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REVIEW

***Porphyromonas gingivalis* is a strong risk factor for Alzheimer's disease**

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23 **ABSTRACT**

24 *Porphyromonas gingivalis* (*P. gingivalis*) is one of the several important bacterial pathogens
25 associated with the sporadic Alzheimer's disease (AD). Different serotypes are either
26 capsulated or are non-capsulated. It has been demonstrated that *P. gingivalis* (non-capsulated)
27 can reproduce the neurodegenerative AD-like changes *in vitro*, and a capsular *P. gingivalis*
28 (strain W83) could reproduce the cardinal hallmark lesions of AD in a wild-type mouse model.
29 All *P. gingivalis* forms express proteolytically active proteases that enable cleavage of the
30 amyloid precursor protein (APP) and tau resulting in the formation of amyloid-beta and
31 neurofibrillary tangles (NFTs). Tau is an established substrate for gingipains, which can cleave
32 tau into various peptides. Some of the *P. gingivalis* fragmented tau protein peptides contain
33 "VQIINK" and "VQIVYK" hexapeptide motifs which map to the flanking regions of the
34 microtubule binding domains and are also found in paired helical filaments that form NFTs. *P.*
35 *gingivalis* can induce peripheral inflammation in periodontitis and can also initiate signaling
36 pathways that activate kinases, which in turn, phosphorylate neuronal tau. Periodontal disease
37 related inflammation has metabolic implications for an individual's peripheral and brain health
38 as patients suffering from generalized periodontitis often have related co-morbidities and are
39 "at risk" of developing AD. The aim here is to discuss the role of *P. gingivalis* behind such
40 associations with the backdrop of huge efforts to test *P. gingivalis* virulence factors clinically
41 (GAIN Trial: Phase 2/3 Study of COR388 in Subjects with AD) with inhibitors, which may
42 lead to an intervention by reducing the pathogenic bacterial load.

43

44 **KEYWORDS:** Alzheimer's disease, gingipains, *Porphyromonas gingivalis*, periodontal
45 disease

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52 **Alzheimer's disease and its association with periodontal disease**

53 The presence of extra neuronal amyloid (A β) plaques and intra-neuronal
54 neurofibrillary tangles (NFTs) in the brain, together with defined clinical signs of cognitive
55 deficit, form the basis of the diagnostic criteria for AD at autopsy [1, 2]. The origins and the
56 roles of A β plaques and NFTs appear quite distinct but together they lead to neurodegeneration.
57 The hippocampus contains abundant intra-neuronal NFTs composed of abnormally
58 phosphorylated tau protein. According to Braak staging of AD; the NFTs spread in the brain
59 in a defined distribution, which allows correlations to be made with the clinical stages [3].
60 Early involvement of tau pathology has been described in the subcortical nuclei such as the
61 locus coeruleus and the pons anatomical areas of the brainstem [3, 4]. This anatomical site of
62 the brain releases norepinephrine in response to emotional and stress related factors. The locus
63 coeruleus also controls the attention and alertness of an individual, so any adverse change in
64 its homeostasis could affect behaviour and mood. Notably, the trigeminal ganglion is located
65 adjacent to the locus coeruleus of the brainstem [5]. It follows that the brainstem and the
66 periodontium communicate via the trigeminal nerve because tooth related pain is registered in
67 the brain. In support, the neurons of the trigeminal nerve are known to be distributed within the
68 periodontal ligament [13]. This provides an important link between periodontitis and the areas
69 of the brain that are affected early in the progression of AD pathology.

70 Periodontitis is a chronic inflammatory disease which damages the tooth supporting
71 tissues i.e. gingivae, periodontal ligament and alveolar bone [6, 7]. Bacteria which can instigate
72 changes in a normally symbiotic microbial community to a dysbiotic state are termed ‘keystone
73 pathogens’. *Porphyromonas gingivalis* (*P. gingivalis*), is a Gram negative coccobacillus
74 shaped bacterium which has long been established as a keystone pathogen for periodontitis [6,
75 7]. This oral commensal becomes pathogenic and can exert its virulence via its endo/exotoxins
76 (surface membrane lipopolysaccharide or LPS, and gingipains) and capsular polysaccharides
77 that allow this bacterium to induce chemokine paralysis of the host and evade immune

