Anaerobic Antimicrobial Therapy After Necrotizing Enterocolitis in VLBW Infants

Julie Autmizguine, MD, MHS^a, Christoph P. Hornik, MD, MPH^{a,b}, Daniel K. Benjamin Jr, MD, PhD, MPH^{a,b}, Matthew M. Laughon, MD, MPH^c, Reese H. Clark, MD^d, C. Michael Cotten, MD, MHS^b, Michael Cohen-Wolkowiez, MD, PhD^{a,b}, Daniel K. Benjamin, PhD^e, P. Brian Smith, MD, MPH, MHS^{a,b}, on behalf of the Best Pharmaceuticals for Children Act—Pediatric Trials Network Administrative Core Committee

OBJECTIVE: To evaluate the effect of anaerobic antimicrobial therapy for necrotizing enterocolitis (NEC) on clinical outcomes in very low birth weight (\leq 1500 g) infants.

METHODS: We identified very low birth weight infants with NEC from 348 US NICUs from 1997 to 2012. Anaerobic antimicrobial therapy was defined by antibiotic exposure on the first day of NEC. We matched (1:1) infants exposed to anaerobic antimicrobial therapy with infants who were not exposed by using a propensity score stratified by NEC severity (medical and surgical). The primary composite outcome was in-hospital death or intestinal stricture. We assessed the relationship between anaerobic antimicrobial therapy and outcome by using a conditional logistic regression on the matched cohort.

RESULTS: A total of 1390 infants exposed to anaerobic antimicrobial therapy were matched with 1390 infants not exposed. Mean gestational age and birth weight were 27 weeks and 946 g, respectively, and were similar in both groups. We found no significant difference in the combined outcome of death or strictures, but strictures as a single outcome were more common in the anaerobic antimicrobial therapy group (odds ratio 1.73; 95% confidence interval, 1.11–2.72). Among infants with surgical NEC, mortality was less common with anaerobic antimicrobial therapy (odds ratio 0.71; 95% confidence interval, 0.52–0.95).

CONCLUSIONS: Anaerobic antimicrobial therapy was not associated with the composite outcome of death or strictures but was associated with an increase in intestinal strictures. This higher incidence of intestinal strictures may be explained by the fact that death is a competing outcome for intestinal strictures, and mortality was slightly lower in the anaerobic cohort. Infants with surgical NEC who received anaerobic antimicrobial therapy had lower mortality.



^aDuke Clinical Research Institute, and ^bDepartment of Pediatrics, Duke University Medical Center, Durham, North Carolina; ^cDivision of Neonatal–Perinatal Medicine, School of Medicine, The University of North Carolina at Chapel Hill, Chapel Hill, North Carolina; ^dPediatrix-Obstetrix Center for Research and Education, Sunrise, Florida; and ^eJohn E. Walker Department of Economics, Clemson University, Clemson, South Carolina

Dr Autmizguine conceptualized and designed the study, carried out the analyses, and drafted the initial manuscript; Drs Hornik, Cohen-Wolkowiez, and Benjamin supervised the analyses and revised the article critically for important intellectual content; Drs Benjamin, Laughon, Clark, and Cotten conceptualized and designed the study and revised the article critically for important intellectual content; Dr Smith conceptualized and designed the study, drafted the initial manuscript, and revised the article critically for important intellectual content; and all authors approved the final manuscript as submitted.

www.pediatrics.org/cgi/doi/10.1542/peds.2014-2141

DOI: 10.1542/peds.2014-2141

Accepted for publication Oct 22, 2014

WHAT'S KNOWN ON THIS SUBJECT: Necrotizing enterocolitis is associated with high mortality and morbidity in premature infants. Anaerobic antimicrobial therapy has been associated with increased risk of intestinal strictures in a small randomized trial. Optimal antimicrobial therapy for necrotizing enterocolitis is unknown.

WHAT THIS STUDY ADDS: Anaerobic antimicrobial therapy was associated with increased risk of stricture formation. Infants with surgical necrotizing enterocolitis treated with anaerobic antimicrobial therapy had lower mortality. For infants with medical necrotizing enterocolitis, there was no added benefit associated with anaerobic antimicrobial therapy. Necrotizing enterocolitis (NEC) is a common and devastating intestinal complication of prematurity. Incidence of NEC ranges from 3% in infants with birth weight (BW) of 1251 to 1500 g to 11% for infants born weighing <750 g.¹ Despite treatment, 15% of infants who develop NEC die, and mortality approaches 50% in infants with surgical NEC.^{2,3} Survivors often suffer from complications including intestinal stricture, short bowel syndrome, and poor neurodevelopmental outcomes.^{4–6}

