crops and weeds can be reliably estimated without
- 562 considering aboveground biomass. Agriculture, Ecosystems & Environment 251, 141-148.
 563 Huggins, D.R., Reganold, J.P., 2008. No-till: the quiet revolution. Scientific American 299,
 564 70-77.
- 565 IP-SUISSE, 2019. IP-SUISSE Richtlinien Gesamtbetrieb. Grundanforderungen für sämtliche 566 IP-SUISSE Labelprogramme. In: IP-SUISSE (Ed.), Zollikofen.
- 567 Jabloun, M., Schelde, K., Tao, F., Olesen, J.E., 2015. Effect of temperature and precipitation 568 on nitrate leaching from organic cereal cropping systems in Denmark. European Journal of
- 569 Agronomy 62, 55-64.
- 570 Janzen, H., 2015. Beyond carbon sequestration: soil as conduit of solar energy. European 571 Journal of Soil Science 66, 19-32.
- 572 Kätterer, T., Bolinder, M.A., Andrén, O., Kirchmann, H., Menichetti, L., 2011. Roots
- 573 contribute more to refractory soil organic matter than above-ground crop residues, as
- 574 revealed by a long-term field experiment. Agriculture, Ecosystems & Environment 141, 184575 192.
- 576 Kell, D.B., 2012. Large-scale sequestration of atmospheric carbon via plant roots in natural
- and agricultural ecosystems: why and how. Philosophical Transactions of the Royal SocietyB: Biological Sciences 367, 1589-1597.
- 579 Kiær, L.P., Weisbach, A.N., Weiner, J., 2013. Root and shoot competition: a meta-analysis.
- 580 Journal of Ecology 101, 1298-1312.

- 581 Kuznetsova, A., Brockhoff, P.B., Christensen, R.H.B., 2017. ImerTest package: tests in linear 582 mixed effects models. Journal of Statistical Software 82.
- 583 Lajtha, K., Bowden, R.D., Nadelhoffer, K., 2014. Litter and root manipulations provide
- insights into soil organic matter dynamics and stability. Soil Science Society of AmericaJournal 78, S261-S269.
- 586 Lazicki, P.A., Liebman, M., Wander, M.M., 2016. Root parameters show how management
- alters resource distribution and soil quality in conventional and low-input cropping systems in
 Central Iowa. PloS one 11.
- Lehmann, J., Kleber, M., 2015. The contentious nature of soil organic matter. Nature 528, 60-68.
- 591 Lenth, R., 2018. emmeans: Estimated Marginal Means, aka Least-Squares Means. R 592 package version 1.2.4.
- Li, H., Mollier, A., Ziadi, N., Shi, Y., Parent, L.-É., Morel, C., 2017. The long-term effects of
- tillage practice and phosphorus fertilization on the distribution and morphology of corn root.Plant and soil 412, 97-114.
- Lin, L., Liu, Q., Shi, J., Sun, J., Liao, M.a., Mei, L., 2014. Intercropping different varieties of
- radish can increase cadmium accumulation in radish. Environmental toxicology andchemistry 33, 1950-1955.
- 599 Lindeman, R.H., 1980. Introduction to bivariate and multivariate analysis.
- Lorenz, K., Lal, R., 2016. Environmental impact of organic agriculture. Advances in
- 601 Agronomy. Elsevier, pp. 99-152.
- Lynch, J., Marschner, P., Rengel, Z., 2012. Effect of internal and external factors on root
- 603 growth and development. Marschner's mineral nutrition of higher plants. Elsevier, pp. 331-604 346.
- Lynch, J.P., Wojciechowski, T., 2015. Opportunities and challenges in the subsoil: pathways
 to deeper rooted crops. Journal of Experimental Botany 66, 2199-2210.
- Malhi, S., Lemke, R., 2007. Tillage, crop residue and N fertilizer effects on crop yield, nutrient uptake, soil quality and nitrous oxide gas emissions in a second 4-yr rotation cycle. Soil and Tillage Research 96, 269-283.
- 610 Marcinkevičienė, A., Velička, R., Raudonius, S., Kosteckas, R., 2013. The relationship
- 611 between root biomass and productivity of spring oilseed rape (Brassica napus L.) as
- influenced by crop density and fertilization. Žemdirbystė= Agriculture/Lietuvos agrarinių ir
 miškų mokslų centras, Aleksandro Stulginskio universitetas. Akademija, (Kėdainių r.). ISSN,
- 614 1392-3196.