78 recognition, even in low abundance. The compromised host innate and adaptive immune
79 responses may be inadequate to control the inflammophilic microbiota and paradoxically, can
80 contribute to connective tissue damage and inflammatory bone loss [8]. Oral microbes can
81 enter the systemic circulation during transient episodes of bacteraemia which can occur with
82 daily oral hygiene activities and dental interventions [9]. Periodontal pathogens including *P.*
83 *gingivalis* have been detected at disparate sites such as the atherosclerotic plaque [10] and AD
84 brains [11, 12]. Pertinent to this, it has been demonstrated in AD transgenic mice that extraction
85 of molar teeth generated release of the cytotoxic A β , and triggered neurodegeneration in the
86 locus coeruleus via its connection with the trigeminal nerve pathway connecting the
87 periodontium [13]. This may provide an explanation for the underappreciated concepts such as
88 missing molar teeth and their contribution to neuronal loss in AD [14-17].

89 This review will cover some salient aspects of *P. gingivalis*, the keystone periodontal
90 disease pathogen, that makes this bacterium “important” as a strong risk factor for developing
91 AD pathophysiology with an emphasis on the pathways that produce hallmark pathology. This
92 is plausible as we now have a better understanding of the *P. gingivalis* secreted exotoxin known
93 as gingipains that expresses cathepsin B proteolytic enzymatic activity, which enables cleavage
94 of amyloid precursor protein (APP), to form A β plaques. Furthermore, gingipains has the
95 potential to disturb tau homeostasis by hyperphosphorylating serine and threonine residues via
96 inflammatory signaling that activate glycogen synthase kinase-3beta (GSK-3 β). In addition,
97 proteolytically active gingipains can hydrolyse tau protein to release fragments [12] with the
98 “VQIINK” and “VQIVYK” hexapeptide motifs that are present in the paired helical filaments
99 that constitute the NFT lesion. These are significant recent leaps in scientific advances in order
100 to understand the risk factor involvement of periodontal disease with the development of AD.

101

102 **What makes *P. gingivalis* a risk factor for AD?**

103 Inflammation is widely thought to contribute to the cognitive decline in AD [18]. However,
104 peripheral episodes of inflammation and/or microbial access to the brain and their impact on
105 triggering cerebral inflammation have largely been ignored. The pathways for microbial access
106 to the brain are many [19] including a leaky blood-brain barrier (BBB). *P. gingivalis* can trigger
107 the peripheral and cerebral immune responses. Periodontitis can exert its influence indirectly
108 by sustaining peripheral inflammation. This together with defective susceptibility genes that
109 normally help with clearance of waste from the brain, can prime microglial cells into a pro-
110 inflammatory phenotype [20].

111 Other major risk factors for AD are comorbid in the presence of the *Apolipoprotein E, allele*
112 *4 (APOE ϵ 4)* susceptibility gene inheritance [21]. One effect is related to the decline of cerebral
113 blood flow across the lifespan of an individual during normal aging but this effect is greater in
114 subjects with *APOE ϵ 4* susceptibility gene inheritance [22]. Lack of an adequate systemic
115 blood flow to the brain in older individuals and those suffering from dementia harbouring the
116 *APOE ϵ 4* susceptibility gene can precipitate cerebrovascular pathologies such as stroke, small
117 vessel arteriosclerosis and others [23-25]. In addition, middle aged hypertension and diabetes
118 also increase the risk of AD [26], this risk may be heightened in the presence of *P. gingivalis*
119 oral infection due to severe periodontitis, which has metabolic implications for an individual's
120 peripheral and brain health [27, 28]. Meta-analysis has shown that there is a 250% higher risk
121 for incident dementia in persons with both *APOE ϵ 4* inheritance and diabetes than those
122 without; and a 35% higher risk for those with *APOE ϵ 4* alone [29]. Figure 1 illustrates the
123 shared risk factors of periodontal disease and AD, which includes the genetic components,
124 comorbidities and environmental factors. This further supports the plausibility of *P. gingivalis*
125 as a risk factor in itself.

