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Microsieves for the detection of circulating tumor cells in leukapheresis product in non-small cell lung cancer patients

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Background: Circulating tumor cells (CTC) in non-small cell lung cancer (NSCLC) patients are a prognostic and possible therapeutic marker, but have a low frequency of appearance. Diagnostic leukapheresis (DLA) concentrates CTC and mononuclear cells from the blood. We evaluated a protocol using two VyCAP microsieves to filter DLA product of NSCLC patients and enumerate CTC, compared with CellSearch as a gold standard.

Methods: DLA was performed in NSCLC patients before starting treatment. DLA product equaling 2×10^8 leukocytes was diluted to 9 mL with CellSearch dilution buffer in a Transfix CTC tube. Within 72 hours the sample was filtered with a 7 μm pore microsieve and subsequently over a 5 μm pore microsieve. CTC were defined as nucleated cells which stained for cytokeratin, but lacked CD45 and CD16. CellSearch detected CTC in the same volume of DLA.

Results: Of 29 patients a median of 1.4 mL DLA product (range, 0.5–4.1) was filtered (2% of total product) successfully in 93% and 45% of patients using 7 and 5 μm pores, respectively. Two DLA products were unevaluable for CTC detection. Clogging of the 5 μm but not 7 μm microsieves was positively correlated with fixation time ($\rho=0.51$, $P<0.01$). VyCAP detected CTC in 44% (12/27) of DLA products. Median CTC count per mL DLA was 0 [interquartile range (IQR): 0–1]. CellSearch detected CTC in 63% of DLA products (median =0.9 CTC per mL DLA, IQR: 0–2.1). CTC counts detected by CellSearch were significantly higher compared with VyCAP ($P=0.05$).

Conclusions: VyCAP microsieves can identify CTC in DLA product, but workflows need to be optimized.

Keywords: Diagnostic leukapheresis (DLA); circulating tumor cell (CTC); non-small lung cancer (NSCLC); liquid biopsy; biomarker; VyCAP microsieves

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Introduction

Circulating tumor cell (CTC) have prognostic value in non-small lung cancer (NSCLC) (1-5). However, in NSCLC CTC are detected in only 30% of metastatic NSCLC patients and in low frequencies of 1–2 CTC per 7.5 mL of blood (1-8). CTC are likely present in the majority of metastatic patients but the current available methods are only capable of screening limited volumes of blood (9,10). As CTC have a similar density as mononuclear cells they can be extracted from the blood by means of a diagnostic leukapheresis (DLA), allowing the screening of a large volume of blood for CTC (10-13).

In the DLA product the CellSearch platform identifies CTC in higher frequencies and in more patients compared to peripheral blood (11-13). CellSearch uses positive immunomagnetic selection to extract the epithelial cell adhesion molecule (EpCAM) positive CTC. Due to the high numbers of leukocytes in the DLA product, which have non-specific interaction with the immunomagnetic particles targeting EpCAM, usage of the CellSearch platform in processing DLA is limited (11). VyCAP microsieves collect CTC by their relative size and rigidity, allowing for the detection of both EpCAM positive and negative cells (1-3). In addition, modified versions of these filters may allow isolation of single CTC for genomic analysis, or to perform functional tests in captured CTC (14-18). In this study we looked into the capability of VyCAP microsieves to filter the DLA product of NSCLC patients to enumerate CTC and compare the CTC counts per patient with those of CellSearch.

Methods

Patient inclusion and clinical data

Consecutive patients with proven NSCLC starting their treatment were prospectively included in an exploratory cohort. Patients who had an Eastern Cooperative Oncology Group Performance Status (PS) of 0–2 and did not use anticoagulation medication or suffer from clotting disorders were selected. All patients gave informed consent. The study was approved by the regional Medical Ethical Committee (NL55754.042.15) and registered in the Dutch trial register (NL5423), conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Diagnostic leukapheresis procedure

DLAs were performed with the Spectra Optia® Apheresis

System using an intermediate density layer set (IDL) and software version 11 (Terumo BCT inc, Lakewood, CO, USA). The standard continuous MNC protocol was used with a packing factor of 4.5, collection pump set to 1 mL per minute, hematocrit minus 3 percent points and a flexible inlet flow. Anticoagulant Citrate Dextrose Solution A (ACDA) was used for anticoagulation in a dilution of 1:11, further adjusted when aggregate formation was observed in the collection chamber.