The pathogenesis of NEC involves a combination of factors, including genetic predisposition, immaturity of the intestinal tract, imbalance in microvascular tone, abnormal microbial intestinal colonization, and infectious agents.7-10 Although no single microorganism has been identified, infection probably plays a key role in the disease process, as demonstrated by bacterial overgrowth in the intestinal mucosa and the occurrence of NEC outbreaks.^{8,11,12} A wide range of pathogens are associated with NEC, including aerobic and anaerobic bacteria.^{10,13–19} Therapy for NEC includes broad-spectrum antibiotics with coverage of bacteria from the intestinal tract. In a small cohort of infants with NEC, a randomized controlled trial with 42 infants observed no difference in mortality or intestinal perforation in those who received an antibiotic regimen of ampicillin, gentamicin, and clindamycin compared with those who received only ampicillin and gentamicin.²⁰ However, there was a higher rate of intestinal strictures in the clindamycin group. Despite this finding, clindamycin is often used for NEC in the nursery, and the safety and efficacy of other antibiotic regimens for NEC have not been established.^{20–22} The objective of the current study was to assess the association of anaerobic antimicrobial therapy and subsequent clinical

outcomes in very low birth weight (VLBW, ≤1500 g BW) infants.

METHODS

Study Design and Setting

We identified all VLBW infants with medical or surgical NEC discharged from 348 NICUs managed by the Pediatrix Medical Group from 1997 to 2012. The Pediatrix Medical Group maintains a data warehouse that is populated from an electronic medical record that prospectively captures information from notes generated by clinicians. Data on multiple aspects of care are entered into the system to generate admission notes, daily progress notes, procedure notes, and discharge summaries. Information is collected on maternal history and demographics, medications, respiratory support, laboratory results, culture results, procedures, and diagnoses. The study was approved by the Duke University Institutional Review Board without the need for written informed consent because the data were collected without identifiers.

Definitions

Antimicrobials were considered to provide anaerobic coverage if they were previously reported as having in vitro activity against the major obligate anaerobic bacilli from the intestinal flora. These antimicrobial agents included metronidazole, clindamycin, cefoxitin, any carbapenem, moxifloxacin, piperacillin-tazobactam, ticarcillin-clavulanate, ampicillin-sulbactam, and amoxicillin-sulbactam. An infant was defined as receiving anaerobic antimicrobial therapy based on antibiotics received on the first day of NEC. The diagnosis and severity of NEC were assigned at each site by the attending neonatologist and included either medical NEC or surgical NEC. Surgical indication included the need for a peritoneal drain. The assessment of NEC severity was not based on standardized

criteria and was assigned daily by the treating physician. NEC severity was defined on the first day of the course of NEC regardless of change in severity thereafter. Infants were excluded if they received antimicrobial agents for <5consecutive days from the start of NEC, unless they died during this 5-day period. Mortality was defined as in-hospital death from any cause. Mortality status was defined as missing for infants with nonconvalescent transfers of care. Presence of intestinal stricture was defined as any diagnosis of noncongenital intestinal obstruction after the start of the NEC episode. The diagnosis of intestinal obstruction was assigned by the treating physician in the electronic medical record, and methods used to assign this diagnosis were not available. If an infant had >1 episode of NEC, only the first episode was considered in the analysis.

Demographic data included gender, race, BW, gestational age (GA), postnatal age, and Apgar score at 5 minutes. Surrogates for severity of illness on the first day of NEC were collected and included ventilator support (yes or no), highest level of fraction of inspired oxygen (FIO₂), and inotropic support (yes or no).

Statistical Methods

The primary outcome was in-hospital death or development of an intestinal stricture. Secondary outcomes consisted of death or strictures analyzed individually. An additional secondary outcome was assessed among the subgroup of infants with medical NEC: the composite of progression from medical to surgical NEC or death within the first 7 days of the NEC episode. Outcomes were compared between infants exposed and not exposed to anaerobic antimicrobial therapy. Because infants with more severe illness are more likely to receive anaerobic antimicrobial therapy, propensity score (PS) 1:1 matching was used to

ensure comparison of similar infants.²³ We used baseline demographics and surrogates for severity of illness that might predict both anaerobic antimicrobial therapy and primary outcome to build the PS model by using a multivariable logistic regression.²⁴ The PS model was stratified by NEC severity and derived from the following covariates: postnatal age, ventilator support, FIO2 requirement, inotropic support on day 1 of NEC, GA, small-for-GA status, gender, race, Apgar score at 5 minutes, discharge year, and site. Because site was analyzed as a fixed effect in the PS model, no PS could be estimated for infants belonging to a site with an insufficient number of observations or a site where every infant had the same anaerobic coverage status. We included the discharge year as a categorical variable in the PS model to adjust for changes in care over the study period. We assessed covariate balance across treatment groups by comparing covariate means. Histograms and kernel density plots of PS across groups were also examined. We performed 1:1 matching by using the nearest neighbor without replacement, and it was allowed only if the difference in PS between case and control was < 0.01. On the PS-matched cohort, we assessed the effect of anaerobic antimicrobial therapy on clinical outcomes by using a logistic regression conditioned on the matched pair.23

Because of previous literature linking clindamycin with intestinal strictures, we investigated the effects of clindamycin specifically, as a secondary analysis. We built a separate PS model estimating the conditional probability of receiving clindamycin among infants who were exposed to clindamycin and those who were not exposed to anaerobic antimicrobial therapy, by using the same covariates as in the primary analysis. We then compared outcomes by using a conditional logistic regression after 1:1 matching based on PS. Finally, we performed a multivariable logistic regression without matching to compare outcomes between infants exposed and not exposed to any anaerobic antimicrobial therapy, adjusting for the same covariates used in the PS of the primary analysis.

