Fine Mapping and Identification of BMI Loci in African Americans

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Genome-wide association studies (GWASs) primarily performed in European-ancestry (EA) populations have identified numerous loci associated with body mass index (BMI). However, it is still unclear whether these GWAS loci can be generalized to other ethnic groups, such as African Americans (AAs). Furthermore, the putative functional variant or variants in these loci mostly remain under investigation. The overall lower linkage disequilibrium in AA compared to EA populations provides the opportunity to narrow in or fine-map these BMI-related loci. Therefore, we used the Metabochip to densely genotype and evaluate 21 BMI GWAS loci identified in EA studies in 29,151 AAs from the Population Architecture using Genomics and Epidemiology (PAGE) study. Eight of the 21 loci (*SEC16B, TMEM18, ETV5, GNPDA2, TFAP2B, BDNF, FTO*, and *MC4R*) were found to be associated with BMI in AAs at 5.8 × 10⁻⁵. Within seven out of these eight loci, we found that, on average, a substantially smaller number of variants was correlated ($r^2 > 0.5$) with the most significant SNP in AA than in EA populations (16 versus 55). Conditional analyses revealed *GNPDA2* harboring a potential additional independent signal. Moreover, Metabochip-wide discovery analyses revealed two BMI-related loci, *BRE* (rs116612809, p = 3.6 × 10⁻⁸) and *DHX34* (rs4802349, p = 1.2×10^{-7}), which were significant when adjustment was made for the total number of SNPs tested across the chip. These results demonstrate that fine mapping in AAs is a powerful approach for both narrowing in on the underlying causal variants in known loci and discovering BMI-related loci.

Introduction

Obesity (MIM 601665) is a major risk factor for a number of chronic diseases, such as type 2 diabetes (MIM 125853), hyperlipidemia (MIM 144250), cardiovascular diseases, and several cancer types.^{1,2} Worldwide obesity prevalence has nearly doubled since 1980, and in 2008 more than 1.4 billion adults worldwide were obese. In the United States, more than one-third of adults (35.7%) were obese in 2010.

Studies have shown that obesity is highly heritable; heritability is estimated to fall in the range of 40%–70%.^{3,4}

Genome-wide association studies (GWAS) have identified numerous loci associated with body mass index (BMI),^{5–7} a common measure of obesity. However, most of these studies were performed among European-ancestry (EA) populations. It is still unclear whether previously identified GWAS loci are population specific or whether they can be generalized to other ethnic groups, such as African Americans (AAs). Furthermore, the overall lower linkage disequilibrium (LD) patterns in AA compared to EA populations can offer opportunities to narrow in or fine-map BMI-related loci.⁸ This will help to reduce the number of variants for functional follow-up studies, which tend to

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be time and labor intensive. In addition, dense genotyping of the GWAS loci could aid the discovery of additional independent signals within the GWAS loci.

In this study, we densely genotyped 21 BMI loci identified in EA studies in 29,151 AAs from the Population Architecture using Genomics and Epidemiology (PAGE) consortium by using the Metabochip.⁹ We aimed to finemap the 21 known BMI loci in the AA population and search for additional independent signals associated with BMI. For validated loci in AAs, we evaluated whether weaker LD patterns in AAs can help narrow in on the underlying potential causal variants. In addition, because Metabochip was developed to test putative association signals for BMI and many obesity-related metabolic and cardiovascular traits and to fine-map established loci,⁹ we also conducted a Metabochip-wide discovery-oriented analysis to search for potential BMI-associated loci.

Subjects and Methods

Study Population

The National Human Genome Research Institute funds the PAGE consortium to investigate the epidemiologic architecture of well-replicated genetic variants associated with human diseases or traits.¹⁰ PAGE consists of a coordinating center and four consortia, Epidemiologic Architecture for Genes Linked to Environment (EAGLE), which is uses data from Vanderbilt University Medical Center's biorepository and links it to deidentified electronic medical records (BioVU); the Multiethnic Cohort Study (MEC); the Women's Health Initiative (WHI); and Causal Variants Across the Life Course (CALiCo), itself a consortium of five cohort studies—the Atherosclerosis Risk in Communities (ARIC) study, Coronary Artery Risk Development in Young Adults (CARDIA), the Cardiovascular Health Study (CHS), the Hispanic Community Health Study/Study of Latinos, and the Strong Heart Study.¹⁰

This PAGE Metabochip study included AA participants from the ARIC, BioVU, CHS, CARDIA, MEC, and WHI studies and from extended collaborations to two additional studies - GenNet and the Hypertension Genetic Epidemiology Network (HyperGen) (Table S1, in the Supplemental Data available with this article online). The detailed description of each study can be found in the Supplemental Data. We excluded underweight (BMI < 18.5 kg/m²) and extremely overweight (BMI > 70 kg/m²) individuals under the assumption that these extremes could be attributable to data-coding errors or an underlying rare condition outside the scope of this investigation. We also limited analysis to adults (defined as having an age > 20 years). The CARDIA participants are young, and the BMI < 18.5 exclusion criterion was not applied in this cohort. All studies were approved by institutional review boards at their respective sites, and all study participants provided informed consent.