We aimed to process the total blood volume (TBV) of the patient, as estimated with the formula of Nadler (19). DLA collection would be performed at a hematocrit around 5%.

DLA procedure efficacy was measured by determining the percentage of lymphocytes that were captured compared to the number that passed through the machine (lymphocytes in DLA product divided by lymphocytes in blood that was processed).

DLA product processing

A DLA aliquot with a volume equaling that of 2×10^8 leukocytes was diluted with CellSearch Circulating Tumor Cell Kit Dilution Buffer (Menarini Silicon Biosystems, Huntingdon Valley, PA, USA) to 9 mL and transferred to a CTC-TVT Transfix tube (Cytomark, Buckingham, United Kingdom) and stored at room temperature for at least 24 h. The sample was filtered within 72 h after completion of the DLA procedure using two VyCAP microsieves. This filtration in two steps was because of the experiences in the CTC-Trap study, where CTC were lost and only a small volume of DLA could be filtered (13). Filtration and CTC detection occurred according to manufacturer instructions [version 2.1, September 2017 (20)]. Firstly, the whole DLA sample is filtered using a microsieve with pores of 7 μm using a pressure of 30–250 mbar. Subsequently, the 7 μm microsieve filtrate, collected in the waste compartment of the VyCAP disposable, was passed through a microsieve with 5 μm pores at a pressure of 150–250 mbar. Immediately after filtration, the cells on the filter are stained with immunofluorescent labels using a permeabilization buffer, followed by the addition of a staining cocktail containing pan-cytokeratin CK-11 (PE, Cell Signaling Technology, #5075S, clone C11), pan-cytokeratin AE1/AE3 (eFluor 570, eBioscience, 41-9003-82, clone AE1/AE3), CD45 (APC, eBioscience, 17-0459-41/42, clone HI30) and CD16 (APC, BioLegend, #302011, clone 3G8). Mounting medium containing DAPI was added followed by the addition of a cover slip. The microsieves were scanned and images of the cells on the microsieves were acquired.