126 In addition, the BBB function becomes inadequate during normal aging and in cognitively
127 impaired individuals carrying the *APOE ϵ 4* genetic susceptibility [30-32]. A disrupted BBB

128 during aging and dementia [33] can potentially facilitate the passage of *P. gingivalis* from the
129 systemic circulation into the brain. The scientific rationale for this statement comes from
130 observations made from *P. gingivalis* mono-infected apolipoprotein null mice models of
131 periodontitis for AD pathophysiology. *P. gingivalis* from the oral cavity accessed the brain and
132 the BBB appeared significantly damaged [34-36]. A mechanistic pathway for bacterial entry
133 into the brain is suggested to be via proteolytically active gingipains (the extracellular cysteine
134 proteinases of *P. gingivalis* strain W83) breaking down the epithelial transmembrane proteins,
135 E-cadherin, β_1 integrin and occludin, thereby disrupting the tight junctions between capillary
136 endothelial cells. This increases the permeability of the BBB [37]; and in addition, systemic
137 proinflammatory cytokines also can potentially disrupt the BBB [38]. It is assumed that when
138 tight junction integrity is sufficiently disturbed, passage of bacteria into the brain is likely to
139 take place. In supporting the adverse effects of the cytokine on the BBB integrity, Vernal et al.
140 [39] have reported that capsular serotype K1 and K2 *P. gingivalis* strains outside the central
141 nervous system induce an enhanced secretion of pro-inflammatory interleukin cytokines (IL)-
142 1β , IL-4, IL-6, IL-10, IL-17, interferon (IFN)- γ , and tumor necrosis factor (TNF)- α in
143 macrophages and dendritic cells, implying capsular *P. gingivalis* strains have the armoury for
144 potentiating BBB permeability through cytokine liberation. Encapsulated bacterial infections
145 for example, *P. gingivalis* W83, harbours an additional virulence factor in the form of the
146 capsular polysaccharide or A-lipopolysaccharide (A-LPS) which is different from the
147 lipopolysaccharide (LPS) of the outer membrane of non-encapsulated bacteria for example *P.*
148 *gingivalis* (strain ATCC 33277) [see 40]. The A-LPS in the capsulated *P. gingivalis* (W83)
149 plays a vital role in this bacterium's virulence development by inducing proinflammatory
150 cytokine paralysis [41] as it is connected to the post-translational additions of Arg-gingipains
151 that make the bacterium more virulent [42, 43].

152 Therefore, a person harbouring capsular forms of *P. gingivalis* could have a higher
153 risk of developing AD, compared with those who carry the non-capsulated bacterium. This
154 may be one reason why not all individuals who suffer from periodontal disease also manifest
155 AD [15, 43]. Prevalence studies among the Dutch population identified K6 serotype (capsular)
156 as the most prevalent (23%) [44], whilst, the K5 serotype (capsular) was frequently observed
157 in 25% of the subgingival isolates examined from Indonesian AD subjects [45].

158
159 ***P. gingivalis* gingipains, the most critical virulence factor**

160 *P. gingivalis* produces gingipains R, (RgpA, RgpB) and gingipain K, (Kgp). Gingipains
161 are classified as collagenases and trypsin-like cysteine proteinases [46], and are secreted by all
162 strains (with/without capsule) of *P. gingivalis* [47, 48]. Together they degrade a variety of
163 proteins involved in the immune response [49-52]. Here, our interest is in the trypsin-like
164 cysteine proteinases, secreted by all strains of *P. gingivalis* [47, 48], which specifically cleave
165 peptides at Arginine-Xaa and Lysine-Xaa (Xaa = any amino acid) from the C-terminus. The
166 Arg-specific proteolytic activity is encoded by *rgpA/B* genes while the Lys-specific activity is
167 encoded by the *kgp* gene. Rgp and Kgp are enzymes responsible for the trypsin-like activity
168 associated with *P. gingivalis* and hence are important in the context of tau NFTs [47].

169 Recent studies have postulated that tau protein (associated with microtubules) and
170 actin are substrates of Kgp gingipain [13, 53]. Neither Lys-gingipain nor Arg-gingipain are
171 inhibited by internal protease inhibitors such as cystatins and α -antichymotrypsin and are
172 therefore, able to diffuse into host tissues. Furthermore, gingipains are thermally stable from
173 0-45°C and over a range of pH fluctuations 5.5 to 10.5 [54]. It is not surprising in view of the
174 sensitivity of tau to trypsin [55], that it is a potential substrate for gingipains *in vivo* [12]. This
175 exciting finding is potentially a major breakthrough for therapy based on low molecular weight
176 small-molecules designed to block the toxic effects of the different types of gingipains secreted

177 by *P. gingivalis*, which are currently in Phase 3 clinical trials. An aspect of the gingipains
178 inhibitor COR388 based therapy is provided elsewhere [56].