Demographic and baseline characteristics were summarized and compared between 2 groups: infants exposed and not exposed to anaerobic antimicrobial therapy on the first day of NEC. A χ^2 test for categorical variables and a Wilcoxon rank-sum test or a *t* test for continuous variables were used to assess differences between groups. We performed statistical analyses by using Stata 12 (Stata Corp, College Station, TX). All statistical tests were 2-sided, with significance defined as P < .05.

RESULTS

We identified 6737 infants meeting the inclusion criteria, of whom 3358 (50%) were exposed to anaerobic antimicrobial therapy and 3379 (50%) were not. Overall, 4958 (74%) had medical NEC, and 1779 (26%) had surgical NEC. The mean GA was 27 weeks (5th, 95th percentile: 23, 31) and 27 weeks (5th, 95th percentile: 24, 31) in the anaerobic antimicrobial therapy and control groups, respectively. The mean BW was 936 g (5th, 95th percentile: 530, 1417) and 952 g (5th, 95th percentile: 540, 1420) in infants exposed to anaerobic antimicrobial therapy and those who were not, respectively. Infants who were exposed to anaerobic antimicrobial therapy were more likely to be on ventilation (2152 [64%] vs 1467 [45%], P < .001) and vasopressor therapy (839 [25%] vs 283 [8%], P < .001) and had a higher median F_{10_2} (30% vs 25%, P < .001) compared with infants not exposed to anaerobic antimicrobial therapy.

After nearest-neighbor PS matching, 1390 infants exposed to anaerobic antimicrobial therapy were matched to infants who were not exposed to anaerobic antimicrobial therapy to yield a final cohort of 2780 infants (41% of the initial cohort) (Fig 1). PS matching provided a well-balanced cohort based on baseline characteristics (Table 1), and PS was equally distributed in both treatment groups (Supplemental Fig 3). The mean GA and BW of the cohort were 27 weeks (23, 32) and 946 g (540, 1421), respectively. In the matched cohort, 75% (n = 2074) of infants had



FIGURE 1 Study population flowchart.

TARI F	1	Demographics	and	Clinical	Characteristics	of	the	Matched	Cohort
IADLL		Demographics	anu	unnuar	01101 00101 101100	υı	LIIG	Matcheu	0011011

	Anaerobic . The	Anaerobic Antimicrobial Therapy		
	No, <i>n</i> = 1390	Yes, <i>n</i> = 1390		
GA, wk			.96	
<25	228 (16)	233 (17)		
25–28	737 (53)	736 (53)		
>28	425 (31)	421 (30)		
BW, g			.39	
<750	428 (31)	406 (29)		
751–1000	414 (30)	391 (28)		
1001–1250	302 (22)	330 (24)		
1251–1500	246 (18)	263 (19)		
Small for GA	275 (20)	270 (19)	.81	
Male	736 (53)	742 (53)	.82	
Day of life ^a			.49	
≤7	124 (9)	107 (8)		
8–30	832 (60)	849 (61)		
≥31	434 (31)	434 (31)		
Race or ethnicity			.91	
White	574 (41)	569 (41)		
African American	425 (31)	415 (30)		
Hispanic	329 (24)	345 (25)		
Other	62 (5)	61 (4)		
5-min Apgar score			.83	
<3	70 (5)	68 (5)		
4–6	293 (21)	306 (22)		
7–10	1027 (74)	1016 (73)		
NEC stage ^a			>.99	
Medical	1037 (75)	1037 (75)		
Surgical	353 (25)	353 (25)		
Mechanical ventilation ^a	747 (54)	743 (53)	.88	
Inotropic support ^a	185 (13)	193 (14)	.66	
Highest fraction of supplemental oxygen, ^a median (5th, 95th percentile)	27 (21, 100)	26 (21, 100)	.81	

Data presented as frequency (%) unless specified otherwise.

^a On the first day of the NEC episode.

medical NEC, and 706 (25%) had surgical NEC. Among infants with anaerobic antimicrobial therapy, clindamycin was the most frequently used anaerobic antibiotic (56%), followed by metronidazole (29%) and piperacillin-tazobactam (9%). However, clindamycin use decreased and metronidazole use increased during the study period (Fig 2).