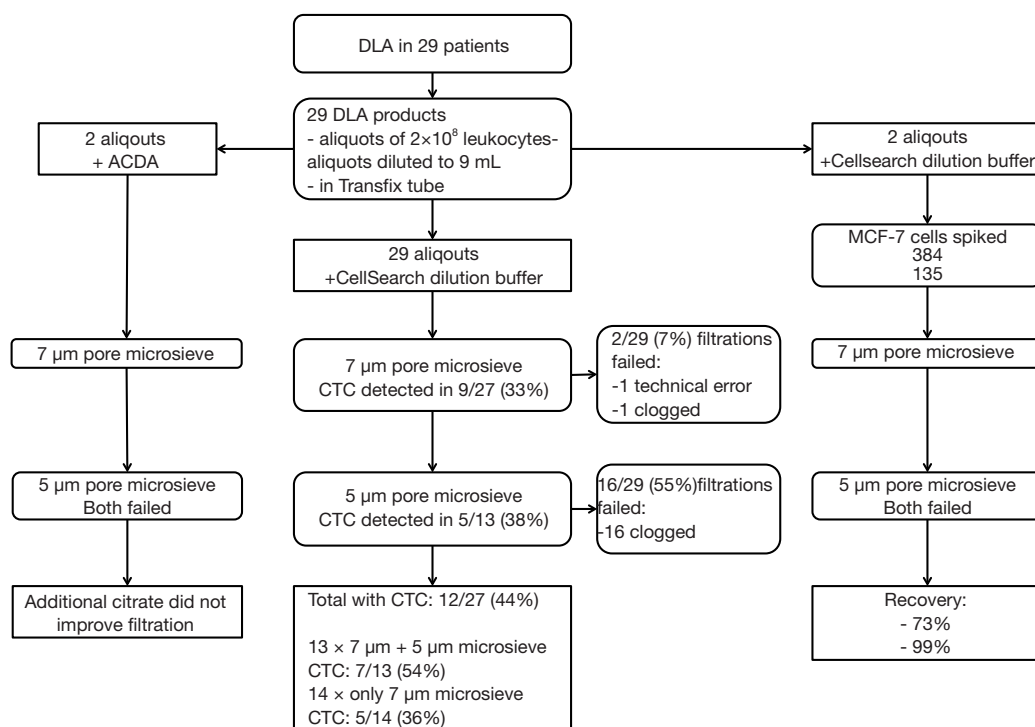


Figure 1 Processing diagnostic leukapheresis (DLA) samples for circulating tumor cells (CTC) filtration.

CTC, defined as cells expressing cytokeratins and containing a nucleus, but lacking CD45 and CD16 were counted. CTC counts are reported per mL DLA product.

Two samples which clogged were additionally labeled with CD41 (FITC, BD Pharmingham, cat#557296) to detect platelets. In two patients a second DLA sample, in which ACDA was used to dilute the sample to 9 mL instead of CellSearch dilution buffer, was used to determine the influence of anticoagulants on the formation cell clusters and cell clumps in the DLA product.

Comparison with CellSearch

We detected CTC in DLA product with CellSearch as previously described (13). In short, a DLA product volume containing in total 2×10^8 leukocytes was diluted to 7.5 mL with CellSearch dilution buffer in a CellSave CTC tube (Menarini siliconbiosystems, Bologna, Italy). CellSearch would then be used to identify CTC according to manufacturer’s instructions.

Spiking tumor cells in DLA product

To determine the recovery of tumor cells in DLA product,

DLA products of two patients were spiked with 384 and 135 MCF-7 cells that were manually counted before transferring these to the DLA sample. MCF-7 cells were exposed to the sample concentration of transfix for the same times as the DLA samples.

Results

NSCLC patients and apheresis

Twenty-nine NSCLC patients underwent an apheresis (Tables S1,S2). The median estimated patient total blood volume was 5.2 L [interquartile range (IQR): 4.7–5.8], of which 4.6 L (IQR: 3.9–5.5 L) was processed in 102 minutes (IQR: 87–110). DLA procedures had a median efficacy (%lymphocytes captured that passed through the machine) of 57% (IQR: 55–65%). The median volume of DLA product was 88 mL (IQR: 70–96), including 12 mL ACDA (IQR: 11–13) for anticoagulation. The total number of cells used for filtration was 2×10^8 , which equaled a median volume of 1.4 mL (IQR: 1.1–2.3) of DLA product, constituting 2.1% (IQR: 1.3–3.0%) of the total DLA product. CTC counts were determined using VyCAP filtration with 7 µm followed by 5 µm microsieves (Figure 1).

Table 1 Diagnostic leukapheresis aliquots (n=29) from non-small cell lung cancer patients, dilution material and volumes and cell counts and final concentrations processed and filtered to detect CTC

Variable	In blood	Vycap	CellSearch
DLA aliquot			
DLA product (mL)	0	1.4 (1.1–2.3)	1.4 (1.1–2.3)
Processed cells (absolute number of cells present in the sample)			
Leukocytes ($\times 10^6$)	67.5 (57.0–87.8)	200*	200*
Lymphocytes ($\times 10^6$)	12 (8.3–15.8)	74.0 (50.8–107.9)	74.0 (50.8–107.9)
Monocytes ($\times 10^6$)	6.0 (5.3–8.3)	42.7 (30.0–49.1)	42.7 (30.0–49.1)
Granulocytes ($\times 10^6$)	49.5 (68.3–41.6)	99.2 (85.5–117.7)	99.2 (85.5–117.7)
Platelets ($\times 10^6$)	2,197 (1,646–2,891)	2,549 (1,964–4,282)	2,549 (1,964–4,282)
Erythrocytes ($\times 10^9$)	35.3 (31.4–36.9)	0.9 (0.5–1.3)	0.9 (0.5–1.3)
Dilution material and volume			
Dilution material	NA	CellSearch buffer	CellSearch buffer
Dilution volume (mL)	0	7.6 (6.7–7.9)	6.1 (5.2–6.4)
Total sample (mL)	7.5	9	7.5
Concentrations per mL filtrate			
Leukocytes ($\times 10^6$ /mL)	9.0 (7.6–11.7)	22.2	26.7
Lymphocytes ($\times 10^6$ /mL)	1.6 (1.1–2.1)	8.2 (5.6–12.0)	9.9 (6.8–14.4)
Monocytes ($\times 10^6$ /mL)	0.8 (0.7–1.1)	4.7 (3.3–5.5)	5.7 (4.0–6.6)
Granulocytes ($\times 10^6$ /mL)	6.6 (5.6–9.1)	11.1 (9.5–13.1)	13.2 (11.4–15.7)
Platelets ($\times 10^6$ /mL)	293.0 (219–385)	283 (218–476)	340 (262–571)
Erythrocytes ($\times 10^9$ /mL)	4.7 (4.2–4.9)	0.1 (0.1–0.1)	0.1 (0.1–0.2)

