

NIH Public Access

Author Manuscript

Lancet Infect Dis. Author manuscript; available in PMC 2013 June 01.

Published in final edited form as:

Lancet Infect Dis. 2012 June ; 12(6): 457–468. doi:10.1016/S1473-3099(12)70055-5.

Impact of haemoglobinopathies on the clinical epidemiology of malaria: a systematic review and meta-analysis

Steve M Taylor, MD^{1,2,*}, Christian M Parobek, BS³, and Rick M Fairhurst, MD⁴

¹Department of Epidemiology, Campus Box 7435, Gillings School of Global Public Health, University of North Carolina, Chapel Hill, NC, 27599, USA

²Division of Infectious Diseases and International Health, DUMC Box 102359, Duke University Medical Center, Durham, NC, 27710, USA

³University of North Carolina School of Medicine, Chapel Hill, USA

⁴Laboratory of Malaria and Vector Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, 12735 Twinbrook Pkwy, Room 3E-10A, Rockville, MD, 20852, USA

SUMMARY

Background—Haemoglobinopathies variously reduce the risk of developing malaria syndromes. Quantifying these relationships may strengthen the foundation for translational studies of malaria pathogenesis and immunity.

Methods—The databases of MEDLINE and Embase (1950 – September 9, 2011) were searched to identify studies that estimated the risk of malaria in patients with and without haemoglobinopathies. Additional studies were identified from reference lists. Included outcomes were *Plasmodium falciparum*-related outcomes of severe malaria, uncomplicated malaria, asymptomatic parasitaemia, or pregnancy-associated malaria, and *P. vivax* malaria. Two independent reviewers identified studies, assessed their quality, and extracted data; data were meta-analyzed when outcomes were reported in more than one study.

Findings—Of 62 identified studies, 44 reported on HbAS, 19 on HbAC and HbCC, and 18 on a-thalassaemia. Case-control studies showed a decreased risk of severe malaria for HbAS (summary

Author contributions

Conflicts of interest

All authors declare that they have no conflicts of interest relevant to the subject of this manuscript.

Additional Contributions

^{© 2012} Elsevier Ltd. All rights reserved.

^{*}Corresponding author: Steve M Taylor, Department of Epidemiology, Campus Box 7435, University of North Carolina, Chapel Hill, NC 27599, USA, taylo115@email.unc.edu, Tel: 919/843-4384, FAX: 919/966-0584.

SMT conceived of and designed the study, conducted the literature search, analyzed and interpreted data, and wrote the manuscript. CMP conducted the literature search, analyzed and interpreted data, and contributed to the drafting of the manuscript. RMF conceived of and designed the study, interpreted the data, drafted the manuscript, and supervised the study conduct. SMT had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

We thank Peter Crompton MD MPH (National Institute of Allergy and Infectious Diseases) and Sunil Parikh MD MPH (University of California, San Francisco) for providing access to unpublished data, and Steven Meshnick MD PhD and Jonathan Juliano MD MSPH (each with the University of North Carolina, Chapel Hill) for providing material support. These individuals received no compensation for their assistance. We also thank our anonymous reviewers whose input greatly enhanced this report. Ultimately, we are indebted to the investigators from whose work we have drawn, and the patients for whom they cared.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

95% CI 0.74 – 0.92). Only HbAS was consistently associated with protection from uncomplicated malaria (summary Incidence Rate Ratio 0.69; 95% CI 0.61 – 0.79); none demonstrated protection from asymptomatic parasitaemia. There was a paucity of clinical studies investigating β -thalassaemia, HbE, *P. vivax* malaria, and pregnancy-associated malaria.

Interpretation—Protection from severe malaria syndromes is significant for HbAS, HbCC, HbAC, and homozygous and heterozygous α -thalassaemia, but these haemoglobinopathies differ substantially in the degrees of protection. Protection from uncomplicated malaria and asymptomatic parasitaemia is mild or absent. By attenuating the severity of malaria, haemoglobinopathies serve as a model for investigating the mechanisms of malaria pathogenesis and immunity.

INTRODUCTION

Haemoglobinopathies are highly prevalent in some human populations currently or historically exposed to the malaria parasite *Plasmodium falciparum*. In the major haemoglobinopathies, adult haemoglobin – normally composed of two α -globin and two β globin chains – is altered by genetic polymorphisms that encode single amino acid substitutions in β -globin (as in HbS, HbC, and HbE) or reduce production of α - and β globin chains (in α - and β -thalassaemia, respectively).¹ These haemoglobin variants and thalassaemias are postulated to have been naturally selected for their protection from malaria, as evidenced by a broad spectrum of investigations. These include experimental *P. falciparum* infection protocols, *in vitro* laboratory experimentation, ecological epidemiologic studies, and cartographic modeling.² Nevertheless, confirmation and quantification of malaria risk reductions due to haemoglobinopathies requires clinical studies.

Correlates of both malaria pathogenesis and immunity to disease can be identified by studying patterns of differential susceptibility to malaria. Investigations of increased susceptibility to *P. falciparum* malaria during pregnancy^{3,4} and resistance to *P. vivax* infection in West Africans lacking erythrocyte expression of Duffy Antigen Receptor for Chemokines (DARC)^{5,6} have unearthed fundamental mechanisms of both malaria pathogenesis and acquired immunity. These molecular mechanisms – adumbrated by careful epidemiologic studies – are foundations for leading vaccine candidates against pregnancy-associated malaria⁷ and vivax malaria.⁸ While some falciparum malaria vaccines are showing partial efficacy,^{9,10} malaria's pathogenic mechanisms are not understood sufficiently to inform the rational design of future therapeutics and preventive measures.

The clinical manifestations of *P. falciparum* malaria display a broad spectrum of severity from asymptomatic parasitaemia to severe malaria syndromes.¹¹ Differential protection from specific syndromes owing to genetic resistance may constitute a "natural experiment" that helps to identify the mechanisms of malaria pathogenesis that cause clinical morbidity. Toward this end, we conducted a systematic review of published studies to estimate the direct clinical effects of haemoglobinopathies on malaria syndromes.

METHODS

Search strategy & review criteria

We performed our review and meta-analysis in accordance with the PRISMA guidelines (Supplementary methods, Table S1).¹² Two authors (SMT and CMP) independently performed the database searches (through September 9, 2011), appraised study quality, and

extracted study data. Additional references were selected from the reference lists of identified studies. To appraise the quality of the observational studies, we adapted the principles of the Newcastle-Ottawa scale;¹³ in order to base analyses on robust data, we only included studies that scored at least seven stars on the scale's assessment of patient selection, comparability, and exposure/outcome. When reported data were not sufficient for estimation of desired comparisons, we contacted study authors. Overall, we selected studies that reported the frequency of clinical outcomes in patients with and without a haemoglobinopathy.

Study participants—We included studies that principally enrolled children; the exceptions were studies that investigated pregnancy-associated malaria. We included studies conducted in any level of malaria endemicity, but did not consider studies of non-immune travelers.

Study designs—For the incident outcomes of severe malaria, uncomplicated malaria, asymptomatic parasitaemia, and vivax malaria, we included data from both prospective cohort and case-control studies. For asymptomatic parasitaemia (with either *Plasmodium* species), we also included data from cross-sectional studies. For pregnancy-associated malaria outcomes, we included data from cross-sectional studies of pregnant women. For case-control studies, we required a clear description of the selection of controls. We excluded case reports.

Exposure assessment—We only considered papers in which haemoglobin typing employed electrophoresis, chromatography, or DNA analysis.

Outcome assessment—We investigated clinical outcomes owing to infection with either *P. falciparum* or *P. vivax. P. falciparum*-related outcomes were severe malaria (including cerebral malaria and severe malarial anaemia),¹⁴ uncomplicated malaria, asymptomatic parasitaemia, and pregnancy-associated malaria; vivax malaria was also included (Supplementary methods).

Definitions

The human genome normally contains four copies of the α -globin gene and two copies of the β -globin gene. Individuals with deletions of one a-globin gene ($-\alpha/\alpha\alpha$) and two α -globin genes ($-\alpha/-\alpha$ or $\alpha\alpha/--$) are referred to as α -thalassaemia heterozygotes and homozygotes, respectively. β -thalassaemia refers to individuals with impaired production of a single β -globin gene (β -thalassaemia trait, or β -thalassaemia minor). We did not investigate HbSS, HbSC, the deletion of three α -globin genes (α -/--), or the impaired production of two β -globin genes (β -thalassaemia major) because these genotypes typically manifest severe clinical sequelae which complicate any assessment of malaria-specific clinical morbidity. Additionally, we did not explore haemoglobin mutations with low global population prevalences, including haemoglobins D, Constant Spring, and Lepore. Odds Ratios (ORs) and Incidence Rate Ratios (IRRs) reflect comparisons between patients with haemoglobin variants and those with HbAA, or between patients with thalassaemias and those without.

Data analysis

For studies that did not report comparisons of interest, we extracted raw data to either 1) compare prevalences of parasitaemia between patient groups with the chi-squared test (in cross-sectional studies); 2) compare prevalences of haemoglobin variants between groups of patients with malaria syndromes with unadjusted ORs (in case-control studies); or 3) compute Risk Ratios (RRs) or IRRs of malaria syndromes between groups of patients with

Because case-control and prospective cohort studies estimate relative risk using distinct statistical methodologies, we employed separate analyses to meta-analyze ORs and IRRs. When individual-level case-control data were available for two or more studies that compared the prevalence of a haemoglobinopathy for the same case and control groups, we meta-analyzed the data to produce summary ORs. Meta-analyses were computed using random-effects models employing the DerSimonian & Laird method (metan in Stata/IC); the I^2 statistic for heterogeneity was calculated using the Mantel-Haenszel method for meta-analyzed data within subgroups (haemoglobinopathy and malaria syndrome). Similarly, when data were available for two or more prospective studies which compared incidence rates of the same outcome, we meta-analyzed the data to produce summary IRRs. Meta-analyses of IRRs were computed using random-effects Poisson meta-regression.¹⁵. We assessed publication bias in case-control studies using funnel plots and Begg's test (Supplementary methods). All single-study and summary analyses were calculated with Stata/IC (version 11, Stata Corp, College Station, TX).