179

180 ***P. gingivalis* and its association with neurodegeneration**

181 AD is a neurodegenerative disease and hence neuronal loss is pivotal to the disease
182 process. To this end, Goto et al. [13], performed surgical extractions of molar teeth in their
183 triple transgenic AD (3xTg-AD) mice and uncovered a neurodegenerative pathway involving
184 the trigeminal distribution within the periodontal ligament to the locus coeruleus. From the
185 locus coeruleus, the tau pathology subsequently spreads to the hippocampus. The loss of molar
186 teeth in AD patients is predominantly associated with periodontitis and in general, tooth loss
187 has been linked to a higher risk of AD [14-17]. *P. gingivalis* (ATCC 33277) has been reported
188 to cause AD-like neurodegeneration in infected neurons derived from induced pluripotent stem
189 cells (iPSC) in an *in-vitro* culture system with persistent expression of active gingipains,
190 resulting in a 25% loss of neurons over three days [57]. The differentiated iPSC neurons can
191 be maintained for months and offer a time efficient *in vitro* analysis of measuring neuronal
192 degeneration compared to examining the years of AD-related neurodegenerative processes in
193 humans. Furthermore, *P. gingivalis* can invade and survive within neurons and generate intra-
194 neuronal gingipains, which are proteolytically active, implying the plausibility of direct
195 neurodegeneration associated with NFT lesion formation in AD taking place [57].

196

197 ***P. gingivalis* infection and periodontitis are linked to deteriorating memory**

198 The first interventional study with human AD subjects was carried out by Rolim et al. [58] in
199 which they demonstrated a beneficial outcome of dental treatment in AD subjects. The reported
200 benefit to the patients with mild AD was in terms of reduced orofacial pain and an improvement
201 in the mandibular function and periodontal indices. These improvements were maintained until

202 the last evaluation after six months, and were followed by a reduction in the functional
203 cognitive impairment. Several proof of concept studies carried out in either mice with
204 experimentally induced periodontal disease with an oral infection with *P. gingivalis*, or with
205 the introduction of its LPS have indicated that inflammatory signaling pathways contribute to
206 a clinical phenotype in which impaired learning and memory is observed [59-61]. The
207 inflammatory basis of deteriorating memory is upheld by the results from clinical trials in
208 human AD subjects with periodontitis [62]. For a more comprehensive overview, see Singhrao
209 and Olsen [63].

210

211 ***P. gingivalis* and its association with AD diagnostic lesion formation**

212 *P. gingivalis* has the potential to induce inflammation peripherally due to periodontitis [64] and
213 subsequently in the brain via its intracerebral entry or entry of its virulence factors (LPS and
214 gingipains) [65-67]. Gingipains released by *P. gingivalis* which have similar APP-cleaving
215 action as cathepsin B (of the host) [12], interact with the APP signaling pathways (amyloid
216 cascade) to release A β [60]. In this process, gingipains together with LPS can proteolytically
217 activate kinases such as GSK-3 β which subsequently phosphorylates neuronal tau [57, 66].

218 Up until now, abnormally phosphorylated tau protein has not featured negatively in
219 the pathophysiology of periodontal disease *per se*. However, Adamowicz et al. [68] implicated
220 the role of GSK-3 β in bacterial-induced periodontitis because its inhibition rescued bone loss.
221 Thus, GSK-3 β may be influencing phosphorylation of brain tau via immune responses
222 mediated by *P. gingivalis*, in the Ilievski et al. [66] and Haditsch et al. [57] studies. The
223 introduction of phosphorylated tau is an important point, especially since the intraneuronal
224 cytoskeletal alterations precede the formation of amyloid in AD locus coeruleus [69]. This is

225 interesting when compared with the Tg mouse tooth extractions where cytotoxic A β , triggered
226 neurodegeneration in the same brain region (i.e locus coeruleus) [13].