Anthropometric Measurements

For individuals from the ARIC, CHS, CARDIA, HyperGEN, GenNet, and WHI studies, BMI was calculated from height and weight measured at the time of study enrollment. For individuals from BioVU, the median height and weight across all visit years were used in BMI calculations. For individuals from MEC, self-reported height and weight were used for calculations of baseline BMI. A validation study within MEC has shown high validity of self-reported height and weight. Specifically, this study showed that BMI was underestimated on the basis of self-reports versus measured weight, but the difference was small (< 1 BMI unit) and was comparable to the findings from national surveys.¹¹

Genotyping and Quality Control

Genotyping was performed with the Metabochip, whose design has been described elsewhere.⁹ In brief, the Metabochip, a custom Illumina iSelect genotyping array of nearly 200,000 SNP markers, is designed to cost-effectively analyze putative association signals identified through GWAS meta-analyses of many obesity-related metabolic and cardiovascular traits and to fine-map established loci.⁹ Metabochip SNPs were selected from the catalogs developed by the International HapMap and 1000 Genomes projects.⁹ More than 122,000 SNPs were included for fine mapping of 257 GWAS loci of 23 traits (including 21 BMI loci).⁹ For determination of the boundaries around each GWAS index SNP, all SNPs with $r^2 \ge 0.5$ with the index SNP were identified, and then initial boundaries were expanded by 0.02 cM in either direction through use of the HapMap-based genetic map. SNPs were excluded if (1) the Illumina design score was <0.5 or (2) SNPs within 15 bp in both directions of the SNP of interest could be found with an allele frequency of >0.02 among Europeans (CEU). SNPs annotated as nonsynonymous, essential splice site, or stop codon were included regardless of allele frequency, design score, or nearby SNPs in the primer.⁹ Twenty-one BMI GWAS loci identified at the time at which the Metabochip was designed were represented for signal fine mapping (Table S2).

Samples were genotyped at the Human Genetics Center of the University of Texas, Houston (ARIC, CHS, CARDIA, GenNet, and HyperGEN), the Vanderbilt DNA Resources Core in Nashville (BioVU), the University of Southern California Epigenome Center (MEC), and the Translational Genomics Research Institute (WHI). Each center genotyped the same 90 HapMap YRI (Yoruba in Ibadan, Nigeria) samples to facilitate cross-study quality control (QC), as well as 2%-3% study-specific blinded replicates to assess genotyping quality. Genotypes were called separately for each study via GenomeStudio with the GenCall 2.0 algorithm. Studyspecific cluster definitions (based on samples with call rate > 95%; ARIC, BioVU, CHS, CARDIA, MEC, and WHI) or cluster definitions provided by Illumina (GenNet and HyperGEN) were used for sample calling, and samples were kept in the analysis if the call rate was >95%. We excluded SNPs with a GenTrain score <0.6 (ARIC, BioVU, CHS, CARDIA, MEC, and WHI) or <0.7 (GenNet and HyperGEN), a cluster separation score <0.4, a call rate <0.95, and a Hardy-Weinberg equilibrium $p < 1 \times 10^{-6}$. We utilized the common 90 YRI samples and excluded any SNP that had more than 1 Mendelian error (in 30 YRI trios), any SNP that had more than two replication errors with discordant calls when comparisons were made across studies in 90 YRI samples, and any SNP that had more than three discordant calls for 90 YRI genotyped in PAGE versus the HapMap database. SNPs were excluded from the meta-analyses if they were present in less than three studies.

For ARIC, BioVU, CHS, CARDIA, MEC, and WHI combined we identified related individuals by using PLINK to estimate identical-by-descent (IBD) statistics for all pairs. When apparent pairs of first-degree relatives were identified, we excluded from each pair the member with the lower call rate. We excluded from further analysis samples with an inbreeding coefficient (F) above 0.15 (ARIC, BioVU, CHS, CARDIA, MEC, and WHI).¹² We determined principal components of ancestry in each study separately by using EIGENSOFT^{13,14} and excluded apparent ancestral outliers from further analysis as described elsewhere.¹⁵

WHI SHARe Imputation

Of the WHI women genotyped on the Metabochip, 1,962 women were part of the group of 8,288 WHI subjects genotyped for the WHI SNP Health Association Resource (SHARe) GWAS via the Affymetrix 6.0 platform. To improve statistical power, we imputed the Metabochip SNPs in the remaining 6,326 SHARe subjects with Affymetrix 6.0 data. Details can be found elsewhere.¹⁶ In brief, we first merged genotypes for the 1,962 subjects genotyped on both the Affymetrix 6.0 platform and the Metabochip and constructed haplotypes (study-specific reference panel). We then phased the haplotypes for samples genotyped on the Affymetrix 6.0 platform only and performed a haplotype-to-haplotype imputation on Metabochip SNPs for the 6,326 target individuals to estimate genotypes (as allele dosages). We used MACH for phasing and Minimac for final imputation. To evaluate the quality of each imputed SNP, we calculated the dosage r². We excluded imputed SNPs with $r^2 < 0.5$ for SNPs with allele frequency < 1% and with $r^2 < 0.3$ for SNPs with allele frequency > 1%. Given the large reference panel and strict QC criteria, this resulted in high imputation quality.¹⁶