Role of the funding source

The sponsor of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding author (SMT) had full access to all the data in the study and had final responsibility for the decision to submit for publication.

RESULTS

The search strategy identified 2664 studies for review, and we selected 62 for inclusion (Figure 1): 44 studies of HbAS, 19 of HbAC, eight of HbCC, 18 of α -thalassaemia (all except one included homozygotes), three of HbE, and two of β -thalassaemia (some studies examined more than one haemoglobinopathy). Of the 62 studies, 18 were prospective cohorts, 15 were case-control, and 31 were cross-sectional studies (two studies reported more than one design). Five studies investigated pregnancy-associated malaria, and two studies included patients with *P. vivax* malaria. There was no evidence of reporting bias amongst comparable studies (Supplementary methods, Figure S1).

Severe P. falciparum malaria syndromes

Haemoglobin S—Compared to healthy controls, the summary OR for severe malaria was 0.09 (95% CI 0.06 – 0.12; I^2 10.6%) across five studies which together enrolled more than 10,000 patients,^{16–20} and was similar to summary ORs for the specific syndromes of cerebral malaria (0.07; 95% CI 0.04 – 0.14; I^2 0%)^{18,19} and severe malarial anaemia (0.09; 95% CI 0.06 – 0.13; I^2 0%)(Figure 2, Table 1).^{18,19} Compared to children with uncomplicated malaria, ORs for severe malaria in three studies^{17,21,22} were heterogeneous and a summary OR estimate was non-significant (0.52; 95% CI 0.20 – 0.38; I^2 50.0%).

Only two cohort studies have reported the incidence of severe malaria (Figure 3, Table 2). The incidence of severe malaria was reduced by 71% (95% CI 38% - 88%)²³ and 83% (95% CI 60% - 93%)²⁴ in similar cohorts of Kenyan children. In the second cohort, the incidence of cerebral malaria was nonsignificantly reduced by 86% (95% CI -17% - 98%), while that of severe malarial anaemia was reduced by 60% (95% CI 40% - 70%)²⁵ and 89% (95% CI 3% - 99%).²⁴ Taken together, data from both case-control and prospective cohort studies indicate that HbAS is consistently associated with large reductions in the risk of severe malaria syndromes.

Haemoglobin C—HbC appears to protect against severe malaria to a lesser degree than HbAS and in proportion to allele frequency (Figure 2). Compared to healthy children in four studies^{16,17,19,20} that together enrolled over 9,000 patients, the summary ORs for severe malaria were 0.27 (95% CI 0.11 – 0.63; I^2 0%) for HbCC and 0.83 (95% CI 0.74 – 0.92; I^2 10.2%) for HbAC (Figure 2; Table S1). Protection from specific severe malaria syndromes has not been fully investigated in HbCC; in one study,¹⁹ HbAC showed mild protection from cerebral malaria (OR 0.64; 95% CI 0.45 – 0.91) and severe malarial anaemia (OR 0.87; 95% CI 0.68 – 0.11). When compared to children with uncomplicated malaria, protection from severe malaria is inconsistent: non-significant protection is reported from severe malaria in three studies^{17,21,26} of HbCC (summary OR 0.12; 95% CI 0.12 – 10.70; I^2 0.7%) and HbAC (summary OR 0.76; 95% CI 0.32 – 0.79; I^2 60.7%), and from severe malarial anaemia in two studies^{21,26} that combined homozygotes and heterozygotes (summary OR 0.35; 95% CI 0.04 – 0.73; I^2 0%). Significant protection from cerebral malaria was reported in one study of Malian children that combined homo- and heterozygotes (OR 0.15; 95% CI

 $0.004 - 0.93)^{21}$

Prospective studies have not reported the incidence of severe syndromes in HbC children (Table 2). Thus, convincing evidence for protection from severe malaria owing to HbC derives largely from few case-control studies.

Haemoglobin E—Meta-analysis of two studies^{27,28} in Myanmar and Thailand that compared the prevalence of HbE in severe and uncomplicated malaria cases demonstrated no evidence of protection (summary OR 0.41; 95% CI 0.04 – 0.95), though this should be interpreted cautiously given the significant heterogeneity of the findings (I^2 70.5%, p=0.027) and the highly-selected settings of the studies.

a-thalassaemia—Four studies^{19,29–31} investigated a-thalassaemia in healthy children and children with severe malaria: summary ORs were 0.63 (95% CI 0.48 – 0.83; I^2 20.6%) for homozygotes and 0.83 (95% CI 0.74 – 0.92; I^2 0%) for heterozygotes. Protection from cerebral malaria was nonsignificant in one study¹⁹ for heterozygotes (OR 0.80; 95% CI 0.64 – 1); protection from severe malarial anaemia was reported in two studies,^{19,29} with summary ORs of 0.50 (95% CI 0.35 – 0.72; I^2 0%) for homozygotes and 0.86 (95% CI 0.75 – 0.996; I^2 0%) for heterozygotes. One prospective study from Kenya documented a decreased incidence of severe disease in a-thalassaemia homozygotes (IRR 0.54; 95% CI 0.30 – 0.99) and heterozygotes (IRR 0.60; 95% CI 0.39 – 0.90) (Table 2; Figure 3).²³ Additionally, protection from severe malarial anaemia among heterozygotes (IRR 0.33; 95% CI 0.14 – 0.78) was similar to protection from cerebral malaria (IRR 0.48; 95% CI 0.24 – 0.97).³²

 β -thalassaemia—No studies have investigated the risk of severe malaria in patients with β -thalassaemia.

Uncomplicated P. falciparum malaria

Haemoglobin S—In two West African studies, ^{17,33} compared to healthy children the summary OR for children with uncomplicated malaria was 0.30 (95% CI 0.20 – 0.45; I^2 0.8%) (Table 1; Figure 2). Multiple prospective studies have characterized the risk reduction in malaria attributable to HbS (Table 2; Figure 3). Meta-analysis of five studies^{23,34–37} produced a summary IRR estimate of 0.69 (95% CI 0.61 – 0.79), which likely approximates the risk reduction owing to HbAS more closely in these malaria hyperendemic settings.³⁸

Haemoglobin C—Few studies have reported the risk of uncomplicated malaria associated with HbC. Two studies in West Africa compared healthy children and children with

uncomplicated malaria: for HbCC, the OR for malaria was 0 (95% CI 0 – 0.41) owing to the absence of HbCC in the case patients, ¹⁷ and for HbAC the summary OR was 0.16 (95% CI 0.26 – 0.23; I^2 80.9%).^{17,33} Three prospective studies have yielded conflicting results (Table 2; Figure 3): meta-analysis of two studies^{35,37} yielded a summary OR of 0.05 (95% CI 0.88 – 0.26). Thus, definitive evidence of protection from uncomplicated malaria afforded by HbCC and HbAC has not been established.

Haemoglobin E-No identified studies quantified susceptibility to malaria by HbE.

α-thalassaemia—Several prospective studies have assessed the incidence of uncomplicated malaria in α-thalassaemic children (Table 2; Figure 3), with conflicting results. In Vanuatu, the incidence of falciparum malaria was higher in α-thalassaemia homozygotes (IRR 0.3; 95% CI 0.32 – 0.07) and heterozygotes (IRR 0.1; 95% CI 0.77 – 0.61);³⁹ in contrast, the incidence of uncomplicated malaria was lower in homozygotes (IRR 0.83; 95% CI 0.70 – 0.97) and heterozygotes (IRR 0.93; 95% CI 0.82 – 0.04) in Kenya,²³ as well as homozygotes (RR 0.12; 95% CI 0.02 – 0.83) and heterozygotes (RR 0.30; 95% CI 0.10 – 0.85) in Tanzania.⁴⁰ Meta-analysis of three studies^{23,35,39} suggests lack of protection for both homozygotes (summary IRR 0.12; 95% C.I. 0.69 – 0.81) and heterozygotes (summary IRR 0.98; 95% C.I. 0.87 – 0.11).

β-thalassaemia—In one case-control study in Liberia, the prevalence of β-thalassaemia was lower in cases of uncomplicated malaria than in community controls (OR 0.56; 95% CI 0.36 - 0.86) (Table 1; Figure 2).³³

P. falciparum parasitaemia

Haemoglobin S—Cross-sectional studies have reported conflicting data on the prevalence of *P. falciparum* parasitaemia in asymptomatic HbAS children (Table S3). Compared with HbAA children, a lower prevalence of parasitaemia in HbAS children was reported in four studies, $^{41-44}$ similar prevalence in ten studies, $^{45-54}$ and higher prevalence in two studies. 55,56 In these surveys, parasite densities were reported in HbAS children as lower 41,46,49,56,57 or similar 45,50,52,55 to those in HbAA children. One case-control study reported similar prevalences of HbAS in parasitized (23%) and unparasitized (24%) asymptomatic children (Table 1).²⁰ In two prospective studies, 58,59 parasitaemia rates were similar in HbAS and HbAA children (Table 2). Taken together, HbAS does not consistently protect from *P. falciparum* parasitaemia.

Haemoglobin C—In cross-sectional surveys of adults and of children, HbC has not been associated with a reduced prevalence of *P. falciparum* parasitaemia^{45–47,49,55,60} or *P. falciparum* density.^{37,45,46,49,55} The incidence of asymptomatic parasitaemia did not differ between HbAC and HbAA children in Mali.³⁷ Thus, HbC does not appear to modify the risk of *P. falciparum* parasitaemia.