227

228 ***P. gingivalis* and the A β plaque lesion**

229 The extra neuronal AD plaque is composed of A β forming the basis of the “Amyloid
230 Hypothesis” [70]. A β is a cleavage product of APP, which is seen in internal vesicular
231 membranes, including the Golgi apparatus and endosomes [63, 71]. Hence, a direct
232 extracellular deposition of soluble/insoluble A β as well as an intra-cellular processed soluble
233 A β [63, 72, 73] both contribute to the extracellular fibrillary/insoluble (diffuse and neuritic)
234 plaques in AD brains. The APP proteolytic cleavage is completed by the proteolytically active
235 secretase enzymes (α -, β - and γ -secretases). In rare familial AD cases however, there is a
236 mutant form of APP (Swedish double mutation APP K670M, N671L) which has a double
237 mutation at the start of the cleavage site of the beta-site of APP cleaving enzyme 1 (BACE -1).
238 BACE-1 has been shown to bind this mutated site with much higher affinity than the wtAPP
239 and thus cleaves to form A β much more readily [73]. Notably, the enzyme cathepsin B, which
240 is expressed in secretory vesicles within neurons, is shown, conversely, to bind with high
241 affinity to wtAPP (and not to the Swedish mutant APP) and has been suggested as a likely
242 candidate for production of A β in the sporadic form of AD [73]. As described earlier,
243 gingipains, the exotoxin of *P. gingivalis*, also has cathepsin B-like activity [12] and may act to
244 cleave APP [73]. Wu et al. [60] demonstrated the host related cathepsin B to interact with APP
245 to liberate A β . In order to determine the intracellular processing of APP by *P. gingivalis*-LPS
246 (Pg-LPS), Wu et al. [60] challenged the wt-APP mice and observed that the host’s cathepsin
247 B, together with inflammatory mediators (IL-1 β), directed APP proteolysis to release soluble
248 amyloid which they interpreted to be A β 42 species. Why the host’s cathepsin B activity and
249 not the *P. gingivalis* cathepsin B-like activity are acting here is because we postulate that the

250 highly purified Pg-LPS used by Wu et al. [60] is likely to have been denatured by the
251 purification process leaving the LPS activity intact. Following *P. gingivalis* infections, an
252 increase in peripheral A β _{1-40/42} accumulation within periodontal tissues have been shown in
253 mice models and in the human gingival tissues and human serum thereby potentially adding to
254 the A β pool in the AD brain [74-76]. To this end, Zeng et al. [77] identified advanced glycation
255 end products (AGE) as a likely receptor for A β _{1-40/42} in cerebral endothelial cells implying AGE
256 products-receptor are a plausible mediator of cerebrovascular-related A β accumulation after *P.*
257 *gingivalis* infection in their mouse model. This supports the hypothesis that patients harboring
258 the generalized form of periodontal disease may be “at risk” of developing AD via multiple
259 pathways. Key evaluations of prospective and retrospective population based data have shown
260 that chronic periodontal disease that exists for longer than ten years results in a doubling of the
261 risk for the sporadic form of AD [78-80]. Putting this information in perspective, the 10 year
262 lag allows amyloid-beta (A β) plaques to reach a plateau in the human host and this is when the
263 host begins to indicate the earliest stage (mild cognitive impairment or MCI or prodromal) of
264 AD. Following this stage, the pathological cascade of progressive AD would take over. This is
265 interesting because the pieces of the jig-saw puzzle that form the picture of AD is beginning to
266 emerge from reported scientific observations regarding the co-morbidity between periodontal
267 disease and AD in some patients. Periodontal disease can start at any age and the time it takes
268 to become chronic may vary from individual to individual. If that individual was vulnerable to
269 manifesting AD later in life, it should be possible to predict the risk age of that individual from
270 the time that their periodontitis became chronic. Taken together, an early detection of disease
271 means patients may benefit from an earlier medical intervention.

272

273 **“Gingipains” interaction with tau protein: what does this mean for neurofibrillary**
274 **tangles?**

275 The NFTs represent hyperphosphorylated tau proteins binding to microtubules.
276 Hyperphosphorylation of microtubules is abnormal because normal tau becomes insoluble and
277 subsequently aggregates. This also causes the collapse of microtubules into non-membrane-
278 bound masses of abnormal paired helical and straight filaments (PHF) which are found in the
279 perinuclear cytoplasm of specific neurons. These constitute NFTs [81, 82]. Understanding the
280 formation of the NFT lesion due to bacterial interaction is important as autopsied brains from
281 AD cases have confirmed the presence of the following microbes (Actinomycetes [83, 84], *P.*
282 *gingivalis* [12, 56], *Helicobacter pylori*,
283 and *Chlamydia pneumoniae* [85], Herpes Simplex type 1 virus (HSV1) [86], select species of
284 oral/non-oral spirochaetes [87], and select fungi [88]). Furthermore it has been reported that
285 *Bacteroides* species such as *P. gingivalis* are more virulent as a result of mixed infection with
286 the *Actinobacillus actinomycetemcomitans* bacteria facilitating their movement between
287 different organs [89, 90].