Statistical Analysis

In each study, we evaluated the association between natural-logtransformed BMI and each SNP. Because the distribution of BMI was not normal (it was skewed toward higher BMI), we used natural-log-transformed BMI,¹⁷ which reduces the influence of potential outlying observations on the analyses. Except for GenNet and HyperGen, linear regression models were used under the assumption of an additive genetic model and with the adjustment for age, sex, study site (as applicable), and ancestry principal components in each study. All models (except WHI) included the interaction term of sex and age so that possible effect modification by sex was accounted for. Family data from GenNet and HyperGen were analyzed with linear mixed models so that relatedness was accounted for. We used fixed-effect models with inverse variance weighting to pool the study-specific association results as implemented in METAL.¹⁸ We used Q-statistics and I² to measure heterogeneity across studies. To determine whether the 21 previously identified GWAS loci were significantly associated with BMI in AA, we used the p value threshold 5.8 \times 10⁻⁵ as an approximate correction for an average of 866 SNPs at each locus (i.e., 0.05/ 866 SNPs).¹⁹ To identify additional independent signals in any of the loci that were significantly associated with BMI in our study, we conducted conditional analyses. We performed linear regression models that included the most significant SNP (i.e., lead SNP) as a covariate and each of the other SNPs at the same locus (two SNPs in each model) to search for additional independent signals. If, after adjustment for the lead SNP, any SNP remained significant at the locus-specific Bonferroni-corrected significance level (i.e., 0.05/number of SNPs tested at a given locus), we defined the SNP as an additional independent signal. When testing all other SNPs on the Metabochip, we used a Bonferroni-adjusted significance level based on the total number of SNPs on the chip, 2.5 × 10^{-7} (0.05/200,000 SNPs), to declare a BMI-associated SNP. No inflation was observed in any analysis (the inflation factor $\lambda = 1.00$) in our Metabochip-wide analysis. For sex-stratified analysis, GenNet and HyperGen were excluded because of the familystudy design.

LD in the AA sample was calculated in 500 kb sliding windows via PLINK.²⁰ Likewise, the Malmo Diet and Cancer Study on 2,143 controls from a Swedish population²¹ provided Metabochip LD and frequency information in Europeans to facilitate the LD pattern comparisons between AA and EA populations. We used LocusZoom plots²² to graphically display the fine-mapping results. SNP positions from NCBI build 37 were used, and recombination rates were estimated from 1000 Genomes Project data.

Functional Annotation

To inform the discussion about the underlying potential functional variants, we made functional hypotheses for each of our most significantly BMI-associated variants by compiling a list of correlated SNPs ($r^2 > 0.5$) genotyped in our AA study populations and annotating each list for potential regulatory evidence consistent with enhancers, promoters, insulators, silencers and other effects related to gene expression. Because our lead SNPs and the SNPs in strong LD with our lead SNPs were in noncoding regions, we hypothesized that the underlying biology behind the signal was likely to impact gene expression through some unknown regulatory mechanism. For each list we aligned correlated SNPs with a combined browser view of all currently available ENCODE tracks in the UCSC Genome Browser and compared each allelic region for altered transcription factor binding site (TFBS) motifs by using JASPAR and ConSite. Given that methylation patterns are highly variable, it is useful to look for, in addition to BMI-relevant tissues, histone modifications in many cell lines to identify regions that are actively regulated, meaning that histone marks are present in some but not all cell lines. Thus, in addition to adipocytes, hepatocytes, and neurons, we used ENCODE's histone modification tracks to query a wide variety of cell lines for all available histone marks to identify SNPs falling in various regulatory regions. The DNase hypersensitivity track provided a more precise demarcation of open chromatin loci, and the ChIP-Seq TFBS track provided evidence for the binding of specific proteins. Although less specific than ChIP-Seq, JASPAR, ConSite, and HaploReg databases were used for querying a larger number of conserved TFBS and predicting alterations in predicted motifs between reference and alternate alleles. A 46-way PhastCons track in the UCSC Genome Browser was used as secondary evidence for a regulatory region, but lack of conservation did not rule out a functional candidate. The most likely functional-candidate SNPs for each locus were evaluated for statistical significance in association with BMI. The detailed list of functional annotation data sets we used is shown in Table S3.

Results

This study consisted of 29,151 AAs from eight studies. Study participants had an average age of 51.2 years (Table S1). Approximately 80% of study participants were women. Within and across studies, men tended to have a lower mean BMI than women. The obesity rate (BMI \geq 30 kg/m²) ranged from 16%–46% in men and 26%–64% in women. After quality control, we tested 18,187 genetic variants across 21 BMI loci and 177,663 variants across the Metabochip.