Haemoglobin E—One cross-sectional study in India reported a significantly lower prevalence of *P. falciparum* parasitaemia in patients with HbE (AE or EE) (0.6%) compared with patients with HbAA (20.5%; p = 0.005 by chi-squared test).⁶¹

 α -thalassaemia—In cross-sectional studies, α -thalassaemia was not associated with the prevalence of parasitaemia in children^{32,62–66} or, in several studies, the density of parasitaemias.^{56,62–64} In one prospective study of children in Papua New Guinea, both α -thalassaemia homozygotes (IRR 0.51; 95% CI 0.32 – 0.81) and heterozygotes (IRR 0.56; 95% CI 0.36 – 0.87) had fewer episodes of PCR-detectable parasitaemia than those without α -thalassaemia,⁶⁷ though this outcome has not been investigated in other studies.

β-thalassaemia—In one cross sectional study in Liberia, *P. falciparum* prevalence was similar in children with (78%) and without (82%) β-thalassaemia.⁶⁸

Pregnancy-associated P. falciparum malaria

Compared to women with HbAA, the prevalence of peripheral *P. falciparum* parasitaemia was similar in women with HbAS among Nigerian primigravidae⁶⁹ and Gabonese primiand secundigravidae,⁷⁰ and significantly higher in Ugandan women of all gravidities (Table S4).⁷¹ In two studies in Ghana there was no association between HbS, HbC, or α -thalassaemia and *P. falciparum* prevalence.⁷² In one study in Papua New Guinea that assessed birth outcomes, α -thalassaemia was not associated with placental malaria, birth weight, placental parasite density, maternal peripheral parasitaemia, or maternal anaemia.⁷³ On the whole, there are few data on the effect of haemoglobin variants on pregnancy-associated malaria or placental parasitization.

P.vivax malaria: Is protection species-specific?

No studies investigated an effect of HbAS, HbAC, or HbCC on *P. vivax* infection. In a prospective study in Vanuatu, the incidence of *P. vivax* malaria was significantly increased in homozygous α -thalassaemic children less than 5 years old (IRR 0.4; 95% CI 0.40 – 0.30) and nonsignificantly increased in children greater than 5 years old (IRR 0.0; 95% CI 0.42 – 0.14) (a similar pattern of increased malaria susceptibility was reported for *P. falciparum* malaria).³⁹ In a cross-sectional study investigating HbE in India, *P. vivax* parasitaemia was significantly less prevalent in HbEE/AE (0.7%) than in HbAA individuals (20.1%; p < 0.001).⁶¹

DISCUSSION

Genetic polymorphisms that affect the structure and production of the β - or α -chains of haemoglobin are variously associated with protection from a range of clinical manifestations of *P. falciparum* infection. The degree of protection varies between haemoglobinopathies, but in general is greatest against severe malaria, moderate against uncomplicated malaria, and absent against asymptomatic *P. falciparum* parasitaemia. The degrees of protection against severe malaria by HbAS (91%; 95% CI 88 – 94), HbCC (73%; 95% CI 37 – 89), and homozygous α -thalassaemia (37%; 95% CI 17 – 52) compare favorably with those reported for current large-scale malaria-control efforts, including intermittent preventive antimalarial therapy in children (87% to 69%)^{74,75} or infants (38%)⁷⁶ and the use of insecticide-treated bed nets (45%).⁷⁷

HbS and to a lesser extent HbC protect from malaria but not from parasitaemia, suggesting that these haemoglobin variants prevent the transition from asymptomatic parasitaemia to malaria. This transition is poorly understood. This protective effect may derive from the abnormal display of parasite virulence factors on the surface of parasitized HbC and HbS erythrocytes,^{78,79} possibly owing to the disruption of the parasite's remodeling of erythrocyte's intracellular trafficking network by HbS and HbC.⁸⁰ Additionally, the age-dependent nature of malaria protection owing to HbAS^{81,82} and α-thalassaemia⁸³ among children in recent reports support a protective mechanism based upon an enhanced acquisition of malaria immunity. Though HbS does not generally enhance IgG responses to a diverse array of *P. falciparum* proteins,⁸⁴ HbS may yet enhance IgG responses specifically to the parasite's major cytoadherence ligand and virulence factor Plasmodium falciparum erythrocyte membrane protein (PfEMP1).⁸⁵ Additional possible mechanisms for protection owing to haemoglobinopathies include an enhanced clearance of parasitized erythrocytes,⁸⁶ impaired parasite growth,⁸⁷ or the induction of protective immunomodulatory mechanisms by parasitized erythrocytes.⁸⁸ Data supporting these various molecular mechanisms are

complex [reviewed in ^{89,90}], and because these possibilities are not mutually-exclusive, the relative contribution of mechanisms may vary between haemoglobinopathies. By allowing parasitization while attenuating the pathogenic mechanisms that lead to disease and fatal outcomes, haemoglobin variants offer a model system to explore the cellular events involved in causing morbidity (Panel 1).

Panel 1

Unanswered questions for future clinical and translational research

- **1.** Does HbCC protect from uncomplicated malaria and asymptomatic parasitaemia, or only from severe falciparum malaria?
- **2.** Does α-thalassaemia reduce the risk of disease from specific non-*Plasmodium* pathogens?
- **3.** Do haemoglobinopathies influence the risk of uncomplicated or severe *P. vivax* malaria?
- 4. Do haemoglobinopathies influence the risk of pregnancy-associated malaria?
- **5.** Do HbE and β-thalassaemia confer protection from uncomplicated or severe falciparum malaria?
- **6.** Does α-thalassaemia exert negative epistatic effects on malaria protection by HbC and HbE?
- 7. Do haemoglobinopathies confer malaria protection to non-immune populations?
- 8. How do co-inherited G6PD deficiency variants and ABO blood groups influence the malaria-protective effects of haemoglobinopathies? 9. Does HbAS confer protection against falciparum malaria outside of sub-Saharan Africa, (e.g., India)?

The attenuation of malaria by haemoglobinopathies has important implications for nonrandomised analyses of clinical malaria studies. While randomised trials may achieve balance of underlying protective polymorphisms, comparisons of non-randomised groups may be compromised by differential prevalences of haemoglobinopathies or other risk modifiers.⁹¹ Such potential bias could impact the differential efficacy of therapies, vaccines, or other preventive measures in ecological analyses that compare populations that are not defined by randomisation and in analyses of predictors of individual-level risk. Our data endorse HbS as an important covariate in such analyses owing to its consistent protection from uncomplicated malaria (IRR 0.69; 95% CI 0.61 – 0.79), which is a common outcome in vaccine trials.^{9,10}

Our review highlights several gaps in our basic understanding of how *Plasmodium* parasites cause the symptoms and life-threatening manifestations of malaria. The paucity of outcome investigations of pregnancy-associated malaria is striking, considering that this disease model has revealed fundamental mechanisms of both parasite virulence and host adaptive immunity.⁹² Similarly, the effect of haemoglobinopathies on *P. vivax* parasitaemia and malaria incidence is relatively unknown despite geographic overlap in South Asia. Additionally, given the measurable incidence of severe *P. vivax* malaria,⁹³ case-control studies may explore associations between haemoglobinopathies and severe vivax malaria syndromes. Finally, clinical investigations have relatively neglected HbE, β -thalassaemia, and HbCC. This is surprising given the high prevalence (up to 50%) of HbE in Cambodia and HbC in parts of West Africa, as well as Haldane's 60-year-old 'malaria hypothesis' that heterozygous β -thalassaemia protects against severe and fatal falciparum malaria.⁹⁴

Two further points merit attention. First, though our systematic review was specifically designed to assess malaria outcomes, within the identified studies we found some evidence that while HbAS conferred malaria-specific protection^{22,24} a-thalassaemia protected against other mild and severe infectious syndromes, including pneumonia.^{29,32} Because malaria itself may counfound the relationship between haemoglobinopathies and other infections as recently reported for the effect of HbAS on bacteremia⁹⁵ – myriad individual and epidemiologic factors could account for this difference, in addition to biological differences in the mechanisms of protection. The identification of these mechanisms may be aided if this phenomenon is confirmed by future clinical studies or meta-analyses. Second, the dissimilarity of estimates from prospective studies of the risk of uncomplicated falciparum malaria in homozygous a-thalassaemic children is striking, with significantly increased risk on the southwestern Pacific island of Vanuatu³⁹ but either slightly decreased or unchanged risk in Africa and Papua New Guinea (Table 2).^{31,35,40,67,96} Other data have suggested an increased Plasmodium prevalence in homozygous a-thalassaemics in Papua New Guinea,97 underscoring that haemoglobinopathies may have variable effects in different settings upon different outcomes. Future studies are needed to more definitively characterize these effects and define their relationship with host genetics, malaria epidemiology, and acquired immunity to malaria.

This systematic review is subject to several limitations. We may have failed to identify relevant studies, though the independent selection of studies by two independent reviewers who each assessed over 2600 studies suggests adequate identification. Secondly, risk estimates for malaria may be influenced by unmeasured or unreported host factors, such as G6PD deficiency and ABO blood groups. Nevertheless, heterogeneity was low for most meta-analyzed comparisons, suggesting a consistent effect of haemoglobinopathies upon malaria risk. Finally, the clinical epidemiology of malaria results from poorly-understood interactions between host, parasite, and environmental factors which vary between included studies. We therefore employed random-effects meta-analysis models, and heterogeneity in risk estimates was generally low.

Despite previous successes in exploiting innate malaria protective-factors to investigate malaria pathogenesis, recent reports highlight the complexity of the co-evolution of host and parasite. *P. vivax* infection is now recognized in Malagasy individuals who lack DARC expression on their erythrocytes that were previously thought to be resistant to vivax malaria,⁹⁸ suggesting alternate erythrocyte invasion pathways. Additionally, a-thalassaemia can attenuate the malaria-protective effect of HbAS when co-inherited,²³ emphasizing the need to integrate investigations of genetic resistance. Nevertheless, by attenuating the virulence of malaria parasites, haemoglobinopathies offer an attractive "natural experiment" to help elucidate malaria's pathogenic mechanisms and potentially translate models of pathogenesis and immunity into clinical application.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Funding. Intramural Research Program, National Institute of Allergy and Infectious Diseases

Funding/Support

This research was supported in part by the Intramural Research Program of the National Institute of Allergy and Infectious Diseases, National Institutes of Health.