288 Dominy et al. [12] demonstrated tau protein as a substrate for *P. gingivalis* protease
289 gingipains and as a consequence, the resultant tau protein fragments can be released into the
290 brain's parenchymal tissues. Depending on their size (Table 1), these extracellular
291 phosphorylated tau fragments may be directly toxic to other neurons. The smaller sized
292 phosphorylated tau fragments may be taken up by other connecting cells at the synaptic clefts
293 during neurotransmitter uptake, thereby facilitating its spread from neuron to neuron and
294 subsequently spreading pathology.

295

296 **Tau cleavage by gingipains and its involvement in paired helical filaments**

297 *P. gingivalis* protease gingipains have been demonstrated to co-localize with
298 microtubules and paired helical filaments (PHF) constituting NFTs in AD brains [12]. As

299 mentioned earlier, tau protein can be hydrolyzed by gingipains into several fragments both
300 from the N and C termini [12]. Six out of several fragments have “VQIINK” and “VQIVYK”
301 hexapeptide common motifs that bind to the microtubule binding domains flanking regions,
302 which support the microtubule filaments (Figure 2, Table 1). This implies that six of the several
303 tau fragments released by gingipains are from the pivotal sites of the functional microtubules
304 and agrees with previously reported observations [12]. Furthermore, PHF filaments also have
305 VQIINK and VQIVYK hexapeptide signatures [see ref 91]. Therefore, tau cleavage at
306 VQIINK and VQIVYK hexapeptides from the regions of considerable functional significance
307 to microtubules “would collapse” the intact microtubule assembly with the said hexapeptide
308 peptide amino acid signatures within tau and PHFs, constituting the NFT lesions [12, 91].
309 Further research is needed to clarify the role of gingipains fragmented tau peptides in the
310 pathogenesis of AD.

311

312 ***P. gingivalis* phosphorylates tau protein**

313 Tau protein binds to microtubules and is prone to hyperphosphorylation.
314 Hyperphosphorylation of tau protein signifies a pathological change, which precedes NFT
315 formation. It is believed that phosphorylated tau proteins accumulate within neurons prior to
316 NFT formation [92]. What causes this shift between bound to free pathological tau in neuronal
317 cells is not clear, but *P. gingivalis* enzyme activity hydrolysing this protein as shown by
318 Dominy et al. [12] cannot be ruled out. Haditsch et al. [57] demonstrated an increased tau
319 phosphorylation at two residues (enhanced tau phosphorylation at Thr231 and at Ser396)
320 following *P. gingivalis* infection in an iPSC differentiated neuronal cell culture model.
321 Furthermore, the capsular serotype K1 *P. gingivalis* W83 strain has been shown to have the
322 potential to contribute to tau phosphorylation at Ser396 in the *in vivo* wild-type mouse model
323 [66]. These are accepted phosphorylation sites as evaluated previously by Hanger et al. [93].

324 However, which kinase may be responsible for phosphorylating Ser396 and Thr231 following
325 *P. gingivalis* infection is not clear, but glycogen synthase kinases-3 beta (GSK-3 β) is a strong
326 candidate [93]. Recently Liu et al. [94] observed in their *gingivalis*-infected microglial cell
327 model that phosphoinositide 3-kinase (Pi3K)/ protein kinase B1 (Akt) and mitogen-activated
328 protein kinase/extracellular signal-regulated kinase (ERK) kinase/ERK pathways were
329 activated. Our study in which purified *P. gingivalis* LPS (Pg-LPS) was applied to a
330 neuroblastoma cell line, *in vitro* cell model also demonstrated that the PI3K/AKT pathways
331 were activated [95]. In addition, Bahar and Singhrao [95] showed GSK-3 β and forkhead box
332 class O1 (FOXO1) and NF-kB mRNA expression was also up-regulated and that this was
333 dependent on the MyD88 pathway [95]. The importance here is that GSK-3 β is one of the
334 enzymes that can phosphorylate tau residues Ser396 and Thr231 [93] and its activation
335 implicates the onset of inflammatory signaling [95] that are known to be involved in AD
336 pathophysiology [96, 97]. This implies that *P. gingivalis* infection plays an important role in
337 the infected cells where balance of inflammation and inflammation responsive kinases (e.g.
338 GSK-3 β) are tipped in favour of phosphorylation, NFT lesion formation and subsequent
339 pathophysiology of AD.