Table 1.	Association R	esults for the N	lost Significa	nt SNP and Al	ll Previously I	dentified GWA	S SNPs for Eight BM	I-Relat	ed Loci v	vith Significant	a Results i	n African Arr	erican	5	
Region	Lead AA SNP on Metabochip	GWAS Index SNP	Position	Number of SNPs	Candidate Gene	R ² Lead SNP with GWAS SNP in AAs	R ² Lead SNP with GWAS SNP in EA Populations	CA ^b	CAF ^c in AAs	CAF in EA Populations	Effect	p Value	Rsq ^d	Het P ^e	l ²
1q25.2	rs543874	rs543874	177889480	765	SEC16B	-	-	Т	0.75	0.81	-0.0110	2.4×10^{-9}	NA	0.04	0.52
2p25.3	rs6548240	-	636929	1,123	TMEM18	-	-	А	0.87	0.83	0.0130	1.1×10^{-7}	0.98	0.96	0
-	-	rs6548238	634905	-	TMEM18	0.86	0.99	А	0.12	0.17	-0.0128	3.8×10^{-7}	1.00	0.82	0
-	-	rs2867125	622827	-	TMEM18	0.80	0.96	А	0.12	0.17	-0.0114	5.6×10^{-6}	1.00	0.79	0
-	-	rs7561317	644953	-	TMEM18	0.49	0.99	А	0.24	0.17	-0.0054	4.8×10^{-3}	1.00	0.89	0
3q27.2	rs7647305	rs7647305	185834290	369	ETV5	-	-	А	0.41	0.23	-0.0069	3.2×10^{-5}	0.95	0.29	0.18
-	-	rs9816226	185834499	-	ETV5	0.37	0.85	А	0.80	0.81	0.0067	9.1×10^{-4}	0.96	0.57	0
4p12	rs10938397	rs10938397	45182527	342	GNPDA2	-	-	А	0.75	0.57	-0.0099	1.7×10^{-7}	0.99	0.24	0.23
6p12.3	rs2744475	-	50784880	1,685	TFAP2B	-	-	С	0.67	0.71	-0.0082	2.8×10^{-6}	0.98	0.39	0.05
-	-	rs987237	50803050	-	TFAP2B	0.23	0.50	А	0.90	0.82	-0.0093	5.2×10^{-4}	NA	0.07	0.46
11p14.1	rs1519480	-	27675712	688	BDNF	-	-	А	0.25	0.68	-0.0095	7.8×10^{-7}	1.00	0.90	0
-	-	rs6265	27667202	-	BDNF	0.14	0.12	А	0.05	0.18	-0.0172	1.8×10^{-5}	NA	0.64	0
-	-	rs925946	27679916	-	BDNF	0.12	0.97	Т	0.27	0.30	-0.0005	7.9×10^{-1}	1.00	0.23	0.24
-	-	rs10767664 ^f	27724745	-	BDNF	0.12	0.13	С	0.93	0.76	0.0111	3.7×10^{-4}	0.99	0.28	0.18
16q12.2	rs62048402	-	53803223	1,814	FTO	-	-	А	0.11	0.41	0.0120	5.1×10^{-6}	1.00	0.14	0.34
-	-	rs1421085	53800954	-	FTO	1.00	1.00	Т	0.89	0.59	-0.0119	6.5×10^{-6}	NA	0.15	0.33
-	-	rs9930506	53830465	-	FTO	0.44	0.81	Т	0.79	0.57	-0.0082	3.9×10^{-5}	NA	0.21	0.27
-	-	rs9941349	53825488	-	FTO	0.52	0.91 ^g	А	0.19	0.41	0.0082	1.1×10^{-4}	NA	0.56	0
-	-	rs1558902	53803574	-	FTO	0.98	1.00	А	0.88	0.58	-0.0132	4.5×10^{-4}	1.00	0.43	0
-	-	rs8050136	53816275	-	FTO	0.15	0.94	А	0.43	0.41	0.0027	1.0×10^{-1}	NA	0.05	0.48
-	-	rs9939609	53820527	-	FTO	0.13	0.94	А	0.47	0.41	0.0027	1.0×10^{-1}	1.00	0.10	0.40
-	-	rs1121980	53809247	-	FTO	0.14	0.90	Т	0.47	0.43	0.0021	1.9×10^{-1}	NA	0.26	0.21
-	-	rs6499640	53769677	-	FTO	0.00	0.09	А	0.65	0.60	-0.0013	4.6×10^{-1}	1.00	0.52	0
18q21.32	rs6567160	-	57829135	1275	MC4R	-	-	А	0.81	0.75	-0.0096	4.7×10^{-6}	0.98	0.03	0.52
-	-	rs17782313	57851097	-	MC4R	0.06	0.99	Т	0.72	0.77	-0.0068	1.5×10^{-4}	1.00	0.50	0
-	-	rs10871777	57851763	-	MC4R	0.05	0.97	А	0.71	0.75	-0.0061	6.7×10^{-4}	1.00	0.40	0.04

(Continued on next page)

		4CIU/82ISI	- 00/100/0	MC4K	0.24	0.77	A	0.14	0.20	conn.n	01 × 6.0	1.UU	17.0	07.0
		rs571312	57839769 -	MC4R	0.03	0.99	А	0.34	0.25	0.0009	6.1×10^{-1}	NA	0.99	0
Results ^a Signific ^b CA: co ^c CAF: co ^d Measu ^e Het p: ^f SNP fai ^g SNP fai	or all 21 loci are sh ance level: 0.05 div ded allele. oded allele frequenc ement of imputatic reterogeneity test ed in guality contro led in genotyping i	iown in Table S2. vided by 866 (avera cy. on accuracy, rangin p value. ol; SNP proxy (r598 in our EA population	age number of SNPs across 19 from 0 (low) to 1 (high); 18748, LD r ² = 1.0 in Haplv 11, and r ² is calculated from	21 BMI loci). ; NA indicates th 1ap YRI) was sub 1 1000 Genomes	at there was no stituted. Project Europea	imputation in a subset n populations.	of 6,326	WHI sam	ples and that all oth	ner sample	s have directly	/ genoty	ped data.	