Role of the Sponsors

Funders had no role in the design or conduct of the study, in the collection, management, analysis, or interpretation of the data, or in preparation, review, or approval of the manuscript.

References

- Weatherall DJ, Provan AB. Red cells I: inherited anaemias. Lancet. Apr 1; 2000 355(9210):1169– 1175. [PubMed: 10791394]
- Weatherall DJ. Genetic variation and susceptibility to infection: the red cell and malaria. Br J Haematol. May; 2008 141(3):276–286. [PubMed: 18410566]
- 3. Fried M, Duffy PE. Adherence of Plasmodium falciparum to chondroitin sulfate A in the human placenta. Science. Jun 7; 1996 272(5267):1502–1504. [PubMed: 8633247]
- Fried M, Nosten F, Brockman A, Brabin BJ, Duffy PE. Maternal antibodies block malaria. Nature. Oct 29; 1998 395(6705):851–852. [PubMed: 9804416]
- Miller LH, Mason SJ, Clyde DF, McGinniss MH. The resistance factor to Plasmodium vivax in blacks. The Duffy-blood-group genotype, FyFy. N Engl J Med. Aug 5; 1976 295(6):302–304. [PubMed: 778616]
- King CL, Michon P, Shakri AR, et al. Naturally acquired Duffy-binding protein-specific binding inhibitory antibodies confer protection from blood-stage Plasmodium vivax infection. Proc Natl Acad Sci U S A. Jun 17; 2008 105(24):8363–8368. [PubMed: 18523022]
- 7. Hviid L. The role of Plasmodium falciparum variant surface antigens in protective immunity and vaccine development. Hum Vaccin. Jan; 2010 6(1):84–89. [PubMed: 19823032]
- 8. Galinski MR, Barnwell JW. Plasmodium vivax: who cares? Malar J. 2008; 7 (Suppl 1):S9. [PubMed: 19091043]
- Bejon P, Lusingu J, Olotu A, et al. Efficacy of RTS,S/AS01E vaccine against malaria in children 5 to 17 months of age. N Engl J Med. Dec 11; 2008 359(24):2521–2532. [PubMed: 19064627]
- 10. The RTS SCTP. First Results of Phase 3 Trial of RTS,S/AS01 Malaria Vaccine in African Children. N Engl J Med. Oct 18.2011
- 11. Warrell, DA.; Gilles, HM. Essential malariology. 4. London ; New York, New York: Arnold; U.S.A: Oxford University Press; 2002.
- Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med. Jul 21.2009 6(7):e1000097. [PubMed: 19621072]
- Wells, GA.; Shea, B.; O'Connell, D., et al. [Accessed 11/7/11, 2011.] The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomized studies in meta-analyses. 2010. http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp
- World Health Organization, Division of Control of Tropical Diseases. Severe and complicated malaria. Trans R Soc Trop Med Hyg. 1990; 84 (Suppl 2):1–65.
- Bagos PG, Nikolopoulos GK. Mixed-effects Poisson regression models for meta-analysis of follow-up studies with constant or varying durations. International Journal of Biostatistics. 2009; 5(1):1–33.
- Gilles HM, Fletcher KA, Hendrickse RG, Lindner R, Reddy S, Allan N. Glucose-6-phosphatedehydrogenase deficiency, sickling, and malaria in African children in South Western Nigeria. Lancet. Jan 21; 1967 1(7482):138–140. [PubMed: 4163314]
- Modiano D, Luoni G, Sirima BS, et al. Haemoglobin C protects against clinical Plasmodium falciparum malaria. Nature. Nov 15; 2001 414(6861):305–308. [PubMed: 11713529]
- Ackerman H, Usen S, Jallow M, Sisay-Joof F, Pinder M, Kwiatkowski DP. A comparison of casecontrol and family-based association methods: the example of sickle-cell and malaria. Ann Hum Genet. Sep; 2005 69(Pt 5):559–565. [PubMed: 16138914]
- May J, Evans JA, Timmann C, et al. Hemoglobin variants and disease manifestations in severe falciparum malaria. JAMA. May 23; 2007 297(20):2220–2226. [PubMed: 17519411]
- Mockenhaupt FP, Ehrhardt S, Cramer JP, et al. Hemoglobin C and resistance to severe malaria in Ghanaian children. J Infect Dis. Sep 1; 2004 190(5):1006–1009. [PubMed: 15295709]

- Agarwal A, Guindo A, Cissoko Y, et al. Hemoglobin C associated with protection from severe malaria in the Dogon of Mali, a West African population with a low prevalence of hemoglobin S. Blood. Oct 1; 2000 96(7):2358–2363. [PubMed: 11001883]
- Hill AV, Allsopp CE, Kwiatkowski D, et al. Common west African HLA antigens are associated with protection from severe malaria. Nature. Aug 15; 1991 352(6336):595–600. [PubMed: 1865923]
- 23. Williams TN, Mwangi TW, Wambua S, et al. Negative epistasis between the malaria-protective effects of alpha+-thalassemia and the sickle cell trait. Nat Genet. Nov; 2005 37(11):1253–1257. [PubMed: 16227994]
- 24. Williams TN, Mwangi TW, Wambua S, et al. Sickle cell trait and the risk of Plasmodium falciparum malaria and other childhood diseases. J Infect Dis. Jul 1; 2005 192(1):178–186. [PubMed: 15942909]
- 25. Aidoo M, Terlouw DJ, Kolczak MS, et al. Protective effects of the sickle cell gene against malaria morbidity and mortality. Lancet. Apr 13; 2002 359(9314):1311–1312. [PubMed: 11965279]
- Guinet F, Diallo DA, Minta D, et al. A comparison of the incidence of severe malaria in Malian children with normal and C-trait hemoglobin profiles. Acta Trop. Nov; 1997 68(2):175–182. [PubMed: 9386792]
- 27. Oo M, Tin S, Marlar T, O'Sullivan WJ. Genetic red cell disorders and severity of falciparum malaria in Myanmar. Bull World Health Organ. 1995; 73(5):659–665. [PubMed: 8846492]
- Hutagalung R, Wilairatana P, Looareesuwan S, Brittenham GM, Aikawa M, Gordeuk VR. Influence of hemoglobin E trait on the severity of Falciparum malaria. J Infect Dis. Jan; 1999 179(1):283–286. [PubMed: 9841856]
- Allen SJ, O'Donnell A, Alexander ND, et al. alpha+-Thalassemia protects children against disease caused by other infections as well as malaria. Proc Natl Acad Sci U S A. Dec 23; 1997 94(26): 14736–14741. [PubMed: 9405682]
- Mockenhaupt FP, Ehrhardt S, Gellert S, et al. Alpha(+)-thalassemia protects African children from severe malaria. Blood. Oct 1; 2004 104(7):2003–2006. [PubMed: 15198952]
- Williams TN, Wambua S, Uyoga S, et al. Both heterozygous and homozygous alpha+ thalassemias protect against severe and fatal Plasmodium falciparum malaria on the coast of Kenya. Blood. Jul 1; 2005 106(1):368–371. [PubMed: 15769889]
- Wambua S, Mwangi TW, Kortok M, et al. The effect of alpha+-thalassaemia on the incidence of malaria and other diseases in children living on the coast of Kenya. PLoS Med. May.2006 3(5):e158. [PubMed: 16605300]
- 33. Willcox M, Bjorkman A, Brohult J, Pehrson PO, Rombo L, Bengtsson E. A case-control study in northern Liberia of Plasmodium falciparum malaria in haemoglobin S and beta-thalassaemia traits. Ann Trop Med Parasitol. Jun; 1983 77(3):239–246. [PubMed: 6354114]
- Parikh S, Dorsey G, Rosenthal PJ. Host polymorphisms and the incidence of malaria in Ugandan children. Am J Trop Med Hyg. Dec; 2004 71(6):750–753. [PubMed: 15642965]
- Crompton PD, Traore B, Kayentao K, et al. Sickle cell trait is associated with a delayed onset of malaria: implications for time-to-event analysis in clinical studies of malaria. J Infect Dis. Nov 1; 2008 198(9):1265–1275. [PubMed: 18752444]
- Clark TD, Greenhouse B, Njama-Meya D, et al. Factors determining the heterogeneity of malaria incidence in children in Kampala, Uganda. J Infect Dis. Aug 1; 2008 198(3):393–400. [PubMed: 18522503]
- Kreuels B, Kreuzberg C, Kobbe R, et al. Differing effects of HbS and HbC traits on uncomplicated falciparum malaria, anemia, and child growth. Blood. Jun 3; 2010 115(22):4551–4558. [PubMed: 20231425]
- Zhang J, Yu KF. What's the relative risk? A method of correcting the odds ratio in cohort studies of common outcomes. JAMA. Nov 18; 1998 280(19):1690–1691. [PubMed: 9832001]
- Williams TN, Maitland K, Bennett S, et al. High incidence of malaria in alpha-thalassaemic children. Nature. Oct 10; 1996 383(6600):522–525. [PubMed: 8849722]
- Enevold A, Lusingu JP, Mmbando B, et al. Reduced risk of uncomplicated malaria episodes in children with alpha+-thalassemia in northeastern Tanzania. Am J Trop Med Hyg. May; 2008 78(5):714–720. [PubMed: 18458302]