340

341 **Conclusions**

342 This review has applied a pathobiome concept in substantiating the link between *P.*
343 *gingivalis* infection and AD lesions. It is clear from the human and proof of concept studies in
344 animal models that whole bacteria and their constituent endo/exotoxins enter the central
345 nervous system. In-situ, this bacterium has a range of enzymes that are shown to interact with
346 APP and tau, deregulating their structure and intracellular processing, with resultant formation
347 of A β and PHF respectively. We appreciate that other bacterial, fungal or viral pathogens

348 implicated in AD may follow different pathways towards AD pathophysiology compared to
349 those described here for *P. gingivalis*.

350 It is evident that *P. gingivalis* is a potentially significant etiological agent for AD
351 pathophysiology. Whilst generalised periodontitis develops around middle age, the sporadic
352 form of AD is of much later onset. The lag phase between the two comorbidities could provide
353 us with a mechanistic association between the two diseases and a window of opportunity to
354 inhibit the toxic insults of gingipains in patients with periodontitis, and its downstream
355 inflammatory effects such as systemic inflammation contributing to the risk of developing AD
356 or increasing the severity of dementia by hyperphosphorylating tau. Hence, the clinical “GAIN
357 Trial”: Phase 2/3 Study of COR388 in Subjects with Alzheimer's disease. (ClinicalTrials.gov
358 Identifier: NCT03823404) is key to testing this hypothesis and advancing the adjunctive
359 management of healthcare landscape in preventing and/or slowing down AD. It is imperative
360 that the oral health component is included as a modifiable risk factor in AD public health
361 messages along with other preventative advice such as keeping active, eating healthily and
362 exercising.

363

364 **Disclosure statement**

365 The authors declare no competing interests.

366

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371

372 **References**

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- 723 Table 1: Peptides of interest taken from reference 12. Amino acids indicated in green colour
724 are putative phosphorylation sites in the tau protein. * indicates peptides of significance to the
725 PHF tau constituting neurofibrillary tangles that bind to microtubule binding domain (MBD).
726

Peptide number	Region in Tau N-C termini	Nucleotide/ Peptide
1 N-terminal projection domain	K87- R126	QAAQPHTEIPEGTTAEEAGIGDTPSLEDEAAAGHVTQAR

Tau(1–165)		
2 Proline rich domain 166-242	R211- R221	TPSLPTPPTR
3* MBD R1/2(274–304)	K259- K290	267 HQPGGGK VQIINKKLDLSNVQ S¹⁸⁴ K287
4* MBD-2 R2(274–304)	K28- K290	275 VQIINKKLDLSNVQ S¹⁸⁴ K285
5* MBD-2/3 305–335	K298- K317	298 HVPGGG SVQIVYK PVDLSK 317
6* MBD-2/3 305–335	K298- K321	298 HVPGGG SVQIVYK PVDLSK VT SK 321
7* MBD-2/3 305–335	K294- K317	294 DNIK HVPGGG SVQIVYK PVDLSK 317
8* MBD-2/3 305–335	K294- K321	294 DNIK HVPGGG SVQIVYK PVDLSK VT SK 321
9 C-terminal domain (368–441)	R406- K438	HLSNV SST⁴¹⁴ GS⁴¹⁶ IDMVD SPQLATL DEV SALAK