Table 1. Continued

Fine Mapping BMI Loci

Among the 21 BMI GWAS loci identified in EA studies, eight loci (SEC16B [MIM 612855], TMEM18 [MIM 613220], ETV5 [MIM 601600], GNPDA2 [MIM 613222], TFAP2B [MIM 601601], BDNF [MIM 113505], FTO [MIM 610966], and MC4R [MIM 155541]) displayed SNPs with significant evidence of association (Table 1; see also Table S2). The lead SNP (the most significant SNP in AAs) in each of these eight loci had a minor-allele frequency >0.05 and showed little evidence of heterogeneity. In these eight loci most GWAS index SNPs previously identified in EA GWASs (20 out of 23) had a consistent direction of the effects reported in original EA studies. Because the results of the FTO locus have been described previously,²³ we focused on the other seven regions. Among those seven loci, the lead SNPs, rs543874 in SEC16B (p = 1.5×10^{-9}), rs7647305 in *ETV5* ($p = 3.2 \times 10^{-5}$), and rs10938397 in GNPDA2 (p = 1.7×10^{-7}), were consistent with the previously identified GWAS SNP (the most significant SNP highlighted in the previous GWAS) in EA populations. For rs543874 and rs10938397, the observed effects on BMI were slightly stronger in AAs than in EA individuals (change in BMI per coded allele: 1.1% in AAs and 0.9% in EA individuals for rs543874; 1.0% in AAs and 0.8% in EAs for rs10938397⁵), whereas they were slightly weaker in AAs than in EA individuals for rs7647305 (change in BMI per coded allele: 0.7% in AAs and 0.9% in EA individuals⁶). The minor-allele frequency (MAF) was higher for rs543874 and rs7647305 and lower for rs10938397 in AAs than in EA individuals (Table 1). In the other four loci, the lead SNPs in AAs differed from the GWAS SNPs from EA populations. The lead SNPs in all four loci (rs6548240 in TMEM18, rs2744475 in TFAP2B, rs1519480 in BDNF, and rs6567160 in MC4R) were modestly to strongly correlated (r^2 ranged from 0.5–1.0) with at least one of the GWAS SNPs on the basis of LD in EA populations. However, when LD was based on AA populations, the correlation was weaker, in several cases substantially weaker (Table 1). For the 13 BMI loci that did not replicate in our AA analysis, most GWAS index SNPs (13 out of 17) from previous GWASs involving EA individuals showed effects in the same direction when results from our AA samples were compared with results from the previous EA studies.

We investigated the question of whether LD patterns in AA studies can narrow previous association signals from EA studies and found that at seven out of the eight significant loci, AA LD patterns assisted with narrowing association signals (Table 2; see also Figure S1). One of the most extreme examples was for *MC4R*. Among EA individuals, 107 and 119 SNPs were correlated ($r^2 > 0.5$) with the lead SNP in our analysis (rs6567160) and with the GWAS index SNPs, respectively; these SNPs represent a region spanning 184 kb and 230 kb, respectively. However, among AAs only five SNPs were correlated with rs6567160 at $r^2 > 0.5$, and these SNPs represented a region spanning 71 kb. In *SEC16B, ETV5, TFAP2B, BDNF,* and *FTO*, the number of

					AA Populations		EA Populations			
Region	Gene	Lead SNP in AAs of PAGE	Region Size	Number of SNPs	Number of SNPs with r ² > 0.5 with Lead SNP	Region size for SNPs with r ² > 0.5 with Lead SNP (bp)	Number of SNPs with r ² > 0.5 with Lead SNP	Region Size for SNPs with r ² > 0.5 with Lead SNP (bp)	Number of SNPs with r ² > 0.5 with GWAS Index SNPs	Region size for SNPs r ² >0.5 with GWAS index SNPs (bp)
1q25.2	SEC16B	rs543874	180 kb	765	2	4,242	39	95,660	39	95,660
2p25.3	TMEM18	rs6548240	250 kb	1,123	81	40,820	104	53,290	106	75,430
3q27.2	ETV5	rs7647305	110 kb	369	5	10,348	28	49,410	29	49,620
4p12	GNPDA2	rs10938397	90 kb	342	9	20,482	5	20,482	5	20,482
6p12.3	TFAP2B	rs2744475	560 kb	1,685	1	1,128	44	15,320	29	152,570
11p14.1	BDNF	rs1519480	300 kb	688	7	21,942	42	22,4200	42 ^b	224,200 ^b
16q12.2	FTO	rs62048402	650 kb	1,814	19	44,529	74	47,575	76 ^c	73,850 ^c
18q21.32	MC4R	rs6567160	360 kb	1,275	5	71,160	107	183,730	119	229610
Average fo	r number of	SNPs or region si	ze		16	26,831	55	86,208	56	115,178
Average fo Significant	r number of ce level: 0.05	SNPs or region si divided by 866 (¿	ize average nun	aber of SNPs	16 across 21 BMI loci).	26,831		55	55 86,208	51 10/ 100/100 11/ 55 86,208 56

SNPs correlated at r^2 >0.5 with the lead SNP and the spanning region size were also substantially reduced and to a limited degree for *TMEM18* (Table 2). Only for *GNPDA2* did the number of correlated SNPs and the spanning region size not reduce, but the number of correlated SNPs in Europeans was already small.

To further refine the regions associated with BMI in AAs, we performed conditional analyses for each of the eight significant loci by including the lead SNP in a locus as a covariate to search for additional independent signals. rs186117327 in *GNPDA2* was borderline significantly associated with BMI when the locus-specific significance level was taken into account (Table 3). No evidence of heterogeneity was observed across studies. In *GNPDA2*, rs186117327 was significantly associated with BMI in the marginal analysis (p = 3.7×10^{-7}),; this association became less significant but remained marginally so when adjustment was made for the lead SNP (p = 1.7×10^{-4}). rs186117327 was weakly correlated with the lead SNP (rs10938397) as well as with the GWAS SNP at this locus (r² = 0.11 in AAs; 0.16 in EA individuals).