*, standardized to a concentration of 200×10^6 leukocytes + maximal capacity of CellSearch.

DLA filtration through 7 μ m sieves

Out of 29 DLA products (Table 1) that were filtered with the 7 μ m pore microsieves, 93% [27] succeeded. One sample failed to filtrate due to a technical issue and one due to clogging (processed at 72 hours after DLA). Failure of the sample to filtrate could not be associated with time between the DLA procedure and filtration (exposure time to fixative) or to cell counts in the DLA product (white blood cells, monocytes, lymphocytes, granulocytes, platelets, hematocrit). CTC were detected in 33% (9/27) successfully filtered samples. CTC counts ranged from 0–3 CTC per mL DLA product (median =0, IQR: 0–0.7).

DLA filtration through 5 μ m filters

Only 45% (13/29) of filtrations through the 5 μ m pore microsieves were successful, the others failed due to

clogging of the filter. Neither cell counts in the DLA product nor DLA volume correlated to filtration failure. Only the time that DLA product was exposed to the fixative in the Transfix tube correlated positively with filtration failure ($\rho=0.51$, $P<0.01$). Out of 29 samples filtered with 5 μ m pore microsieves, 19 were processed 24 hours after DLA. Seven of these nineteen failed (36%). Seven samples were processed 48 hours after DLA, of which 6 failed (86%). Two samples were processed at 72 hours and both clogged. CTC were detected in 38% (5/13) of successfully filtered samples. CTC count ranged from 0–1.8 CTC per mL DLA product (median =0, IQR: 0–0.9).

CD41 staining

On several filters cellular aggregates were observed (Figure 2A). To study the presence of platelets immunofluorescent staining with CD41 was performed. Accumulation was observed in the