Overall, 26% (n = 725) of matched cohort infants died before discharge or developed intestinal strictures; 23% of infants (n = 645) died, and 3% (n = 84) developed strictures (4 infants had strictures before death). Fewer infants experienced the composite outcome of death or strictures in the anaerobic antimicrobial therapy group, but the risk was not significantly different (odds ratio [OR] 0.96; 95% confidence interval [CI], 0.80–1.14; Table 2). We observed similar results for the individual outcome of death (OR 0.87; 95% CI, 0.72–1.05). Conversely, more infants developed strictures in the anaerobic antimicrobial therapy group compared with the control group (OR 1.73; 95% CI, 1.11–2.72).

Among infants with medical NEC (n = 2074), all covariates used in the PS model were well balanced after matching (Supplemental Table 4). In this subgroup, we observed a nonsignificant increase in death or strictures in infants treated with anaerobic antimicrobial therapy (OR 1.09; 95% CI, 0.87–1.37; Table 2). Death rates were similar in both treatment groups (OR 0.99; 95% CI,

0.78–1.26). Strictures were more common in infants exposed to anaerobic antimicrobial therapy (OR 1.60; 95% CI, 0.97–2.64), although this result was not statistically significant. The number of infants with medical NEC who progressed to surgical NEC or died within the first 7 days of the episode was similar in both treatment groups, with 121 (12%) and 126 (12%) infants in the nonanaerobic and anaerobic antimicrobial treatment groups, respectively (OR 1.05; 95% CI, 0.80–1.37).

Among infants with surgical NEC (n = 706), baseline clinical characteristics were well balanced across treatment groups after PS matching (Supplemental Table 4). In this subgroup, fewer infants either died or developed strictures in the anaerobic antimicrobial therapy group, although this result was not statistically significant (OR 0.77; 95% CI, 0.57-1.03; Table 2). Death was significantly less common in infants exposed to anaerobic antimicrobial therapy (OR 0.70; 95% CI, 0.52-0.95), whereas we observed a nonsignificant increase in strictures in exposed infants (OR 2.40; 95% CI, 0.85-6.81).

When we restrict the anaerobic cohort to the infants who received only clindamycin, the matched cohort included 1922 infants, of whom 961 (50%) were exposed to clindamycin and 961 (50%) were not exposed to any anaerobic antimicrobial therapy. Baseline characteristics used in the PS were well balanced in both treatment groups (Supplemental Table 5). The composite outcome of death or stricture was similar in the clindamycin and control groups, as were the outcomes of death alone and stricture alone (Table 3).

In addition to our primary analysis, the multivariable logistic regression model developed in the full, prematched cohort (N = 6737) yielded similar results to those obtained under matching but with



FIGURE 2

Distribution of therapy among infants receiving anaerobic antimicrobial therapy on the first day of NEC. Cohort after PS matching. Others = moxifloxacin, ticarcillin-clavulanate, cefoxitin, and ampicillin-sulbactam.

a statistically significant difference in death and strictures between treatment groups (OR of death or strictures 0.90; 95% CI, 0.76–1.07; OR of death 0.80; 95% CI, 0.67–0.97; OR of strictures 1.67; 95% CI, 1.16–2.39).

DISCUSSION

We found no significant difference in the risk of the composite outcome of death or strictures in all infants with NEC exposed or not exposed to anaerobic antimicrobial therapy. We did observe lower mortality in infants with surgical NEC treated with anaerobic antimicrobial therapy and higher risk of strictures among all infants with NEC who received anaerobic antimicrobial therapy.

The overall mortality we observed (26%) is consistent with previous data from cohorts combining infants with medical and surgical NEC (15%-25%),^{20,21,25} but the incidence of intestinal strictures is in the lower range reported in the literature (3% vs 4%-30%).^{4,20,21,26-30} The wide range of stricture incidence reported is probably a result of inconsistent definitions leading to ascertainment bias; some studies might report all strictures as diagnosed with radiologic tests, whereas others

TABLE 2 Anaerobic Antimicrobial Therapy and Clinical Outcomes

Outcomes	Anaerobic / The	Antimicrobial erapy	OR (95% CI)	Р
	No, n (%)	Yes, n (%)		
Overall, $N = 2780^{a}$				
Death or strictures	368 (26)	357 (26)	0.96 (0.80-1.14)	.62
Death	338 (24)	307 (22)	0.87 (0.72-1.05)	.14
Strictures	31 (2)	53 (4)	1.73 (1.11-2.72)	.02
Medical NEC, $N = 2074$				
Death or strictures	186 (18)	199 (19)	1.09 (0.87-1.37)	.45
Death	162 (16)	161 (16)	0.99 (0.78-1.26)	.95
Strictures	25 (2)	40 (4)	1.60 (0.97-2.64)	.07
Surgical NEC, $N = 706$				
Death or strictures	182 (52)	158 (45)	0.77 (0.57-1.03)	.08
Death	176 (50)	146 (41)	0.71 (0.52-0.95)	.02
Strictures	6 (2)	13 (4)	2.40 (0.85-6.81)	.10

^a Four infants were diagnosed with strictures before death.

might report only symptomatic strictures. The diagnosis of stricture in our data is limited to those reported by the treating physician in the electronic medical record. The methods used to assign the stricture diagnosis were not standardized and may have varied between infants.