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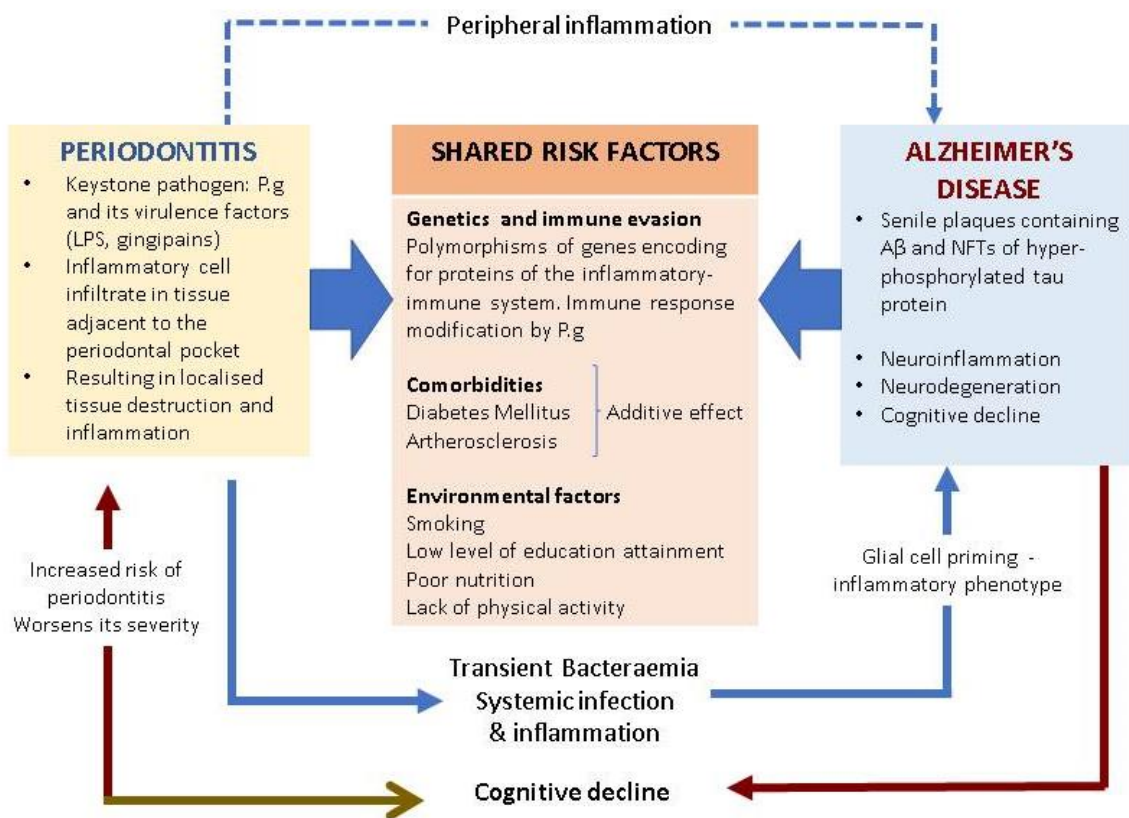
733 **Figure Legends**

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736 Figure 1: The keystone periodontal pathogen, *P. gingivalis* (P.g) can enter the bloodstream
 737 during episodes of transient bacteraemia and gain access to the brain via multiple routes
 738 including a leaky blood-brain barrier (BBB). The microbial invasion triggers the cerebral
 739 immune response resulting in neuroinflammation and in the formation of the two diagnostic
 740 lesions of AD. Periodontitis can exert its influence indirectly by sustaining peripheral
 741 inflammation (blue dotted arrow). This can affect glial cells by priming them into a pro-
 742 inflammatory phenotype. In addition, this could also overload and overwhelm the clearing of
 743 toxic neuropeptides from the central nervous system (CNS). The potential causal relationship
 744 of periodontitis and AD is further supported by the shared risk factors.
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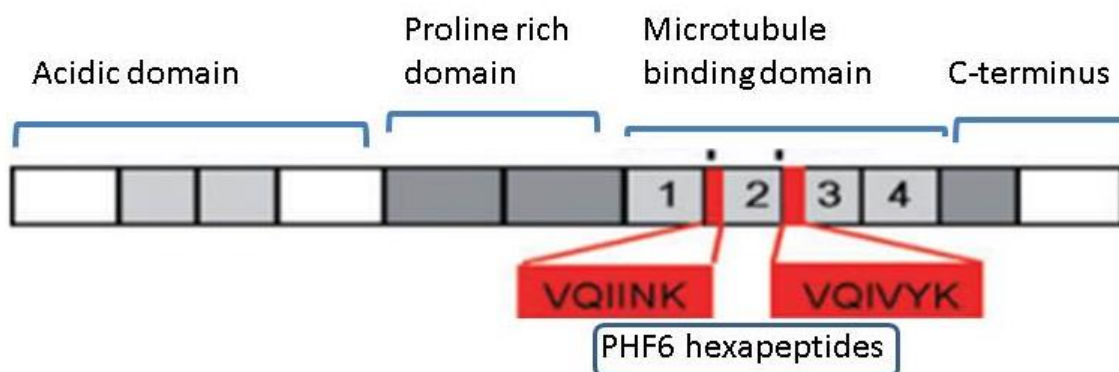
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752 Fig 2. Schematic of tau protein and domain organisation to help understand the position of
753 peptides in Table 1 and the 1-4 microtubule binding domains of Tau to microtubules.

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