Identification of Two BMI-Related Loci in AAs by Metabochip-wide Analysis

In the Metabochip-wide analysis (excluding the SNPs in the 21 BMI-related loci), we identified five SNPs in two loci (these SNPs were rs116612809, rs114584581, rs74941130, and rs79329695 in 2p23.2/*BRE* [MIM 610497] and rs4802349 in 19q13.32/*DHX34*) as being associated with BMI in AAs at a Metabochip-wide significance level ($p < 2.5 \times 10^{-7}$). Two of the SNPs in 2p23.2/*BRE* (rs116612809 and rs79329695) reached the conventional genome-wide significance level ($p < 5.0 \times 10^{-8}$) (Table 4 and Figure 1). Furthermore, with $p = 3.7 \times 10^{-7}$, rs57813622 at the 7p21.2/*DGKB* [MIM 604070] locus approached the Metabochip-wide significance level.

In 2p23.2/BRE, all four SNPs are rare variants in Europeans (MAF = 0.1% in the 1000 Genomes Project) but common in AAs (MAF = 10%). The most significant SNP, rs116612809, is located in the intronic region of BRE and is highly correlated with the three other significant SNPs $(r^2 ranges from 0.98-1.00, Figure 2)$. In the conditional analysis for all other SNPs at the BRE locus, we did not observe evidence for an additional independent signal associated with BMI after conditioning on rs116612809, suggesting that all four SNPs point to the same functional variant. The 2p23.2/BRE was included on the Metabochip as a part of a ~1.3-Mb-long region so that a GWAS locus associated with blood triglyceride (TG) concentrations could be fine-mapped.^{24,25} Within this region, the four BMI-associated SNPs were more than 500 kb away from the TG GWAS index SNPs (rs1260326, rs1260333, rs780093, and rs780094; r² ranged from 0.02-0.05 in AAs). To examine the association between rs116612809 and blood TG concentrations in our population, we used ARIC and WHI, for which the TG individual data were available (n = 11,680). We did not observe a significant

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				Number of			Marginal	Results ^a		Condition	al Results ^b		
Region	Gene	SNP	Position	Tested SNPs	Coded Allele	CAF	Effect	p Value	Het p ^d	Effect	p Value	Het p	r ² with Lead SNP in AAs
4p12	GNPDA2	rs186117327	45,101,187	285	А	0.75	-0.0095	3.7×10^{-7}	0.54	-0.0074	1.7×10^{-4}	0.91	0.11
-	-	rs10938397 ^c	45,182,527	-	А	0.75	-0.0099	1.7×10^{-7}	0.24	-0.0068	1.6×10^{-3}	0.37	-

^aMarginal results represent results when only the single variant was in the model. ^bConditional analysis represents the result for the SNP when adjustment was made for the most significant lead SNP and vice versa. ^cThe most significant SNP in a locus. ^dHet p: heterogeneity-test p value.

Region	SNP	Position	Candidate Gene	Coded Allele	CAF in AAs	CAF in EA Individuals	Effect	p value	Rsqª	Het p	r ² with Lead SNP in AAs
2p23.2	rs116612809	28,301,171	BRE	A	0.90	0.001	-0.0151	3.6×10^{-8}	0.99	0.48	-
-	rs114584581	28,304,380	BRE	А	0.10	0.001	0.0148	5.9×10^{-8}	0.99	0.43	0.98
-	rs74941130	28,306,293	BRE	А	0.10	0.001	0.0148	6.9×10^{-8}	0.99	0.44	1.00
-	rs79329695	28,319,874	BRE	А	0.10	0.001	0.0151	3.7×10^{-8}	0.99	0.56	0.99
19q13.32	rs4802349	47,874,510	DHX34	А	0.48	0.12	-0.0087	1.2×10^{-7}	1.00	0.46	-



Figure 1. Manhattan Plot for Metabochip-Wide Analysis of BMI in African Americans

The –log10 of p values for each SNP on the Metabochip is plotted against physical chromosomal positions. Green dots represent the SNPs in 21 BMI-related loci that were previously identified in European populations and fine-mapped on the Metabochip. Blue line: $p = 2.5 \times 10^{-7}$; red line: $p = 5 \times 10^{-8}$.

rs6548240 is in a region of open chromatin that binds multiple transcription factors, and elevated levels of active histone marks associated with promoters were detected in several

association (p = 0.71) with blood TG concentrations, which did not change after adjustment for BMI. Furthermore, the association between rs116612809 and BMI did not change after adjustment for TG (Table S4).

In 19q13.32/DHX34, rs4802349 is located in an intron of DHX34 and has a MAF of 0.48 in AAs and 0.12 in EA individuals. Only ten SNPs surrounding rs4802349 (\pm 200 kb) were genotyped; none was correlated with rs4802349 in Aas, and none was associated with BMI. Previously, rs4802349 was reported to be a putative association signal for high-density lipoprotein (HDL) cholesterol at a moderate p value of 0.005.²⁶ We examined the association between rs4802349 and blood HDL concentrations with and without BMI adjustment in ARIC and WHI (n = 11,680). We observed that the p value changed from 0.25 to 0.04 after BMI adjustment. Furthermore, the association between rs4802349 and BMI did not change after adjustment for HDL (Table S4).

Sex-Stratification Analyses

We did not observe any additional loci when we stratified the analysis by sex (Tables S5 and S6 and Figures S2 and S3). Also, the results for the SNPs that were significant in the fine-mapping analyses and the two BMI loci results were consistent across sex (p-heterogeneity ≥ 0.29 , Table S7). However, as a result of the relatively small sample of men in this study, we have limited power to detect a difference-of-sex effect.