- Allison AC. Protection afforded by sickle-cell trait against subtertian malareal infection. Br Med J. Feb 6; 1954 1(4857):290–294. [PubMed: 13115700]
- 42. Colbourne MJ, Edington GM. Sickling and malaria in the Gold Coast. Br Med J. 1956; 4970(1): 784–786. [PubMed: 13304332]
- Fleming AF, Storey J, Molineaux L, Iroko EA, Attai ED. Abnormal haemoglobins in the Sudan savanna of Nigeria. I. Prevalence of haemoglobins and relationships between sickle cell trait, malaria and survival. Ann Trop Med Parasitol. Apr; 1979 73(2):161–172. [PubMed: 315211]
- Ntoumi F, Mercereau-Puijalon O, Ossari S, et al. Plasmodium falciparum: sickle-cell trait is associated with higher prevalence of multiple infections in Gabonese children with asymptomatic infections. Exp Parasitol. Sep; 1997 87(1):39–46. [PubMed: 9287956]
- 45. Edington GM, Laing WN. Relationship between haemoglobins C and S and malaria in Ghana. Br Med J. Jul 20; 1957 2(5037):143–145. [PubMed: 13436882]
- 46. Thompson GR. Significance of haemoglobins S and C in Ghana. Br Med J. Mar 10; 1962 1(5279): 682–685. [PubMed: 13920874]
- 47. Labie D, Richin C, Pagnier J, Gentilini M, Nagel RL. Hemoglobins S and C in Upper Volta. Hum Genet. 1984; 65(3):300–302. [PubMed: 6538181]
- Aluoch JR. Higher resistance to Plasmodium falciparum infection in patients with homozygous sickle cell disease in western Kenya. Trop Med Int Health. Jun; 1997 2(6):568–571. [PubMed: 9236824]
- Danquah I, Ziniel P, Eggelte TA, Ehrhardt S, Mockenhaupt FP. Influence of haemoglobins S and C on predominantly asymptomatic Plasmodium infections in northern Ghana. Trans R Soc Trop Med Hyg. Nov; 2010 104(11):713–719. [PubMed: 20800861]
- Bernstein SC, Bowman JE, Kaptue Noche L. Population studies in Cameroon: hemoglobin S, glucose-6-phosphate dehydrogenase deficiency and falciparum malaria. Hum Hered. 1980; 30(4): 251–258. [PubMed: 6993342]
- 51. Willcox MC, Beckman L. Haemoglobin variants, beta-thalassaemia and G-6-PD types in Liberia. Hum Hered. 1981; 31(6):339–347. [PubMed: 7333623]
- Motulsky AG, Vandepitte J, Fraser GR. Population genetic studies in the Congo. I. Glucose-6phosphate dehydrogenase deficiency, hemoglobin S, and malaria. Am J Hum Genet. Nov; 1966 18(6):514–537. [PubMed: 5333143]
- Bienzle U, Guggenmoos-Holzmann I, Luzzatto L. Plasmodium falciparum malaria and human red cells. I. A genetic and clinical study in children. Int J Epidemiol. 1981; 10(1):9–15. [PubMed: 7016778]
- 54. Jeremiah ZA, Jeremiah TA, Emelike FO. Frequencies of some human genetic markers and their association with Plasmodium falciparum malaria in the Niger Delta, Nigeria. J Vector Borne Dis. Mar; 2010 47(1):11–16. [PubMed: 20231768]
- Ringelhann B, Hathorn MK, Jilly P, Grant F, Parniczky G. A new look at the protection of hemoglobin AS and AC genotypes against plasmodium falciparum infection: a census tract approach. Am J Hum Genet. May; 1976 28(3):270–279. [PubMed: 773175]
- 56. Allen SJ, Bennett S, Riley EM, et al. Morbidity from malaria and immune responses to defined Plasmodium falciparum antigens in children with sickle cell trait in The Gambia. Trans R Soc Trop Med Hyg. Sep-Oct;1992 86(5):494–498. [PubMed: 1475814]
- Guggenmoos-Holzmann I, Bienzle U, Luzzatto L. Plasmodium falciparum malaria and human red cells. II. Red cell genetic traits and resistance against malaria. Int J Epidemiol. 1981; 10(1):16–22. [PubMed: 7016776]
- 58. Stirnadel HA, Stockle M, Felger I, Smith T, Tanner M, Beck HP. Malaria infection and morbidity in infants in relation to genetic polymorphisms in Tanzania. Trop Med Int Health. Mar; 1999 4(3): 187–193. [PubMed: 10223213]
- 59. Le Hesran JY, Personne I, Personne P, et al. Longitudinal study of Plasmodium falciparum infection and immune responses in infants with or without the sickle cell trait. Int J Epidemiol. Aug; 1999 28(4):793–798. [PubMed: 10480713]
- 60. Storey J, Fleming AF, Cornille-Brogger R, Molineaux L, Matsushima T, Kagan I. Abnormal haemoglobins in the Sudan savanna of Nigeria. IV. Malaria, immunoglobulins and antimalarial

antibodies in haemoglobin AC individuals. Ann Trop Med Parasitol. Aug; 1979 73(4):311–315. [PubMed: 496483]

- Kar S, Seth S, Seth PK. Prevalence of malaria in Ao Nagas and its association with G6PD and HbE. Hum Biol. Apr; 1992 64(2):187–197. [PubMed: 1559689]
- Mockenhaupt FP, Bienzle U, May J, et al. Plasmodium falciparum infection: influence on hemoglobin levels in alpha-thalassemia and microcytosis. J Infect Dis. Sep; 1999 180(3):925–928. [PubMed: 10438396]
- 63. Fowkes FJ, Michon P, Pilling L, et al. Host erythrocyte polymorphisms and exposure to Plasmodium falciparum in Papua New Guinea. Malar J. 2008; 7:1. [PubMed: 18173836]
- 64. Veenemans J, Andang'o PE, Mbugi EV, et al. Alpha+-thalassemia protects against anemia associated with asymptomatic malaria: evidence from community-based surveys in Tanzania and Kenya. J Infect Dis. Aug 1; 2008 198(3):401–408. [PubMed: 18582194]
- 65. Shekalaghe S, Alifrangis M, Mwanziva C, et al. Low density parasitaemia, red blood cell polymorphisms and Plasmodium falciparum specific immune responses in a low endemic area in northern Tanzania. BMC Infect Dis. 2009; 9:69. [PubMed: 19460160]
- 66. Allen SJ, Rowe P, Allsopp CE, et al. A prospective study of the influence of alpha thalassaemia on morbidity from malaria and immune responses to defined Plasmodium falciparum antigens in Gambian children. Trans R Soc Trop Med Hyg. May-Jun;1993 87(3):282–285. [PubMed: 8236392]
- 67. Lin E, Tavul L, Michon P, et al. Minimal association of common red blood cell polymorphisms with Plasmodium falciparum infection and uncomplicated malaria in Papua New Guinean school children. Am J Trop Med Hyg. Oct; 2010 83(4):828–833. [PubMed: 20889874]
- Willcox M, Bjorkman A, Brohult J. Falciparum malaria and beta-thalassaemia trait in northern Liberia. Ann Trop Med Parasitol. Aug; 1983 77(4):335–347. [PubMed: 6357119]
- 69. Fleming AF, Harrison KA, Briggs ND, et al. Anaemia in young primigravidae in the guinea savanna of Nigeria: sickle-cell trait gives partial protection against malaria. Ann Trop Med Parasitol. Aug; 1984 78(4):395–404. [PubMed: 6383238]
- Bouyou-Akotet MK, Ionete-Collard DE, Mabika-Manfoumbi M, et al. Prevalence of Plasmodium falciparum infection in pregnant women in Gabon. Malar J. Jun 25.2003 2:18. [PubMed: 12919637]
- Hamilton PJ, Gebbie DA, Wilks NE, Lothe F. The role of malaria, folic acid deficiency and haemoglobin AS in pregnancy at Mulago hospital. Trans R Soc Trop Med Hyg. 1972; 66(4):594– 602. [PubMed: 5071088]
- Mockenhaupt FP, Rong B, Gunther M, et al. Anaemia in pregnant Ghanaian women: importance of malaria, iron deficiency, and haemoglobinopathies. Trans R Soc Trop Med Hyg. Sep-Oct;2000 94(5):477–483. [PubMed: 11132370]
- 73. O'Donnell A, Raiko A, Clegg JB, Weatherall DJ, Allen SJ. Alpha+-thalassaemia and pregnancy in a malaria endemic region of Papua New Guinea. Br J Haematol. Oct; 2006 135(2):235–241. [PubMed: 16939488]
- 74. Dicko A, Diallo AI, Tembine I, et al. Intermittent preventive treatment of malaria provides substantial protection against malaria in children already protected by an insecticide-treated bednet in Mali: a randomised, double-blind, placebo-controlled trial. PLoS Med. 2011; 8(2):e1000407. [PubMed: 21304923]
- Konate AT, Yaro JB, Ouedraogo AZ, et al. Intermittent preventive treatment of malaria provides substantial protection against malaria in children already protected by an insecticide-treated bednet in Burkina Faso: a randomised, double-blind, placebo-controlled trial. PLoS Med. 2011; 8(2):e1000408. [PubMed: 21304925]
- 76. Aponte JJ, Schellenberg D, Egan A, et al. Efficacy and safety of intermittent preventive treatment with sulfadoxine-pyrimethamine for malaria in African infants: a pooled analysis of six randomised, placebo-controlled trials. Lancet. Oct 31; 2009 374(9700):1533–1542. [PubMed: 19765816]
- Lengeler C. Insecticide-treated bed nets and curtains for preventing malaria. Cochrane Database Syst Rev. 2004; (2):CD000363. [PubMed: 15106149]