For adults and older children, evidence strongly supports the recommendation of anaerobic antimicrobial therapy as part of the empirical regimen for complicated intra-abdominal infections.³¹ On the other hand, optimal antimicrobial therapy for NEC is unknown, including whether to use empirical anaerobic antimicrobial therapy. One trial randomly assigned 42 infants with NEC receiving ampicillin and gentamicin to clindamycin or no additional therapy.²⁰ Clindamycin did not affect mortality, intestinal perforation, or gangrene but was associated with significantly more intestinal strictures (6/15 survivors [40%] vs 1/18 [5%] in the control group; P = .02). Of note, infants in this previous trial were excluded if intestinal perforation occurred <12 hours after randomization and therefore are most comparable to our subgroup of infants with medical NEC. For infants with medical NEC, our findings are consistent with previous results demonstrating no added benefit of anaerobic antimicrobial therapy on mortality, but we observed only a slight nonsignificant increased risk of stricture.²⁰ In addition, anaerobic antimicrobial therapy did not prevent progression to surgical NEC or mortality within 7 days of the NEC episode. In contrast, anaerobic antimicrobial therapy was associated with lower mortality in infants with surgical NEC.

Post-NEC stricture is an intestinal obstruction resulting from wound healing, most prominently in the intestinal submucosa.²⁶ Stricture is probably a marker of severity of NEC, and it is possible that the lower rate

TABLE 3	Outcomes	and	Clindamycin	Therapy
---------	----------	-----	-------------	---------

Outcomes	No Anaerobic Antimicrobial Therapy, <i>n</i> (%)	Clindamycin, <i>n</i> (%)	OR (95% CI)	Р
Overall, $N = 1922^{a}$				
Death or strictures	291 (30)	279 (29)	0.94 (0.76-1.15)	.53
Death	267 (28)	242 (25)	0.87 (0.70-1.07)	.18
Strictures	27 (3)	42 (4)	1.56 (0.96–2.52)	.07

^a Eight infants were diagnosed with strictures before death.

of strictures in the nonanaerobic treatment group was a function of the death of infants with the most severe cases of NEC, which precluded the occurrence of strictures. Strictures may also be directly related to a specific drug such as clindamycin, although the biological mechanism is unknown. Moreover, other than the study from Faix et al,²⁰ we found no report in the literature linking clindamycin (or any antibiotics) to intestinal strictures. The fact that we observed similar results when we limited our analysis to infants treated with clindamycin compared with infants with no anaerobic treatment suggests that clindamycin may drive this effect, but the number of infants receiving each anaerobic antimicrobial therapy was insufficient to compare outcomes for each individual therapy.

The differential effect of anaerobic antimicrobial therapy on mortality in medical and surgical NEC that we observed suggests that anaerobic bacteria play a more prominent role in the disease process of infants with surgical NEC. A wide range of pathogens are associated with NEC.¹⁵ Several case series of infants with NEC have reported the presence of anaerobic bacteria, including Clostridium perfringens and *Bacteroides fragilis,* in the blood or peritoneal fluid.^{17,19,32-36} A study including 25 infants with NEC showed that infants who had *Clostridium* spp. recovered from peritoneal fluid had more severe disease with more extensive pneumatosis intestinalis, higher incidence of portal venous gas, and more extensive gangrene.33 Although

the exact relationship between anaerobic bacteria and the pathophysiology of NEC has not been established, our findings suggest they are contributing to the disease process, especially in infants presenting with surgical NEC.

Anaerobic antimicrobial use changed dramatically over the study period (Fig 2); clindamycin use decreased, whereas metronidazole and piperacillin-tazobactam use increased. Factors that led to these findings might include safety concerns linking clindamycin to intestinal stricture in the literature,²⁰ increasing clindamycin resistance in anaerobic bacteria such as *B fragilis*,³⁷ and growing clinical experience with other therapeutic options. Our data do not provide sufficient information on specific agents for us to conclude which anaerobic antimicrobial should be preferred for empirical therapy. This question may be answered by an ongoing phase II/III clinical trial in infants with complicated intraabdominal infection (NCT01994993).