Functional Annotation

In silico analysis assessed whether each BMI-associated locus was in an "open," transcriptionally permissive conformation, as would be expected of a functional locus. Using ENCODE data sets, we found each signal to be in a region consistent with regulatory evidence, such as active histone marks, open chromatin structure (DNase hypersensitivity), or regions experimentally shown to bind one or more transcription factors (Table S8). Bioinformatics analyses revealed that the lead SNP, rs6548240 in the *TMEM18* locus, was the strongest functional candidate. cell lines (Figure S4). The functional properties of TMEM18 are obscure, although a recent study indicated that TMEM18 plays a regulatory role in adipocyte differentiation and biology.²⁷ For GNPDA2, the potential additional independent signal rs186117327 could be tagging rs7659184 ($r^2 = 0.6$ with rs186117327 in AAs), which falls in a region of open chromatin, and relative to the reference allele, the alternate allele reduces the binding affinity of GATA2 and EN1 transcription factors. In 2p23.2/ BRE, bioinformatics analyses indicated that the lead SNP, rs116612809, tags another intronic SNP, rs78003529 $(r^2 = 0.65 \text{ in AAs})$, which falls in a region of open chromatin with elevated enhancer histone marks and is in a region that binds both Pol2 and c-Jun. Furthermore, evidence from scans of positional weight matrices (PWMs) suggests that the alternate allele of rs78003527 has a much higher binding affinity for Nkx3 and Nkx2. In 19q13.32/DHX34, the DHX34 signal rs4802349 and two SNPs highly correlated with it are located within putative regulatory regions. rs4802349 falls within histone-modification marks associated with enhancer activity and alters the Mtf1 transcription-factor binding motif. In addition, two other functional candidates, rs2547369 and rs2341878 ($r^2 = 0.7$ and 0.6, respectively in African populations) were highly correlated with this signal and fall in regions of open chromatin having methylation patterns associated with promoter or enhancer activity. Furthermore, rs25476369 ($r^2 = 0.65$ with rs4802349 in African populations) falls in a region that binds six transcription factors, whereas rs2341878 is of particular interest because it falls in a strong promoter region in a number of relevant tissues. The function of GNPDA2, BRE, and DHX34 is unknown, which makes it difficult to link the underlying biological mechanisms of these genetic variants to BMI.

Discussion

In this study, encompassing close to 30,000 AAs, we used the Metabochip to systematically evaluate 21 BMI loci



discovered among European descent populations in previous GWASs and to search for potential BMI loci. Eight of the 21 loci (*SEC16B*, *TMEM18*, *ETV5*, *GNPDA2*, *TFAP2B*, *BDNF*, *FTO*, and *MC4R*) were found to be associated with BMI in our AA study population. Further conditional analyses indicated that *GNPDA2* contained an additional independent signal. Moreover, Metabochip-wide analyses revealed two BMI-associated loci: the *BRE* locus at genome-wide significance ($p < 5 \times 10^{-8}$) and the *DHX34* locus at Metabochip-wide significance ($p < 2.5 \times 10^{-7}$).

Among the eight BMI loci that were significant in the AA population, the lead SNPs in SEC16B, ETV5, and GNPDA2 were the same as the GWAS SNPs identified in previous EA studies, which provided further support for the idea that the three SNPs (rs543874, rs7647305, and rs10938397) are proxies of causal variants influencing BMI. The most significant (lead) SNPs in the other five loci differed from the previously reported GWAS SNPs, although, as expected, all were moderately to highly correlated with the GWAS SNPs in European populations (r² ranged from 0.5 to 1.0). In TFAP2B and MC4R, the GWAS SNPs were not significantly associated with BMI in AAs, suggesting that the lead SNPs in these two loci are better proxies for the underlying functional variants. We showed that the weaker LD patterns in AA populations than in EA populations substantially reduced the number of functional-variant proxies in six of the eight BMI loci that were significant in AAs (and to a limited extent in TMEM18). Our results illustrate the important contribution of AAs to systematic fine-mapping of GWAS loci originally reported in EA populations. In addition, the lead SNPs that were either consistent or correlated with the GWAS SNPs at these replicated loci in EA populations might indicate that EA populations and AA populations share the underlying causal variants at these loci.

Bioinformatics analyses revealed that the lead SNP, rs6548240 in the *TMEM18* locus, was the strongest

Figure 2. Regional-Association Plot for the New BMI-Related Locus at 2p23.2/BRE The -log10 of p values (left *y* axis) is plotted against the SNP genomic position based on NCBI build 37 (*x* axis); the estimated recombination rate from the 1000 Genomes Project for African populations is on the right *y* axis and is plotted in blue. The most significant SNP is denoted with a purple diamond. SNPs are colored to reflect correlation with the most significant SNP. Gene annotations are from the UCSC Genome Browser.

functional candidate. Although rs6548240 is located in an intergenic region 31 kb downstream of *TMEM18*, ChIP-seq evidence indicates that CTCF looping might anchor this distant putative enhancer to the promoter of *TMEM18*. Taken

together, these pieces of evidence make rs6548240 an interesting functional candidate for future laboratory follow-up. This example shows that combining fine mapping with bioinformatics analysis can help to narrow in on the putative functional variants for further follow-up studies.