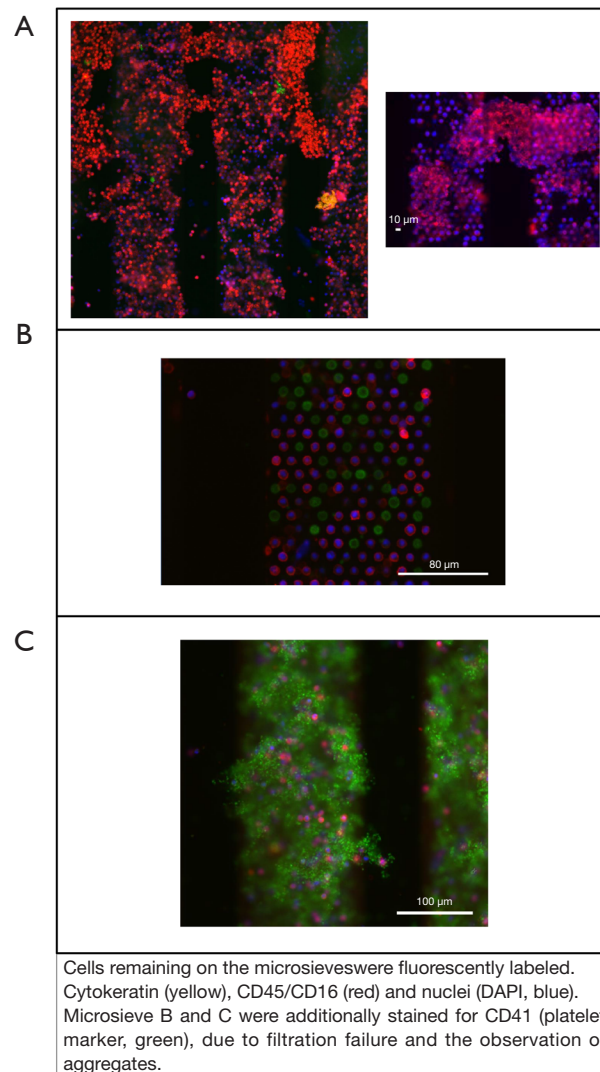


Figure 2 Clogged VyCAP microfilters: aggregates of cells (A), accumulation of CD41 (platelet marker) staining in pores of a sample processed without additional ACDA (B) and with additional ACDA (C).

pores of the filters (*Figure 2B*), indicating aggregation of platelets.

Additional anticoagulant

For two patients 2 DLA product samples of 2×10^8 leukocytes were investigated. Each patient had one sample diluted with CellSearch dilution buffer and one sample with ACDA for additional anticoagulation. The samples were processed at 24 hours after the DLA procedure. Both samples, with and without extra ACDA, could only be

filtered using the 7 µm pore microsieves. The 5 µm pore filters clogged. The number of detected CTC did not differ. The additional ACDA did not prevent the formation of clumps and clusters (*Figure 2C*).

Spiked CTC

For both spiked samples, only the filtration with the 7 µm microsieves was successful at 24 hours after DLA. Recovery percentage of MCF-7 cells was 73% and 99%.

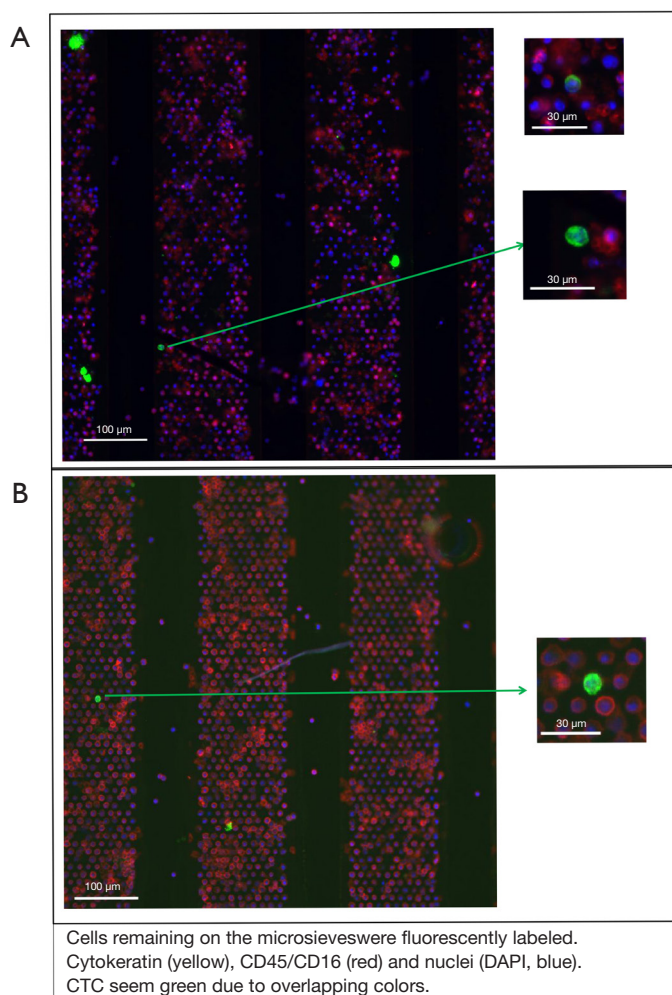


Figure 3 Circulating tumor cells identified on VyCAP filters.