- Fairhurst RM, Baruch DI, Brittain NJ, et al. Abnormal display of PfEMP-1 on erythrocytes carrying haemoglobin C may protect against malaria. Nature. Jun 23; 2005 435(7045):1117–1121. [PubMed: 15973412]
- 79. Cholera R, Brittain NJ, Gillrie MR, et al. Impaired cytoadherence of Plasmodium falciparuminfected erythrocytes containing sickle hemoglobin. Proc Natl Acad Sci U S A. Jan 22; 2008 105(3):991–996. [PubMed: 18192399]
- 80. Cyrklaff M, Sanchez CP, Kilian N, et al. Hemoglobins S and C Interfere with Actin Remodeling in Plasmodium falciparumâ]"Infected Erythrocytes. Science. Nov 10.2011
- Williams TN, Mwangi TW, Roberts DJ, et al. An immune basis for malaria protection by the sickle cell trait. PLoS Med. May.2005 2(5):e128. [PubMed: 15916466]
- 82. Gong L, Maiteki-Sebuguzi C, Rosenthal PJ, et al. Evidence for both innate and acquired mechanisms of protection from Plasmodium falciparum in children with sickle cell trait. Blood. Feb 10.2012
- Veenemans J, Jansen EJ, Baidjoe AY, et al. Effect of alpha(+)-thalassaemia on episodes of fever due to malaria and other causes: a community-based cohort study in Tanzania. Malar J. 2011; 10:280. [PubMed: 21939508]
- 84. Tan X, Traore B, Kayentao K, et al. Hemoglobin S and C Heterozygosity Enhances Neither the Magnitude nor Breadth of Antibody Responses to a Diverse Array of Plasmodium falciparum Antigens. J Infect Dis. Dec; 2011 204(11):1750–1761. [PubMed: 21998476]
- Cabrera G, Cot M, Migot-Nabias F, Kremsner PG, Deloron P, Luty AJ. The sickle cell trait is associated with enhanced immunoglobulin G antibody responses to Plasmodium falciparum variant surface antigens. J Infect Dis. May 15; 2005 191(10):1631–1638. [PubMed: 15838789]
- 86. Ayi K, Turrini F, Piga A, Arese P. Enhanced phagocytosis of ring-parasitized mutant erythrocytes: a common mechanism that may explain protection against falciparum malaria in sickle trait and beta-thalassemia trait. Blood. Nov 15; 2004 104(10):3364–3371. [PubMed: 15280204]
- Fairhurst RM, Fujioka H, Hayton K, Collins KF, Wellems TE. Aberrant development of Plasmodium falciparum in hemoglobin CC red cells: implications for the malaria protective effect of the homozygous state. Blood. Apr 15; 2003 101(8):3309–3315. [PubMed: 12480691]
- Ferreira A, Marguti I, Bechmann I, et al. Sickle hemoglobin confers tolerance to Plasmodium infection. Cell. Apr 29; 2011 145(3):398–409. [PubMed: 21529713]
- Williams TN. How do hemoglobins S and C result in malaria protection? J Infect Dis. Dec; 2011 204(11):1651–1653. [PubMed: 21998475]
- 90. Lopez C, Saravia C, Gomez A, Hoebeke J, Patarroyo MA. Mechanisms of genetically-based resistance to malaria. Gene. Nov 1; 2010 467(1–2):1–12. [PubMed: 20655368]
- Parikh S, Rosenthal PJ. Human genetics and malaria: relevance for the design of clinical trials. J Infect Dis. Nov 1; 2008 198(9):1255–1257. [PubMed: 18752442]
- Rogerson SJ, Hviid L, Duffy PE, Leke RF, Taylor DW. Malaria in pregnancy: pathogenesis and immunity. Lancet Infect Dis. Feb; 2007 7(2):105–117. [PubMed: 17251081]
- Bassat Q, Alonso PL. Defying malaria: Fathoming severe Plasmodium vivax disease. Nat Med. Jan; 2011 17(1):48–49. [PubMed: 21217683]
- 94. Haldane JB. The rate of mutation of human genes. Hereditas. 1949; 35(S1):267-273.
- 95. Scott JA, Berkley JA, Mwangi I, et al. Relation between falciparum malaria and bacteraemia in Kenyan children: a population-based, case-control study and a longitudinal study. Lancet. Oct 8; 2011 378(9799):1316–1323. [PubMed: 21903251]
- 96. Migot-Nabias F, Pelleau S, Watier L, et al. Red blood cell polymorphisms in relation to Plasmodium falciparum asymptomatic parasite densities and morbidity in Senegal. Microbes Infect. Aug; 2006 8(9–10):2352–2358. [PubMed: 16859949]
- Oppenheimer SJ, Hill AV, Gibson FD, Macfarlane SB, Moody JB, Pringle J. The interaction of alpha thalassaemia with malaria. Trans R Soc Trop Med Hyg. 1987; 81(2):322–326. [PubMed: 3617198]
- Menard D, Barnadas C, Bouchier C, et al. Plasmodium vivax clinical malaria is commonly observed in Duffy-negative Malagasy people. Proc Natl Acad Sci U S A. Mar 30; 2010 107(13): 5967–5971. [PubMed: 20231434]

- Jallow M, Teo YY, Small KS, et al. Genome-wide and fine-resolution association analysis of malaria in West Africa. Nat Genet. Jun; 2009 41(6):657–665. [PubMed: 19465909]
- 100. Lell B, May J, Schmidt-Ott RJ, et al. The role of red blood cell polymorphisms in resistance and susceptibility to malaria. Clin Infect Dis. Apr; 1999 28(4):794–799. [PubMed: 10825041]
- 101. Rihet P, Flori L, Tall F, Traore AS, Fumoux F. Hemoglobin C is associated with reduced Plasmodium falciparum parasitemia and low risk of mild malaria attack. Hum Mol Genet. Jan 1; 2004 13(1):1–6. [PubMed: 14613965]

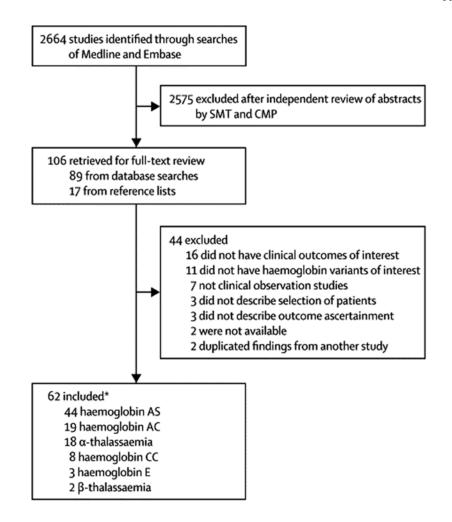


Figure 1. Study identification flowchart

	Odds ratio (95%	CI) Weight
Haemoglobin AS		
Severe malaria		
Gilles et al (1967) ¹⁶	0·19 (0·07-0·55)	7-81
Modiano et al (2001) ¹⁷	0.10 (0.04-0.26)	8-95
Mockenhaupt et al (2004)20	0.02 (0-0.28)	1-12
Ackerman et al (2005)18	0.08 (0.03-0.21)	10-65
May et al (2007) ¹⁹	0.08 (0.06-0.11)	71-47
Jallow et al (2009)21*	0.09 (0.05-0.16)	NA
Summary I2=0%, p=0-456	0.09 (0.06-0.12)	
Cerebral malaria	•	
Ackerman et al (2005)18	0.09 (0.03-0.24)	35-78
May et al (2007) ¹⁹	0.07 (0.03-0.14)	64-22
Jallow et al (2009)21*	0.12 (0.07-0.21)	NA
Summary I2=0%, p=0.667	0.07 (0.04-0.14)	
Severe malarial anaemia		100
Ackerman et al (2005)18	0.07 (0.01-0.47)	4-03
May et al (2007) ¹⁹		
Jallow et al (2009) ^{21*}	0.10 (0.04-0.24)	
Summary /2=0%, p=0.741	0.09 (0.06-0.13)	100
Uncomplicated malaria		
Willcox et al (1983) ²²	0.23 (0.10-0.44)	34-54
Modiano et al (2001) ¹⁷	0-35 (0-20-0-56)	65-46
Summary I ² =3·8%, p=0·308	0-30 (0-20-0-45)	100
laemoglobin CC		
Severe malaria		
Modiano et al (2001) ³⁷	0.14 (0.02-1.01)	18-47
Mockenhaupt et al (2004)20	0.08 (0-1.38)	8-61
May et al (2007) ¹⁹	0-37 (0-14-0-99)	72-92
Summary I2=0%, p=0-424	0-27 (0-11-0-63)	100
Incomplicated malaria	02/(011-0-03)	100
Modiano et al (2001) ³⁷	0 (0-0.41)	NA
Haemoglobin AC	0(0-0-41)	10/0
ievere malaria		
	0-85 (0-32-2-28)	
Gilles et al (1967)16		3-30
Modiano et al (2001) ²⁷		
Mockenhaupt et al (2004) ²⁰	0.73 (0.47-1.12)	15-71
May et al (2007) ¹⁹	0.92 (0.75-1.13)	49-78
Summary /2=17-2%, p=0-305	• 0.80 (0.67-0.96)	100
Cerebral malaria		
May et al (2007) ¹⁹	0.64 (0.45-0.91)	NA
Severe malarial anaemia		
May et al (2007)19	0-87 (0-68-1-11)	NA
Uncomplicated malaria		
Willcox et al (1983)22	2.80 (0.76-11.30) 43-18
Modiano et al (2001)37		56-82
Summary /2=84.9%, p=0.010	1.16 (0.26-5-23)	100
łomozygous α-thalassaemia		
ievere malaria		
Allen et al (1997) ²³	0·50 (0·30-0·85)	19-92
Mockenhaupt et al (2004) ²⁴	1.27 (0.60-2.68)	
Williams et al (2005)25		
May et al (2007) ¹⁹	- - 0.58 (0.40-0.85)	
Summary I ² =29-6%, p=0-234	0.63 (0.48-0.83)	100
evere malarial anaemia		
Allen et al (1997) ²³		31-43
May et al (2007) ¹⁹		68-57
Summary /2=0%, p=0-592	0-50 (0-33-0-84)	100
Heterozygous α-thalassaemia		
ievere malaria		
Allen et al (1997)23	0.77 (0.44-1.34)	3.76
Mockenhaupt et al (2004) ²⁴		
Williams et al (2005) ²⁵		
May et al (2007) ¹⁹	0.88 (0.77-1.01)	66-24
Summary I ² =0%, p=0.402	 → 0.83 (0.74-0.92) 	100
Sommary F=0%, p=0-402 Cerebral malaria	0-83 (0-74-0-92)	100
May et al (2007) ¹⁹	•• 0.80 (0.64–1.00)	NA
evere malarial anaemia		
Allen et al (1997) ²³	1.00 (0.51-1.95)	4-73
May et al (2007) ¹⁹	0-85 (0-74-0-99)	
Summary /2=0%, p=0.651	0.86 (0.74-1.00)	100
3-thalassaemia		
Uncomplicated malaria		
Willcox et al (1983) ²²	0.56 (0.36-0.86)	NA

Figure 2.