This study is the largest evaluation of antibiotic treatment in VLBW infants with NEC. Strengths of this report include its large sample size, representing a large number of NICUs. However, despite our large cohort, matching resulted in the exclusion of nearly 60% of the sample population, which might have resulted in loss of power to detect differences between the 2 treatment groups. As a secondary analysis, a multivariable logistic regression without matching yielded similar results, but differences between groups (lower mortality and more strictures in the anaerobic antimicrobial therapy group) were statistically significant. Nevertheless, we believe matching based on PS provides more robust results by limiting the analysis to a cohort of infants who had similar conditional probabilities of receiving anaerobic antimicrobial therapy, given their clinical characteristics.³⁸ Our study is limited by its cohort design and lack of randomization; therefore, we could not completely avoid the risk of unobserved confounders. For example, documentation of clinical signs is lacking. There are also limitations surrounding diagnosis definitions. The diagnosis was not based on standardized criteria but was assigned by the treating physician. Another limitation is the potential overlap of spontaneous intestinal perforation and NEC diagnosis in the data set. Although these 2 conditions represent separate diagnoses in the data set, differentiating spontaneous intestinal perforation from NEC is difficult clinically, and the diagnosis is often not confirmed until laparotomy. Despite these limitations, this large observational study based on electronic medical records is an efficient way to compare treatment strategies in infants with NEC. A randomized controlled trial is unlikely because a sample size of >7000 VLBW infants would be necessary to detect a difference of 3% in outcomes if the incidence of such outcomes was 30% in the susceptible population (power of 80%; .05 2-sided significance level).

Our study demonstrates differential effects of empirical anaerobic antimicrobial therapy in infants with medical compared with surgical NEC. Infants with surgical NEC treated with anaerobic antimicrobial therapy had lower mortality. For infants with medical NEC, there was no survival benefit associated with anaerobic antimicrobial therapy.

ACKNOWLEDGMENTS

The Pediatric Trials Network Administrative Core Committee:

Jeffrey Barrett, Children's Hospital of Philadelphia, Philadelphia, PA; Katherine Y. Berezny, Duke Clinical Research Institute, Durham, NC; Edmund Capparelli, University of California-San Diego, San Diego, CA; Gregory L. Kearns, Children's Mercy Hospital, Kansas City, MO; Andre Muelenaer, Virginia Tech Carilion School of Medicine, Roanoke, VA; T. Michael O'Shea, Wake Forest Baptist Medical Center, Winston Salem, NC; Ian M. Paul, Penn State College of Medicine, Hershey, PA; John van den Anker, George Washington University School of Medicine and Health, Washington, DC; Kelly Wade, Children's Hospital of Philadelphia, Philadelphia, PA; and Thomas J. Walsh, Weill Cornell Medical College of Cornell University, New York, NY.

The Eunice Kennedy Shriver National Institute of Child Health and Human Development: Alice Pagan, David Siegel, Perdita Taylor-Zapata, and Anne Zajicek.

The EMMES Corporation (Data Coordinating Center): Ravinder Anand, Traci Clemons, and Gina Simone.

Dr Hornik receives salary support for research from the National Center for Advancing Translational Sciences of the National Institutes of Health (NIH) (UL1TR001117). Dr Benjamin receives support from the US government for his work in pediatric and neonatal clinical pharmacology (1R01HD057956-05, 1K24HD058735-05, UL1TR001117, and National Institute of Child Health and Human Development [NICHD] contract HHSN275201000003I). Dr Laughon receives support from the US government for his work in pediatric and neonatal clinical

pharmacology (government contract HHSN267200700051C, PI: Benjamin under the Best Pharmaceuticals for Children Act) and from the NICHD (K23HD068497). Dr Cotten receives salary support for research on neonatal health and outcomes from the NICHD (5U10 HD040492-10) and the National Heart, Lung, and Blood Institute (1R01 HL105702 01A1). Dr Cohen-Wolkowiez receives support for research from the National Center for Advancing **Translational Sciences** (UL1TR001117), the Food and Drug Administration (1U01FD004858-01), and the Biomedical Advanced **Research and Development Authority** (HHS0100201300009C). Dr Smith receives salary support for research from the NIH and the National Center for Advancing Translational Sciences (HHSN267200700051C, HHSN275201000003I, and UL1TR001117).

Address correspondence to P. Brian Smith, MD, MPH, MHS, Department of Pediatrics, Duke University, Duke Clinical Research Institute, PO Box 17969, Durham, NC 27715. E-mail: brian.smith@duke.edu

PEDIATRICS (ISSN Numbers: Print, 0031-4005; Online, 1098-4275).

Copyright © 2015 by the American Academy of Pediatrics

FINANCIAL DISCLOSURE: Dr Benjamin receives support from the nonprofit organization Thrasher Research Fund for his work in neonatal candidiasis (www. thrasherresearch.org); he also has consulted for Astellas, Cubist, Johnson & Johnson, Merck, and Pfizer. Dr Laughon has consulted and served on the Data Safety Monitoring Board for Pfizer. Dr Cohen-Wolkowiez receives support for research from the nonprofit organization Thrasher Research Fund (www.thrasherresearch.org); he has also consulted for GlaxoSmithKline. Dr Smith has consulted for GlaxoSmithKline, Astellas, and Pfizer. The other authors have indicated they have no financial relationships relevant to this article to disclose.