CTC detection

In 27 out of 29 patients the DLA product was successfully filtered; in 2 patients filters could not be evaluated. Out of 27 filtered DLA products, 44% [12] contained CTC (*Figure 3A,B*). The 7 µm pore microsieves identified CTC in 33% (9/27) filtered DLA products. The 5 µm pore microsieves successfully processed 13 samples, of which 38% (n=5) contained CTC. In the whole population, 41% had CTC detected (12/27). The combined CTC count from both filters in all patients ranged from 0–3 CTC per mL DLA product (median=0, IQR: 0–1). Out of the 13 samples filtered by both filters, CTC were detected in 54% (7/13).

Comparison with CellSearch

CellSearch detected CTC in 63% (17/27) of the matched

samples that successfully filtered with VyCAP (*Table S3*); Both samples that failed to filter had CTC detected by CellSearch. There was a low concordance between both methods. CellSearch detected CTC in 10 patients who were negative when processed by VyCAP, while VyCAP detected CTC in 5 patients that did not have CTC detected by CellSearch (*Table S3*). The proportion of patients with CTC detected did not differ significantly between CellSearch (63%) and VyCAP (44%, $P=0.3$). Combining the CellSearch and Vycap results, CTC were identified in 81% (22/27). Median CTC count detected by CellSearch was 0.9 per mL DLA product (IQR: 0–2.1). This was significantly higher than CTC counts detected by VyCAP ($P=0.05$).

Discussion

The VyCAP microsieves filtration system is developed for

7.5 mL blood samples to enumerate CTC. However, CTC are hardly detected in the peripheral blood of even advanced stage NSCLC patients. DLA has been shown to increase CTC yield (11-13). The current golden standard for CTC detection, the FDA cleared CellSearch system, is limited by the number of leukocytes it can process (11). In blood this poses no limitation, but in DLA product it severely restricts the volumes of DLA product that can be screened for CTC. Therefore, we attempted to use the VyCAP system to filter CTC from DLA product, which differentiates CTC based on their size and rigidity. We expected it to be less limited in the volume of DLA that could be processed. However, while the 7 μm microsieves filtered the majority of samples, the 5 μm VyCAP microsieves had problems with the formation of cell aggregates and the different composition of the DLA product, resulting in clogging of the microsieves. Fixation time was the only factor found to be associated with failure of samples to filter. The fixative is necessary to avoid the necessity to process the samples immediately after the DLA procedure. Although the underlying fixation process of the used fixative (Transfix) is unknown, it was demonstrated that the time the DLA samples was exposed to the fixative influences the ability of VyCAP microsieves to filtrate DLA product and detect CTC. Other parameters may also be of influence as well, e.g., the presence of debris in the Vycap disposable waste compartment, or temperature and leukapheresis composition. Though we found no association of filtration failure with platelet counts, we did observe platelets in the microsieves. Therefore, other anticoagulants may have beneficial effects on clogging. However, increasing the amount of ACDA for anticoagulation did not improve filtration. We believe that more research is required to optimize the protocol for processing DLA product with VyCAP microsieves before they can be effectively used.

While not yet optimized, we did observe a high recovery percentage of the spiked MCF-7 cells using only the 7 μm microsieve. Yet CTC were detected in less patients with the microsieves (44%) compared to CellSearch (63%). The number of detected CTC was also significantly lower by VyCAP, reinforcing results found by Andree *et al.* (13). When combining CTC counts from CellSearch and VyCAP, CTC detection increased to 81% of patients.

In conclusion, the used VyCAP filtration is not suitable for DLA product due to clogging of the 5 μm microsieves. This appears to be caused by the time the DLA product is exposed to the fixative in the Transfix tubes. Workflows need to be optimized to be able to process DLA product

and adjustments are necessary for filtration of larger DLA product volumes.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tlcr-19-413>). MT reports grants from IMI during the conduct of the study. TJNH reports grants from IMI during the conduct of the study; grants and other from AstraZeneca, Roche, BMS, other from Pfizer, Boehringer - Ingelheim, MSD outside the submitted work. KCA reports grants from IMI CANCER-ID project during the conduct of the study. ES reports grants from IMI during the conduct of the study; grants, personal fees and non-financial support from Biocartis, personal fees and non-financial support from BMS, Astrazeneca, Pfizer, Bayer, Illumina, Jansen Cilag, Agena Biosciences, Novartis, grants and non-financial support from Roche, grants from CC Diagnostics, Boehringer Ingelheim, personal fees from MSK/Merck outside the submitted work. LWMMT reports to be inventor of issued patents related to the CellSearch system (for example US patent: 6,365,362). HJMG reports grants from IMI during the conduct of the study. The other authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All patients gave informed consent. The study was approved by the regional Medical Ethical Committee (NL55754.042.15) and registered in the Dutch trial register (NL5423), conducted in accordance with the Declaration of Helsinki (as revised in 2013).