Unadjusted individual and summary Odds Ratios for specific malaria syndromes by haemoglobin variants

Abbreviations: CI, confidence interval; Hb, haemoglobin

^a Not included in meta-analyses because prevalences of HbAS in cases and controls were not reported. All Odds Ratios (ORs) are for comparison with healthy controls. The data markers represent either ORs from individual studies (circles) or summary ORs for subgroup data (diamonds) that were generated by random-effects meta-analysis of individual studies (squares). For individual studies included in meta-analyses, the size of the square data marker is relative to the weight of the study. The I^2 statistic is a measure of the heterogeneity of the individual study estimates which were meta-analyzed, and was calculated using the Mantel-Haenszel method. ORs for individual studies may differ from those in Table S1 or in the original published studies because they were calculated from raw data and are thus unadjusted.

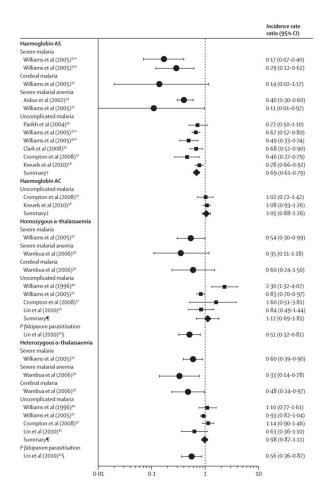


Figure 3.

Individual and summary incidence rate ratios of *P. falciparum* syndromes in prospective studies of children with haemoglobinopathies

Abbreviations: CI, confidence interval; Hb, haemoglobin. All incidence rate ratios (IRRs) compare patients with the variant listed to patients with either HbAA or the aa/aa genotype. Summary measures (diamonds) were computed using random-effects Poisson meta-regression of individual studies (squares).

^a Studies had overlapping cohorts. Because the cohort in Williams et al (2005)²³ subsumes that of Williams et al (2005),²⁴ only data from Williams et al (2005)²³ was included in the meta-analyzed summary IRR for uncomplicated malaria in HbAS children.

^b Summary IRR of uncomplicated malaria in HbAS compared with HbAA children from five studies.^{23,34-37}

^c Summary IRR of uncomplicated malaria in HbAC compared with HbAA children from two studies.^{35,37}

^d Summary IRR of uncomplicated malaria in homozygous or heterozygous α-thalassaemic compared with non-thalassaemic children from three studies [Williams 1996, Williams 2005 (NG), Crompton 2008].^{23,35,39}

^e Detected by polymerase chain reaction.

NIH-PA Author Manuscript

Taylor et al. . .

Table 1

Case-control studies investigating the effect of haemoglobinopathies on P. falciparum malaria

Source	Location	Syndrome ^a	No. patients	Age	Comment	% Haemoglobin opathy
Haemoglobin AS						
Gilles et al (1967) ¹⁶	Nigeria	Severe malaria	100	Range 6m - 4y		4.0
		Healthy	100	Range 6m - 4y	Aparasitaemic outpatients	18.0
Willcox et al (1983) ³³	Liberia	Uncomplicated malaria	518	Range 2m - >15y	Outpatients	1.9
		Healthy	1027	Range 2m - >15y	Community surveys	7.9
Hill et al (1991) ²²	The Gambia	Severe malaria	619	Median 3.0y Range <10y		1.2
		Severe non-malaria illness	332	Median 2.2y Range <10y	Inpatients with severe non- malaria illness	10.9
		Mild non-malaria illness	510	Median 1.9y Range <10y	Mild outpatient illness without malaria parasites	12.9
		Uncomplicated malaria	354	Median 3.0y Range <10y		2.8
Agarwal et al (2000) ²¹	Mali	Severe malaria	67	Mean 4.3y		4.5
		Cerebral malaria	34	Mean 3.8y	Subset of severe malaria patients with coma, obtundation, or convulsions	5.9
		Severe malarial anaemia	12	Mean 1.0y	Subset of severe malaria patients with Hct <15%	0
		Uncomplicated malaria	391	Mean 9.1y		2.7
Modiano et al (2001) ¹⁷	Burkina Faso	Severe malaria	359	Mean 4y Range 0.5 -15y		1.1
		Uncomplicated malaria	476	Mean 4y Range 0.5 -15y		4.0
		Healthy	3513	Mean 11y Range 1- 40y	Community cross-sectional survey	9.5
Mockenhaupt et al (2004) ²⁰	Ghana	Severe malaria	290	Median 24m Range 6 - 102m		0
		Healthy, parasitaemic	290	NA	Community cross-sectional survey, matched to severe cases	7.9
		Healthy aparasitaemic	290	NA	Community cross-sectional survey, matched to severe cases	8.3

_
Z
T
- T
~
7
-
<u> </u>
=
້
thor
2
\geq
B
D
$\overline{\mathbf{o}}$
õ
anuscri
5
¥

Source	Location	Syndrome ^a	No. patients	Age	Comment	% Haemoglobin opathy
Ackerman et al (2005) ¹⁸	The Gambia	Severe malaria	315	NA		1.1
		Cerebral malaria	235	NA	Subset of severe malaria patients with BCS <3	1.3
		Severe malarial anaemia	80	NA	Subset of severe malaria patients with Hb $<5g/dL$	0.6
		Newborns	583	NA	Cord blood specimens	8.7
May et al (2007) ¹⁹	Ghana	Severe malaria	2591	Median 20m		1.4
		Cerebral malaria	581	NA	Subset of severe malaria patients with BCS <3	1.2
		Severe malarial anaemia	1649	NA	Subset of severe malaria patients with Hb <5g/dL	1.6
		Healthy	2048	Median 30m	Community cross-sectional survey, matched to severe cases	14.8
Jallow et al (2009) ⁹⁹	The Gambia	Severe malaria	1060	NA		NA
		Cerebral malaria	869	NA	Subset of severe malaria patients with BCS <3	NA
		Severe malarial anaemia	318	NA	Subset of severe malaria patients with Hb <5g/dL	NA
		Newborns	1500	NA	Cord blood specimens	8.0
Haemoglobin C						
Gilles et al (1967) ¹⁶	Nigeria	Severe malaria	100	Range 6m - 4y		He: 6.0
		Healthy	200	Range 6m - 4y	Clinic attendance, aparasitaemic	He: 7.0
Willcox et al (1983) ³³	Liberia	Uncomplicated malaria	518	Range 2m - >15y		He: 1.3
		Healthy	1027	Range 2m - >15y	Community surveys	He: 0.4
Guinet et al (1997) ²⁶	Mali	Severe malaria	110	Range 6m - 9y		10.0
		Severe malarial anaemia	9	Range 6m - 9y	Subset of severe malaria patients with Hct ${<}15\%$ or Hb ${<}5g/dL$	0
		Uncomplicated malaria	55	Range 6m - 9y	Matched to severe cases	9.0
Agarwal et al (2000) ²¹	Mali	Severe malaria	67	Mean 4.3y		Ho: 0 He: 4.5
		Cerebral malaria	34	Mean 3.8y	Subset of severe malaria patients with coma, obtundation, or convulsions	Ho: 0 He: 2.9
		Severe malarial anaemia	12	Mean 1.0y	Subset of severe malaria patients with $Hct < 15\%$	Ho: 0 He: 0
		Uncomplicated malaria	391	Mean 9.1y		Ho: 1.5 He: 15.9
Modiano et al (2001) ¹⁷	Burkina Faso	Severe malaria	359	Mean 4y		Ho: 0.3 He: 17.6

Lancet Infect Dis. Author manuscript; available in PMC 2013 June 01.

_
_
T
the second s
0
~
Autho
a l
-
_
0
-
~
\leq
a
ЧР
_
-
<u> </u>
Jscri
~
0
-
0
+

NIH-PA Author Manuscript

Taylor et al.