- Mathew, I., Shimelis, H., Mutema, M., Clulow, A., Zengeni, R., Mbava, N., Chaplot, V., 2019.
- 616 Selection of wheat genotypes for biomass allocation to improve drought tolerance and
- 617 carbon sequestration into soils. Journal of Agronomy and Crop Science 205, 385-400.
- 618 Menichetti, L., Ekblad, A., Kätterer, T., 2015. Contribution of roots and amendments to soil
- 619 carbon accumulation within the soil profile in a long-term field experiment in Sweden.
- 620 Agriculture, Ecosystems & Environment 200, 79-87.
- Mondal, S., Chakraborty, D., Bandyopadhyay, K., Aggarwal, P., Rana, D.S., 2020. A global
- analysis of the impact of zero-tillage on soil physical condition, organic carbon content, and
- 623 plant root response. Land Degradation & Development 31, 557-567.
- 624 Moreau, D., Abiven, F., Busset, H., Matejicek, A., Pagès, L., 2017. Effects of species and 625 soil-nitrogen availability on root system architecture traits–study on a set of weed and crop
- 626 species. Annals of Applied Biology 171, 103-116.
- Ninkovic, V., 2003. Volatile communication between barley plants affects biomass allocation.
 Journal of Experimental Botany 54, 1931-1939.
- Nitsch, H., Osterburg, B., 2005. Cross Compliance (CC) in der EU und Ökologischer
- 630 Leistungsnachweis (ÖLN) in der Schweiz: eine vergleichende Analyse. Arbeitsberichte des 631 Bereichs Agrarökonomie.
- Nkurunziza, L., Chongtham, I.R., Watson, C.A., Marstorp, H., Öborn, I., Bergkvist, G.,
- Bengtsson, J., 2017. Understanding effects of multiple farm management practices on barley
- 634 performance. European Journal of Agronomy 90, 43-52.

- 635 Paustian, K., Lehmann, J., Ogle, S., Reay, D., Robertson, G.P., Smith, P., 2016. Climate-636 smart soils. Nature 532, 49-57.
- 637 Pierret, A., Maeght, J.-L., Clément, C., Montoroi, J.-P., Hartmann, C., Gonkhamdee, S.,
- 638 2016. Understanding deep roots and their functions in ecosystems: an advocacy for more 639 unconventional research. Annals of botany 118, 621-635.
- 640 Plaza-Bonilla, D., Álvaro-Fuentes, J., Hansen, N.C., Lampurlanés, J., Cantero-Martínez, C.,
- 641 2014. Winter cereal root growth and aboveground–belowground biomass ratios as affected
- by site and tillage system in dryland Mediterranean conditions. Plant and soil 374, 925-939.
- 643 Poorter, H., Niklas, K.J., Reich, P.B., Oleksyn, J., Poot, P., Mommer, L., 2012. Biomass
- allocation to leaves, stems and roots: meta-analyses of interspecific variation andenvironmental control. New Phytologist 193, 30-50.
- 646 Poorter, H., Sack, L., 2012. Pitfalls and possibilities in the analysis of biomass allocation 647 patterns in plants. Frontiers in plant science 3, 259.
- Powlson, D.S., Stirling, C.M., Jat, M.L., Gerard, B.G., Palm, C.A., Sanchez, P.A., Cassman,
- 649 K.G., 2014. Limited potential of no-till agriculture for climate change mitigation. Nature 650 Climate Change 4, 678-683.
- Qin, R., Noulas, C., Herrera, J.M., 2018. Morphology and distribution of wheat and maize
- roots as affected by tillage systems and soil physical parameters in temperate climates: an
 overview. Archives of Agronomy and Soil Science 64, 747-762.
- 654 R Core Team, 2019. R: A language and environment for statistical computing. R Foundation 655 for Statistical Computing, Vienna, Austria.
- 656 Rasse, D.P., Rumpel, C., Dignac, M.-F., 2005. Is soil carbon mostly root carbon?
- 657 Mechanisms for a specific stabilisation. Plant and soil 269, 341-356.
- Rich, S.M., Watt, M., 2013. Soil conditions and cereal root system architecture: review and
- 659 considerations for linking Darwin and Weaver. Journal of experimental botany 64, 1193-660 1208.
- Richner, W., Sinaj, S., 2017. Principles of Agricultural Crop Fertilisation in Switzerland
 (PRIF). Agrarforschung Schweiz 8, 276.
- 663 Ritchie, H., Roser, M., 2020. Land Use. Our World in Data.