FUNDING: Funded under National Institute of Child Health and Human Development contract HHSN275201000003I for the Pediatric Trials Network. Research reported in this publication was also supported by the National Center for Advancing Translational Sciences of the National Institutes of Health under award UL1TR001117. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. Funded by the National Institutes of Health (NIH).

POTENTIAL CONFLICT OF INTEREST: Dr Benjamin receives support from the nonprofit organization Thrasher Research Fund for his work in neonatal candidiasis (www.thrasherresearch.org); he also has consulted for Astellas, Cubist, Johnson & Johnson, Merck, and Pfizer. Dr Laughon has consulted and served on the Data Safety Monitoring Board for Pfizer. Dr Cohen-Wolkowiez receives support for research from the nonprofit organization Thrasher Research Fund (www. thrasherresearch.org); he has also consulted for GlaxoSmithKline. Dr Smith has consulted for GlaxoSmithKline, Astellas, and Pfizer. The other authors have indicated they have no potential conflicts of interest to disclose.

REFERENCES

- Fanaroff AA, Stoll BJ, Wright LL, et al; NICHD Neonatal Research Network. Trends in neonatal morbidity and mortality for very low birthweight infants. *Am J Obstet Gynecol.* 2007; 196(2):e1–e8
- Holman RC, Stoll BJ, Curns AT, Yorita KL, Steiner CA, Schonberger LB. Necrotising enterocolitis hospitalisations among neonates in the United States. *Paediatr Perinat Epidemiol.* 2006;20(6): 498–506
- Blakely ML, Lally KP, McDonald S, et al; NEC Subcommittee of the NICHD Neonatal Research Network. Postoperative outcomes of extremely low birth-weight infants with necrotizing enterocolitis or isolated intestinal perforation:

a prospective cohort study by the NICHD Neonatal Research Network. *Ann Surg.* 2005;241(6):984–989, discussion 989–994

- Janik JS, Ein SH, Mancer K. Intestinal stricture after necrotizing enterocolitis. *J Pediatr Surg.* 1981;16(4):438–443
- Hintz SR, Kendrick DE, Stoll BJ, et al; NICHD Neonatal Research Network. Neurodevelopmental and growth outcomes of extremely low birth weight infants after necrotizing enterocolitis. *Pediatrics.* 2005;115(3):696–703
- Salhab WA, Perlman JM, Silver L, Sue Broyles R. Necrotizing enterocolitis and neurodevelopmental outcome in extremely low birth weight infants <1000 g. J Perinatol. 2004;24(9): 534–540
- Lee JS, Polin RA. Treatment and prevention of necrotizing enterocolitis. Semin Neonatol. 2003;8(6):449–459
- Ballance WA, Dahms BB, Shenker N, Kliegman RM. Pathology of neonatal necrotizing enterocolitis: a ten-year experience. *J Pediatr*: 1990;117(1 pt 2): S6–S13
- Bhandari V, Bizzarro MJ, Shetty A, et al; Neonatal Genetics Study Group. Familial and genetic susceptibility to major neonatal morbidities in preterm twins. *Pediatrics*. 2006;117(6):1901–1906
- Mai V, Young CM, Ukhanova M, et al. Fecal microbiota in premature infants prior to necrotizing enterocolitis. *PLoS ONE*. 2011; 6(6):e20647
- Boccia D, Stolfi I, Lana S, Moro ML. Nosocomial necrotising enterocolitis outbreaks: epidemiology and control measures. *Eur J Pediatr*: 2001;160(6): 385–391
- Stuart RL, Tan K, Mahar JE, et al. An outbreak of necrotizing enterocolitis associated with norovirus genotype GII.3. *Pediatr Infect Dis J.* 2010;29(7): 644–647
- Kliegman RM, Fanaroff AA. Necrotizing enterocolitis. N Engl J Med. 1984;310(17): 1093–1103
- Blot S, De Waele JJ, Vogelaers D. Essentials for selecting antimicrobial therapy for intra-abdominal infections. *Drugs.* 2012;72(6):e17–e32
- Coates EW, Karlowicz MG, Croitoru DP, Buescher ES. Distinctive distribution of pathogens associated with peritonitis in

neonates with focal intestinal perforation compared with necrotizing enterocolitis. *Pediatrics*. 2005;116(2). Available at: www.pediatrics.org/cgi/ content/full/116/2/e241