Source	Location	Syndrome ^a	No. patients	Age	Comment	% Haemoglobin opathy
				Range 0.5 – 15y		
		Uncomplicated malaria	476	Mean 4y Range 0.5 – 15y		Ho: 0 He: 15.6
		Healthy	3513	Mean 11y Range 1- 40y	Community cross-sectional survey	Ho: 1.7 He: 21.7
Mockenhaupt et al (2004) ²⁰	Ghana	Severe malaria	290	Median 24m Range 6 – 102m		Ho: 0 He: 16.6
		Healthy, parasitaemic	290	NA	Community cross-sectional survey, matched to severe cases	Ho: 1.0 He: 24.5
		Healthy, aparasitaemic	290	NA	Community cross-sectional survey, matched to severe cases	Ho: 1.7 He: 19.0
May et al (2007) ¹⁹	Ghana	Severe malaria	2591	Median 20m		Ho: 0.2 He: 9.4
		Cerebral malaria	581	NA	Subset of severe malaria patients with BCS <3	Ho: NA He: 7.2
		Severe malarial anaemia	1649	NA	Subset of severe malaria patients with Hb $<5g/dL$	Ho: NA He: 9.2
		Healthy	2048	Median 30m	Community cross-sectional survey, matched to severe cases	Ho: 0.5 He: 8.7
Haemoglobin E						
Oo et al (1995) ²⁷	Myanmar	Severe malaria	200	19 - 45y		28.0
		Uncomplicated malaria	109	19 - 45y	Hospitalized	27.5
Hutagalung et al (1999) ²⁸	Thailand	Severe malaria	33	14 - 45y		3.0
		Uncomplicated malaria	184	14 - 45y	Hospitalized	22.3
a-thalassaemia						
Allen et al (1997) ²⁹	Papua New Guinea	Severe malaria	249	NA		Ho: 44.6 He: 37.3
		Severe malarial anaemia	155	Median 2.1y	Subset of severe malaria patients with Hb $<5g/dL$	Ho: 36.1 He: 43.9
		Healthy	249	NA	Community survey, matched to severe cases	Ho: 57.0 He: 31.3
Lell et al (1999) ¹⁰⁰	Gabon	Severe malaria	100	Mean 44m		Ho: 10.0 He: 37.0
		Uncomplicated malaria	100	Mean 44m		Ho: 10.0 He: 43.0

Lancet Infect Dis. Author manuscript; available in PMC 2013 June 01.

Page 21

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Source	Location	Syndrome ^a	No. patients	Age	Comment	% Haemoglobin opathy
Mockenhaupt et al (2004) ³⁰	Ghana	Severe malaria	261	<5y		Ho: 5.0 He: 23.7
		Healthy, parasitaemic	614	<5y	Community cross-sectional survey, matched to severe cases	Ho: 3.3 He: 33.2
		Healthy, aparasitaemic	479	<5y	Community cross-sectional survey, matched to severe cases	Ho: 3.6 He: 31.9
Williams et al (2005) ³¹	Kenya	Severe malaria	655	NA		Ho: 12.7 He: 47.0
		Fatal malaria	72	NA	Death in hospital	Ho: 8.4 He: 10.4
		Healthy	648	<7y	Community survey, matched	Ho: 16.7 He: 50.6
May et al (2007) ¹⁹	Ghana	Severe malaria	2591	Median 20m		Ho: 2.0 He: 25.2
		Cerebral malaria	581	NA	Subset of severe malaria patients with BCS <3	Ho: 2.2 He: 23.8
		Severe malarial anaemia	1649	NA	Subset of severe malaria patients with Hb $<5g/dL$	Ho: 1.8 He: 24.7
		Healthy	2048	Median 30m	Community cross-sectional survey, matched to severe cases	Ho: 2.0 He: 27.3
β -thalassaemia						
Willcox et al (1983) ³³	Liberia	Uncomplicated malaria	526	2m - >15y		5.7
		Healthy	1132	2m - >15y	Community surveys	9.7
Abbreviations: Hb: haemos	elohin: NA: data not a	Abbreviations: Hb: haemoelohin: NA: data not available: Ho: homozyeote: He: heterozyeote: BCS: Blantyre Coma Score: Hc: hematocrit.	heterozvgote: BC3	S: Blantvre Coma	Score: Hct: hematocrit.	

, Acc Da A ž

^aUnless otherwise stated, patients with severe malaria, severe malarial anaemia, cerebral malaria and uncomplicated malaria met definitions described in the Methods section. Healthy patients were recruited as described in the Comments column.

Taylor et al.

_
T
_
0
-
-
~
_
<u> </u>
The second secon
_
uthor
0
<u> </u>
_
2
01
<u>u</u>
<u> </u>
<u> </u>
5
S
S
0
_
$\overline{\mathbf{D}}$
<u> </u>

Table 2

Prospective studies investigating the effect of haemoglobinopathies on P. falciparum malaria

Source	Outcome	Location	u	Ages	Incidence in unexposed ^a	Incidence Rate Ratio (95% C.I.)
Haemoglobin AS						
Williams et al (2005) ^{23,b}	Severe malaria	Kenya	2655	< 5y	0.022/y	$0.17\ (0.07-0.40)$
Williams et al (2005) ^{24,b}	Severe malaria	Kenya	2104	< 5y	0.020/y	0.29 (0.12 - 0.62)
Williams et al (2005) ²³	Cerebral malaria	Kenya	2655	< 5y	0.004/y	$0.14 \ (0.02 - 1.17)$
Aidoo et al (2002) ²⁵	Severe malaria anaemia	Kenya	1022	< 5y	0.048/y	0.40 (0.30 – 0.60)
Williams et al (2005) ²³	Severe malarial anaemia	Kenya	2655	< 5y	0.005/y	0.11 (0.01 – 0.97)
Parikh et al $(2004)^{34}$	Uncomplicated malaria	Uganda	307	6m – 5y	2.04/y	0.72 (0.5 - 1.1)
Williams et al (2005) ^{23,b}	Uncomplicated malaria	Kenya	370	< 11y	2.36/y	$0.67\ (0.57-0.80)$
Williams et al (2005) ^{24,b}	Uncomplicated malaria	Kenya	323	< 8y	3.06/y	0.49 (0.33 – 0.74)
Migot-Nabias et al $(2006)^{96}$	Uncomplicated malaria	Senegal	169	2 – 10y	NA	Reduced incidence in HbAS (IRR not reported)
Clark et al (2008) ³⁶	Uncomplicated malaria3	Uganda	558	1 - 10y	0.82/y	0.68 (0.52 – 0.90)
Crompton et al (2008) ³⁵	Uncomplicated malaria3	Mali	176	2 – 10y	1.76/y	0.46 (0.27 – 0.79)
Enevold et al (2008) ⁴⁰	Uncomplicated malaria	Tanzania	159	6m – 19y	35%/6m	RR: 1.55 (0.51 – 4.77)
Kreuels et al $(2010)^{37}$	Uncomplicated malaria3	Ghana	852	3m – 2y	1.2/y	0.78 (0.66 – 0.92)
<i>Summary</i> ^{23,34-37}	Uncomplicated malaria					0.69 (0.61 – 0.79)
Le Hesran et al (1999) ⁵⁹	<i>P. falciparum</i> parasitaemia	Cameroon	240	< 5y	NA	No difference (IRR not reported)
Stimadel et al (1999) ⁵⁸	<i>P. falciparum</i> parasitaemia	Tanzania	204	< 1y	NA	No difference (IRR not reported)
Haemoglobin C						
Rihet et al $(2004)^{101}$	Uncomplicated malaria3	Mali	256	1 – 24y	NA	Reduced incidence with HbC (IRR not reported)
Crompton et al (2008) ³⁵	Uncomplicated malaria3	Mali	176	2 – 10y	1.76/y	He: 1.02 (0.72 – 1.42)
Kreuels et al $(2010)^{37}$	Uncomplicated malaria3	Ghana	852	3m - 2y	1.2/y	He: 1.08 (0.93 – 1.26)
<i>Summary</i> ^{35,37}	Uncomplicated malaria					He: 1.05 (0.88 – 1.26)
a-thalassaemia						
Williams et al (2005) ³¹	Severe malaria	Kenya	2104	< 5y	0.061/y	Ho: 0.54 (0.30 – 0.99) He: 0.60 (0.39 – 0.90)
W_{0} mbin of all (2006) 32	Cavara malarial anaamia	Кепуа	2104	< 5v	0.010/v	H_{G} : 0 35 (0 11 – 1 18)

Source	Outcome	Location	u	Ages	Incidence in unexposed ^a	Incidence Rate Ratio (95% C.I.)
						He: 0.33 (0.14 – 0.78)
Wambua et al (2006) ³²	Cerebral malaria	Kenya	2104	< 5y	0.002/y	Ho: 0.60 (0.24 – 1.50) He: 0.48 (0.24 – 0.97)
Williams et al (1996) ³⁹	Uncomplicated malaria3	Vanuatu	544	< 5y	0.7/y	Ho: 2.30 (1.32 – 4.07) He: 1.10 (0.77 – 1.61)
Williams et al $(2005)^{31}$	Uncomplicated malaria	Kenya	370	<11y	2.46/y	Ho: 0.83 (0.70 – 0.97) He: 0.93 (0.82 – 1.04)
Migot-Nabias et al (2006) ⁹⁶	Uncomplicated malaria3	Senegal	169	2 - 10y	NA	No difference (IRR not reported)
Enevold et al (2008) ⁴⁰	Uncomplicated malaria	Tanzania	159	6m - 19y 45%/6m	45%/6m	Ho: RR 0.12 (0.02 – 0.83) He: RR 0.30 (0.10 – 0.85)
Crompton et al (2008) ³⁵	Uncomplicated malaria3	Mali	176	2 – 10y	1.76/y	Ho: 1.60 (0.51 – 3.81) He: 1.14 (0.90 – 1.46)
Lin et al (2010) ⁶⁷	Uncomplicated malaria3	Papua New Guinea	206	5 – 14y	NA	Ho: 0.84 (0.49 – 1.44) He: 0.63 (0.36 – 1.10)
Summary ^{23,35,39}	Uncomplicated malaria					Ho: 1.12 (0.69 – 1.81) He: 0.98 (0.87 – 1.11)
Lin et al (2010) ⁶⁷	<i>P. falciparum</i> parasitaemia (PCR)	Papua New Guinea	206	5 – 14y	NA	Ho: 0.51 (0.32 – 0.81) He: 0.56 (0.36 – 0.87)

tts of HbE or β -thalassaemia on malaria.

 a Annual incidence rate in the unexposed population (i.e., HbAA or $\alpha\alpha/\alpha\alpha$ genotype) of the indicated outcome.

b Studies with overlapping cohorts. Because the cohort in Williams et al (2005)²³ subsumes that of Williams et al (2005),²⁴ only data from Williams et al (2005)²³ was included in the meta-analyzed summary IRR for uncomplicated malaria in HbAS children.

NIH-PA Author Manuscript

NIH-PA Author Manuscript