- 664 Rumpel, C., Chabbi, A., Marschner, B., 2012. Carbon storage and sequestration in subsoil
- horizons: Knowledge, gaps and potentials. Recarbonization of the Biosphere. Springer, pp.
 445-464.
- 667 Russell, A.E., Cambardella, C.A., Laird, D.A., Jaynes, D.B., Meek, D.W., 2009. Nitrogen
- fertilizer effects on soil carbon balances in Midwestern US agricultural systems. Ecologicalapplications 19, 1102-1113.
- 670 Sanaullah, M., Chabbi, A., Maron, P.-A., Baumann, K., Tardy, V., Blagodatskaya, E.,
- 671 Kuzyakov, Y., Rumpel, C., 2016. How do microbial communities in top-and subsoil respond 672 to root litter addition under field conditions? Soil Biology and Biochemistry 103, 28-38.
- 673 Sarker, J.R., Singh, B.P., He, X., Fang, Y., Li, G.D., Collins, D., Cowie, A.L., 2017. Tillage
- and nitrogen fertilization enhanced belowground carbon allocation and plant nitrogen uptake in a semi-arid canola crop–soil system. Scientific reports 7, 1-13.
- 676 Schenk, H.J., Jackson, R.B., 2005. Mapping the global distribution of deep roots in relation to 677 climate and soil characteristics. Geoderma 126, 129-140.
- 678 Schloerke, B., Crowley, J., Cook, D., Briatte, F., Marbach, M., Thoen, E., Elberg, A.,
- 679 Larmarange, J., 2018. GGally: Extension to 'ggplot2'. R package version 1.4.0.
- 680 Schuurman, J.J., Goedewaagen, M.A.J., 1971. Methods for the examination of root systems 681 and roots. Centre for agricultural publications and documentation Wageningen.
- 682 Smith, P., Bustamante, M., Ahammad, H., Clark, H., Dong, H., Elsiddig, E.A., Haberl, H.,
- 683 Harper, R., House, J., Jafari, M., 2014. Agriculture, forestry and other land use (AFOLU).
- 684 Climate change 2014: mitigation of climate change. Contribution of Working Group III to the
- Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge
- 686 University Press, pp. 811-922.
- 687 Smucker, A.J.M., McBurney, S.L., Srivastava, A.K., 1982. Quantitative Separation of Roots
- 688 from Compacted Soil Profiles by the Hydropneumatic Elutriation System 1. Agronomy 689 Journal 74, 500-503.

- 690 Steingrobe, B., Schmid, H., Gutser, R., Claassen, N., 2001. Root production and root
- 691 mortality of winter wheat grown on sandy and loamy soils in different farming systems. 692 Biology and fertility of soils 33, 331-339.
- 693 Swiss Federal Council, 1997. Ordinance on Organic Farming and the Labelling of
- 694 Organically Produced Products and Foodstuffs (Organic Farming Ordinance). In: Council, 695 S.F. (Ed.), 910.18.
- 696 Swiss Federal Council, 2013. Verordnung über die Direktzahlungen an die Landwirtschaft 697 (Direktzahlungsverordnung, DZV). In: Council, S.F. (Ed.), 910.13.
- Taghizadeh-Toosi, A., Christensen, B.T., Glendining, M., Olesen, J.E., 2016. Consolidating
- 699 soil carbon turnover models by improved estimates of belowground carbon input. Scientific 700 reports 6, 32568.
- 701 Watt, M., Magee, L.J., McCully, M.E., 2008. Types, structure and potential for axial water
- flow in the deepest roots of field-grown cereals. New Phytologist 178, 135-146.
- Wickham, H., 2016. ggplot2: elegant graphics for data analysis. Springer.
- Williams, J.D., McCool, D.K., Reardon, C.L., Douglas, C.L., Albrecht, S.L., Rickman, R.W.,
- 2013. Root: shoot ratios and belowground biomass distribution for Pacific Northwest drylandcrops. Journal of soil and water conservation 68, 349-360.
- 707 Zhang, W., Liu, K., Wang, J., Shao, X., Xu, M., Li, J., Wang, X., Murphy, D.V., 2015. Relative
- contribution of maize and external manure amendment to soil carbon sequestration in a long term intensive maize cropping system. Scientific reports 5, 10791.
- 710 Zuber, V., Strimmer, K., 2011. High-dimensional regression and variable selection using
- 711 CAR scores. Statistical Applications in Genetics and Molecular Biology 10.
- 712