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References

- de Wit S, Rossi E, Weber S, et al. Single tube liquid biopsy for advanced non- small cell lung cancer. *Int J Cancer* 2019;144:3127-37.
- de Wit S, Dalum G van, Lenferink ATM, et al. The detection of EpCAM+ and EpCAM- circulating tumor cells. *Sci Rep* 2015;5:12270-9.
- Manicone M, Scaini MC, Rodriquenz MG, et al. Liquid biopsy for monitoring anaplastic lymphoma kinase inhibitors in non-small cell lung cancer: Two cases compared. *J Thorac Dis* 2017;9:S1391-6.
- Krebs MG, Hou JM, Sloane R, et al. Analysis of circulating tumor cells in patients with non-small cell lung cancer using epithelial marker-dependent and -independent approaches. *J Thorac Oncol* 2012;7:306-15.
- Hofman V, Long E, Ilie M, et al. Morphological analysis of circulating tumour cells in patients undergoing surgery for non-small cell lung carcinoma using the isolation by size of epithelial tumour cell (ISET) method. *Cytopathology* 2012;23:30-8.
- Tamminga M, De Wit S, Hiltermann TJN, et al. Circulating tumor cells in advanced non-small cell lung cancer patients are associated with worse tumor response to checkpoint inhibitors. *J Immunother Cancer* 2019;7:173.
- Tamminga M, Groen HJM. Circulating tumor cells are prognostic in SCLC, but still lack clinical application. *Ann Oncol* 2019;30:1031-3.
- Tamminga M, de Wit S, Schuurings E, et al. Circulating tumor cells in lung cancer are prognostic and predictive for worse tumor response in both targeted- and chemotherapy. *Transl Lung Cancer Res* 2019;8:854-61.
- Coumans FAW, Ligthart ST, Uhr JW, et al. Challenges in the enumeration and phenotyping of CTC. *Clin Cancer Res* 2012;18:5711-8.
- Stoecklein NH, Fischer JC, Niederacher D, et al. Challenges for CTC-based liquid biopsies: low CTC frequency and diagnostic leukapheresis as a potential solution. *Expert Rev Mol Diagn* 2015;7159:14737159.2016.1123095.
- Fischer JC, Niederacher D, Topp SA, et al. Diagnostic leukapheresis enables reliable detection of circulating tumor cells of nonmetastatic cancer patients. *Proc Natl Acad Sci U S A* 2013;110:16580-5.
- Fehm TN, Meier-Stiegen F, Driemel C, et al. Diagnostic leukapheresis for CTC analysis in breast cancer patients: CTC frequency, clinical experiences and recommendations for standardized reporting. *Cytometry A* 2018;93:1213-9.
- Andree KC, Mentink A, Zeune LL, et al. Toward a real liquid biopsy in metastatic breast and prostate cancer: Diagnostic LeukApheresis increases CTC yields in a European prospective multicenter study (CTCTrap). *Int J Cancer* 2018;143:2584-91.
- Andree KC, Abali F, Oomens L, et al. Self-Seeding Microwells to Isolate and Assess the Viability of Single Circulating Tumor Cells. *Int J Mol Sci* 2019;20:477.
- Abali F, Stevens M, Tibbe AGJ, et al. Isolation of single cells for protein therapeutics using microwell selection and Surface Plasmon Resonance imaging. *Anal Biochem* 2017;531:45-7.
- Wang C, Yang L, Wang Z, et al. Highly multiplexed profiling of cell surface proteins on single circulating tumor cells based on antibody and cellular barcoding. *Anal Bioanal Chem* 2019;411:5373-82.
- Stevens M, Oomens L, Broekmaat J, et al. VyCAP's puncher technology for single cell identification, isolation, and analysis. *Cytometry A* 2018;93:1255-9.
- Swennenhuis JF, Tibbe AGJ, Stevens M, et al. Self-seeding microwell chip for the isolation and characterization of single cells. *Lab Chip* 2015;15:3039-46.
- Nadler SBSB. Prediction of blood volume in normal human adults. *Surgery* 1962;51:224-32.
- Vycap. Filtration and staining protocol Optimized for whole blood samples. 2017. Available online: <https://www.vycap.com/inhoud/uploads/Filtration-and-staining-protocol-2.1.pdf>