There are multiple reasons that 13 BMI loci originally identified in EA populations were not significantly associated with BMI in this sample of AAs. Limited statistical power could be an important reason for this observation. Statistical power is impacted by the variance of BMI, MAF, effect size, and sample size. Compared to populations in some large European-focused studies, our AA population had larger variance in BMI⁷ (standard deviation: 6.2 kg/m² versus 4.2 kg/ m^2), which reduces the statistical power. Also compared with primarily very large European-focused studies with sample sizes from ~90,000 to ~250,000,⁵⁻⁷ our study was relatively small. Nonreplication might arise because of different causal variants between EA individuals and AAs, a weak AA LD pattern, which leads to weak correlation between causal variants and marker SNPs on the Metabochip, and limited statistical power. All of this might explain that we did not observe all loci significantly associated with BMI in AAs, and it emphasizes the need for a larger sample size. We used a uniform p value threshold of 5.8 \times 10⁻⁵ (0.05/average number of SNPs per locus) as an approximate correction for the average of 866 SNPs across these loci. However, if we use Bonferroni correction (i.e., 0.05/the number of SNPs at a given locus) or correction by the effective number of SNPs at each locus after accounting for LD patterns, one more locus, SH2B1 [MIM 608937], would be indicated to be significant in AAs (Table S2).

Conditional analysis indicated that the *GNPDA2* locus contained a potential additional independent signal. The potential additional independent signal, rs186117327, is physically relatively close (80 kb) to the lead SNP,

rs10938397, but is only weakly correlated $\left(r^2=0.11\right)$ with it.

We identified two BMI-related loci, 2p23.2/BRE and 19p13.32/DHX34, in the Metabochip-wide analysis. We evaluated whether African ancestry plays role in these two BMI-related loci among 8,310 AAs from a GWAS in WHI. We didn't observe a significant effect of ancestry at these two loci (p > 0.5; Table S9). At 2p23.3/BRE, the most significant SNP, rs116612809, reached the conventional genome-wide significance level (5 \times 10⁻⁸) and was surrounded (3-18 kb) by three highly correlated SNPs ($r^2 = 0.98-1.00$) showing very similar results. All four SNPs are common (MAF = 0.1), but they all are rare variants in EA populations $(MAF = 0.001)^{28}$ and hence it is unlikely that these SNPs would be identified in EA GWASs unless they have much stronger effects than the moderate effects observed in our study or are tested in very large sample sets. The lead SNP in 2p23.3/BRE (rs116612809) is located within an intron of BRE, which is stress responsive and highly expressed in brain and reproductive organs.²⁹ No previous study reported an association with BMI or any other trait; however, in support of our finding, the GIANT consortium reported that 82 of 200 SNPs within *BRE* had a p value < 0.05 (min. p = 2.5×10^{-4}) for a BMI association in EA populations.⁵ Accordingly, our results showed that varying allele frequency in different ancestral groups significantly contributes to the statistical power and that studying different ancestral groups helps to identify potentially functional loci.³⁰

We found another potential BMI-related locus at 19q13.32/DHX34. The SNP rs4802349, located in an intron DHX34, was marginally significantly associated with BMI at a Metabochip-wide significance level (p = 1.2×10^{-7}), but because the finding did not reach the conventional genome-wide significance level of 5 \times 10⁻⁸, additional replication studies in AAs are warranted. DHX34 is a putative RNA helicase and has not been reported to be associated with any diseases or traits in humans before, except for a suggestive association with HDL cholesterol (p = 0.005).²⁶ A recent GWAS in EA populations showed an association with BMI in a neighboring region, 19q13.32/ZC3H4; rs3810291 was the most significant SNP.5 However, rs4802349 is physically 300 kb away from rs3810291, and a recombination hotspot lies between ZC3H4 and DHX34 in AAs and Europeans (HapMap phase II YRI and CEU), resulting in a low correlation between both SNPs in African populations $(r^2 = 0.04)$ and Europeans ($r^2 < 0.001$). This observation suggests that these two associations are independent from each other. Our results along with the evidence from functional annotation warrant additional validation studies for this locus.

In conclusion, we observed that eight BMI-associated GWAS loci identified from EA populations were significantly associated with BMI in AAs and identified a potential additional independent signal in one locus. In addition, we discovered two potential BMI-related loci through Metabochip-wide analysis; one of these loci which reached the conventional genome-wide significance level. Importantly, our study demonstrated that fine mapping in AA populations in combination with bioinformatics analyses is a valuable and effective way to narrow in on the underlying causal variants in GWAS loci discovered in EA populations and that studying minority populations can contribute to loci discovery.

Supplemental Data

Supplemental data include four figures, nine tables, and supplemental text, including acknowledgments and can be found with this article online at http://www.cell.com/AJHG/.

Acknowledgments

See Supplemental Data.

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Web Resources

The URLs for data presented herein are as follows:

- Center for Disease Control and Prevention (CDC), Adult Obesity Facts, http://www.cdc.gov/obesity/data/adult.html
- GIANT consortium data files, http://www.broadinstitute.org/ collaboration/giant/index.php/GIANT_consortium_data_files
- MACH, http://www.sph.umich.edu/csg/abecasis/mach
- Minimac, http://genome.sph.umich.edu/wiki/Minimac
- Online Mendelian Inheritance in Man (OMIM), http://www. omim.org
- The Population Architecture using Genomics and Epidemiology (PAGE) Study, http://www.pagestudy.org
- World Health Organization (WHO) (2011) Obesity and Overweight Fact Sheet, http://www.who.int/mediacentre/ factsheets/fs311/en/index.html

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