- Brook I, Frazier EH. Aerobic and anaerobic microbiology in intraabdominal infections associated with diverticulitis. *J Med Microbiol.* 2000; 49(9):827–830
- 17. Alfa MJ, Robson D, Davi M, Bernard K, Van Caeseele P, Harding GK. An outbreak of necrotizing enterocolitis associated with a novel clostridium species in a neonatal intensive care unit. *Clin Infect Dis.* 2002;35(suppl 1):S101–S105
- Howard FM, Flynn DM, Bradley JM, Noone P, Szawatkowski M. Outbreak of necrotising enterocolitis caused by *Clostridium butyricum. Lancet.* 1977; 2(8048):1099–1102
- Sturm R, Staneck JL, Stauffer LR, Neblett WW III. Neonatal necrotizing enterocolitis associated with penicillin-resistant, toxigenic *Clostridium butyricum. Pediatrics.* 1980;66(6):928–931
- 20. Faix RG, Polley TZ, Grasela TH. A randomized, controlled trial of parenteral clindamycin in neonatal necrotizing enterocolitis. *J Pediatr*. 1988; 112(2):271–277
- Hansen TN, Ritter DA, Speer ME, Kenny JD, Rudolph AJ. A randomized, controlled study of oral gentamicin in the treatment of neonatal necrotizing enterocolitis. *J Pediatr*. 1980;97 (5):836–839
- Shah D, Sinn JK. Antibiotic regimens for the empirical treatment of newborn infants with necrotising enterocolitis. *Cochrane Database Syst Rev.* 2012;(8): CD007448
- 23. Rosenbaum PR, Rubin DB. Constructing a control-group using multivariate matched sampling methods that incorporate the propensity score. *Am Stat.* 1985;39(1):33–38
- Brookhart MA, Schneeweiss S, Rothman KJ, Glynn RJ, Avorn J, Stürmer T. Variable selection for propensity score models. *Am J Epidemiol.* 2006;163(12): 1149–1156
- Yee WH, Soraisham AS, Shah VS, Aziz K, Yoon W, Lee SK; Canadian Neonatal Network. Incidence and timing of presentation of necrotizing enterocolitis in preterm infants. *Pediatrics*. 2012;

129(2). Available at: www.pediatrics.org/ cgi/content/full/129/2/e298

- Bell MJ, Ternberg JL, Askin FB, McAlister W, Shackelford G. Intestinal stricture in necrotizing enterocolitis. *J Pediatr Surg.* 1976;11(3):319–327
- Yeh TC, Chang JH, Kao HA, Hsu CH, Hung HY, Peng CC. Necrotizing enterocolitis in infants: clinical outcome and influence on growth and neurodevelopment. *J Formos Med Assoc*. 2004;103(10): 761–766
- Lemelle JL, Schmitt M, de Miscault G, Vert P, Hascoet JM. Neonatal necrotizing enterocolitis: a retrospective and multicentric review of 331 cases. *Acta Paediatr Suppl.* 1994;396:70–73
- Schullinger JN, Mollitt DL, Vinocur CD, Santulli TV, Driscoll JM Jr. Neonatal necrotizing enterocolitis. Survival, management, and complications: a 25-year study. *Am J Dis Child.* 1981; 135(7):612–614
- Arnold M, Moore SW, Sidler D, Kirsten GF. Long-term outcome of surgically managed necrotizing enterocolitis in a developing country. *Pediatr Surg Int.* 2010;26(4):355–360
- 31. Solomkin JS, Mazuski JE, Bradley JS, et al. Diagnosis and management of complicated intra-abdominal infection in adults and children: guidelines by the Surgical Infection Society and the Infectious Diseases Society of America [published correction appears in *Clin Infect Dis* 2010;50(12):1695]. *Clin Infect Dis*. 2010; 50(2):133–164
- Kliegman RM, Fanaroff AA, Izant R, Speck WT. *Clostridia* as pathogens in neonatal necrotizing enterocolitis. *J Pediatr*. 1979; 95(2):287–289
- Kosloske AM, Ulrich JA. A bacteriologic basis for the clinical presentations of necrotizing enterocolitis. *J Pediatr Surg.* 1980;15(4):558–564
- 34. Mollitt DL, String DL, Tepas JJ III, Talbert JL. Does patient age or intestinal pathology influence the bacteria found in cases of necrotizing enterocolitis? *South Med J.* 1991;84(7):879–882
- Mollitt DL, Tepas JJ III, Talbert JL. The microbiology of neonatal peritonitis. *Arch Surg.* 1988;123(2):176–179
- 36. Noel GJ, Laufer DA, Edelson PJ. Anaerobic bacteremia in a neonatal intensive care

unit: an eighteen-year experience. *Pediatr Infect Dis J.* 1988;7(12):858-862

37. Karlowsky JA, Walkty AJ, Adam HJ, Baxter MR, Hoban DJ, Zhanel GG. Prevalence of antimicrobial resistance among clinical isolates of *Bacteroides fragilis* group in Canada in 2010–2011: CANWARD surveillance study. *Antimicrob Agents Chemother*. 2012; 56(3):1247–1252 D'Agostino RB Jr. Propensity score methods for bias reduction in the comparison of a treatment to a nonrandomized control group. *Stat Med.* 1998;17(19):2265–2281