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Supplementary

Table S1 Characteristics of non-small cell lung cancer patients (n=29) undergoing apheresis

Characteristics	Values
Age (years)	64±11
Gender, n [%]	
Male	19 [66]
Female	10 [34]
ECOG PS, n [%]	
0	16 [55]
1	8 [28]
2	4 [14]
3	1 [3]
Smoking status, n [%]	
Smokers	15 [52]
Previous	8 [21]
Non smokers	6 [27]
Stage, n [%]	
I	2 [7]
II	1 [3]
III	4 [14]
IV	22 [76]
Histology, n [%]	
Adenocarcinoma	23 [79]
Squamous cell	6 [21]
Therapy line, n [%]	
0	5 [16]
1	10 [35]
2	10 [35]
≥3	4 [14]
Treatment, n [%]	
Surgery	3 [10]
Chemo(radio)therapy	3 [10]
Immunotherapy	17 [59]
Targeted therapy	6 [21]
Blood, median [IQR]	
TBV (L)	5.2 [4.7–5.8]
Processed volume (L)	4.6 [3.9–5.5]
Percentage processed	97% [46–100%]
DLA product, median [IQR]	
mL	88 [70–96]
ACDA	12 [11–13]
mL DLA processed	1.4 [1.1–2.3]

ECOG PS, Eastern Cooperative Oncology Group Performance Score; DLA, diagnostic leukapheresis procedure; SD, standard deviation; TBV, total blood volume; IQR, inter quartile range; ACDA, Anticoagulant Citrate Dextrose Solution A.

Table S2 Blood cell counts in DLA product per liter

Type blood cell	Mean	SD
Red blood cells ($\times 10^{12}/L$)	0.6	0.28
Leukocytes ($\times 10^9/L$)	135	76
Lymphocytes ($\times 10^9/L$)	51.8	22
Monocytes ($\times 10^9/L$)	29	10
Granulocytes ($\times 10^9/L$)	62	30
Platelets ($\times 10^9/L$)	1540	377
Hemoglobin (mmol/L)	0.91	0.28
Hematocrit (%)	0.07	0.04

Table S3 Number of circulating tumor cells detected on Vycap microsieves and by CellSearch in non-small cell lung cancer patients (n=29)

Patient	mL DLA	CellSearch	Vycap total	Vycap 7 μ m pore	Vycap 5 μ m pore
1	2.17	2	4	0	4
2	1.00	12	3	3	NA
3	2.99	0	3	2	1
4	1.16	0	3	1	2
5	1.03	1	2	2	NA
6	1.09	1	2	2	NA
7	1.40	3	1	1	0
8	1.24	0	1	1	NA
9	1.88	2	1	1	NA
10	0.75	0	1	0	1
11	1.53	11	1	1	0
12	2.41	0	1	0	1
13	0.98	0	0	0	0
14	1.37	0	0	0	0
15	1.86	1	0	0	0
16	2.52	3	0	0	0
17	0.54	2	0	0	0
18	0.56	2	0	0	0
19	1.09	0	0	0	NA
20	2.59	13	0	0	NA
21	1.67	0	0	0	NA
22	1.09	2	0	0	NA
23	1.31	1	0	0	NA
24	4.10	0	0	0	NA
25	2.16	6	0	0	NA
26	1.23	2	0	0	NA
27	2.37	4	0	0	NA
28	2.17	26	NA	NA	NA
29	2.74	1	NA	NA	NA